



Supporting Information

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Enantioselective Organocatalytic Hydrophosphination of α,β -Unsaturated Aldehydes

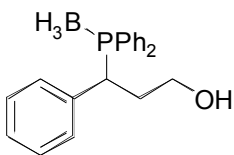
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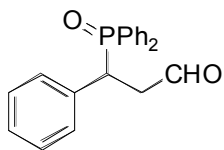
General. Chemicals and solvents were either purchased *puriss p.A.* from commercial suppliers or purified by standard techniques. Catalysts **11** and **12** were synthesized according to literature procedures.¹ For thin-layer chromatography (TLC), silica gel plates Merck 60 F254 were used and compounds were visualized by irradiation with UV light and/or by treatment with a solution of phosphomolybdic acid (25 g), Ce(SO₄)₂·H₂O (10 g), conc. H₂SO₄ (60 mL), and H₂O (940 mL) followed by heating or by treatment with a solution of *p*-anisaldehyde (23 mL), conc. H₂SO₄ (35 mL), acetic acid (10 mL), and ethanol (900 mL) followed by heating. Flash chromatography was performed using silica gel Merck 60 (particle size 0.040-0.063 mm), ¹H NMR, ¹³C NMR and ³¹P NMR spectra were recorded on Varian AS 400. Chemical shifts are given in δ relative to tetramethylsilane (TMS) and to external standard 85% H₃PO₄ (³¹P NMR), the coupling constants *J* are given in Hz. The spectra were recorded in CDCl₃ as solvent at room temperature, TMS served as internal standard (δ = 0 ppm) for ¹H NMR, and CDCl₃ was used as internal standard (δ = 77.0 ppm) for ¹³C NMR. HPLC was carried out using a Waters 2690 Millennium with photodiode array detector. Optical rotations were recorded on a Perkin Elemer 241 Polarimeter (λ = 589 nm, 1 dm cell). High-resolution mass spectra were recorded on a Bruker MicrOTOF spectrometer.

1. a) M. Marigo, T. C. Wabnitz, D. Fielenbach, K. A. Jørgensen, *Angew. Chem. Int. Ed.* **2005**, *44*, 794. b) M. Marigo, D. Fielenbach, A. Braunton, A. Kjaersgaard, K. A. Jørgensen, *Angew. Chem. Int. Ed.* **2005**, *44*, 3703. c) J. Franzén, M. Marigo, D. Fielenbach, T. C. Wabnitz, A. Kjaersgaard, K. A. Jørgensen, *J. Am. Chem. Soc.* **2005**, *127*, 18296. d) Y. Hayashi, H. Gotoh, T. Hayashi, M. Shoji, *Angew. Chem. Int. Ed.* **2005**, *44*, 4212. e) M. Marigo, J. Franzén, T. B. Poulsen, W. Zhuang, K. A. Jørgensen, *J. Am. Chem. Soc.* **2005**, *127*, 6964. g) H. Sundén, I. Ibrahim, A. Córdova, *Tetrahedron Lett.* **2006**, *47*, 99. h) I. Ibrahim, A. Córdova, *Chem. Commun.* **2006**, 1760.

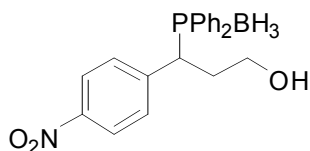
Typical experimental procedure for the organocatalytic AHP reactions. To a stirred solution of catalyst (20 mol%) and 2-fluorobenzoic acid (10 mol%) in CHCl₃ (1.0 mL) at 4 °C, was added 0.25 mmol (1.0 equiv) of α,β -unsaturated aldehyde **1**. The reaction was then flushed with Ar, and 0.3 mmol (1.2 equiv) of diphenylphosphine was added. The reaction was stirred at 4 °C for the time reported in Table 2. Next, the product was oxidized or reduced *in situ*. Thus, 1.5 equiv of I₂ in 2% H₂O in pyridine (1.2 mL) was added. The crude mixture was purified by column chromatography to afford the pure phosphineoxide aldehydes **5**. In order to determine the ee, the crude was reduced “in situ” by NaBH₄ in MeOH, and the crude was purified by column chromatography to afford the corresponding phosphine alcohol derivatives **4**.



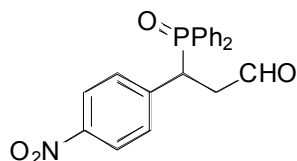
4a: Colorless oil. IR (KBr): 3226, 3080, 3029, 2970, 1593, 1495, 1454, 1437, 1178, 1048, 752, 697 cm⁻¹ ³¹P (162 MHz, CDCl₃): 24.5 (m). ¹H NMR (400 MHz, CDCl₃): δ = 8.00-7.93 (m, 2H), 7.56-7.48 (m, 3H), 7.36-7.29 (m, 3H), 7.22-7.18 (m, 2H), 7.17-7.12 (m, 5H), 4.10-3.99 (m, 1H), 3.62-3.56 (m, 1H), 3.39-3.31 (m, 1H), 2.34-2.26 (m, 1H), 2.16-2.06 (m, 1H), 1.7-0.9 (bs, 3H); ¹³C NMR (100 MHz, CDCl₃): 135.3, 133.4, 133.3, 132.9, 132.8, 131.7, 131.6, 131.0, 130.9, 130.3, 130.2, 129.2, 129.1, 128.3, 128.2, 127.5, 127.4, 125.8, 60.1 (d, J_{C-P} = 12.9 Hz), 39.2 (d, J_{C-P} = 31.8 Hz), 33.0 (d, J_{C-P} = 3.7 Hz), [α]_D = -13.2 (c = 1.0, CHCl₃). HRMS (ESI): calcd. for [M+Na]⁺(C₂₁H₂₄BOP) requires m/z 357.1550, found 357.1565. The enantiomeric excess (83%) was determined by HPLC with an AD column. (*n*-hexane: *i*-PrOH = 95:5, λ = 250 nm), 1.0 mL/min; t_R = major enantiomer 29.77 min, minor enantiomer 25.3 min.



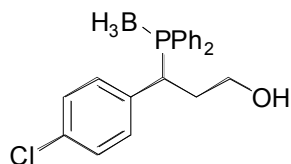
5a: Colorless solid. IR (KBr): 3080, 3029, 2970, 1966, 1723, 1593, 1495, 1454, 1437, 1178, 1048, 752, 697 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 34.5 (s). ^1H NMR (400 MHz, CDCl_3): δ = 9.60 (t, J =1.47Hz, 1H), 7.92-7.87 (m, 2H), 7.55-7.13 (m, 13H), 4.23-4.16 (m, 1H), 3.38-3.28 (m, 1H), 3.01-2.92 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): 199.1 (d, $J_{\text{C-P}}$ =13.7Hz), 135.4, 135.3, 132.4, 132.3, 131.8, 131.7, 131.5, 131.4, 131.3, 131.2, 129.9, 129.8, 129.2, 129.1, 128.6, 128.5, 127.6, 127.5, 125.9, 44.0, 40.2 (d, $J_{\text{C-P}}$ =68.3Hz). Enantiomeric excess: 83%. $[\alpha]_{\text{D}}$ = -44.2 (c = 1.0, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{21}\text{H}_{20}\text{O}_2\text{P})$ requires m/z 335.1195, found 335.1189.



4b: Colorless oil. IR (KBr): 3397, 2928, 2392, 1722, 1597, 1519, 1346, 1261, 694 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 25.2 (m). ^1H NMR (400 MHz, CDCl_3): δ = 8.05-7.95 (m, 4H), 7.60-7.47 (m, 3H), 7.35-7.05 (m, 7H), 4.24 (ddd, J =2.8Hz, 12Hz, 15.2Hz, 1H), 3.66-3.60 (m, 1H), 2.80 (td, J = 3.2Hz, 10Hz, 1H), 2.36-2.26 (m, 1H), 2.23-2.10 (m, 1H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, CDCl_3): 147.0, 141.5, 133.0, 132.9, 132.5, 132.4, 131.8, 131.8, 131.4, 131.3, 130.9, 130.8, 129.2, 129.1, 128.6, 128.5, 128.1, 127.5, 127.2, 126.7, 123.1, 123.1, 59.4 (d, $J_{\text{C-P}}$ =11.4Hz), 39.9 (d, $J_{\text{C-P}}$ =31.1Hz) 32.6 (d, $J_{\text{C-P}}$ =3.0Hz). $[\alpha]_{\text{D}}$ = -25.7 (c = 1.0, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{Na}]^+(\text{C}_{21}\text{H}_{23}\text{NPO}_3)$ requires m/z 402.1401, found 402.1407. The enantiomeric excess (99%) was determined by HPLC with an ODH column. (*n*-hexane: *i*-PrOH = 90:10, λ =250 nm), 1.0 mL/min; t_{R} = major enantiomer 14.9 min, minor enantiomer 20.9 min.

