



Supporting Information

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Catalytic Enantioselective Additions of Allyl- and Crotylboronates to Aldehydes Using Chiral Brønsted Acids. Efficient Catalyst-Controlled Diastereofacial-Selective Formation of Dipropionate Units.

Vivek Rauniyar and Dennis. G. Hall*

Department of Chemistry, University of Alberta, Edmonton, AB, T6G 2G2, Canada

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1. General information

Unless otherwise noted, all reactions were performed under an argon atmosphere using flame-dried glassware. Toluene, hexanes and CH_2Cl_2 were distilled over CaH_2 . THF and Et_2O were distilled over sodium / benzophenone ketyl. Aldehydes were purified by Kugelrohr distillation, prior to use. Molecular sieves were prepared by heating under vacuum at 130°C (over-night) and then stored inside oven maintained at 125°C . Thin layer chromatography (TLC) was performed on Merck Silica Gel 60 F254 plates and, visualized with UV light and KMnO_4 and 5% phosphomolybdic acid / EtOH (PMA). NMR spectra were recorded on Varian INOVA-300, INOVA-400, INOVA-500 or Unity 500 instruments. The residual solvent protons (^1H) or the solvent carbons (^{13}C) were used as internal standards. Boron NMR spectra are referenced to external $\text{BF}_3\cdot\text{OEt}_2$; ^{19}F spectra are referenced to external CFCl_3 . ^1H NMR data are presented as follows: chemical shift in ppm downfield from tetramethylsilane (multiplicity, coupling constant, integration). The following abbreviations are used in reporting NMR data: s, singlet; br s, broad singlet; d, doublet; t, triplet; q, quartet; qt, quartet of triplets; dd, doublet of doublets; dt, doublet of triplets; AB, AB quartet; m, multiplet. High-resolution mass-spectra was recorded by the University of Alberta Mass Spectrometry Services Laboratory, using either electron impact (EI) or electrospray (ES) ionization techniques. Infrared-spectra and optical rotations were recorded by University of Alberta Spectral Services and combustion analyses were performed by the University of Alberta Micro-Analytical Lab.

Preparation of Diols: (*R, R*)-1,2-Diarylethane-1,2-diols were either commercially available or prepared¹ by Sharpless *syn*-dihydroxylations of the corresponding (*E,E*)-1,2-diarylethanes, which in turn were obtained by McMurry coupling of the corresponding aldehydes¹. 1,2-Bis-(3,4-bis-trifluoromethyl-phenyl)-ethane-1,2-diol¹⁴ was generously gifted by Prof. Hisashi Yamamoto (at University of Chicago).

(*R,R*)-1,2-Diphenyl-ethane-1,2-diol (1a): Commercially available from Aldrich.

(*R,R*)-2-Methoxy-1,2-diphenyl-ethanol (1b)²: Prepared by monomethylation of **1a**. To a solution of (*R,R*)-(+)-Hydrobenzoin 100 mg (0.466 mmol, 1 equiv) in 2 ml of DMF under argon was added Ag₂O 108 mg (0.466 mmol, 1 equiv) and MeI 32 μ l (0.513 mmol, 1.1 equiv) and this mixture was stirred under dark for 24 hrs. This resulting solution was then filtered over a short pad of celite and the celite was subsequently washed with DCM. The resulting solution was evaporated under vacuo and purified by flash chromatography (5% EtOAc / Hexanes) to give the title compound in 68% yield. The ¹H NMR, ¹³C NMR, IR, HRMS properties were identical to those reported.

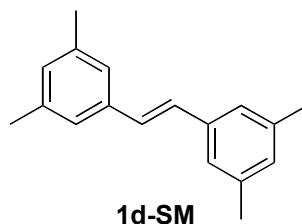
(*R,R*)-2-Benzyloxy-1,2-Diphenyl-ethanol (1c)³: To a solution of **1a** 200mg (0.934 mmol, 1 Equiv) in 10 ml of reagent grade benzene was added 17.7 mg (0.09 mmol, 0.1 Equiv) of *p*-TsOH and to this mixture was added freshly distilled benzaldehyde 113.8 μ l (1.12 mmol, 1.2 Equiv) and the reaction mixture was refluxed for 3 hrs using a dean-stark trap to remove water, after which, the reaction mixture was concentrated under vacuo. The resulting solid was dissolved in 2 ml of toluene and cooled to 0⁰ C, after which 2.8 ml of 1.0 M DIBAL-H in Toluene (3 Equiv) was added and the reaction mixture was stirred at 0⁰C for 2 hours. The reaction was quenched with 2 ml of MeOH

and this was followed by dilution with Et₂O (25ml) and addition of 10 % NaOH and then the resulting mixture was transferred to a separatory funnel and washed with brine followed by drying over Na₂SO₄. The resulting solution was concentrated under vacuo and purified by flash chromatography (5% EtOAc/Hexane) to yield 241 mg (84% yield) of the title compound. The ¹H NMR, ¹³C NMR, IR, HRMS properties were identical to those reported.

1,2-Bis-(3,4-bis-trifluoromethyl-phenyl)-ethane-1,2-diol (1e)¹⁴: Gift from Prof. H. Yamamoto at University of Chicago.

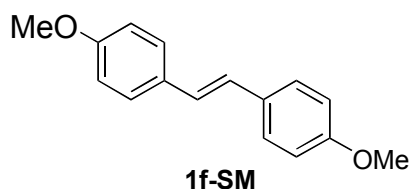
Diols **1e**, **1f**, **1g**, **1h**, **1i** were prepared by the highly enantioselective sharpless syn-dihydroxylation of the corresponding (*E*)-1,2-diarylethenes following the literature procedure.¹

Preparation of the (*E*)-1,2-diarylethene starting materials:

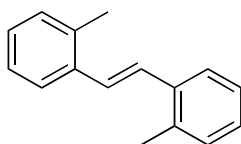


(*E*)-1,2-Di-(3,5-bis(trimethyl)phenyl)-ethene (1d-SM): Into a 250 ml 2-neck round bottom flask equipped with a condenser and a magnetic stirrer under argon was added 2.02 g (45 mmol, 3.0 equiv) of Zn dust and 150 ml of freshly distilled THF. To this suspension was carefully added 2.47 ml (22.5 mmol, 1.5 equiv) of TiCl₄ (to avoid plugging of the needle, the syringe needle should be immediately rinsed with dilute HCl after the addition of TiCl₄). This mixture was stirred for 30 min until a dark green color

was observed after which, 2.017 ml (15 mmol, 1 equiv) of freshly distilled 3,5-dimethylbenzaldehyde was added and the reaction was refluxed overnight. The next day, after the reaction was judged complete by TLC, the reaction mixture was poured over ice cold solution of NaHCO₃ and this solution was filtered through a pad of celite and the resulting mixture was extracted with DCM, washed with brine and dried over Na₂SO₄ and concentrated under vacuo and purified by flash chromatography (1-5% EtOAc/hexanes). Further recrystallization from MeOH gave 1.5 g (85%) of isomerically pure (*E*)-alkene. ¹H NMR (500 MHz, CDCl₃) δ 2.351 (s, 12H), 6.914 (s, 2H), 7.045 (s, 2H), 7.146 (s, 4H) ¹³C NMR (125.7 MHz, CDCl₃) δ 21.348, 124.418, 128.535, 129.307, 137.475, 138.113. IR Cast Film: 697.0, 847.0, 961.9, 1558.8, 1599.3, 2859.1, 2912.0, 3020.8. HRMS (EI) Calcd. 236.1565, found 236.1568. Anal. Calcd for C₁₈H₂₀: C, 91.47, H, 8.53, found C, 91.62, H, 8.54.

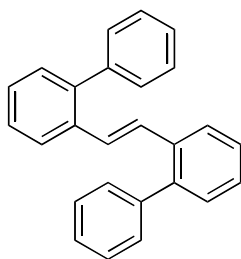


(*E*)-1,2-Di-(4-methoxyphenyl)-ethene (1f-SM): Following general procedure for **1e-SM**, Yield (40%) ¹H NMR δ (400 MHz, CDCl₃) 3.829 (s, 6H), 6.892 (d, *J* = 8.8 Hz, 4H), 6.933 (s, 2H), 7.428 (d, *J* = 8.8 Hz), ¹³C NMR δ (100.58 MHz, CDCl₃) 55.367, 14.166, 126.249, 127.462, 130.556, 159.076, IR Cast Film: 833.8, 1029.5, 1269.8, 1517.2, 1608.0, 2837.7, 2911.3, 2937.0, 2955.6, 3020.7, 3093.6. HRMS (EI) Calcd: 240.11503, found 240.11502, Anal. Calcd for C₁₆H₁₆O₂: C, 79.97, H, 6.71, found C, 79.82, 6.71.



1g-SM

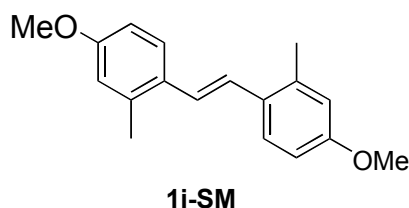
(*E*)-1,2-Di-(2-methylphenyl)-ethene (1g-SM): Following general procedure for **1e-SM**, the product was isolated as 17:14 mixture of *E/Z* alkenes as determined by ^1H NMR, and this isomeric mixture was subjected to iodine catalyzed isomerization¹ in refluxing xylenes for 24 hr. Yield (50%) ^1H NMR (400 MHz, CDCl_3) δ 2.509 (s, 12H), 7.259-7.328 (m, 6H), 7.688-7.691 (d, 1.2 Hz) ^{13}C NMR δ (100.58 MHz, CDCl_3) δ 19.967, 125.604, 126.229, 127.560, 128.058, 130.421, 135.854, 136.853. IR Cast Film: 722.8, 763.2, 978.3, 1492.2, 1954.1, 2860.0, 2946.4, 3014.5, 3061.7, 3094.2. HRMS (EI) Calcd. 208.1252, found 208.1251. Anal. Calcd for $\text{C}_{16}\text{H}_{16}$: C, 92.26, H, 7.74, found C, 92.06, H, 7.77.



1h-SM

(*E*)-1,2-Di-(2-phenylphenyl)-ethene (1h-SM): Following general procedure for **1e-SM**, the product was isolated as a 5.7 : 1 mixture of *E / Z* alkenes and the mixture was subjected to iodine catalyzed isomerization in refluxing xylenes¹ for 24 hr to give isomerically pure *E*-alkene which was purified by recrystallization from DCM/MeOH. Yield (60%). ^1H NMR (400 MHz, CDCl_3) δ 7.118 (s, 2H), 7.301-7.382 (m, 6H), 7.414-

7.561 (m, 12H), ^{13}C NMR (100.58 MHz, CDCl_3) δ 125.932, 127.081, 127.498, 128.130, 129.862, 130.210, 135.660, 140.930, 141.077, IR Cast Film: 700.9, 720.2, 741.7, 761.1, 774.6, 965.0, 1072.7, 1478.7, 1498.2, 1595.3, 3019.9, 3055.9, HRMS (EI) Calcd. 332.15650, found 332.15657. Anal. Calcd. for $\text{C}_{26}\text{H}_{20}$: C, 93.94, H, 6.06, found 93.22, H, 6.05

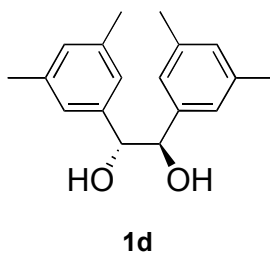


(*E*)-1,2-Di-(2-methyl-4-methoxy-phenyl)ethene (1i-SM): Following general procedure for **1e-SM**. The product was purified by recrystallization to give isomerically pure *E*-alkene. Yield (70%). ^1H NMR (400 MHz, CDCl_3) δ 2.411 (s, 6H), 3.826 (s, 6H), 6.742-6.796 (m, 4H), 7.046 (s, 1H), 7.526 (d, $J = 8.8$ Hz), ^{13}C NMR (100.58 MHz, CDCl_3) δ 20.251, 55.297, 111.786, 115.682, 125.719, 126.598, 129.952, 137.161, 158.924, IR Cast Film: 800.7, 1053.9, 1212.4, 1256.8, 1500.1, 1606.9, 2835.5, 2955.2, 3033.2, HRMS(EI) Calcd. 268.14633, found 268.14648 Anal. Calcd. for $\text{C}_{18}\text{H}_{20}\text{O}_2$: C, 80.56, H, 7.51, O, 11.92, found 80.40, H, 7.52

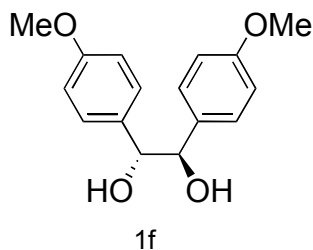
Preparation of Diols via Sharpless Asymmetric Dihydroxylation of *E*-Olefins.

General Procedure for sharpless AD for 1 mmol of olefin¹: In a round bottom flask equipped with a magnetic stirrer was charged 3 equiv of $\text{K}_3\text{Fe}(\text{CN})_6$, 3 equiv. of K_2CO_3 , and 0.01 equiv. of $(\text{DHDQ})_2\text{PHAL}$, 5 ml H_2O and 5 ml *t*BuOH, and this mixture was cooled to 0°C and vigorously stirred for 10 min. after which a thick orange slurry is

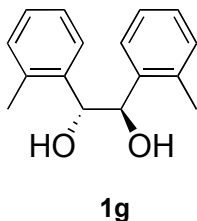
obtained. To above at 0⁰C is added 0.002 mmol of K₂OsO₂(OH)₄ followed by 1 mmol of MeSO₂NH₂ and 1 mmol of *E* olefin, and the reaction mixture is allowed to warm up to room temperature overnight. The progress of the reaction was, monitored by TLC and upon completion of the reaction, to the reaction mixture is added 1.5 gram of Na₂SO₃ and after 1 hr, the reaction mixture is extracted with EtOAc and washed with 1N KOH and then with brine and dried over Na₂SO₄ and concentrated under vacuo. The resulting product is purified by flash chromatography, and recrystallized twice from hot CCl₄ to give the requisite diol. Model diols are obtained by this method in greater than 95 % ee (Reference)¹ and were directly used for catalytic asymmetric allylboration reaction.



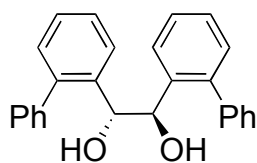
(*R,R*)-(+)-1,2-Bis-(3,5-dimethyl-phenyl)-ethane-1,2-diol (1d) : Yield (75%). ¹H NMR (400 MHz, CDCl₃) δ 2.271 (s, 12H), 2.673 (brs, 2H), 4.650 (s, 1H), 6.822 (s, 4H), 6.895 (s, 2H), ¹³C NMR (100.58 MHz, CDCl₃) δ 21.291, 78.441, 124.527, 129.422, 137.634, 140.158, IR (cast film): 704.0, 726.9, 848.2, 1072.8, 1159.4, 1464.4, 1607.9, 2906.6, 3003.6, 3293.0, 3373.1, HRMS (EI) [M-H₂O]⁺ 252.15162, Anal. Calcd. for C₁₈H₂₂O₂: C, 79.96, H, 8.20, found C, 79.61, H, 8.17, [α]_D = 66.36, (c = 2.97, CHCl₃).



(*R,R*)-(+)-1,2-Bis-(4-methoxy-phenyl)-ethane-1,2-diol (1f): Yield (72%) ^1H NMR (300 MHz, CDCl_3) δ 2.748 (brs, 2H), 3.771 (s, 6H), 4.643 (s, 2H), 6.768 (d, $J = 9$ Hz), 7.051 (d, $J = 8.7$ Hz), ^{13}C NMR (100.58 MHz, CDCl_3) δ 55.207, 78.786, 113.512, 128.197, 132.163, 159.179, IR (cast film): 831.0, 1032.7, 1247.5, 1513.2, 1611.5, 2835.6, 2929.3, 3405.1, HRMS (EI) calcd. 274.12051, found 274.12051, Anal. Calcd. for $\text{C}_{16}\text{H}_{18}\text{O}_4$: C, 70.06, H, 6.61, found 71.11, H, 6.47, $[\alpha]_{\text{D}} = 101.56$ ($c = 1.23$, CHCl_3).

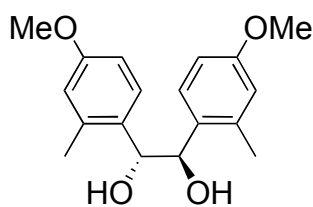


(*R,R*)-(+)-1,2-Bis-(2-methyl-phenyl)-ethane-1,2-diol (1g): Yield (80%) ^1H NMR (500 MHz, CDCl_3) δ 1.680 (s, 6H), 2.905 (brs, 2H), 4.991 (s, 2H), 6.93 (d, $J = 7.5$ Hz), 7.113-7.262 (m, 4H), 7.628 (d, $J = 7.5$ Hz), ^{13}C NMR (125.7 Hz, CDCl_3) δ 18.761, 74.652, 125.970, 127.216, 127.744, 130.211, 135.934, 138.015, IR (cast film) 762.2, 791.7, 1042.5, 1197.9, 1490.6, 1604.4, 2928.2, 3023.2, 3062.3, 3388.3, HRMS (EI) calcd. 242.13068, found 242.13068, Anal. Calcd for $\text{C}_{16}\text{H}_{18}\text{O}_2$, C, 79.31, H, 7.49, found C, 79.22, H, 7.44, $[\alpha]_{\text{D}} = 63.87$ ($c = 1.43$, CHCl_3).