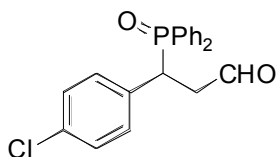


5b: white solid. IR (KBr): 3079, 2970, 2618, 1966, 1723, 1601, 1491, 1485, 1260, 1048, 751 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 34.7 (s). ^1H NMR (400 MHz, CDCl_3): δ = 9.60 (m, 1H), 8.02 (d, $J=8.75\text{Hz}$, 2H), 7.72-7.61 (m, 2H), 7.46-7.43 (m, 3H), 7.32-7.28 (m, 3H), 7.19-7.15 (m, 4H), 4.30-4.21 (m, 1H), 3.17-3.10 (m, 1H), 2.87-2.80 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): 199.3 (d, $J_{\text{C-P}}=12.2\text{Hz}$), 148.7, 148.6, 134.9, 134.8, 134.1, 134.0, 133.9, 133.5, 133.4, 130.3, 130.0, 129.8, 129.6, 129.3, 129.2, 128.6, 128.5, 47.2 (d, $J_{\text{C-P}}=19.7\text{Hz}$), 38.8 (d, $J_{\text{C-P}}=15.0\text{Hz}$). Enantiomeric Excess: 99%. $[\alpha]_{\text{D}} = -176.5$ ($c = 1.0$, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{21}\text{H}_{19}\text{NO}_4\text{P})$ requires m/z 380.1046, found 380.1039.

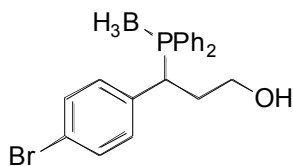


4c: Colorless oil. IR (KBr): 3217 (bs), 2928, 2616, 1965, 1732, 1682, 1591, 1488, 1411, 1216, 1150, 842 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 24.8 (m). ^1H NMR (400 MHz, CDCl_3): δ = 7.98-7.92 (m, 2H), 7.57-7.50 (m, 3H), 7.37-7.30 (m, 3H), 7.24-7.20 (m, 2H), 7.13 (d, $J=8.4\text{Hz}$, 2H), 7.06 (d, $J=8.4\text{Hz}$, 2H), 4.08-3.99 (m, 1H), 3.64-3.58 (m, 1H), 3.36-3.29 (m, 1H), 2.29-2.20 (m, 1H), 2.14-2.06 (m, 1H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, CDCl_3): 134.1, 134.0, 133.4, 133.3, 133.0, 132.8, 132.0, 131.8, 131.6, 131.5, 131.3, 131.2, 129.3, 129.2, 129.0, 128.9, 128.8, 128.6, 128.5, 128.4, 127.9, 59.9 (d, $J_{\text{C-P}}=12.2\text{Hz}$), 38.5 (d, $J_{\text{C-P}}=32.7\text{Hz}$), 32.3 (d, $J_{\text{C-P}}=4.6\text{Hz}$). $[\alpha]_{\text{D}} = -56.9$ ($c = 1.0$, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{Na}]^+(\text{C}_{21}\text{H}_{23}\text{BClOP})$ requires m/z 391.1160, found 391.1174. The enantiomeric excess (92%) was determined by HPLC with an ODH column. (*n*-

hexane: *i*-PrOH = 93:7, λ =250 nm), 1.0 mL/min; t_R = major enantiomer 13.3 min, minor enantiomer 18.0 min.

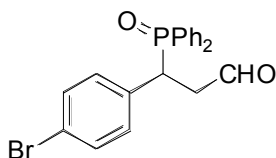


5c: Colorless solid. IR (KBr): 3061, 3029, 2020, 2970, 1965, 1847, 1723, 1601, 1592, 1436, 1259, 1118, 1047 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 34.9 (s). ^1H NMR (400 MHz, CDCl_3): δ = 9.59 (m, 1H), 7.93-7.88 (m, 2H), 7.58-7.35 (m, 8H), 7.24 (d, J =8.4Hz, 2H), 7.14 (d, J =8.4Hz, 2H), 4.21-4.15 (m, 1H), 3.36-3.26 (m, 1H), 3.03-2.93 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): 198.7 (d, $J_{\text{C-P}}$ =13.6Hz), 134.2, 134.1, 133.5, 133.4, 132.6, 132.5, 132.1, 132.0, 131.5, 131.4, 131.3, 131.2, 131.1, 131.0, 129.3, 129.2, 128.9, 128.8, 128.6, 128.5, 44.2, 39.8 (d, $J_{\text{C-P}}$ =68.3Hz). Enantiomeric Excess = 92%. $[\alpha]_D = -56.8$ (c = 1.0, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{21}\text{H}_{19}\text{ClO}_2\text{P})$ requires m/z 369.0806, found 369.0812.

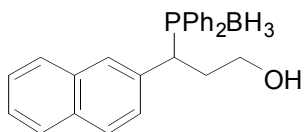


4d: Colorless oil. IR (KBr): 3302, 3080, 2958, 2932, 2382, 1725, 1663, 1628, 1488, 1437, 1063, 738, 693 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 24.8 (m). ^1H NMR (400 MHz, CDCl_3): δ = 8.00-7.92 (m, 2H), 7.56-7.52 (m, 3H), 7.37-7.33 (m, 3H), 7.28 (d, J =8.4Hz, 2H), 7.24-7.22 (m, 2H), 7.01 (d, J =8.4Hz, 2H), 4.07-3.98 (m, 1H), 3.64-3.57 (m, 1H), 3.36-3.28 (m, 1H), 2.30-2.19 (m, 1H), 2.15-2.06 (m, 1H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, CDCl_3): 134.5, 134.4, 133.4, 133.3, 132.9, 132.8, 132.0, 131.9, 131.8, 131.7, 131.5, 131.4, 131.3, 131.2, 129.3, 129.2, 128.6, 128.5, 121.6, 121.5, 59.9 (d, $J_{\text{C-P}}$ =12.1Hz), 38.6

(d, J_{C-P} =32.7Hz), 32.9 (d, J_{C-P} =4.6Hz). $[\alpha]_D = -44.8$ (c = 1.0, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{Na}]^+(\text{C}_{21}\text{H}_{23}\text{BBr}^{79}\text{OP})$ requires m/z 435.0655, found 435.0676. The enantiomeric excess (90%) was determined by HPLC with an AD column. (*n*-hexane: *i*-PrOH = 92:8, $\lambda = 250$ nm), 1.0 mL/min; t_R = major enantiomer 16.7 min, minor enantiomer 18.3 min.

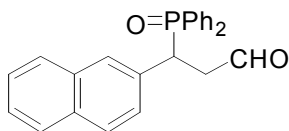


5d: Colorless solid. IR (KBr): 3063, 3028, 2967, 1966, 1843, 1726, 1602, 1587, 1422, 1259, 1115, 1047, 738 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 34.5 (s). ^1H NMR (400 MHz, CDCl_3): δ = 9.58 (m, 1H), 7.92-7.86 (m, 2H), 7.60-7.36 (m, 8H), 7.29 (d, J =8.4Hz, 2H), 7.17 (d, J =8.4Hz, 2H), 4.20-4.14 (m, 1H), 3.36-3.26 (m, 1H), 3.03-2.93 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): 198.6 (d, J_{C-P} =13.6 Hz), 135.0, 134.7, 133.0, 132.8, 132.5, 132.4, 132.1, 131.6, 131.5, 131.4, 131.3, 131.2, 131.1, 131.0, 130.9, 129.3, 129.2, 128.6, 128.5, 44.1, 39.5 (d, J_{C-P} =67Hz). Enantiomeric Excess = 90%. $[\alpha]_D = -67.0$ (c = 1.0, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{21}\text{H}_{19}\text{Br}^{81}\text{O}_2\text{P})$ requires m/z 415.0283, found 415.0264.

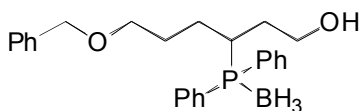


4e: Colorless oil. IR (KBr): 3365, 3060, 2935, 2880, 2390, 1600, 1486, 1436, 1217, 1062, 738, 600 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 24.6 (m). ^1H NMR (400 MHz, CDCl_3): δ = 8.14-8.12 (m, 1H), 8.04-7.98 (m, 2H), 7.84-7.80 (m, 2H), 7.78-7.74 (m, 1H), 7.64-7.60 (m, 2H), 7.54-7.24 (m, 7H), 7.18-7.14 (m, 2H), 4.82 (bs, 1H), 4.25-4.16 (m, 1H), 3.65-3.60 (m, 1H), 3.40-3.33 (m, 1H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, CDCl_3): 133.6, 133.2, 133.1, 133.0, 132.7, 132.6, 131.5, 131.4, 130.8, 130.2, 129.3, 129.2, 129.0, 128.9, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 126.2, 126.0, 59.9 (d, J_{C-P} =12.2Hz), 39.1 (d, J_{C-P} =31.9Hz), 32.8 (d, J_{C-P} =3.8Hz). $[\alpha]_D = -15.2$ (c = 0.5, CHCl_3).

HRMS (ESI): calcd. for $[M+Na]^+(C_{25}H_{26}BOP)$ requires m/z 407.1707, found 407.1712. The enantiomeric excess (98%) was determined by HPLC with an AD column. (*n*-hexane: *i*-PrOH = 95:5, λ =250 nm), 1.0 mL/min; t_R = major enantiomer 41.6 min, minor enantiomer 70.2 min.

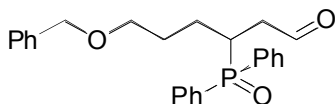


5e: Yellow solid. IR (KBr): 2919, 2851, 1437, 1788, 1721, 1626, 1599, 1437, 1354, 913, 744 cm^{-1} . ^{31}P (162 MHz, $CDCl_3$): 34.1 (s). 1H NMR (400 MHz, $CDCl_3$): δ = 9.61 (m, 1H), 8.00-7.14 (m, 17H), 4.40-4.34 (m, 1H), 3.50-3.40 (m, 1H), 3.12-2.02 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$): 198.8 (d, J_{C-P} =13.5), 132.5, 132.2, 132.1, 131.6, 131.6, 131.4, 131.3, 131.2, 131.1, 131.0, 129.1, 129.0, 128.9, 128.8, 128.7, 128.2, 128.1, 128.1, 127.9, 127.6, 127.6, 127.2, 127.5, 126.1, 126.0, 44.1, 40.1 (d, J_{C-P} =68.0Hz). Enantiomeric Excess: 98%. $[\alpha]_D = -45.7$ (c = 1.0, $CHCl_3$). HRMS (ESI): calcd. for $[M+H]^+(C_{25}H_{21}O_2P)$ requires m/z 385.1352, found 385.1357.

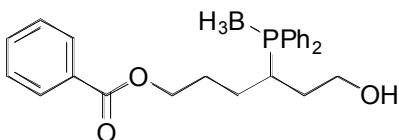


4f: Colorless oil: IR (KBr): 3056, 2920, 2857, 1718, 1663, 1589, 1437, 1357, 1278, 1161, 1118, 913, 747 cm^{-1} . ^{31}P (162 MHz, $CDCl_3$): 24.0 (m). $[\alpha]_D = -11.0$ (c =1.0). 1H NMR (400 MHz, $CDCl_3$): δ = 7.79-7.73 (m, 3H), 7.48-7.28 (m, 12H), 4.44 (q, J =7.6Hz, 11.6Hz, 2H), 3.59-3.47 (m, 2H), 3.44-3.33 (m, 2H), 2.86-2.76 (m, 1H), 1.96-1.56 (m, 6H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, $CDCl_3$): 138.3, 132.8, 132.8, 131.3, 129.4, 128.9, 128.8, 128.6, 127.9, 127.8, 73.1, 70.3, 60.8 (d, J_{C-P} =10.0Hz), 32.8 (d, J_{C-P} =3.0Hz), 29.2 (d, J_{C-P} =34.9Hz), 28.4 (d, J_{C-P} =9.2Hz), 27.0 (d, J_{C-P} =3.0Hz). HRMS (ESI): calcd. for $[M+Na]^+(C_{25}H_{32}BNO_2P)$ requires m/z 429.2125, found 429.2134. The enantiomeric

excess (91%) was determined by HPLC with an ODH column. (*n*-hexane: *i*-PrOH = 97:3, λ = 210 nm), 1.0 mL/min t_R = major enantiomer 60.3 min, minor enantiomer 66.3 min.

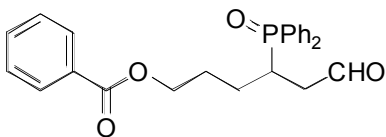


5f: Colorless oil: IR (KBr): 3391, 2925, 2860, 2380, 1719, 1437, 1279, 1105, 1064, 739 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 36.7 (s). Enantiomeric Excess = 91%. $[\alpha]_D = -33.1$ ($c=1.0$). ^1H NMR (400 MHz, CDCl_3): δ = 9.67 (s, 1H), 7.84-7.75 (m, 3H), 7.52-7.43 (m, 6H), 7.33-7.23 (m, 6H), 4.41 (d, $J=11.2\text{Hz}$, 1H), 4.39 (d, $J=11.2\text{Hz}$, 1H), 3.42-3.33 (m, 2H), 3.21-3.12 (m, 1H), 2.83 (dt, $J=3.6\text{Hz}$, 16.4Hz, 1H), 2.68 (dt, $J=4.0\text{Hz}$, 17.2Hz, 1H), 1.87-1.46 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3): 199.5 (d, $J_{C-P}=9.9\text{Hz}$), 138.7, 132.1, 132.0, 131.3, 131.2, 131.1, 129.1, 129.0, 128.9, 128.8, 128.7, 128.5, 127.7, 73.0, 69.8, 42.4, 30.8 (d, $J_{C-P}=72.0\text{Hz}$), 27.9 (d, $J_{C-P}=11.3\text{Hz}$), 25.6. HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{25}\text{H}_{27}\text{NO}_3\text{P})$ requires m/z 407.1771, found 407.1790.



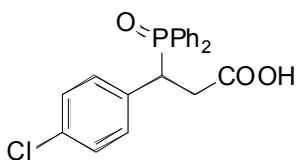
4g: Colorless oil. IR (KBr): 3307 (bs), 2924, 2381, 1752, 1714, 1698, 1606, 1457, 1276, 1194, 913, 744 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 24.7 (m). ^1H NMR (400 MHz, CDCl_3): δ = 7.92-7.84 (m, 2H), 7.82-7.72 (m, 4H), 7.60-7.56 (m, 1H), 7.50-7.30 (m, 8H), 4.22-4.18 (m, 2H), 3.64-3.56 (m, 2H), 2.86-2.78 (m, 1H), 2.00-1.50 (m, 6H), 1.7-0.9 (bs, 3H); ^{13}C NMR (100 MHz, CDCl_3): 166.5, 132.9, 132.7 (d, $J_{C-P}=4.3\text{Hz}$), 132.5 (d, $J_{C-P}=3.9\text{Hz}$), 131.3, 131.2 ($J_{C-P}=2.3\text{Hz}$), 129.5, 128.9 (d, $J_{C-P}=17.6\text{Hz}$), 128.7, 128.5 (d, $J_{C-P}=23.3\text{Hz}$), 64.2, 60.4, 32.5 (d, $J_{C-P}=3.3\text{Hz}$), 28.9 (d, $J_{C-P}=35.1\text{Hz}$), 27.6 (d, $J_{C-P}=8.7\text{Hz}$), 26.2 (d, $J_{C-P}=3.1\text{Hz}$). $[\alpha]_D = -11.4$ ($c = 1.0$, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{Na}]^+(\text{C}_{25}\text{H}_{30}\text{BO}_3\text{P})$ requires m/z 443.1918, found 443.1922. The enantiomeric excess

(95%) was determined by HPLC with an AD column. (*n*-hexane: *i*-PrOH = 92:8, λ =250 nm), 1.0 mL/min; t_R = major enantiomer 16.7 min, minor enantiomer 18.3 min.