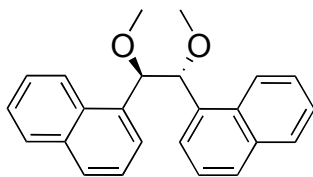
1h

(*R,R*)-(+)-1,2-Bis-(2-phenyl-phenyl)-ethane-1,2-diol (1h): Yield (56%) ^1H NMR (400 MHz, CDCl_3) δ 2.610 (brs, 2H), 4.944 (s, 2H), 6.77 (brs, 4H), 6.990-7.331 (m, 14H), ^{13}C NMR (125.7 MHz, CDCl_3) δ 74.436, 126.725, 127.386, 127.402, 127.936, 129.208, 129.676, 136.629, 140.597, 141.951, IR (cast film) 707.47, 744.71, 997.69, 1074.65, 1189.90, 1436.67, 1451.60, 1596.94, 2929.78, 2976.37, 3025.38, 3066.08, 3348.87, 3468.23, HRMS (EI) Calcd. 366.16198, found 366.16179. Anal. Calcd for $\text{C}_{26}\text{H}_{22}\text{O}_2$: C, 85.22, H, 6.05, found C, 84.65, H, 6.14, $[\alpha]_{\text{D}} = 88.68$ ($c = 0.98$, CHCl_3).



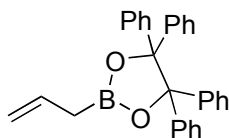
1i

(*R,R*)-(+)-1,2-Bis-(4-methoxy-2-methyl-phenyl)-ethane-1,2-diol (1g): Yield (77%) ^1H NMR (500 MHz, CDCl_3) δ 1.689 (s, 6H), 3.009 (brs, 2H), 3.745 (s, 6H), 4.867 (s, 2H), ^{13}C NMR (125.7 MHz, CDCl_3) δ 19.079, 55.108, 74.497, 111.294, 115.388, 128.404, 130.460, 137.453, 158.755, IR (cast film), 1045.6, 1197.2, 1252.6, 1289.0, 1504.2, 1608.6, 2835.5, 2852.2, 2925.1, 2999.8, 3414.0, HRMS (EI) $[\text{M}-\text{H}_2\text{O}]^+$ 284.14210, Anal. Calcd. for $\text{C}_{18}\text{H}_{22}\text{O}_4$: C, 71.50, H, 7.33, found 71.03, 7.45, $[\alpha]_{\text{D}} = 104.60$ ($c = 0.55$, CHCl_3).

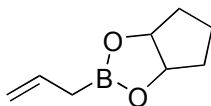


Preparation of (*R,R*)-(+)-Di-(1-Naphthyl)-1,2-dimethoxy-ethane: Into a flame-dried 25 ml RB flask is added 50.0 mg (0.16 mmol, 1.0 equiv) of diol **1j** and the diol is dissolved in 2 ml of freshly distilled THF after which is added 20.3 μ l (0.33 mmol, 2.05 equiv) of MeI and this mixture is cooled to 0⁰C and to above is added in portions, 22.0 mg (0.56 mmol, 3.5 equiv) of NaH (60 % in mineral oil) in portions, and the reaction mixture is sealed and stirred at RT over-night after which the reaction is quenched with addition of water the product is extracted with Et₂O and washed with brine and dried over Na₂SO₄ and filtered and concentrated under vacuo and finally, purified by column chromatography (0-5% EtOAc/Hexane) to give 55.0 mg of product (>99% yield). % ¹H NMR (400 MHz, CDCl₃) δ 3.335 (s, 6H), 5.347 (s, 2H), 7.100-7.158 (m, 4H), 7.276-7.360 (m, 4H), 7.539 (d, 2H, *J* = 8.4 Hz), 7.667 (d, *J* = 8.0 Hz), 8.150 (brs, 2H), ¹³C NMR (100.58 Hz, CDCl₃) δ 57.266, 123.869, 124.732, 125.144, 125.472, 128.312, 128.493, 131.670, 133.590, 134.042, IR (cast film) 767.7, 801.9, 1227.7, 1447.0, 1509.3, 1595.7, 2820.3, 2925.5, 3046.2, HRMS (EI) Calcd. 342.16198, found 342.15982. Anal. Calcd. for C₂₄H₂₂O₂: C, 84.18, H, 6.48, O, 9.34, found C, 83.39, H, 6.54, [α]_D = 158.88 (c = 0.11, CHCl₃).

Allyl (3), Cis-crotyl (7) and Trans-crotyl pinacol boronic esters (5) were prepared by literature method⁴.



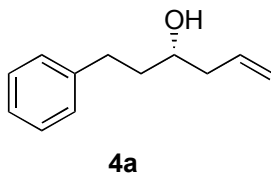
Benzopinacol-allylboronate⁵: Into a flame dried RB flask equipped with a magnetic stir bar was added 3.68g (10.0 mmol, 1 equiv) of 1,1,2,2-Tetraphenylethylene diol and to this was added 10.0 ml of toluene followed by 2.31 ml (10.0 m mol, 1 equiv) of B(OⁱPr)₃. The RB flask was then assembled into a dean-stark trap and refluxed for 24 hr at 105⁰C after which, toluene was distilled off to give a colorless glassy solid, which, was put under high vaccum overnight. To the resulting solid under argon was added 10.0 ml of anhydrous ether and the resulting mixture was cooled to -78⁰C followed by addition of 10.0 ml (10.0 mmol, 1.0 equiv) of allylmagnesium bromide (1.0 M in Et₂O), which results in a white suspension and this mixture was stirred at -78⁰C for 2 hr and this was followed by additional stirring at 0⁰C, after which, the reaction was quenched with 8 ml (1.0 N) HCl and the mixture was then extracted with DCM (2 X 50 ml) and the organic extracts were washed with water (25 ml) and finally brine (25 ml) and dried over Na₂SO₄, filtered, and concentrated under vacuo to give the crude product which, was purified by flash chromatography (5% EtOAc/Hexane) to give 3.0 g (72%) of white solid. The product is very stable to air and moisture and remains intact even on bench. ¹H NMR (400 MHz, CDCl₃) δ 2.251-2.281 (m, 2H), 5.131-5.172 (m, 1H), 5.258-5.318 (m, 1H), 6.160-6.270 (m, 1H), 7.134-7.252 (m, 20H), ¹³C NMR (100.58 MHz, CDCl₃) δ 17.949, 95.982, 115.614, 126.913, 127.194, 128.347, 133.515, 142.401, ¹¹B NMR (128.3 MHz) δ 33.176, IR (Cast film) 696.9, 768.1, 1000.1, 1190.3, 1327.9, 1445.3, 1634.9, 2935.9, 2974.2, 3039.0, 3056.6, HRMS (EI) calcd : 416.19476, found 416.19382. Anal Calcd. for C₂₉H₂₅BO₂: C, 83.66, H, 6.05, found C, 83.02, H, 6.16.



Allylboronic cyclopentane-diol ester: Into a flame dried 100 ml RB flask under argon was added 2.31 ml (10 mmol, 1 equiv) of $B(O^iPr)_3$ and diluted with 10 ml of anhydrous Et_2O , and cooled to $-78^{\circ}C$. To above is added 11.0 ml (11 mmol, 1.1 equiv) of allylmagnesium bromide (1.0 M in Et_2O) and the reaction mixture is stirred at $-78^{\circ}C$ for 3 hrs, after which 20 ml of 1N HCl is added to the reaction mixture and the reaction mixture is brought to room temperature and stirred for 1 hr, and extracted with Et_2O and washed with brine and finally concentrated under vacuo (to leave a minimal amount of solvent,. To this crude mixture is added 714 mg (7.0 mmol, 0.7 equiv) of cis-1,2-cyclopentane diol along with 5 grams of anhydrous $MgSO_4$, and 15 ml of THF and the reaction mixture is stirred under argon overnight, followed by filtration and concentration under vacuo (10 mm Hg, bath temperature $20^{\circ}C$) and, the allylboronate product is purified by flash chromatography (5% Et_2O /Pentane) to provide the requisite product 530 mg (50% yield). 1H NMR (400 MHz, $CDCl_3$) δ 1.492-1.644 (m, 4H), 1.746 (d, $J = 7.2$ Hz), 1.869-1.933 (m, 2H), 4.809-4.911 (m, 2H), 4.911-5.025 (m, 2H), 5.810-5.915 (m, 1H), ^{13}C NMR (100.58 MHz, $CDCl_3$) δ 21.473, 34.622, 82.366, 114.911, 134.209, ^{11}B NMR (128.3 MHz, $CDCl_3$) 32.893, IR (Cast film) 1033.0, 1307.0, 1329.4, 1361.1, 1385.8, 1441.8, 1637.7, 2876.3, 2964.7, 3077.3, HRMS (EI) calcd: 152.10086, found 152.10081. Anal Calcd for $C_8H_{13}BO_2$: C, 63.21, H, 8.62, found C, 63.82, H, 8.72.

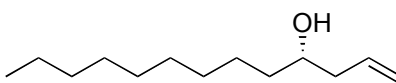
Determination of absolute configuration of homoallylic alcohol products: This was done by comparing the signs of rotation of compound **8a** with literature.¹³ All other absolute stereochemistry were deduced by analogy.

General procedure for catalytic enantioselective and diastereoselective allylboration:



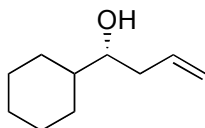
(3S)-1-Phenyl-hex-5-en-3-ol (4a)⁶: Into a flame dried 25 ml round bottom flask equipped with a rice needle stir bar was added 8.65 mg (0.025 mmol, 0.11 equiv) of (*R,R*) 1,2-di-naphthyl-ethanediol, 5.6 mg (0.05 mmol, 0.05 equiv) of anhydrous Na₂CO₃ and 50 mg of activated 4 Å⁰ Molecular sieves, and to this mixture, under argon was added 0.5 ml of freshly distilled toluene. To above was added 25 µl (0.025 mmol, 0.10 equiv) of 1.0 M solution of SnCl₄ in DCM (in order to get reproducible results, it is necessary to use a gas tight 25µl syringe, and that all of the SnCl₄ drops into the diol-solution without touching the side walls, and hence, pyrex test-tubes were not good for this purpose). The resulting mixture was stirred for 5 minutes and cooled to -78⁰C and maintained at this temperature for 15 minutes after which 46.2 mg of allyl boronic pinacol ester **3** (1.1 Equiv) dissolved in 0.5 ml of toluene was added via syringe and this mixture was maintained at -78⁰C for 15 minute, after which, 32.6 µl (0.25 mmol, 1.0 equiv) of freshly distilled hydrocinnamaldehyde was added. The reaction mixture was stirred at -78⁰C for 12 hrs after which 0.5 ml (0.50 mmol, 2.0 equiv) of DIBAL-H in toluene was added to quench any remaining aldehyde and after 30 minutes, 2 ml of 1 N HCl was added (to neutralize DIBAL-H and hydrolyze the borate ester to corresponding alcohol), after which the reaction mixture was brought to room temperature and stirred for 1 hr. At this point, dark brown biphasic solution is obtained which upon extraction with (2 X 25 ml)

of Et₂O gives a clear ethereal solution which is then washed with brine and dried over Na₂SO₄ and concentrated under vacuo to give the crude product (TLC of which gives two spots, one of the diol, and the other of the product, which, are easily separated), which, was purified by flash chromatography (5% EtOAc/Hexanes) to give 37.4 mg of the corresponding homoallylic alcohol in 85 % yield. Analytical data of this product are in complete accordance with literature.⁶ HPLC analysis with chiracel-OD column (5% IPA/Hexane, 254 nm) at 0.5ml / min gives the major isomer (89%, t_r = 26.19 min) and the minor isomer (11 % t_r = 37.62 min). [α]_D = - 4.54 (c = 0.41, CHCl₃).



4b

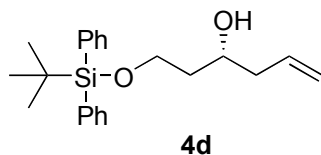
(4S)-Tridecen-4-ol (4b)⁷: Following general procedure for **4a**, yield (76%). The spectroscopic data of this are identical with those reported in literature.⁷ HPLC analysis of the *p*-Nitro benzoyl derivative with chiracel-OD column (100% hexane) 1ml / minute gave the major isomer (90%, t_r = 24.83 min) and the minor isomer (10%, t_r = 28.57 min). [α]_D = -6.64 (c = 0.39, CHCl₃)



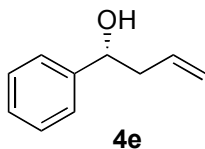
4c

(1R)-1-Cyclohexyl-3-buten-1-ol (4c)⁸: Following general procedure for **4a**, except that the product was purified by 5% Et₂O/Pentane eluent system, yield (90%). The spectroscopic data of the product are identical with those reported in literature.⁸ Enantiomeric excess was determined from the integration of the diastereomeric peaks in the ¹⁹F NMR of the corresponding (*R*) Mosher ester, major (71.544 ppm, 85%), minor

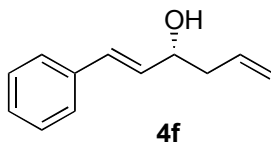
(71.605 ppm, 15%). $[\alpha]_D = -0.43$ ($c = 0.46$, CHCl_3)



(3R)-1-(*tert*-Butyl-diphenyl-silanoxy)-hex-5-en-3-ol (4d)⁶: Following general procedure for **4a**, except, that the aldehyde was dissolved in 0.5 ml of toluene, and injected into the reaction mixture, and reaction time was 24 hrs, yield (90%). The spectroscopic data of this are identical with those reported in literature.⁶ HPLC analysis using chiracel OD (2.5% IPA/hexane, 1ml/min, 254 nm) gave the minor isomer (17.1%, $t_r = 24.3$ min), and the major isomer (82.9%, $t_r = 29.6$ min). $[\alpha]_D = 1.47$, ($c = 0.58$, CHCl_3)

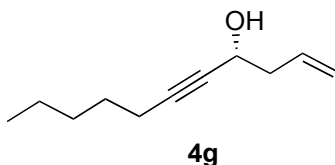


(1R)-1-phenyl-3-butene-1-ol (4e)⁶: Following general procedure for **4a**. Yield (99%). The spectroscopic data of this are identical with those reported in literature.⁶ HPLC analysis using chiracel OD (5% IPA/hexanes, 0.5 ml/min, 254 nm) gave the minor isomer (48.9%, $t_r = 29.8$ min), and the major isomer (51.1%, $t_r = 31.9$ min). $[\alpha]_D = -4.9$, ($c = 0.49$, CHCl_3)

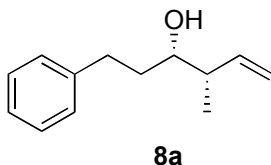


(1E,3R)-1-phenyl-1,5-hexadien-3-ol (4f)⁹: Following general procedure for **4a**. Yield (72%). The spectral properties of the obtained compound are identical with those reported in literature. HPLC analysis of the product using Chiracel OD (5%

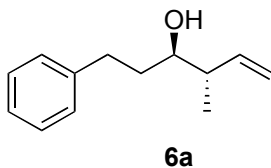
IPA/hexanes, 0.5 ml/ml, 254 nm) gave the major isomer (59.5%, t_r = 42.6 min) and the minor isomer (41.5%, t_r = 75.3 min). $[\alpha]_D = -5.69$, ($c = 0.13$, CHCl_3)



(4R)-Undec-1-en-5-yn-1-ol (4g)¹⁰: Following general procedure for **4a**. Yield (99%). The spectroscopic data of this are identical with those reported in literature. Enantiomeric excess was determined by integration of the diastereomeric peaks of the corresponding (*S*)- Mosher-ester. ^{19}F NMR (376.141 MHz) Major 71.961 ppm (55.98%) and minor 72.183 ppm (44.02%). $[\alpha]_D = -4.0$, ($c = 0.40$, CHCl_3)

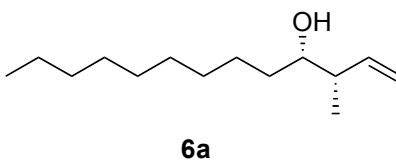


(3S,4S)-4-Methyl-1-Phenyl-5-hexen-3-ol (8a)¹¹: Following general procedure for **4a**, except that cis-crotyl boronic pinacol ester **7** was used and the reaction time was 24 hr. Yield (87%). The spectroscopic data of this are identical with those reported in literature⁶. HPLC analysis of the product using Chiracel OD (10% IPA/Hexane, 1 ml/min, 250 nm) gave the major isomer (70%, t_r = 10.8 min), and minor isomer (30%, t_r = 15.8 min). $[\alpha]_D = -20.36$, ($c = 0.72$, CHCl_3).