5g: Colorless oil. IR (KBr): 3057, 2920, 1717, 1438, 1275, 1117, 713 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 37.7 (s). ^1H NMR (400 MHz, CDCl_3): δ = 9.71 (m, 1H), 7.94-7.80 (m, 5H), 7.60-7.30 (m, 10H), 4.23-4.15 (m, 2H), 3.22-3.18 (m, 1H), 2.94-2.60 (m, 2H), 1.90-1.80 (m, 2H), 1.70-1.60 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3): 199.1 (d, $J_{\text{C-P}}$ =10.6 Hz), 166.4, 132.8, 132.0, 131.9, 131.0, 130.9, 129.5, 128.9, 128.9, 128.8, 128.7, 128.3, 64.0, 42.3, 30.4 (d, $J_{\text{C-P}}$ =72.1Hz), 26.8 (d, $J_{\text{C-P}}$ =10.7Hz), 25.1(d, $J_{\text{C-P}}$ =1.5Hz). Enantiomeric Excess = 95%. $[\alpha]_D = -34.2$ ($c = 1.0$, CHCl_3). HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+$ ($\text{C}_{25}\text{H}_{25}\text{O}_4\text{P}$) requires m/z 421.1563, found 421.1575.

Typical experimental procedure for the direct enantioselective catalytic hydrophosphination of α,β unsaturated aldehydes and *in situ* oxidation to acid. To a stirred solution of catalyst (20 mol%) and 2-fluorobenzoic acid (10 mol%) in CHCl_3 (1.0 mL) at 4 $^\circ\text{C}$, was added 0.25 mmol (1.0 equiv) of α,β -unsaturated aldehyde. The reaction was then flushed with Ar, and 0.3 mmol (1.2 equiv) of diphenylphosphine was added. The reaction was stirred at 4 $^\circ\text{C}$ for the time reported in Table 2. Next, isobutene (0.1 mL), *tert*-butanol (0.4 mL), H_2O (0.2 mL) KH_2PO_4 (54.4 mg, 4 mmol), and NaClO_2 (36 mg, 4mmol) were added sequentially. The reaction was allowed to reach room temperature and was stirred overnight. The crude was purified by column chromatography (pentane/Ethyl Acetate mixtures) to afford the desired acid.



13c: Colorless solid. IR (KBr): 3057, 3025, 2969, 2916, 2533, 2227, 1717, 1486, 1438, 1164, 840, 726 cm^{-1} . ^{31}P (162 MHz, CDCl_3): 35.9 (s). $[\alpha]_{\text{D}} = -35.2$ ($c=1.0$). ^1H NMR (400 MHz, CDCl_3): δ = 8.65 (s, 1H), 7.90-7.85 (m, 2H), 7.60-7.40 (m, 2H), 7.17 (d, $J=8.5\text{Hz}$, 2H), 7.06 (d, $J=8.5\text{Hz}$, 2H), 4.16-4.10 (m, 1H), 3.12-3.02 (m, 1H), 2.90-2.81 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): 172.9, 172.7, 134.0, 133.9, 133.3, 133.2, 133.0, 132.9, 132.5, 132.0, 131.7, 131.5, 131.4, 131.3, 131.2, 131.1, 131.0, 130.3, 130.1, 129.3, 129.2, 128.6, 128.5, 42.6, 42.0, 35.1. HRMS (ESI): calcd. for $[\text{M}+\text{H}]^+(\text{C}_{21}\text{H}_{18}\text{ClO}_3\text{P})$ requires m/z 407.0580, found 407.0583.

Cartesian coordinates of optimized transition states:

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C	-3.046222	2.220684	0.079563
C	-3.551260	2.553004	1.347753
C	-4.410095	3.637941	1.511595
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C	-4.306538	4.072568	-0.863048
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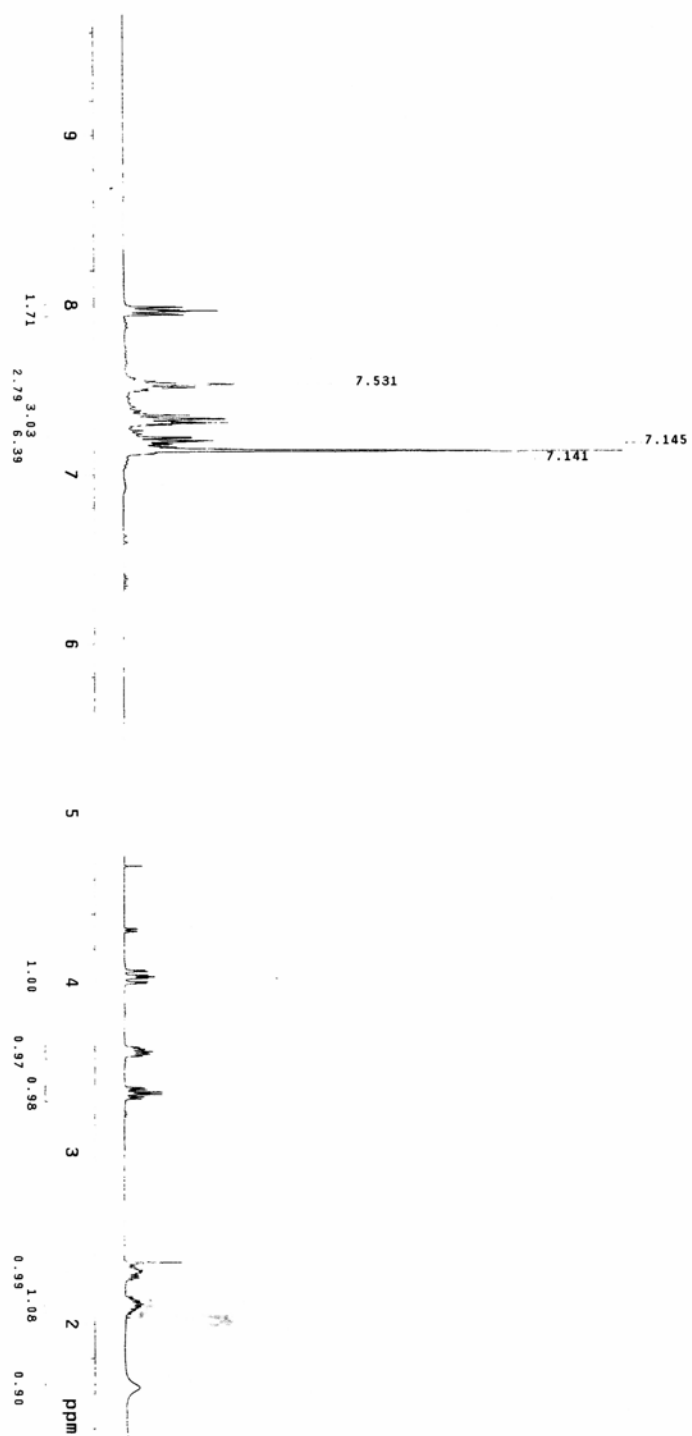
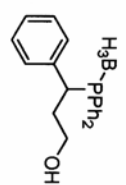
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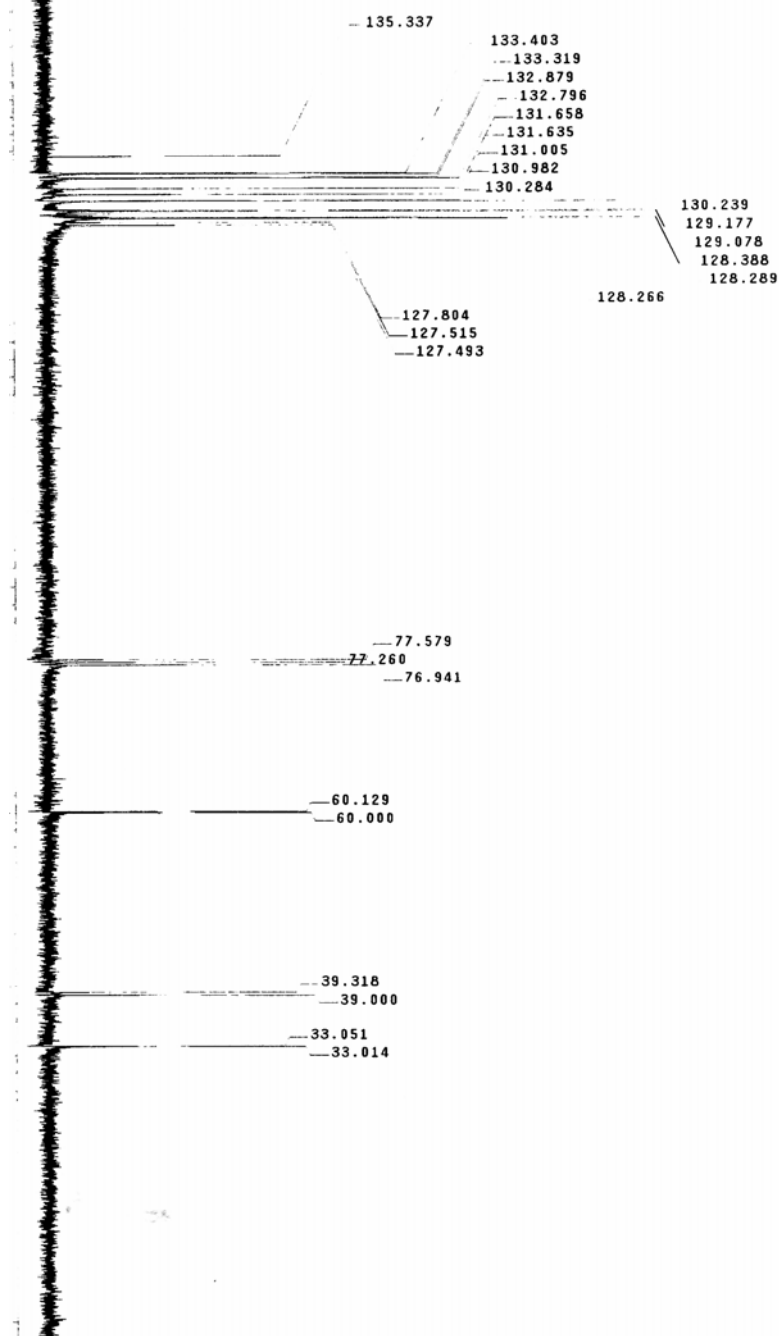
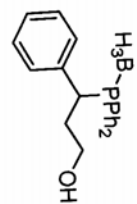
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C	3.954655	-2.010099	-1.883682
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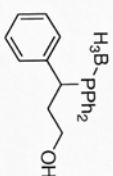
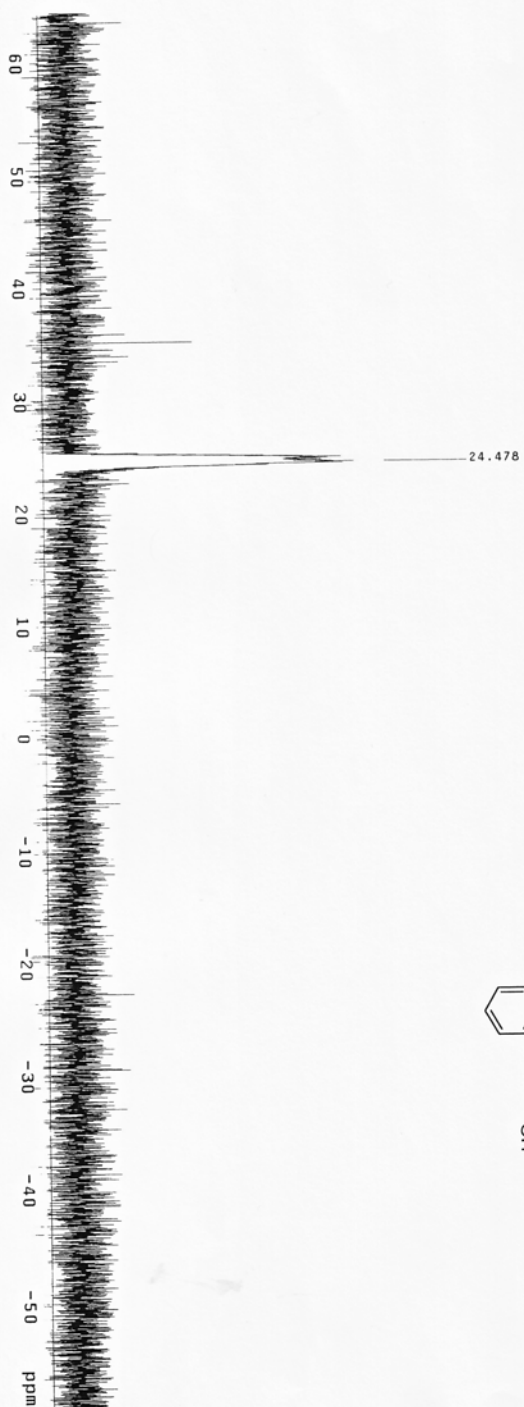
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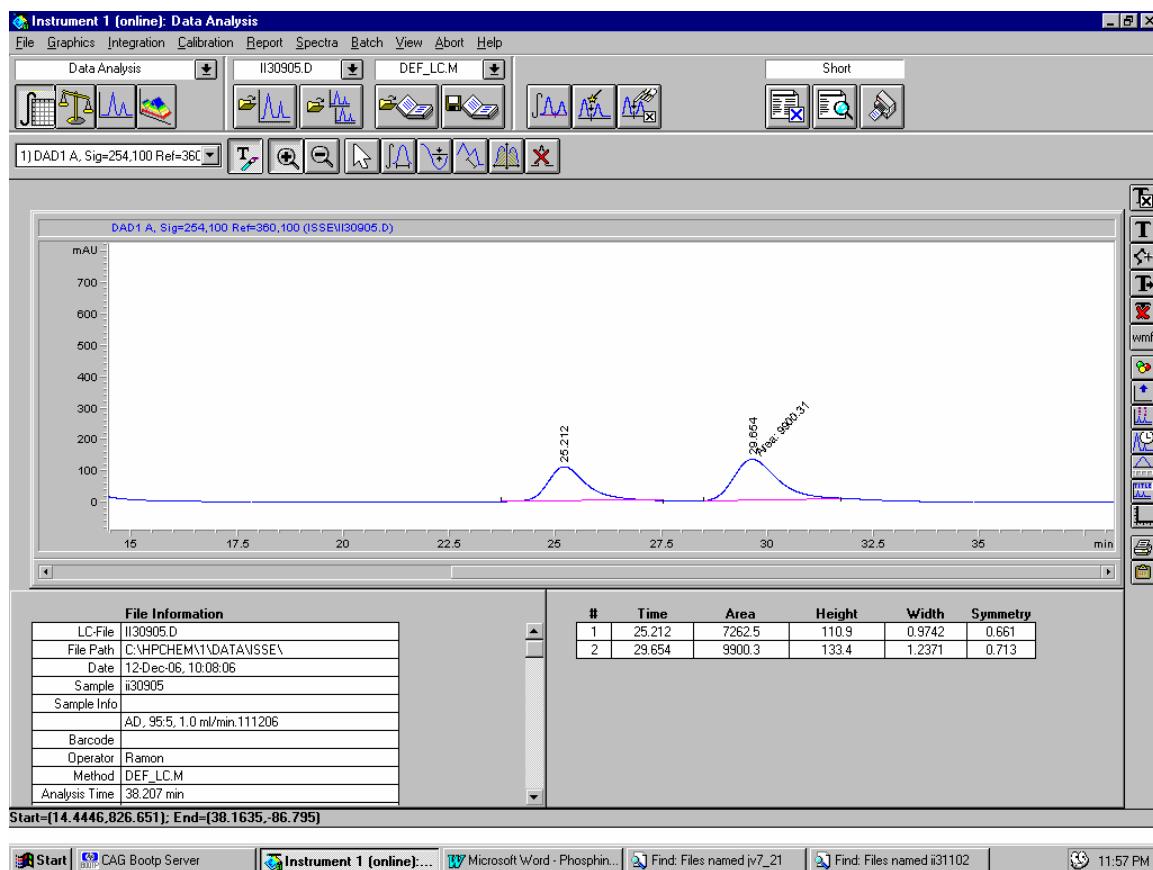
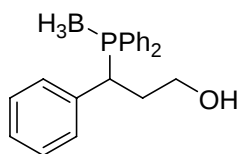