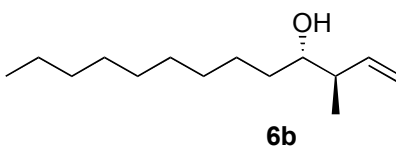


(3R,4S)-4-Methyl-1-Phenyl-5-hexen-3-ol (6a)¹¹: Following general procedure for **8a**,

except that trans-crotyl boronic pinacolester **5** and the isomer (S,S)-(-)-1,2-Di(1-Naphthyl)-1,2-ethane diol of **1j** was used and the reaction time was 24 hr. Yield (99%). The spectroscopic data of this are identical with those reported in literature.¹¹ HPLC analysis of the product using Chiracel OD (10% IPA/Hexane, 1 ml/min, 220 nm) gave the major isomer (86%, t_r = 12.6 min), and minor isomer (14%, t_r = 18.5 min). $[\alpha]_D$ = 13.20, (c = 0.45, CHCl₃).



(S)-3-Methyl-tridec-1-en-4-ol (6a)⁷: Following procedure for **8a**, Yield (70%). The spectroscopic data of this are identical with those reported in literature¹² Enantiomeric excess was determined by integration of the diastereomeric peaks of the corresponding (S)- Mosher-ester. ¹⁹F NMR (376.141 MHz) minor -71.426 ppm (27%), major 71.517 ppm (73%). $[\alpha]_D$ = -15.92, (c = 0.23, CHCl₃).



3-Methyl-tridec-1-en-4-ol (2k)⁷: Following general procedure for **6a**, yield (70%). The spectroscopic data of this are identical with those reported in literature.⁷ Enantiomeric excess was determined by integration of the diastereomeric peaks of the corresponding (S)- Mosher-ester. ¹⁹F NMR (376.141 MHz) minor -71.467 ppm (14%), major 71.530 ppm (86%). $[\alpha]_D$ = -23.94, (c = 0.75, CHCl₃).

Double diastereoselective reactions¹²:

General: (S)-2-Methyl-3-[(tert-butyldimethylsilyl)oxy]propionaldehyde **9** was prepared following literature procedure¹². The crude aldehyde that is obtained (after the work-up, and evaporation of the solvent) of swern oxidation is filtered through a short plug (~10 cm, 1 inch diameter) of silica using 30% EtOAc / Hexanes to free the aldehyde from any residual Et₃N-HCl and other inorganic impurities. Allyl and crotyl reagents were freshly prepared and used for the following reactions. Temperature control during reaction was provided by cryo-cool systems.

Procedure for the reaction of allyl or cis- crotyl boronic pinacol esters with aldehyde 3: Into a flame-dried 25 ml RB flask was charged 17.3mg (0.055 mmol, 0.11 equiv) of diol **1j** or its isomer **1i**, 10.3 mg of Na₂CO₃, and 50 mg of 4 Å⁰ activated molecular sieves and the flask was capped with a septa and put under argon. To this mixture is added 0.75 ml of anhydrous toluene followed by 50 µl of SnCl₄ (1.0 M in DCM) (0.05mmol, 0.10 equiv) and the resulting mixture is stirred for 5 min, after which, it is cooled to -78⁰C and maintained at this temperature for 15 minutes, after which 84 mg of allylboronic pinacol ester **3** or 100.1 mg of freshly prepared cis-crotyl boronic pinacol ester **7** (0.55 mmol, 1.1 equiv) in 0.5 ml of toluene is slowly added and this mixture is maintained at -78⁰C for another 15 minutes. After this, 101 mg (0.50 mmol, 1.0 equiv) of freshly prepared aldehyde **3** dissolved in 0.25 ml of toluene is added dropwise and the reaction is maintained for 24 hrs at -78⁰C. After the elapsed time, 1.0 ml of DIBAL-H (2 equiv, 2 mmol) 1.0 M in toluene, is slowly added to the reaction mixture (in-order to quench any unreacted aldehyde) and the mixture is stirred for 30 min at -78⁰C, after which 2.0 ml of 1 N HCl is added in one portion, and the reaction mixture is gradually brought to room

temperature and stirred for additional 1 hr. At this point, the biphasic reaction mixture appears dark brown. This mixture is extracted for product with 2 X 25 ml portions of Et₂O and washed with 10 ml of water and 10 ml of brine and finally dried over Na₂SO₄. The ethereal extract is then filtered and concentrated under vacuo to give the crude mixture, which was analyzed for diastereomeric composition by ¹H-NMR. The diastereomeric mixture is determined by integration of the doublet signals of the methyl groups of the diastereomers. (See attached NMR insets). The product is purified by column chromatography using 5% EtOAc/Hexane.

Compound **10**, **11** elute together and give a combined yield of 83% for matched-case using diol **1j**, and give 84 :16 ratio of **10**: **11**.

In the mis-matched case using the (**S,S**) isomer of diol **1j**, the combined yield is 60 % and gives a ratio of 64 :36 of **11** : **10**. The spectral characteristics of these mixture is identical with reported literature.¹²

Compounds **12** and **13** in the diastereomeric crude mixture is are purified by the same system as above, and in this case, care is made in order to get both diastereomers into same fractions in order to get accurate ratio of the diastereomers. In the case of diol **1j**, the isolates yield is 77% and the ratio of **12** : **13** is 95 : 5.

In the case of the (**S,S**) isomer of diol **1j**, the ratio of 12 : 13 is **68** : **32**. In this case, the results are analogous to Ref 12 as it is difficult to get 1,2,3 *syn* relationship in such type of cis- crotylboration. Spectral characteristic of these mixtures are in accordance with literature.¹²

For SnCl₄ catalyzed reaction of allyl **3**, cis- crotyl **5** pinacol boronic esters procedure is

analogous except that no molecular sieves, or Na_2CO_3 or the diol auxiliary is used. The product is purified by column chromatography using 5% EtOAc/Hexanes as eluent to give 70% combined yield of diastereomers in the ratio of 54: 46 of **10** : **11**.

For cis-crotylation with SnCl_4 , similar purification as the one described above is used to give a combined yield of 81% and a ratio of 66 : 34 for **12** : **13**.

NMR Studies: Into a flame dried NMR tube was charged 31.4 mg (0.1 mmol, 1 equiv) of diol **1j**, and 21.2 mg (0.20 mmol, 2 equiv) of anhydrous Na_2CO_3 and the NMR tube was sealed and flushed with argon, and 0.8 ml of toluene- d_8 from a new bottle was added to the above mixture, followed by 11.7 μl (0.1 mmol, 1.0 equiv) of SnCl_4 . The minimally soluble diol **1j** is immediately taken into solution via complex formation. This tube is cooled to -78°C and analyzed by ^1H NMR and ^{119}Sn NMR at -78°C . The results indicate presence of 3 alcoholic protons that have shifted downfield, and the ^{119}Sn NMR indicates three peaks in the region of hexa-coordinate Sn complexes. The same result is obtained in the absence of Na_2CO_3 . In the case of a complex between diol **1a**, and SnCl_4 , only one alcoholic proton is noticed, which is shifted downfield and only one complex of hexa-coordinated Sn is observed. Possible scenarios are listed in the attached NMR spectra copies.

Also, allylboration under the general conditions for diol **1j**, (page 14-15) except, using the (*R,R*)-(+)-1,2-Di-(1-Naphthyl)-1,2-dimethoxy ethane (page 11), gave no catalysis and the minimal amount of product that was formed was racemic. As such, the necessity for protons on the chiral auxiliary is demonstrated.

References:

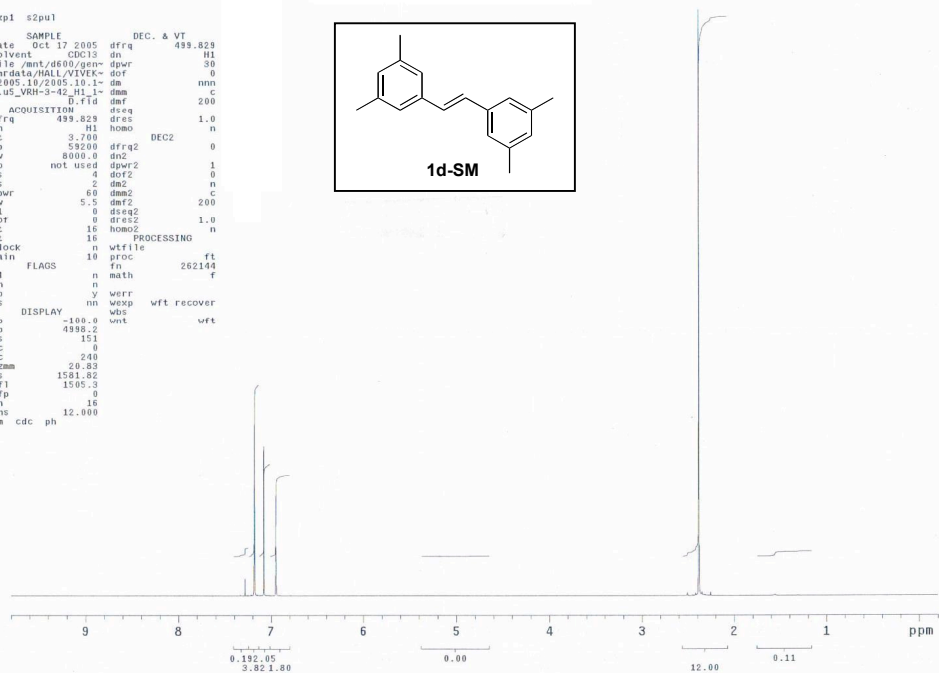
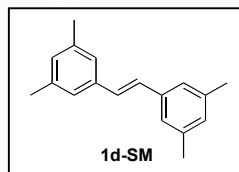
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Vivek VRH-3-42
499.8 MHz 1H in CDCl3

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sw 8000.0 dn2 0
fb not used dpwr2 1
ls 4 dof2 0
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pw 5.5 dm2 200
d1 0 dres2 1.0
TOT 0 dres2 1.0
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ct 16 PROCESSING
alock n vtfile ft
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in n
dp y verr wft recover
hs nm wexp wbs
DISPLAY nm wnt wft
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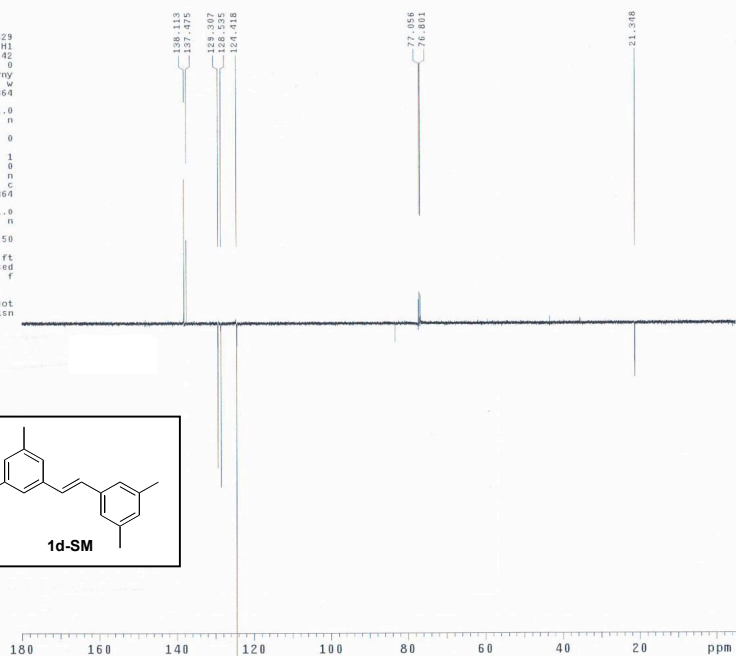
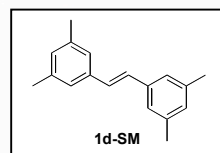


Vivek VRH-3-42
125.7 MHz 13C APT in CDCl3

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sw 30007.5 dn2 1
fb 16000 dpwr2 0
ls 4 dof2 0
ss 2 dm2 n
tpwr 57 dm2 c
pw 3.0 dm2 11364
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d2 0.007 homo2 n
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ct 298 proc not used
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il n verr
in n wexp
dp y wbs
hs nm wnt
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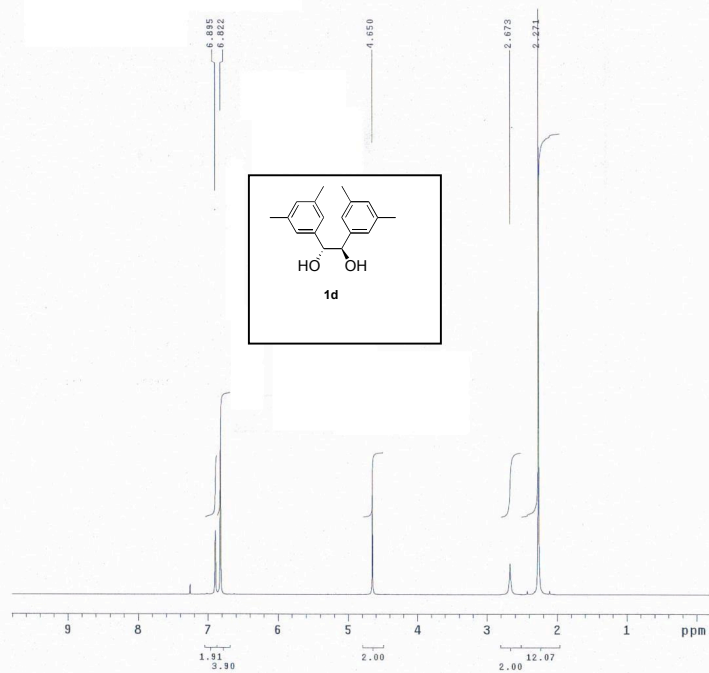
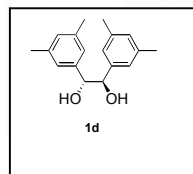
Vivek, VRH-5-14
399.85 MHz in CDCl3

exp11 etdih

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np        44932      dfrq2      0
sw        6000.6      dn2      1
fb        not used      dpr2      1
bs        4      dof2      0
ss        2      de2      n
tpwr      60      dnm2      c
pw        3.6      dar2      200
d1        0      dseq2      1.0
tof      10      dres2      n
nt        16      homo2      n
ct        16      PROCESSING
atock      n      vtfille
gain      20      proc      lp
          FLAGS      fn      not used
il        n      math      f
ln        n      verr
dp        y      werr
hs        nn      wexp      wft recover
          DISPLAY      vbi
sp        -77.0      wnt
wp        399.85
vs        151
sc        0
wc        180
hzmm      22.21
ls        4509.76
rfl        3900.5
rfp        2902.6
th        2.000
ins      nm cdc ph

```



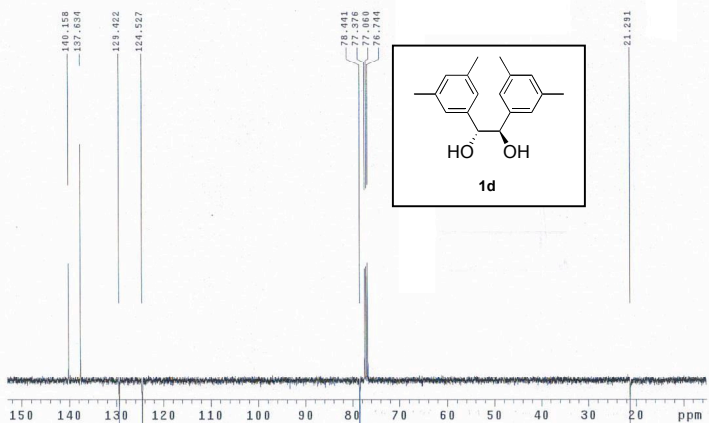
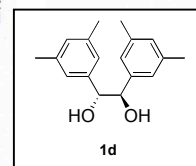
Vivek, VRH-5-14
100.58 MHz APT in CDCl3

exp10 apt