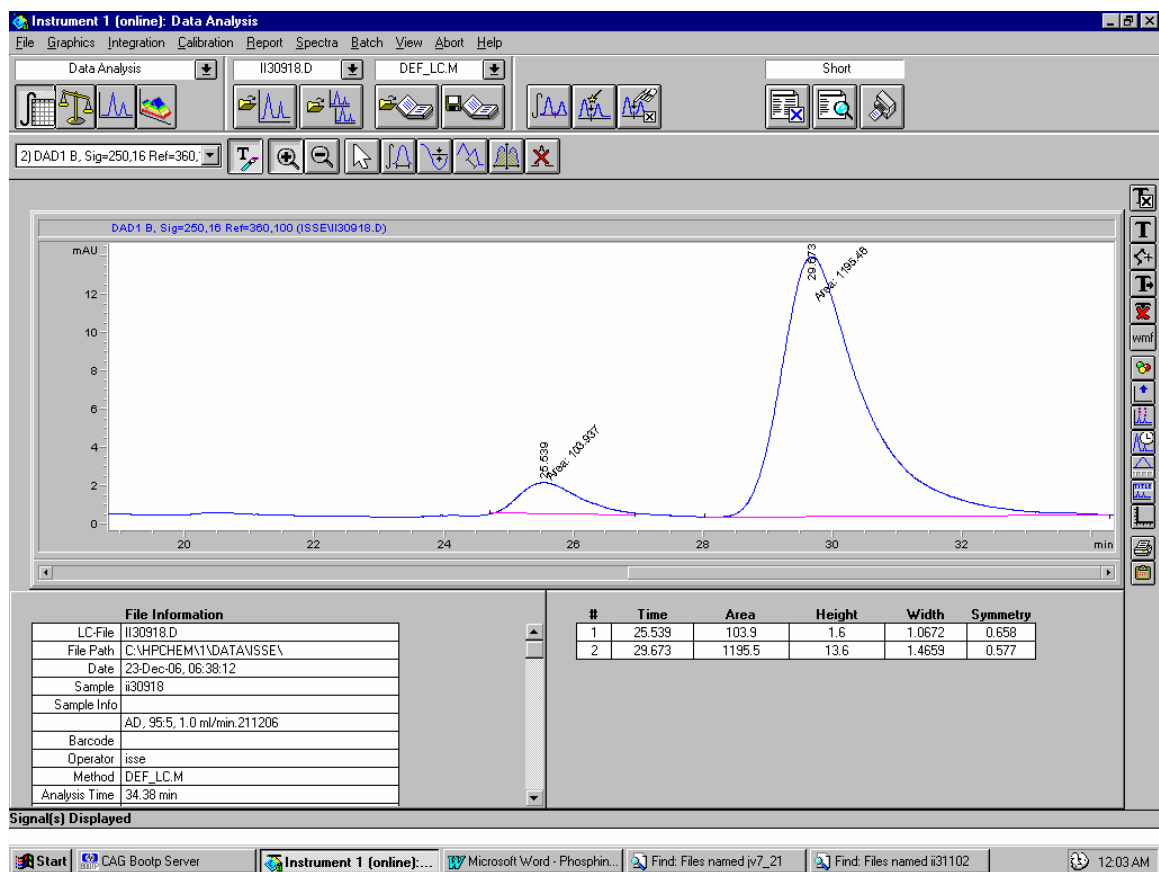
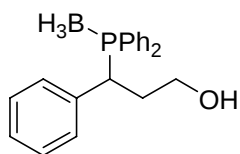


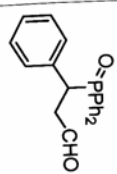
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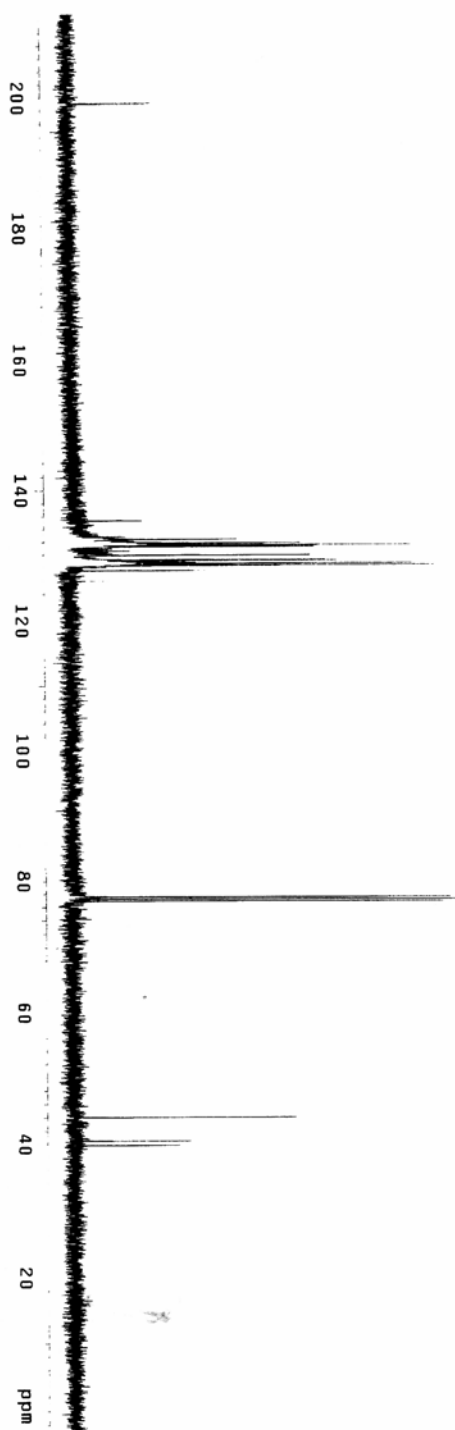
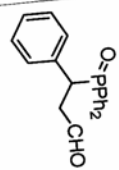
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F1 size 32768
Total time 0 min, 51 sec



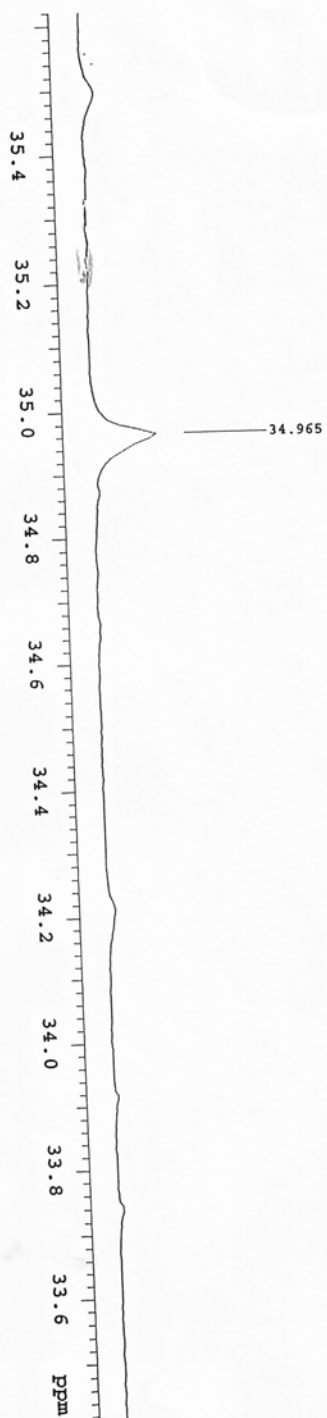
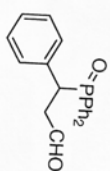