```

SAMPLE          DEC. & VT
date    Oct 19 2005   dfrq    399.855
solvent  CDCl3      dn      H1
file    /mnt/d600/gen- dpwr    40
nmrdata/HALL/VIVEK- dor      0
/2005.10/2005.10.1- da      yny
9.84_VRH-5-14_C13- dnm      10582
          apt. rfd      def
ACQUISITION      dseq      1.0
sfreq    100.555      dres      n
tn        C13      homo
at        2.005      DEC2      0
np        108586      dfrq2      0
sw        27081.9      dn2      1
fb        not used      dpr2      1
bs        4      dof2      0
tpwr      55      dnm2      c
pw        3.2      dar2      10582
p1        17.6      dseq2      1.0
d2        0.007      dres2      n
d3        0.000      homo2      n
tof      2048.3      PROCESSING
nt        50000      lb      1.00
ct        228      vtfille
alock      n      proc      ft
gain      not used      fn      not used
il        n      math      f
ln        n      verr
dp        y      wexp      procplot
hs        nn      wnt      teston
          DISPLAY      vbi
sp        519.3      wnt
wp        14862.1
vs        80
sc        0
wc        180
hzmm      82.57
ls        500.00
rfl        9746.6
rfp        7747.3
th        100.000
ins      nm no ph

```



Vivek, VRH-3-28
399.85 MHz 1H in CDCl3

exp10 std1h

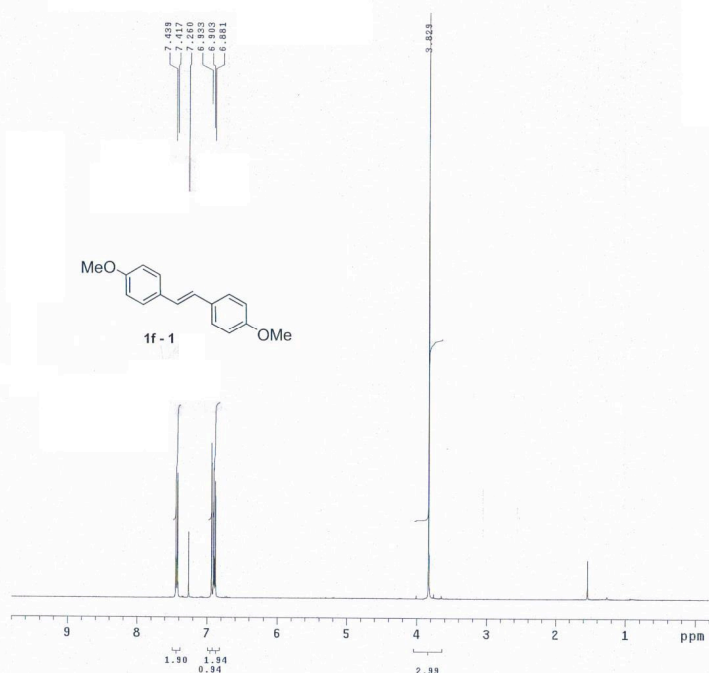
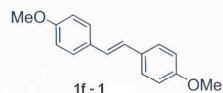
```

SAMPLE      DEC. & VT
date Oct 17 2005 dfrq 399.855
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 30
nmrdata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- dm nmh
7.s4_VRH-3-28_H1_1- dnm c
0.fid daf 200

ACQUISITION dseq 1.0
sfrq 399.855 dres n
tn H1 homo n
at 5.000 DEC2 0
np 6000.6 dfrq2 0
sv 6000.6 dn2 1
fb not used dpwr2 0
bs 4 dof2 0
ss 2 dm2 c
tpwr 6.0 dnm2 200
pw 3.6 dmf2 1.0
d1 0 dseq2 1.0
tof 0 dres2 n
nt 16 homo2 n
ct 16 PROCESSING n
alock n wtrfile 1p
gain 2.0 proc fn 131072
f

FLAGS n math
in n
dn n
hs nm wexp wft recover
DISPLAY -80.0 wnt
sp 3998.5
vs 840
sc 0
wc 180
h2mm 22.21
ls 15658.56
rf1 999.1
rfp 0
th 12
ins 3.000
at cdc ph

```



Vivek, VRH-3-28
100.58 MHz 13C[1H] in CDCl3

exp10 apt

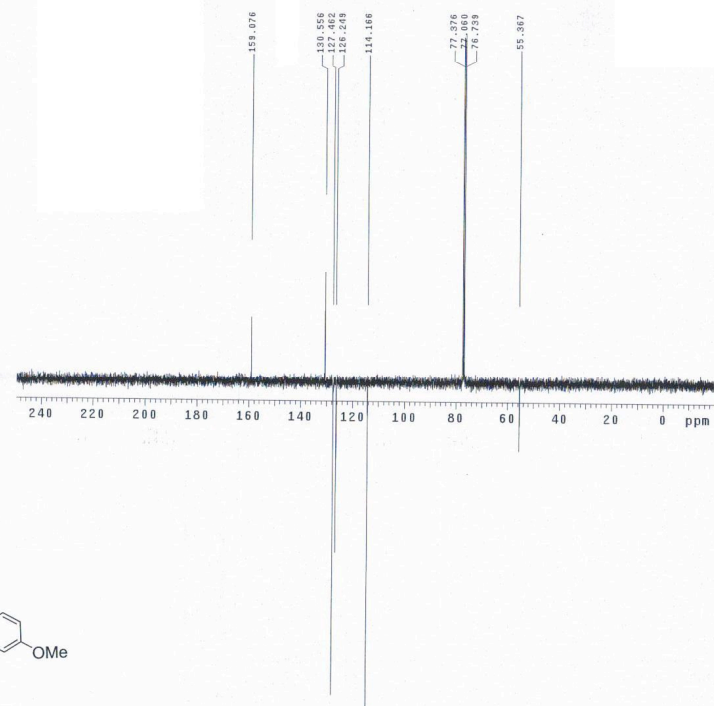
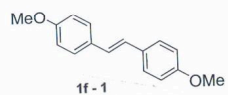
```

SAMPLE      DEC. & VT
date Oct 17 2005 dfrq 399.855
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 40
nmrdata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- dm yny
7.s4_VRH-3-28_C13- dnm 10582
0.fid daf 10582

ACQUISITION dseq 1.0
sfrq 100.555 dres n
tn C13 homo DEC2 0
at 2.000
np 108328 dfrq2 0
sv 27081.9 dn2 1
fb not used dpwr2 0
bs 4 dof2 0
ss 55 dm2 c
tpwr 3.2 dnm2 10582
pw 17.6 dmf2 1.0
d1 0.500 dseq2 1.0
d2 0.007 dres2 n
d3 0.000 homo2 n
tof 2040.3 PROCESSING 1.00
nt 50000 lb
ct 768 wtrfile ft
alock n proc fn not used
gain not used fn not used
f

FLAGS n math
in n werr autosave_fid
dn n y wexp procplot
hs nm vbs autosave_fid
DISPLAY -1995.0 wnt
sp 27081.9
vs 30
sc 0
wc 180
h2mm 2.50
ls 500.00
rf1 9743.3
rfp 7747.3
th 11
ins 100.000
nm no ph

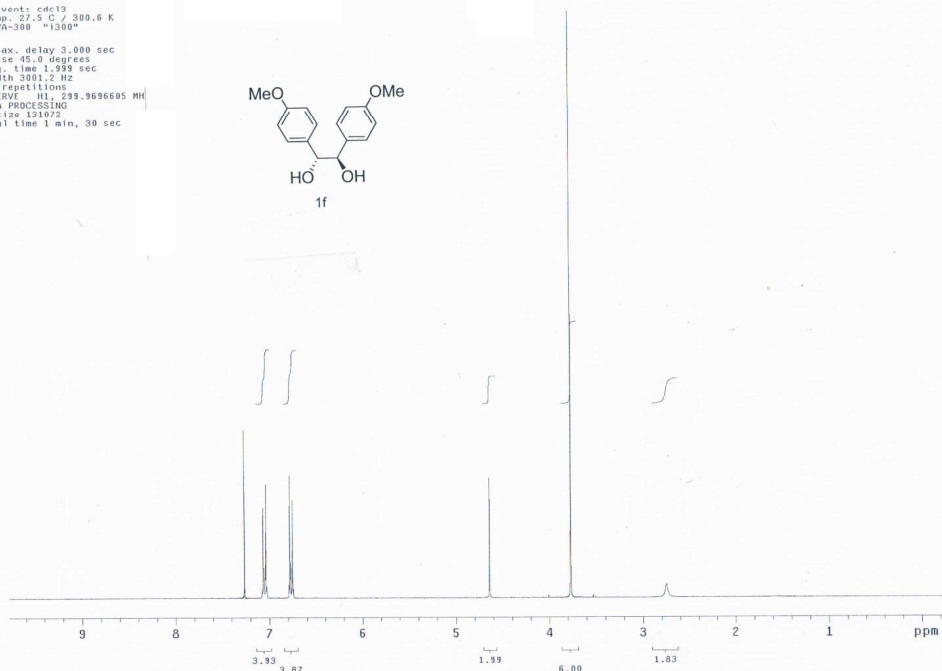
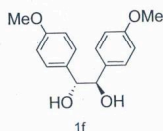
```



VRH-3-298
300 MHz 1D in CDCl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.5 C -> actual temp = 27.0 C, id probe

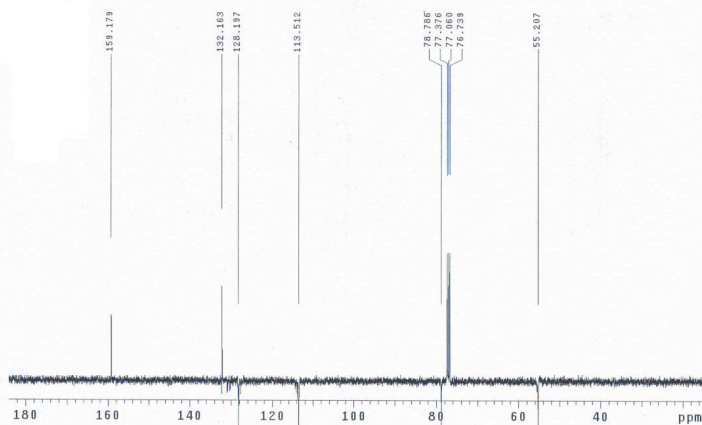
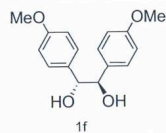
Pulse Sequence: s2pu1

Solvent: cdc13
Temp: 27.5 C / 300.6 K
INOVA-300 "1300"
Relax. delay 3.000 sec
Pulse 45.0 degrees
Acq. time 1.999 sec
Width 3001.2 Hz
16 repetitions
OBSERVE H1 299.965605 MHz
DATA PROCESSING
FT size 131072
Total time 1 min, 30 sec



Vivek, VRH-3-29
100.58 MHz 13C{1H} in CDCl3
exp11 apt

SAMPLE DEC. & VT 399.855
date Oct 17 2005 dfreq H1
solvent CDCl3 dn 40
file /mnt/d600/gsm- dper 0
nadata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- de vny
7.54_VRH-3-29_C13- dma W
apt- f1d def 10582
ACQUISITION dseq
sfrq 100.555 dras 1.0
in C13 homo n
at 2.000 DEC2 0
np 108328 dfreq2
sw 27081.3 dn2 1
fb not used dper2 0
bs 4 dof2 0
tpwr 55 dm2 n
pw 3.2 dm2 c
p1 17.6 dm2 10582
d1 0.500 dseq2 1.0
d2 0.007 dres2
d3 0.007 homo2 n
tof 2048.3 PROCESSING
nt 50000 lb 1.00
ct 285 vfile
alock n proc ft
gain not used fn not used f
FLAGS n
in n verr
dp y vexp
hs nm vbi
DISPLAY 1422.4
wp 17073.7
vs 9
sc 180
wc 94.85
is 500.00
rfl 9746.5
rfp 7747.3
th 7
ins 100.000
na no ph

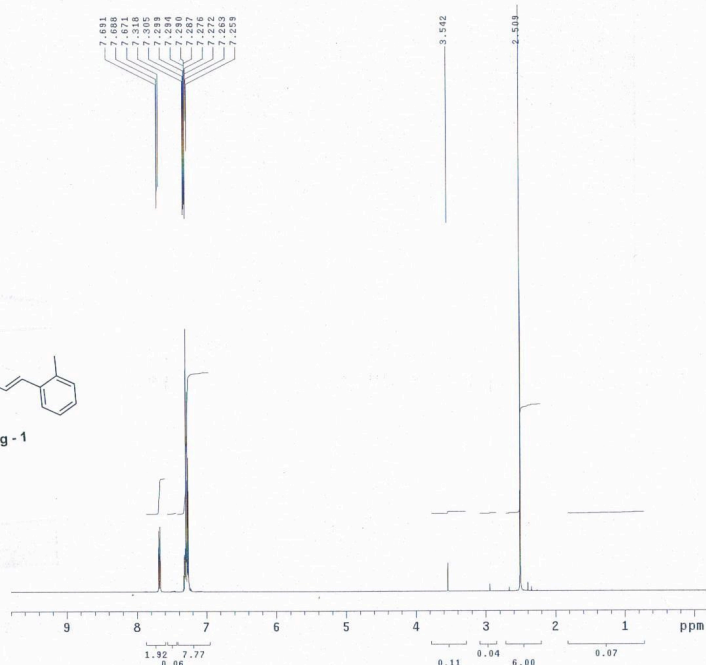
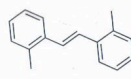


Vivek, VRH-3-17
399.85 MHz 1H in CDCl3

```

exp10 std1h
SAMPLE
date Oct 17 2005 dfrq 399.855
solvent CDCl3 dn H1
file /mnt/d600/gen- dpr 30
nmrdata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- dm nm
7-s4_VRH-3-17_11-1- dam c
D.fid def 200
ACQUISITION
sfrq 399.855 dres 1.0
tn H1 homo DEC2 n
at 5.000 dfrq2 0
np 60006 dno 0
sw 6000.6 dpr2 1
fb not used dpr2 1
bs 4 dot2 0
ss 2 dm2 n
tpwr 60 dm2 c
pw 3.6 dm2 200
d1 0 dseq2 1.0
tor 0 dres2 n
nt 16 homo2 n
ct 16 PROCESSING
alock n wtf11e lp
gain 20 proc fn 131072
FLAGS n math f
il n
dp y verr wft recover
hs nm wexp vbs
DISPLAY
sp -88.0
vp 3998.5
vs 107
sc 0
wc 180
hzmm 22.51
ls 957.46
rfi 398.1
rfp 0
ths 5
ins 6.000
al cdc ph

```

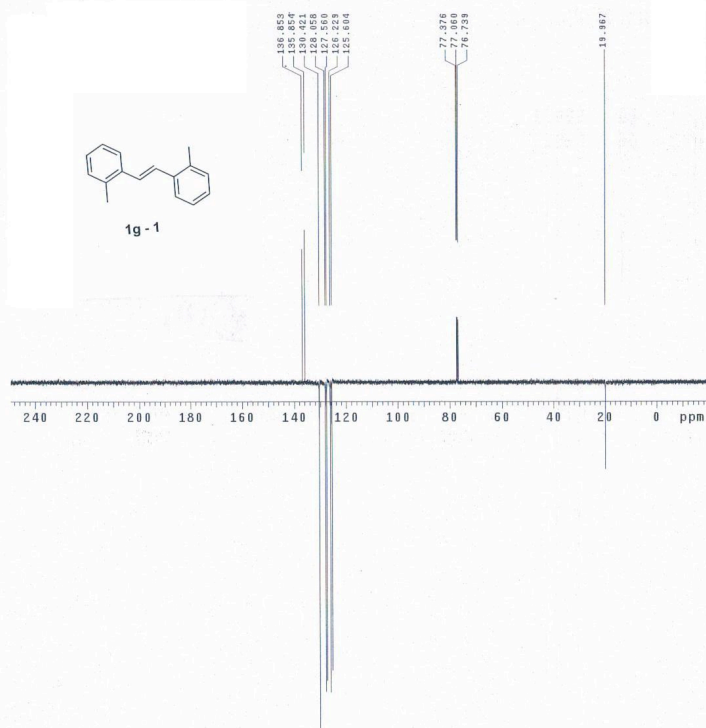
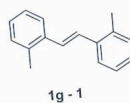


Vivek, VRH-3-17
100.58 MHz 13C[1H] in CDCl3

```

exp11 apt
SAMPLE
date Oct 17 2005 dfrq 399.855
solvent CDCl3 dn H1
file /mnt/d600/gen- dpr 30
nmrdata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- dm yny
7-s4_VRH-3-17_11-1- dam w
apt.fid def 10582
ACQUISITION
sfrq 100.555 dres 1.0
tn C13 homo DEC2 n
at 2.000 dfrq2 0
np 108328 dno 0
sw 27081.9 dpr2 1
fb not used dpr2 1
bs 4 dot2 0
ss 55 dm2 n
tpwr 3.2 dm2 c
pw 17.6 dm2 10582
d1 0.500 dseq2 1.0
d2 0.007 dres2 n
d3 0.009 homo2 n
tor 2048.9 PROCESSING
nt 50000 lb wtf11e 1.00
ct 148 n math ft
alock not used n not used
gain FLAGS n math f
il n
dp y verr procplot
hs nm wexp testin
DISPLAY
sp -2086.2
vp 27081.5
vs 30
sc 0
wc 180
hzmm 6.22
ls 9754.5
rfi 7747.3
rfp 4
ths 100.000
nm no ph

```

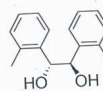


Vivek VRH-3-23
499.8 MHz 1H in CDCl3

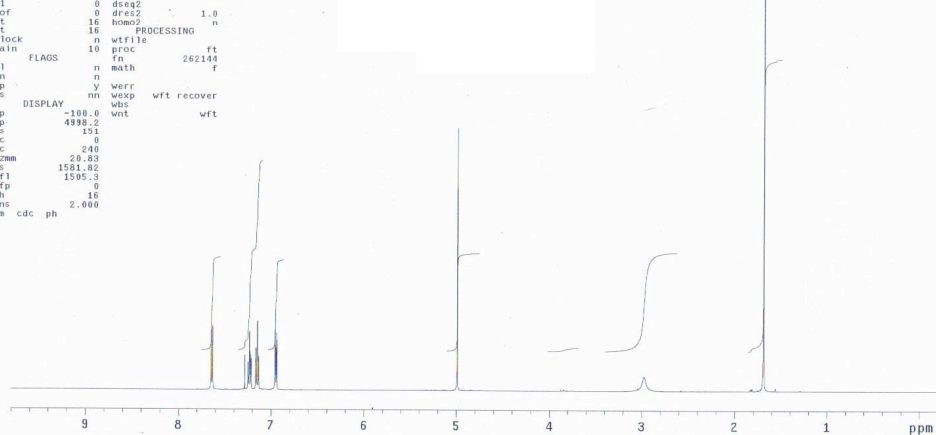
```

exp1 s2pul
SAMPLE
date Oct 17 2005 dfrq 499.829
solvent CDCl3 dn H1
file /mnt/ds00/gen- dpwr 30
nmrdata/HALL/VIVEK- dcf 0
/2005.10/2005.10.1- dm nnn
7-us_VRH-3-23_H1- dam c
0. f1d daf 200
ACQUISITION
sfrq 499.829 dres 1.0
tn H1 homo n
at 3.700 DEC2 0
np 58280 dfrq2 0
sw 8000.0 dn2 0
fb not used dpwr2 1
ls 4 dof2 0
ss 2 dm2 n
tpwr 60 dm2 c
pw 5.5 daf2 200
d1 0 dres2 1.0
tof 0 dres2 n
nt 16 homo2 n
ct 16 PROCESSING
alock n wfile ft
gain 10 proc tn 262146
il n math f
in n
dp y verr wft recover
hs nm wss wft
DISPLAY
sp -100.0 wnt wft
wp 499.829
vs 151
sc 9
wc 240
h2ma 20.83
ls 1581.82
rf1 1585.3
rtp 9
th 16
nm cdc ph 2.000

```



1g

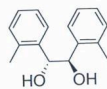


Vivek VRH-3-23
125.7 MHz 13C APT in CDCl3

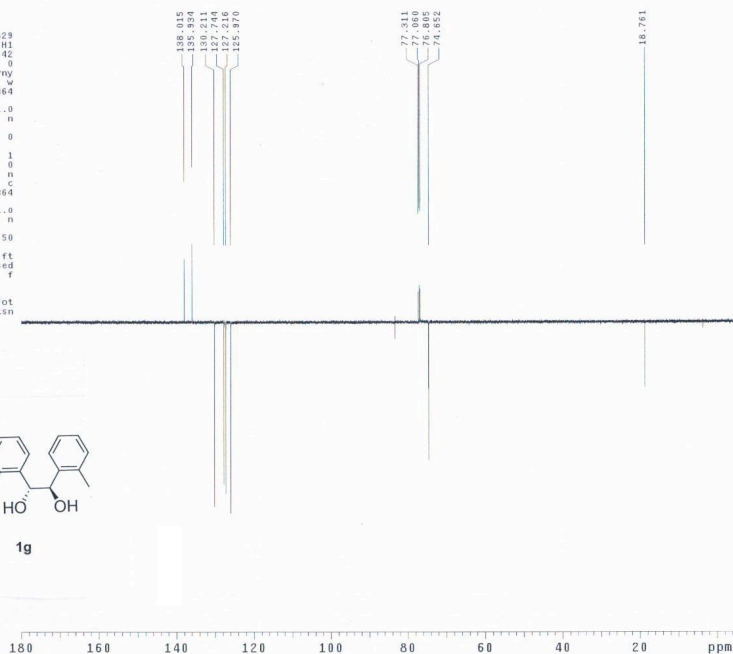
```

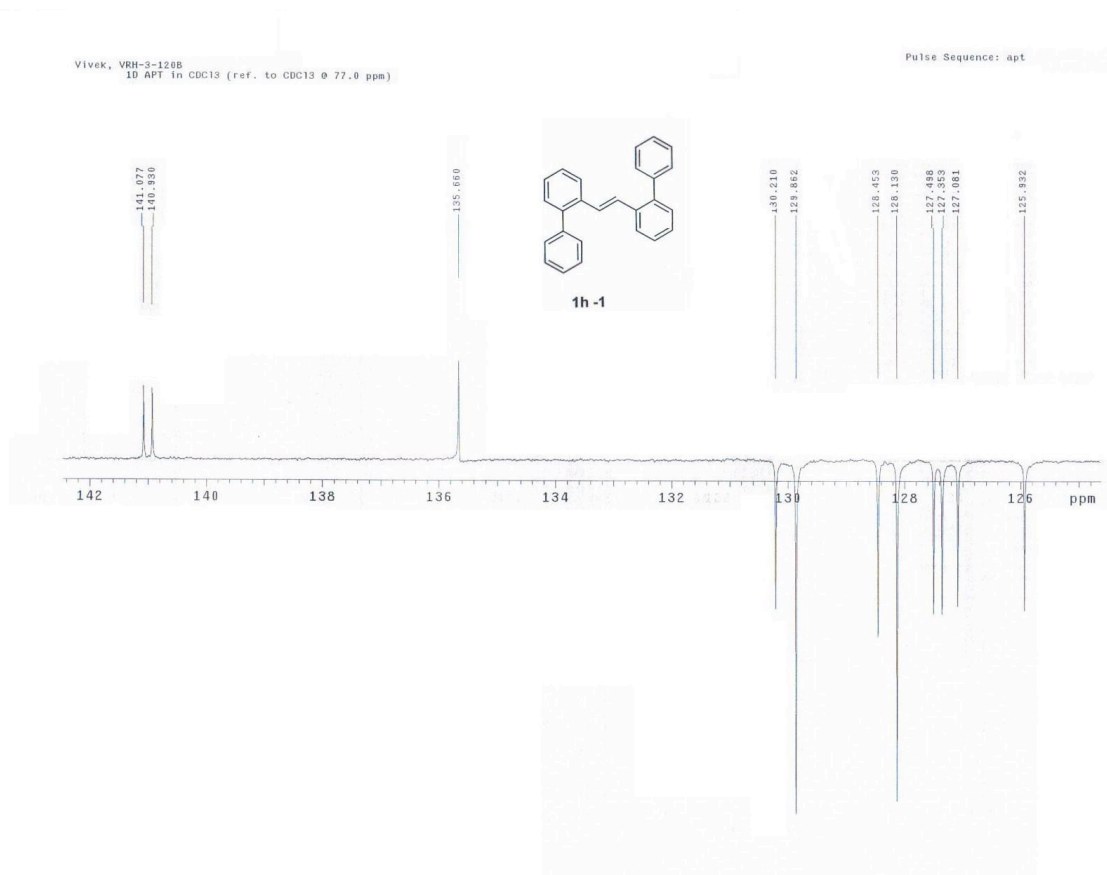
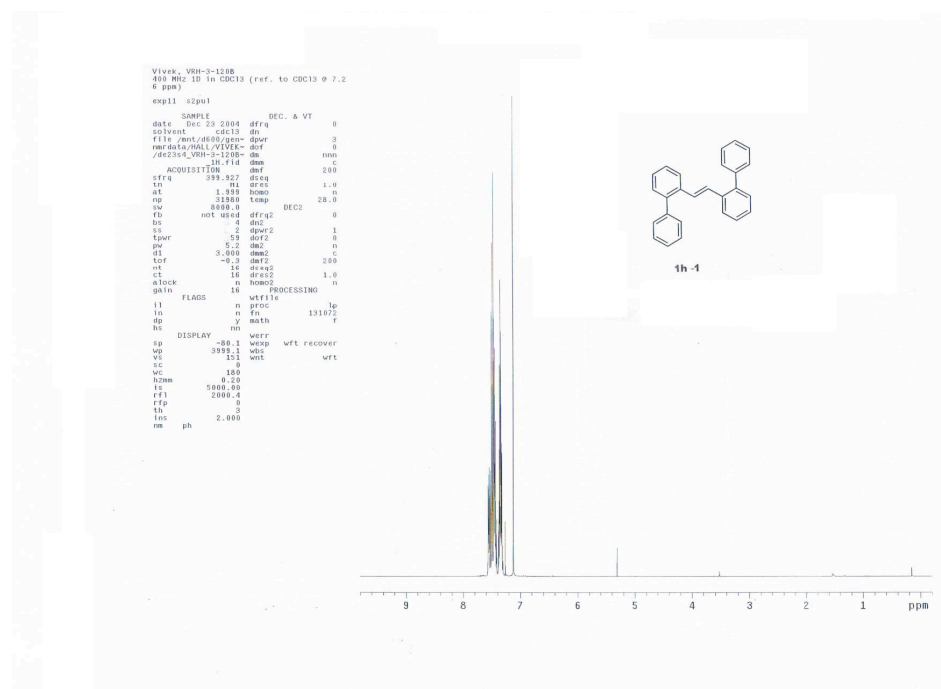
exp2 apt
SAMPLE
date Oct 17 2005 dfrq 499.829
solvent CDCl3 dn H1
file /mnt/ds00/gen- dpwr 42
nmrdata/HALL/VIVEK- dcf 0
/2005.10/2005.10.1- dm yny
7-us_VRH-3-23_C13- dam w
0. f1d daf 11364
ACQUISITION
sfrq 125.694 dres 1.0
tn C13 homo n
at 2.000 DEC2 0
np 120030 dfrq2 0
sw 30007.5 dn2 0
fb 10000 dpwr2 1
ls 4 dof2 0
ss 2 dm2 n
tpwr 57 dm2 c
pw 13.8 daf2 11364
d1 0 dres2 1.0
d2 0.007 homo2 n
d3 0.000 PROCESSING
tof 630.0 lb 0.50
nt 50000 wfile ft
ct 252 proc not used
alock n tn f
gain 40 math f
il n verr
in nm wss
dp y vss
hs nm wnt
DISPLAY
sp -628.8
wp 23250.9
vs 49
sc 0
wc 185
h2ma 125.68
ls 500.00
rf1 12146.8
rtp 3655.0
th 7
nm cdc ph 100.000

```



1g

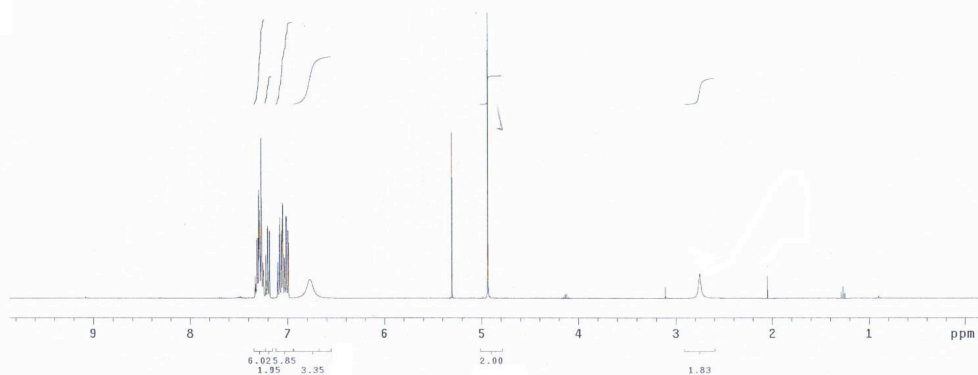
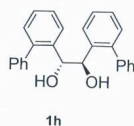




VRH-3-1218
400 MHz 1D in CDCl3 (ref. to CDCl3 @ 7.26 ppm), temp 26.8 C -> actual temp = 27.0 C, asw400 probe

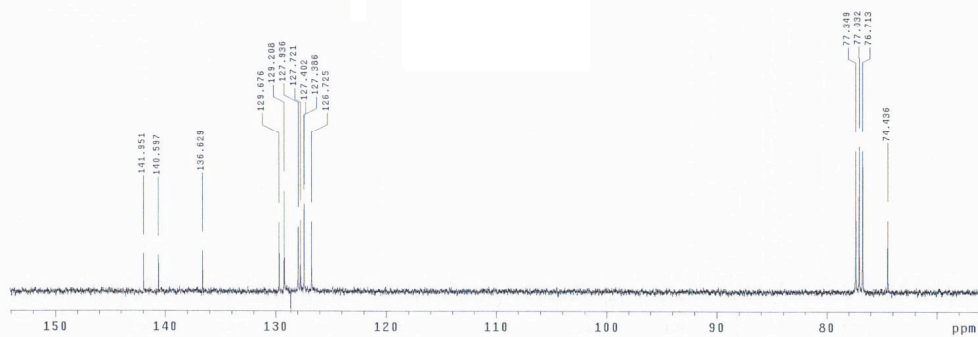
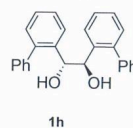
Pulse Sequence: s2pu1

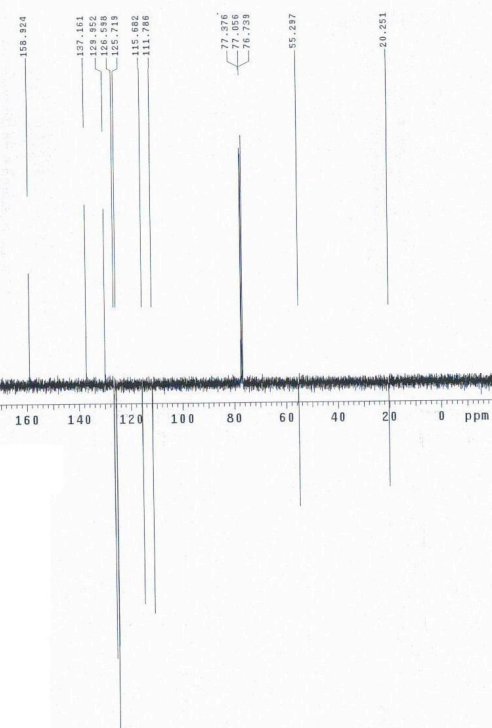
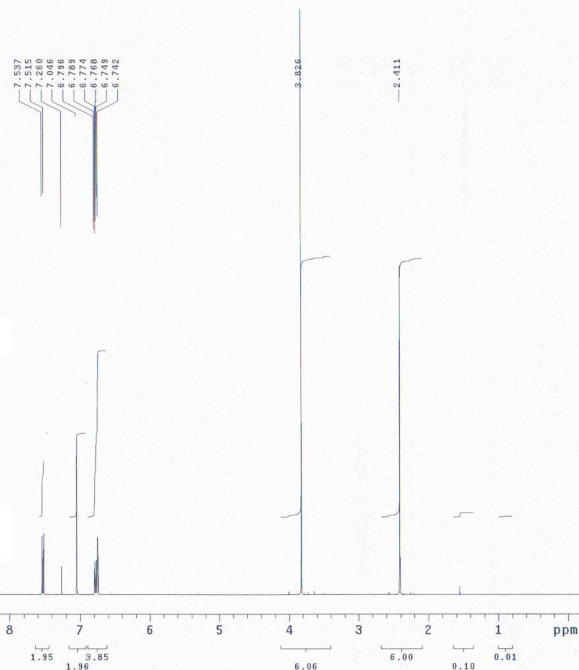
Solvent: cdcl3
Temp: 26.8 C / 299.9 K
INNOVA-400 "1400"
Relax. delay 3.000 sec
Pulse 45.0 degrees
Acq. time 1.959 sec
Width 4001.6 Hz
16 Repetitions
OBSERVE H1, 399.7923305 MHz
DATA PROCESSING
FT size 131072
Total time 1 min, 30 sec



VRH-3-1218
100 MHz 1D C13 in CDCl3 (ref. to CDCl3 @ 77.0 ppm), temp 26.8 C -> actual temp = 27.0 C, asw400 probe

Pulse Sequence: s2pu1

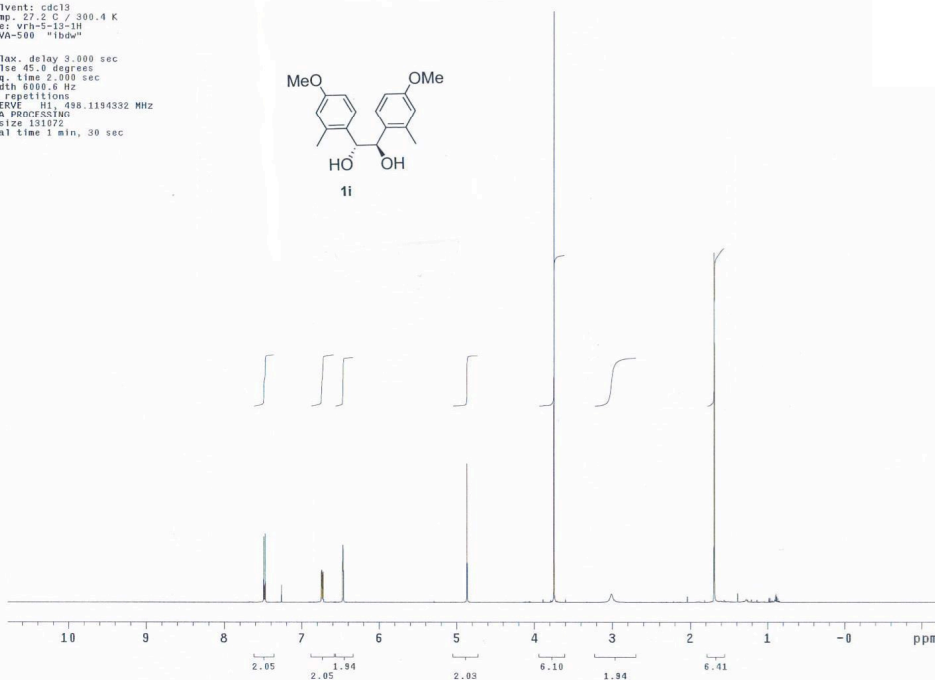
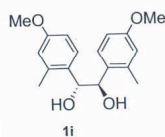




VRH-5-13
500 MHz 1D in CDCl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.2 → actual temp = 27.0 C, sw500 probe

Pulse Sequence: s2pu1

Solvent: cdcl3
Temp: 27.2 C / 300.4 K
File: vrh-5-13-1H
INDVA-500 "1bdw"
Relax. delay 3.000 sec
Pulse 45.0 degrees
Acq. time 2.000 sec
Width 8000.0 Hz
16 repetitions
OBSERVE H1: 498.1194392 MHz
DATA PROCESSING
F1 size 131072
Total time 1 min, 30 sec

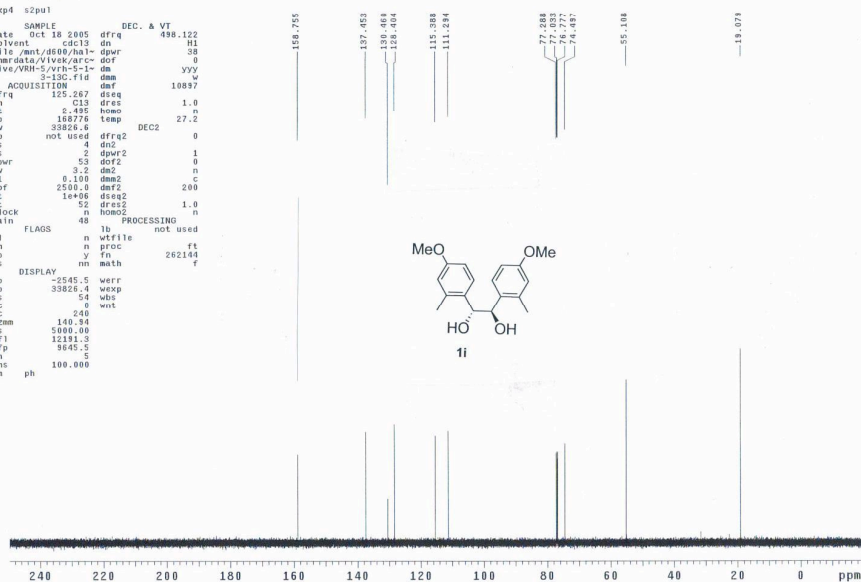
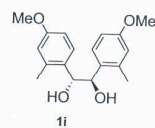


125 MHz 1D C13 in CDCl3 (ref. to CDCl3 @ 77.0 ppm), temp 27.2 C → actual temp = 27.0 C, sw probe
125 MHz 1D C13 in CDCl3 (ref. to CDCl3 @ 77.0 ppm), temp 27.2 C → actual temp = 27.0 C, sw probe

Pulse Sequence: s2pu1

exp4 s2pu1

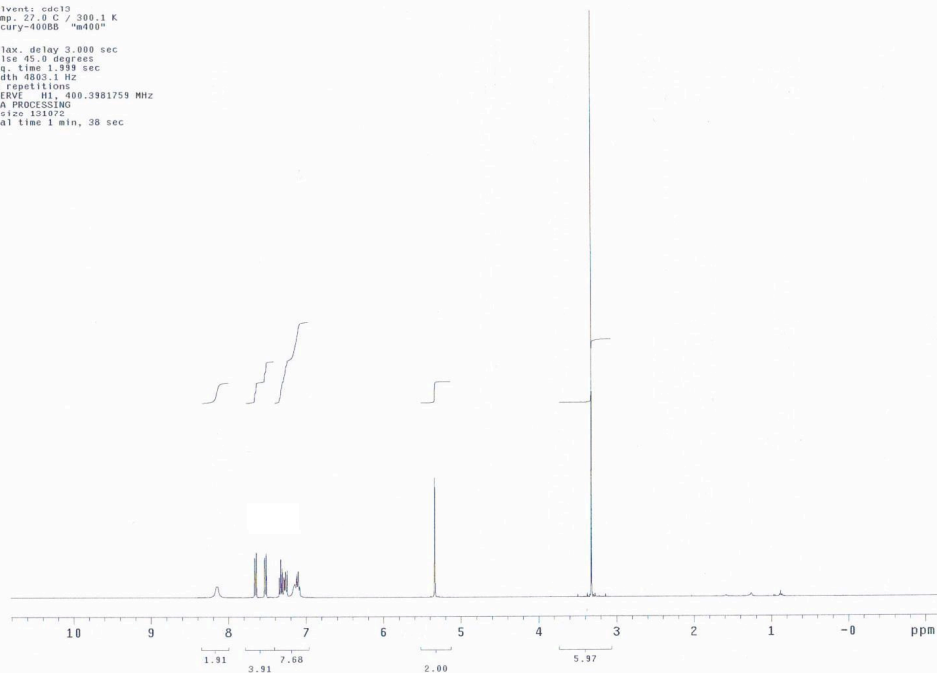
SAMPLE DEC. & VT 496.122
date Oct 18 2005 dfrq H1
solvent cdcl3 dn H1
file /mnt/d600/hal-dpur 38
lnmrdata/vivek/arcc- dcf 0
hive/vrh-5/vrh-5-1- dm 55V
3-13C.fid dm 10897
ACQUISITION
sfreq 125.267 dfeq 1.0
tn C13 dres 1.0
wt 1.495 homo n
np 168776 temp 27.2
sw 33926.6 DEC2
fb not used dfr2 0
bs 4 dm2 1
ss 2 dpwr2 0
tqwr 53 dcf2 0
pw 3.2 dm2 n
d1 0.100 dm2 c
tof 2500.0 dm2 200
nt 1e+60 dfeq2 1.0
ct 52 dres2
atock n homo2
gain 48 PROCESSING
il n lb not used
in n wfile
dp y fn 202144
hs nm math f
DISPLAY
sp -2545.5 werr
wp 33926.4 wexp
vs 54 wbs
rc 0 wnt
wc 240
h2nm 160.84
is 5000.00
rf1 12191.3
rfp 9645.5
th 5
ins 100.000
nm ph



VRH-5-15B
400 MHz 1D in CDCl3 (ref. to CDCl3 @ 7.26 ppm), temp 27.0 C -> actual temp = 27.0 C, m400gz probe

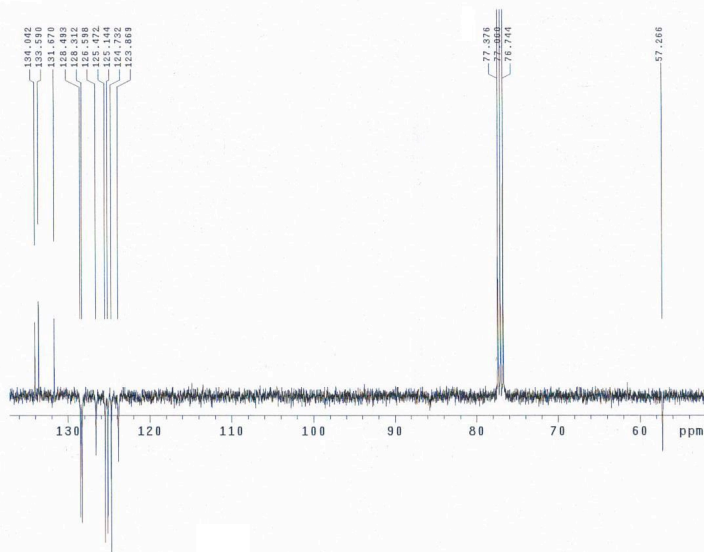
Pulse Sequence: s2pu1

Solvent: cdc13
Temp: 27.0 C / 300.1 K
Mercury-40000 "m400"
Relax. delay 3.000 sec
Pulse 45.0 degrees
Acq. time 1.939 sec
Width 4000.1 Hz
12 repetitions
OBSERVE H1: 400.3981759 MHz
DATA PROCESSING
FT size 131072
Total time 1 min, 38 sec



Vivek, VRH-5-15B
100.58 MHz APT in CDCl3

expt10 apt
SAMPLE DEC. & VT 399.855
date Oct 19 2005 dfrs H1
solvent CDCl3 dn
file /mnt/d600/gpr-dpwr 40
nmrdata/HALL/VIVEK- dof 0
/2005.10/2005.10.1- dm yny
S-st_VRH-5-15B_C13- dm w
apt.fid dmf 10582
ACQUISITION
sfrs 100.555 dres 1.0
ln C13 homo n
at 2.005 DEC2 0
np 180586 dfrq2 0
sw 27091.9 dms
fb not used dpwr2 1
ls 4 dof2 0
tpwr 55 dm2 n
p1 17.6 dm2 10582
d1 0.500 dseq2 c
d2 0.007 dres2 1.0
d3 0.000 homo2 n
vof 2040.3 PROCESSING 1.00
nt 50000 lb
ct 728 vtrile
clock 728 n proc ft
gain not used fn not used
flags FLAOS math f
il n
in n werr
dp y wexp procplot
hs nn testn
wnt
DISPLAY
sp 5197.8
vp 8593.6
vs 111
sc 0
wc 180
hzam 47.72
ls 500.00
rfl 9743.7
rfo 7747.3
th 5
ins 100.000
nm no ph



VRH-5-Benzopinacotester
400.1 MHz 1H in CDCl3

exp3 std1h

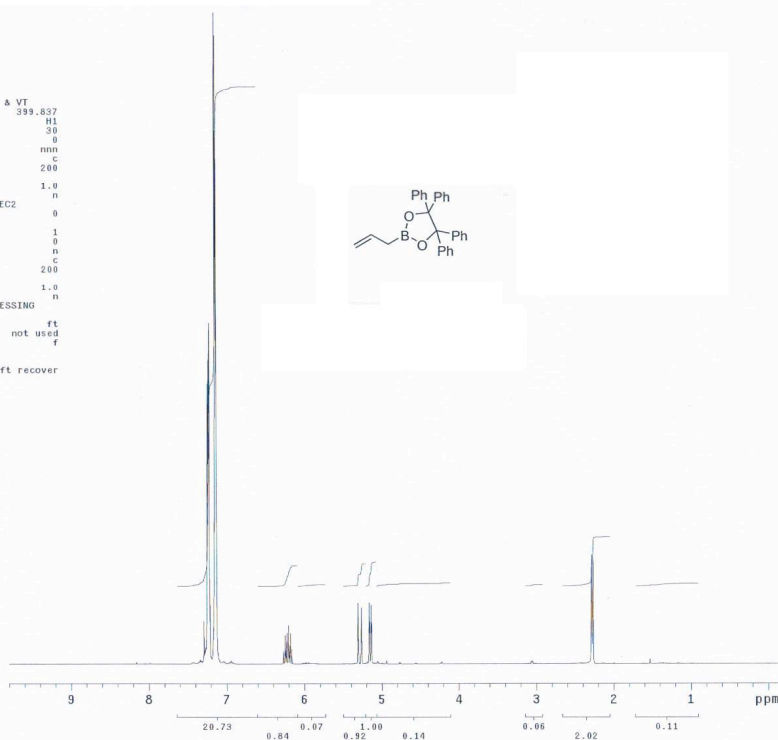
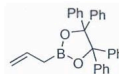
```