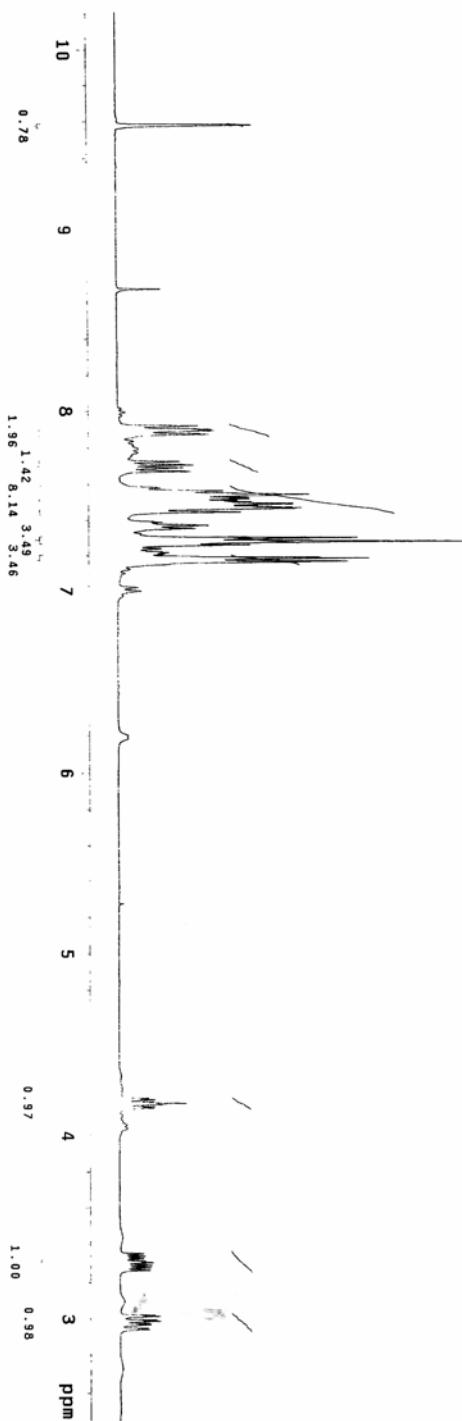
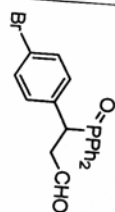




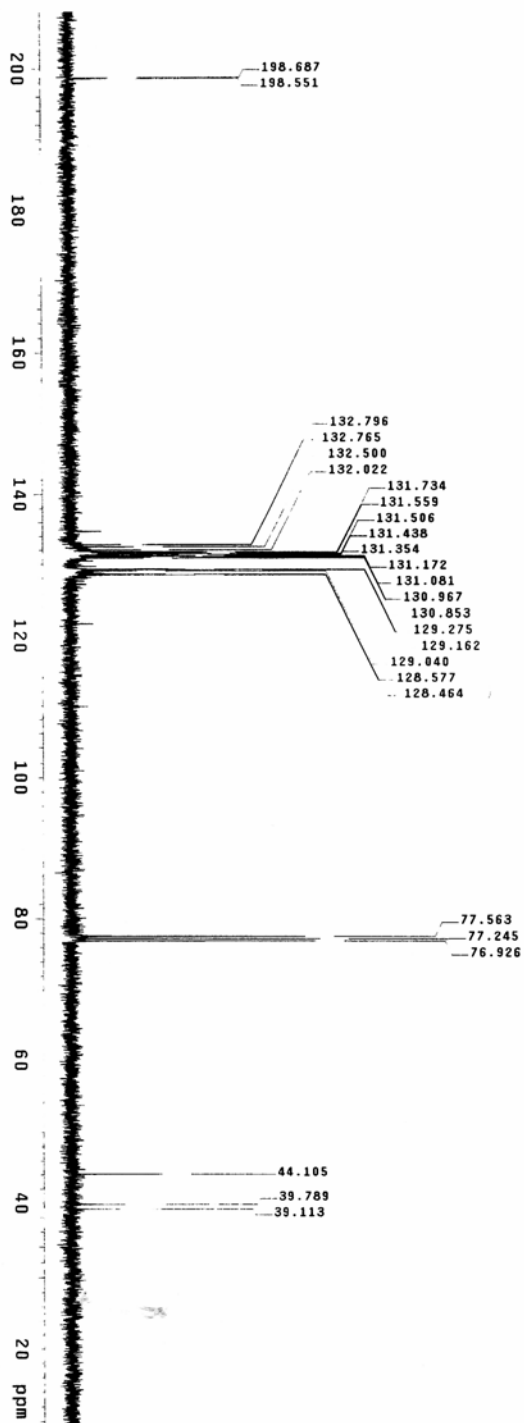
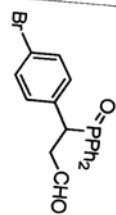
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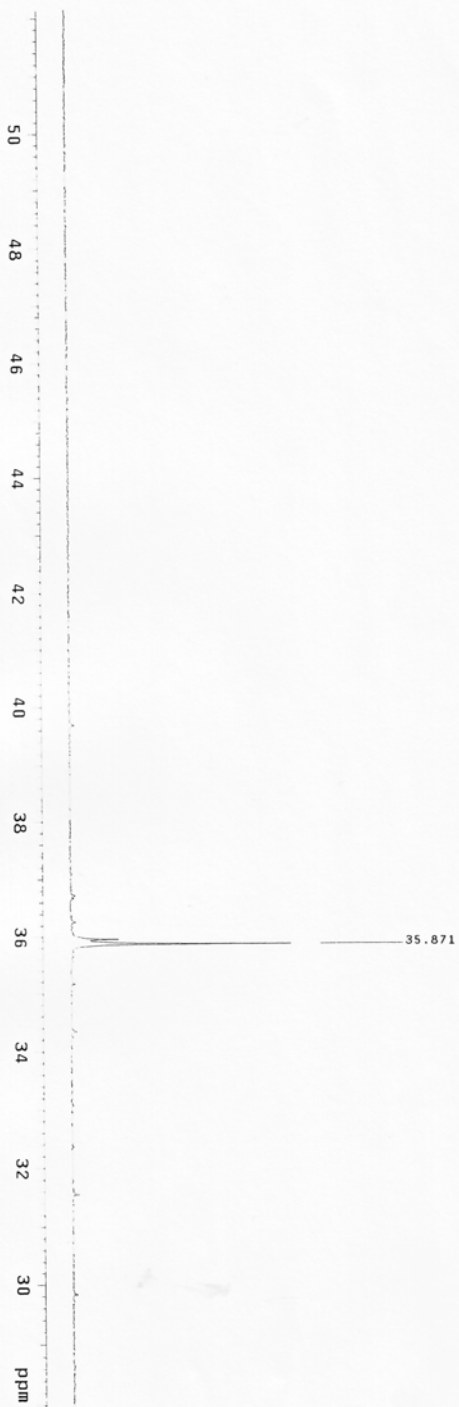
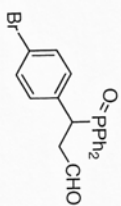
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 Acquisition 2 Hz
 16 repetitions
 OBSERVE H1 399.936500 MHz
 DATA PROCESSING
 Line broadening 0.1 Hz
 FT size 65536
 Total time 1 min, 2 sec

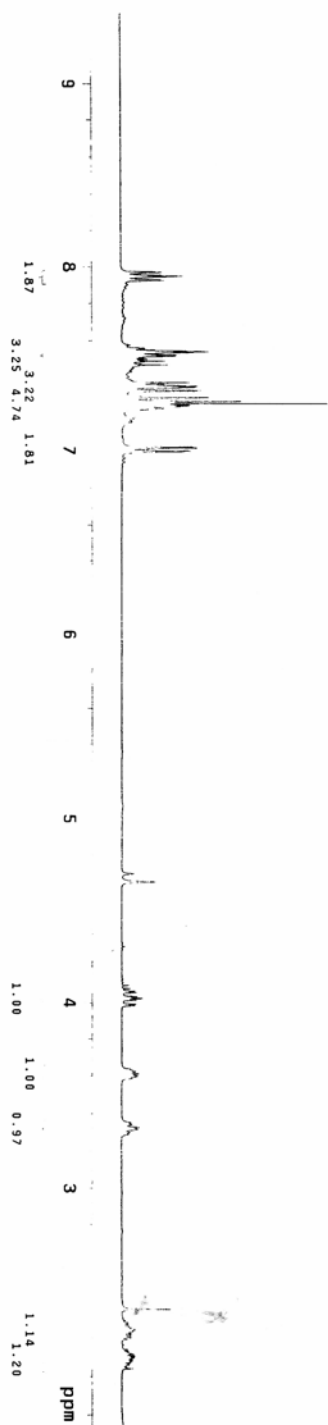
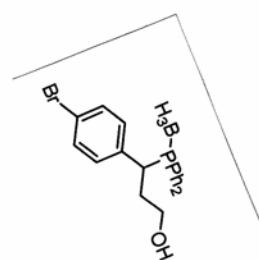