SAMPLE          DEC. & VT
date Dec 5 2005 dfrq 399.837
solvent CDCl3   dn    H1
file /mnt/d600/gen- dpr 30
nmrdata/HALL/VIVEK- dof 0
/2005.12/2005.12.0- dm  mn
5.54_VRH-5-Benzopi- dam  c
nacoester_H1_ID_1- dmf 200

ACQUISITION
sfrq 399.837 dres 1.0
tn      H1      hmo  n
at      3.744 dfrq2 DEC2 0
np      4432 dn2  0
sw      6000.6 dpwr2 1
fb      not used dof2 0
bs      4 dm2    n
tpwr    60 dam2  c
pw      3.6 dmf2  200
d1      0 dseq2  1.0
tcf      0 drac2  n
nt      16 hmo2  n
ct      16 PROCESSING
alock   n wtfle  ft
gain    20 proc  f
        fn      not used
        n math   f
        n werr
        nm wexp  wft recover
        wbs
        wnt

DISPLAY
sp      -80.1
vp      3998.3
va      195
sc      0
wc      200
hzmm    19.39
ls      1733.56
rf1     3859.5
rfp     2858.8
th      20
ins     1.000
nm cdc ph

```



VRH-5-Benzopinacotester. APT
100.58 MHz 13C[M] in CDCl3

exp12 apt

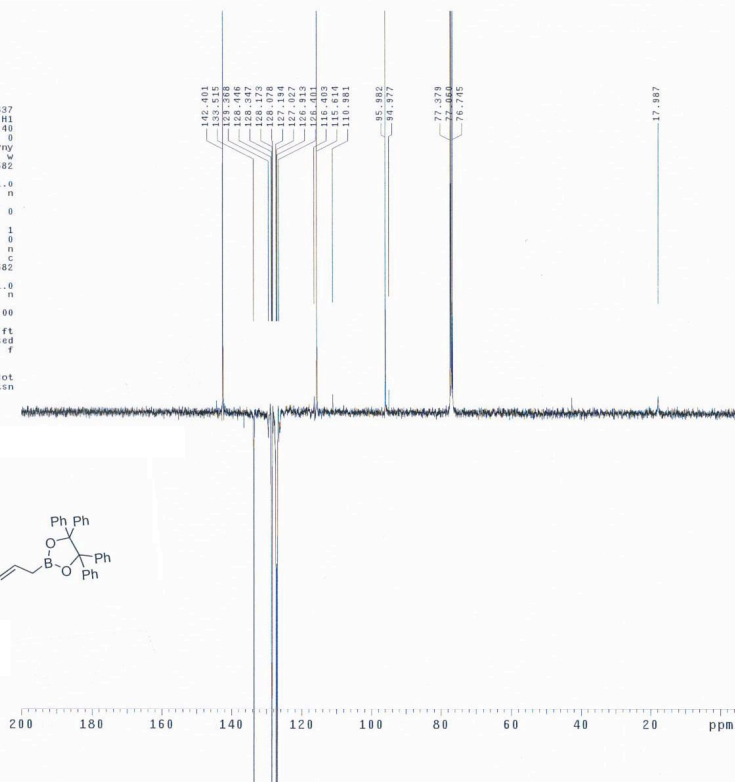
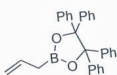
```

SAMPLE          DEC. & VT
date Dec 5 2005 dfrq 399.837
solvent CDCl3   dn    H1
file /mnt/d600/gen- dpr 40
nmrdata/HALL/VIVEK- dof 0
/2005.12/2005.12.0- dm  yny
5.54_VRH-5-Benzopi- dam  w
nacoester_C13_apt- dmf 10582

ACQUISITION
sfrq 100.548 dres 1.0
tn      C13      hmo  n
at      2.000 dfrq2 DEC2 0
np      100000 dn2  0
sw      25000.0 dpwr2 1
fb      not used dof2 0
bs      4 dm2    n
tpwr    55 dam2  c
pw      3.2 dmf2  10582
p1      17.6 dseq2  1.0
d1      2.000 dres2  n
d2      0.007 hmo2  n
d3      0.000 PROCESSING
tcf      0 lb      2.00
nt      50000 wtfle  ft
ct      2000 proc  f
alock   n fn      not used
gain    not used math  f
        n werr
        n wexp  procplot
        y wbs   testen
        nm wnt

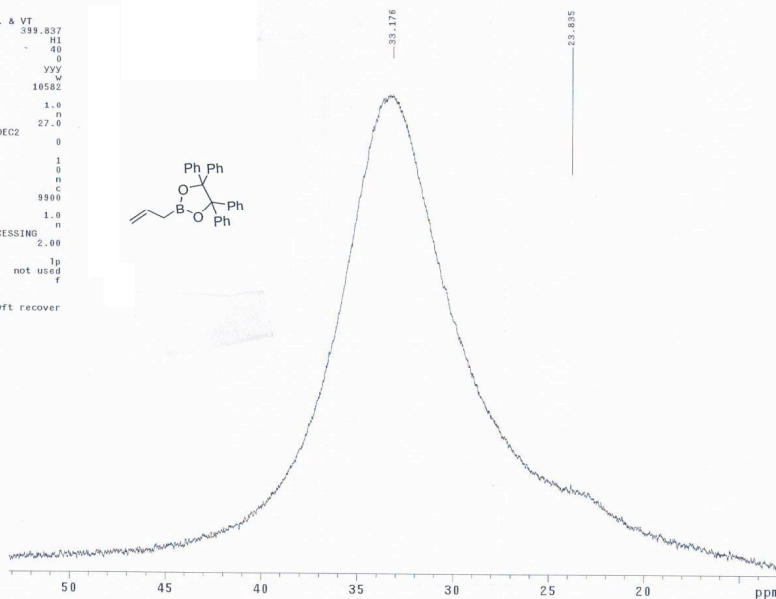
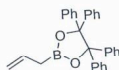
DISPLAY
sp      -502.9
vp      20610.0
va      1456
sc      0
wc      185
hzmm    111.41
ls      500.00
rf1     19743.2
rfp     7747.5
th      100.000
ins     100.000
nm no ph

```



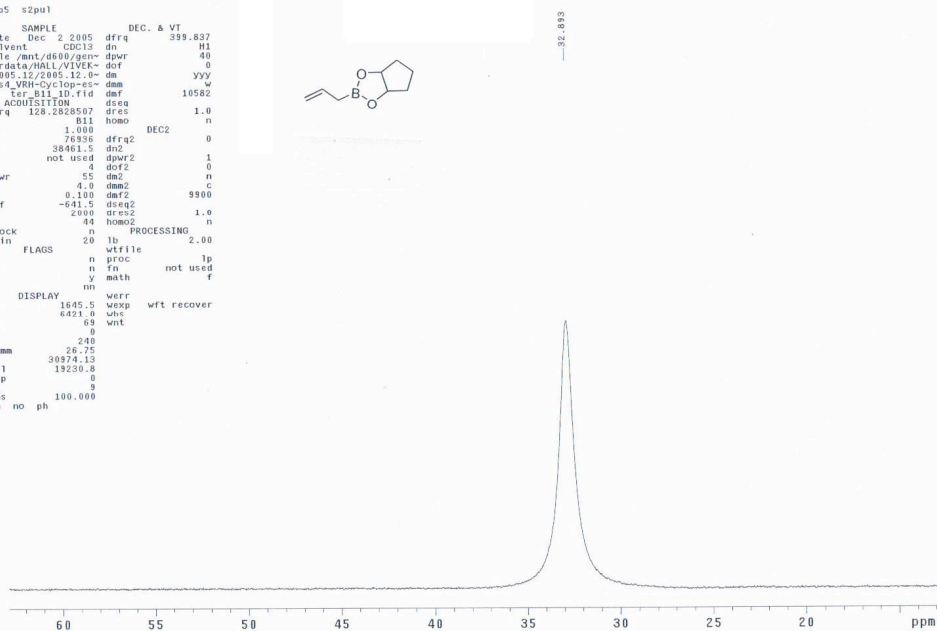
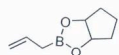
Vivek, VRH-5-Benzopinacotester
128.3 MHz 1H[1H] in CDCl3

```
exp5 s2pu1
SAMPLE
date Dec 5 2005 dfrq DEC. & VT 399.837
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 40
nmrdata/HALL/VIVEK- dof 0
/2005.12/2005.12- de yyy
5.s4_VRH-5-Benzopi- dnm w
nacotester_B11_10- der 10582
ACQUISITION f1d dseq
sfrq 128.2828507 dres 1.0
tn B11 homo n
at 1.000 DEC2 0
np 76900 dfrq2 0
sw 38461.5 dn2 1
fb not used dpwr2 1
bs 4 dof2 0
tpwr 55 dm2 n
pw 4.0 dnm2 c
d1 0.100 dm2 9900
tof -641.6 dseq2 1.0
nt 2000 dres2 n
ct 604 homo2 n
alock n PROCESSING 2.00
gain 20 lb
FLAGS n wfile lp
in n fn not used
dp y math f
hs nm
DISPLAY werr
sp 1622.1 wexp wft recover
vp 5136.2 wds
vs 119 wnt
sc 0
wc 240
hzmm 25.98
ls 38974.13
rf1 19230.8
rfp 0
th 15
ins 100.000
nm no ph
```



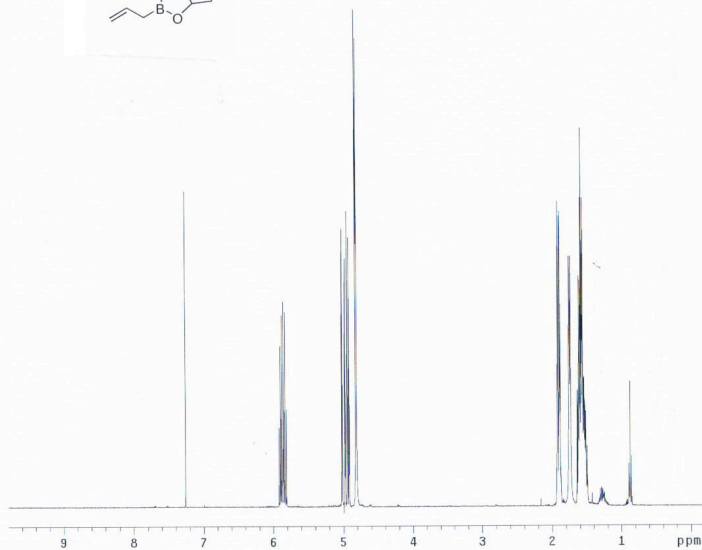
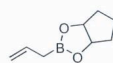
Vivek, VRH-Cyclop-ester
128.3 MHz 1H[1H] in CDCl3

```
exp5 s2pu1
SAMPLE
date Dec 2 2005 dfrq DEC. & VT 399.837
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 40
nmrdata/HALL/VIVEK- dof 0
/2005.12/2005.12- de yyy
2.s4_VRH-Cyclop-ester- dnm w
ter_B11_10- f1d der 10582
ACQUISITION f1d dseq
sfrq 128.2828507 dres 1.0
tn B11 homo n
at 1.000 DEC2 0
np 76900 dfrq2 0
sw 38461.5 dn2 1
fb not used dpwr2 1
bs 4 dof2 0
tpwr 55 dm2 n
pw 4.0 dnm2 c
d1 0.100 dm2 9900
tof -641.5 dseq2 1.0
nt 2000 dres2 n
ct 44 homo2 n
alock n PROCESSING 2.00
gain 20 lb
FLAGS n wfile lp
in n fn not used
dp y math f
hs nm
DISPLAY werr
sp 1645.5 wexp wft recover
vp 6431.0 wds
vs 69 wnt
sc 0
wc 240
hzmm 26.75
ls 38974.13
rf1 19230.8
rfp 0
th 9
ins 100.000
nm no ph
```



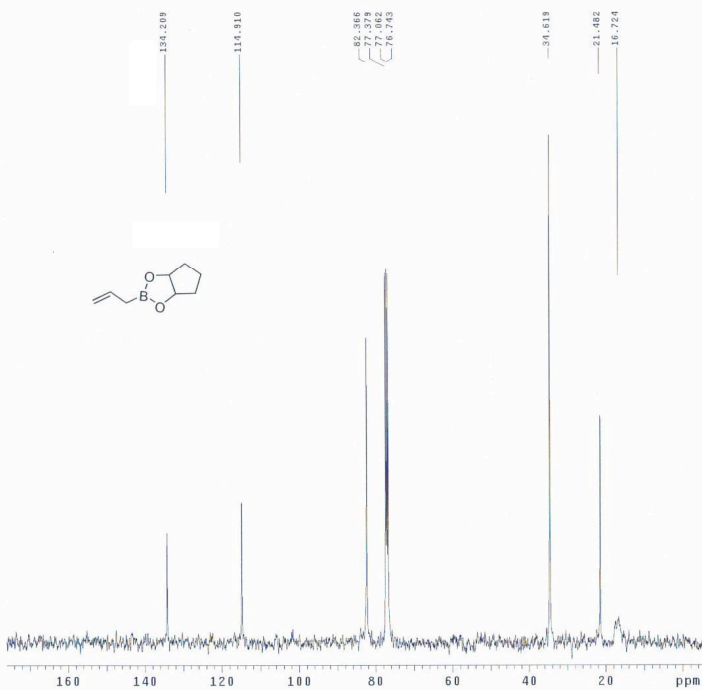
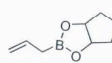
Vivek, VRH-Cyclop-ester
400.1 MHz 1H in CDCl3

```
exp9 std1h
SAMPLE          DEC. & VT
date Dec 2 2005 dfrq 399.827
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 18
nmrdata/HALL/VIVEK- dof -54.2
/2005.12/2005.12.0- dm n
2.s4_VRH-5-Cyclop- dnm c
ester_H1_10.fid dmf 200
ACQUISITION dseq
sfrq 399.837 dres 1.0
tn H1 homo DEC2 n
at 3.744
np 44932 dfrq2 0
sw 6000.6 dn2
fb not used dpwr2 1
bs 2 dof2 0
ss 2 dm2 n
tpwr 60 dnm2 c
pw 3.6 dm2 200
d1 0 dseq2 1.0
cor 0 dres2 n
nt 16 homo2 n
ct 16 PROCESSING
alock n wtf file 1p
gain 20 proc not used f
l1 n math
ln n
dp y werr
hs DISPLAY nn wexp wft recover
sp -83.6 wnt
vp 3998.3
vs 128
sc 0
wc 180
hzmm 22.21
ls 3760.00
rfi 3907.1
rfp 2902.8
th 20
ins 100.000
nm cdc ph
```

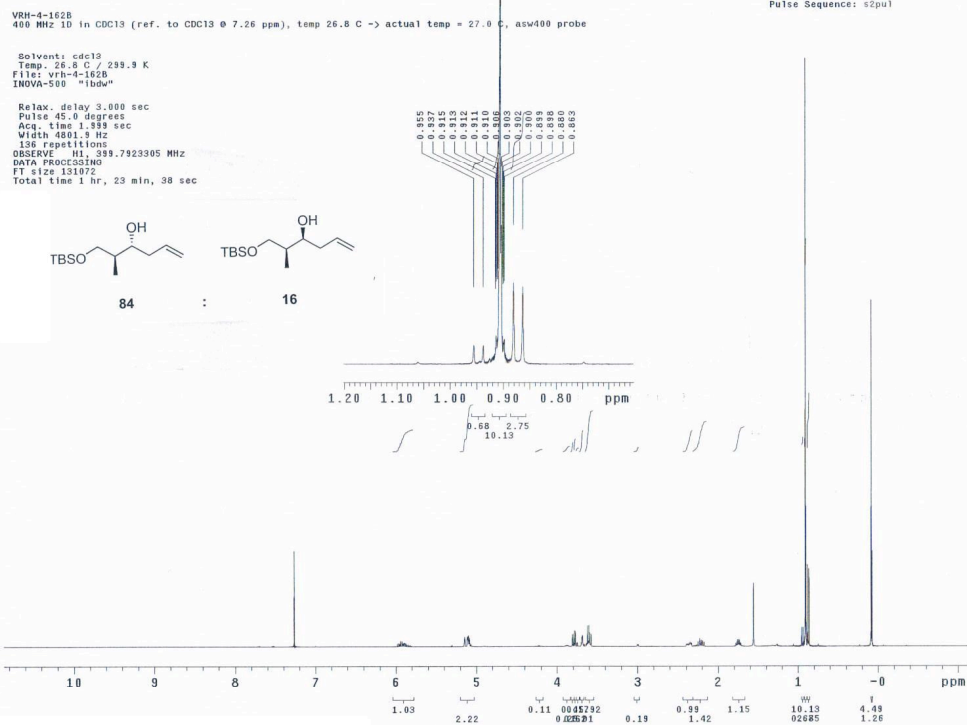


Vivek, VRH-Cyclop-ester
100.55 MHz 13C[1H] in CDCl3

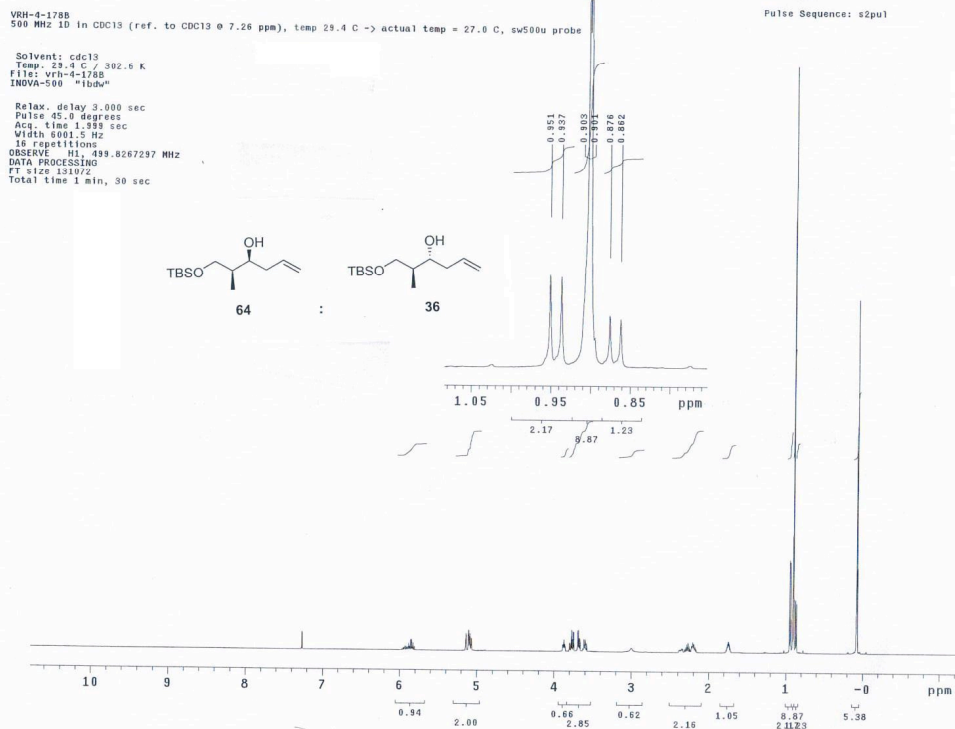
```
exp11 std13c
SAMPLE          DEC. & VT
date Dec 2 2005 dfrq 399.837
solvent CDCl3 dn H1
file /mnt/d600/gen- dpwr 40
nmrdata/HALL/VIVEK- dof 0
/2005.12/2005.12.0- dm yyy
2.s4_VRH-Cyclop-5- dnm w
Ter_c15_10.fid dmf 10502
ACQUISITION dseq
sfrq 100.548 dres 1.0
tn C13 homo n
at 2.000
np 100000 dfrq2 0
sw 25000.0 dn2 DEC2
fb not used dpwr2 1
bs 4 dof2 0
tpwr 55 dm2 n
pw 3.2 dnm2 c
d1 0 dm2 9900
cor 0 dseq2 1.0
nt 100000 dres2 n
ct 152 homo2 n
alock n PROCESSING
gain not used lb 10.00
l1 n wtf file
ln n proc ft
dp y math 26210d
hs nm f
DISPLAY
sp -565.1 werr
vp 10054.7 wexp
vs 132 wnt
sc 0
wc 180
hzmm 3.56
ls 500.00
rfi 10752.5
rfp 7747.5
th 5
ins 100.000
nm no ph
```



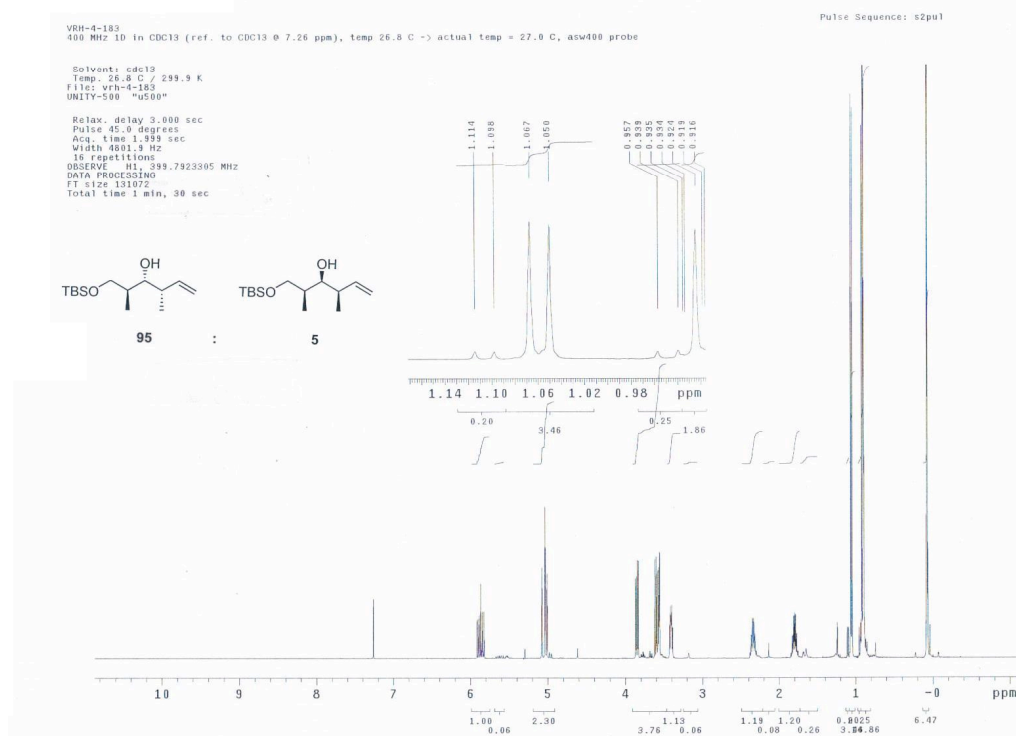
Diastereoselective Allylation with (*R,R*)-diol 1J and SnCl₄



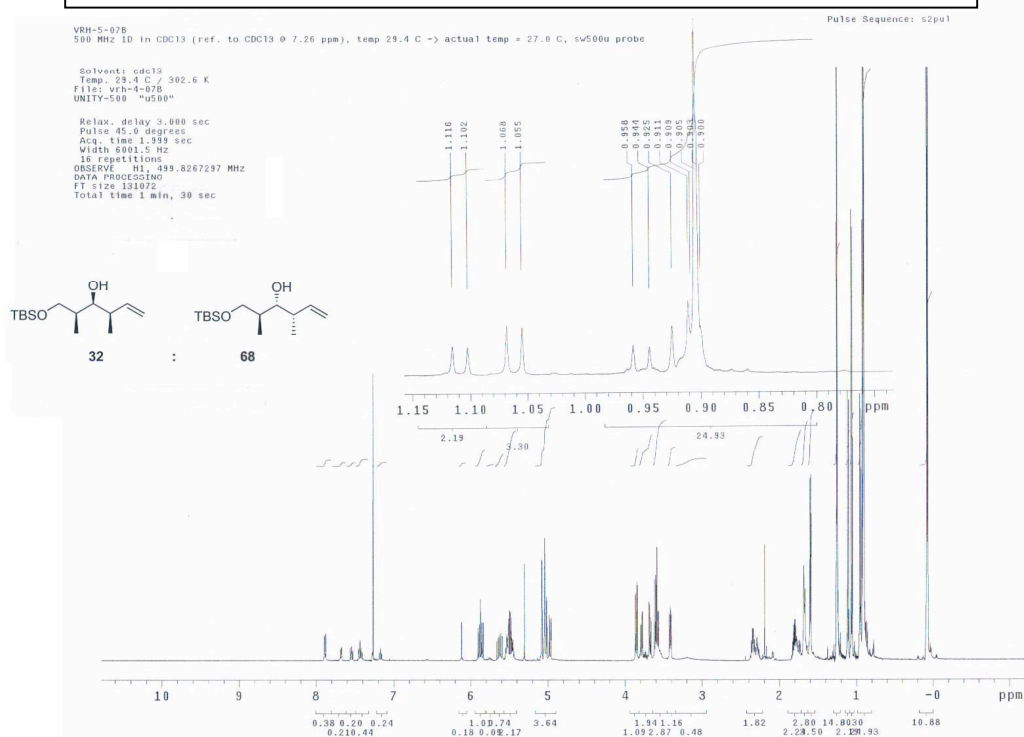
Diastereoselective Allylation with (*S,S*)-diol 1j and SnCl₄



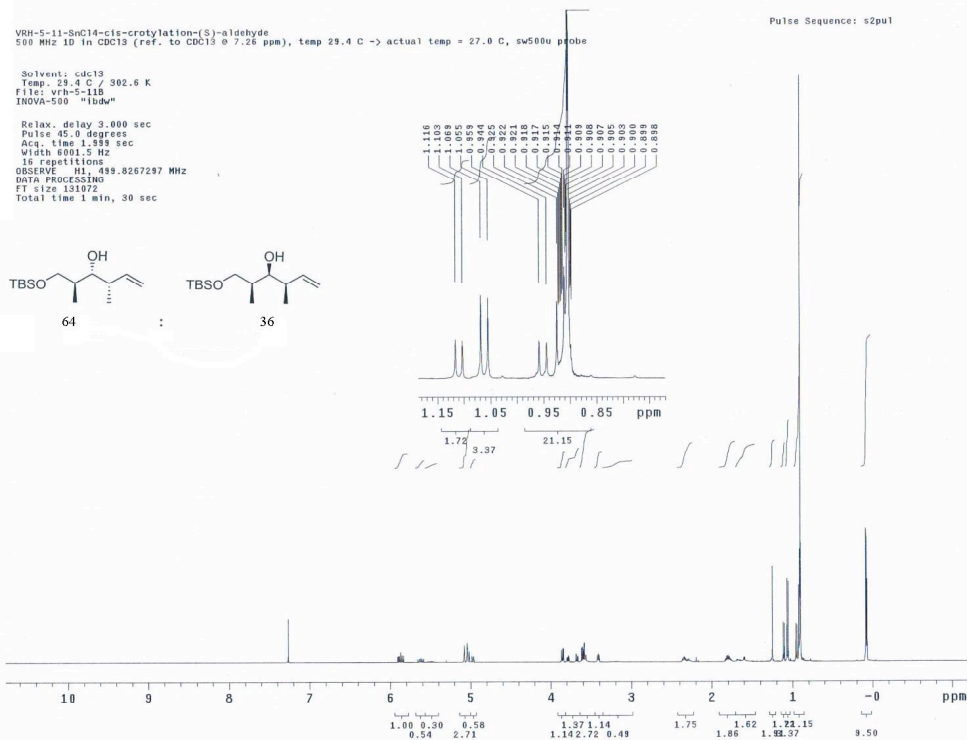
Diastereoselective cis-crotylation with (*R,R*)-diol 1j and SnCl₄



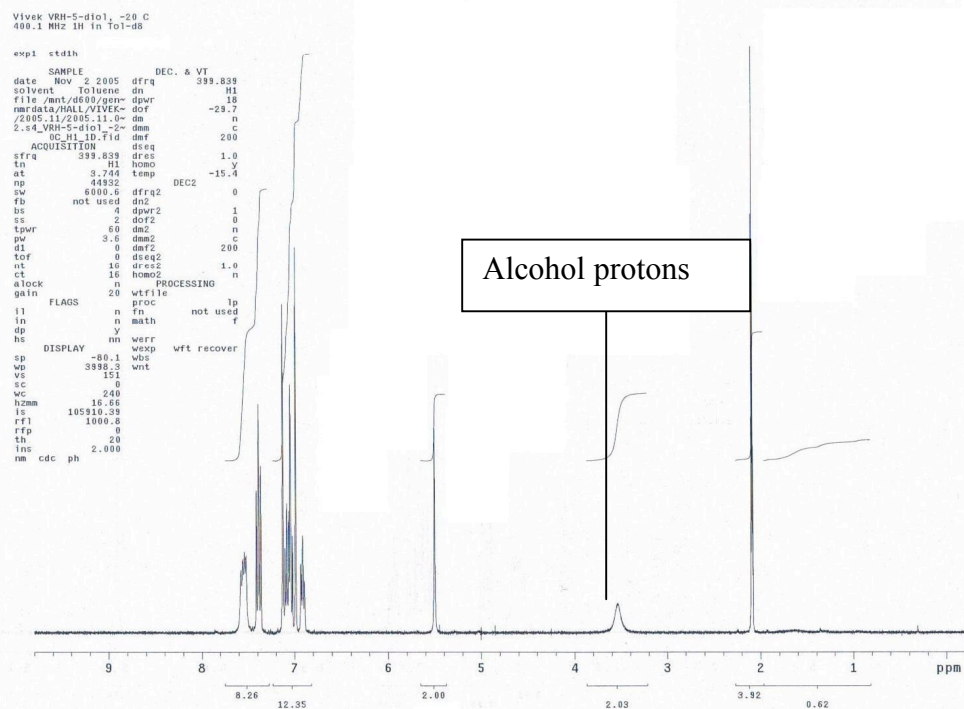
Diastereoselective cis-crotylation with (*S,S*)-diol 1j and SnCl₄



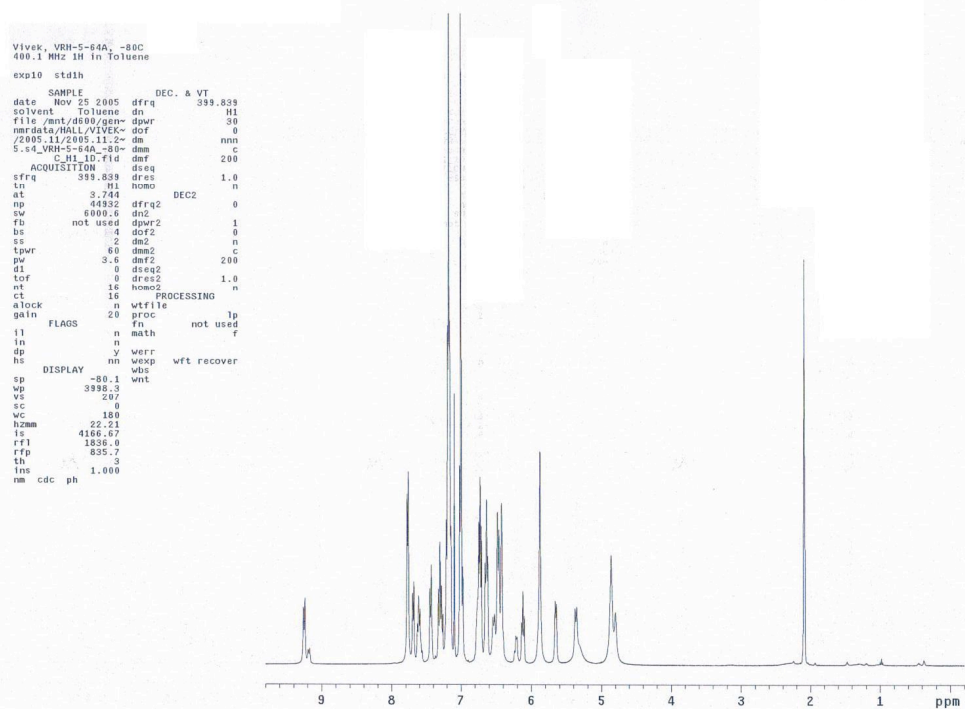
SnCl₄ catalyzed cis-crotylation of aldehyde 3



¹H NMR of diol **1j** in toluene d-8 at -20°C



¹H NMR of diol **1j** and SnCl₄ complex at -80°C in toluene d-8



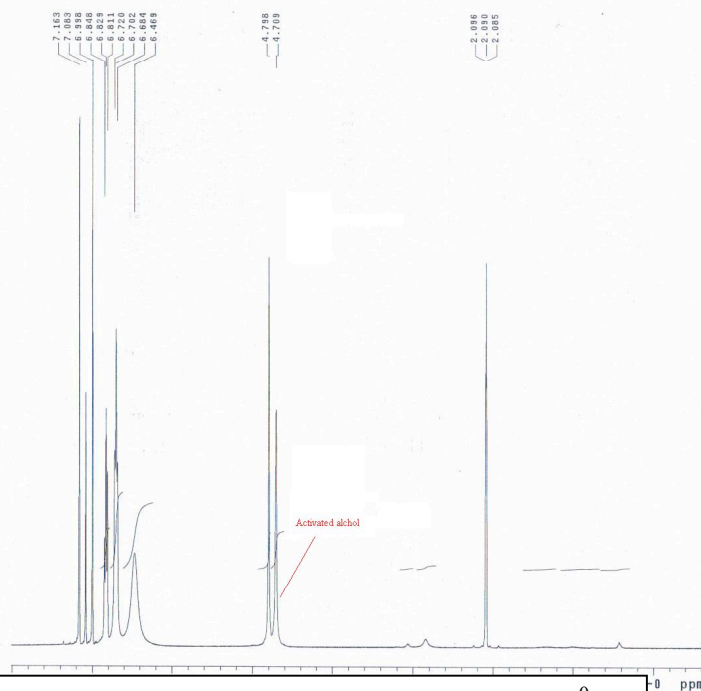
Vivek, VRH-5-64B, -80C
400.1 MHz 1H in Toluene

exp10 std1h

```

SAMPLE          DEC. & VT
date Nov 25 2005 dfrq 399.839
solvent Toluene dn H1
file /mnt/d600/gen- dpwr 30
nmdata/HALL/VIVEK- dof 0
/2005.11/2005.11.2- dm nnn
5-s4_VRH-5-64B_-30- dam c
C-H1_10.fid daf 200
ACQUISITION      dseq 1.0
sfrq 399.839      dres homo n
tn H1            DEC2 0
at 3.744          dfrq2 0
np 44932          dno 1
sw 6000.6         dpr2 0
fb not used       dm2 0
ss 2             dmf2 0
tpwr 60          dseq2 c
pw 3.6           dmf2 200
d1 0             dseq2 1.0
tof 0            dres2 n
nt 16            hnm2 n
ct 16            vtrfile n
alock n          proc lp
gain 20          fn not used
il n            math f
in n            verr f
dp y            wexp wft recover
hs nn          wnt
DISPLAY
sp -277.3        wnt
wp 2479.0
vs 151
sc 0
wc 180
hzm 41.39
ls 4166.67
rfi 1000.8
rfp 0
th 10
ins 2.000
nm cdc ph

```



^{119}Sn -NMR of (*R,R*)-hydrobenzoin and SnCl_4 complex at -80°C in toluene d-8

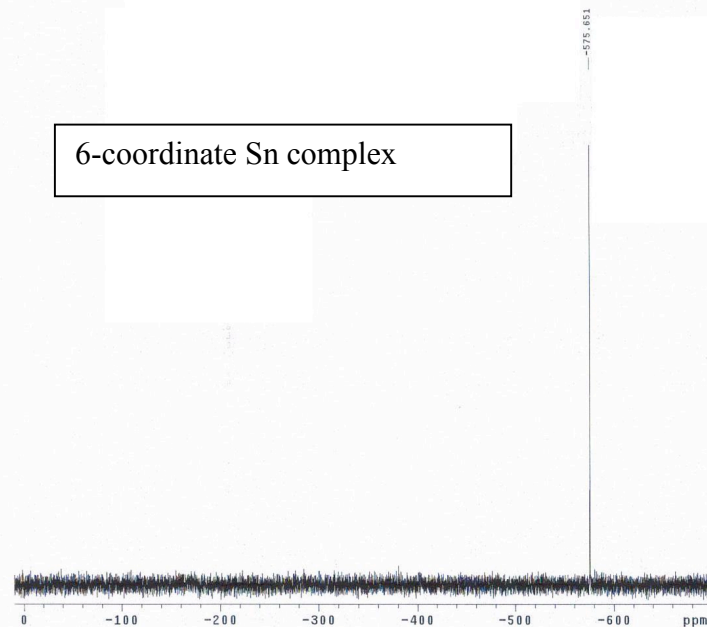
Vivek, VRH-5-64B, -80C
149.05 MHz ^{119}Sn in Toluene

exp14 s2pul

```

SAMPLE          DEC. & VT
date Nov 25 2005 dfrq 399.839
solvent Toluene dn H1
file /mnt/d600/gen- dpwr 30
nmdata/HALL/VIVEK- dof 0
/2005.11/2005.11.2- dm yyy
5-s4_VRH-5-64B_-30- dam v
C-Sn119_10.fid daf 10582
ACQUISITION      dseq 1.0
sfrq 149.050      dres homo n
tn Sn119          DEC2 0
at 1.998          dfrq2 0
np 42292          dno 1
sw 195820.1       dpr2 0
fb not used       dm2 0
ss 4             dmf2 0
tpwr 55          dseq2 c
pw 3.0           dmf2 10582
d1 0             dseq2 1.0
tof -51976.9      dres2 n
nt 10000          hnm2 n
ct 100           vtrfile n
alock n          proc ft
gain 20          fn not used
il n            math f
in n            verr f
dp y            wexp wft recover
hs nn          wnt
DISPLAY
sp -104349.8      wnt
wp 105819.7
vs 113
sc 0
wc 180
hzm 41.39
ls 500.00
rfi 104350.2
rfp 0
th 20
ins 100.000
nm no ph

```



6-coordinate Sn complex