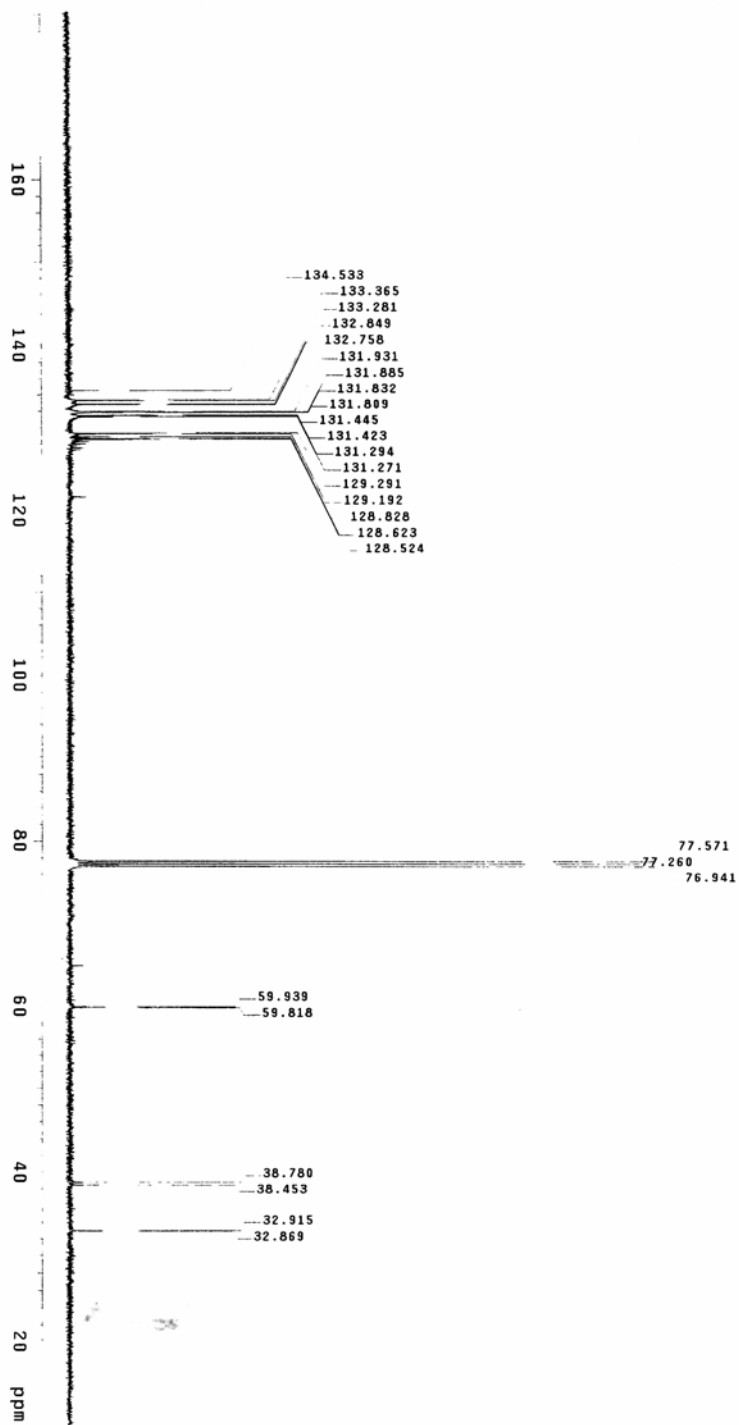
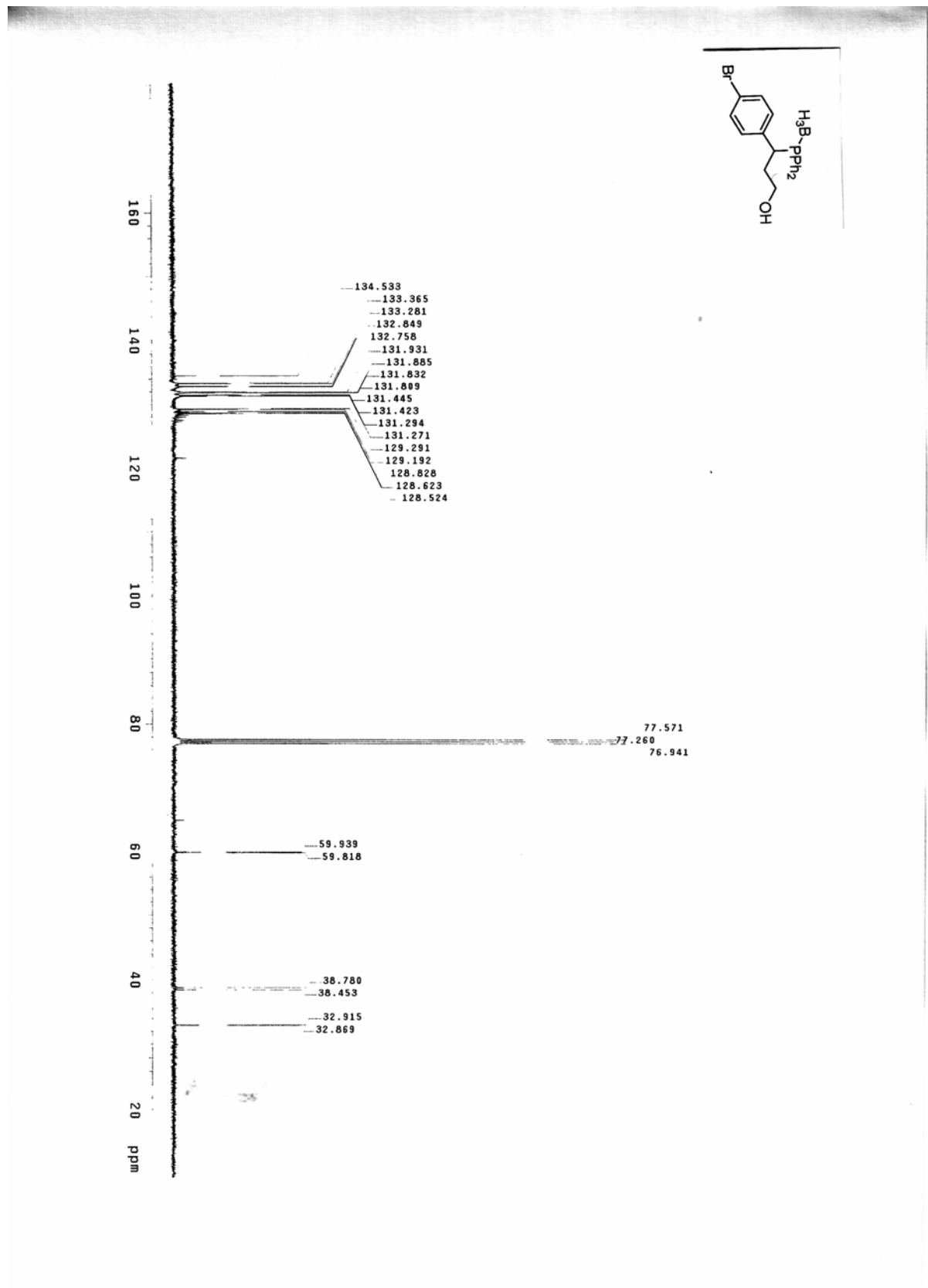
1131209_flash_13C
Pulse Sequence: szpul
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
Mercury-40088 "nmr4"
Relax: delay 1.000 sec
Pulse 48.9 degrees
Acq: time 1.199 sec
Width 25000.0 Hz
F2 100.626130 MHz
Observed C13 100.626130 MHz
Decouple H1 399.9375154 MHz
Power 42 dB
continuously on
VAlTz-16 modulated
DATA PROCESSING
FID size 65536
F2 size 65536
Total time 137 hr, 1 min, 31 sec



1131106_flash_31P
Pulse Sequence: szpul
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
File: 1131106_flash_31P
Mercury-400BB 7mm 4
Pulse 27.6 degrees
P1 1.500 sec
Width 20000.0 Hz
16 repetitions
OBSERVE P31, 161.8967379 MHz
DECOUPLE H1, 399.3376154 MHz
Power 42.0 dB
NUC1 31P
NUC2 1H
WALTZ16 modulated
DATA PROCESSING
Line broadening 1.0 Hz
FT size 32768
Total time 0 min, 15 sec

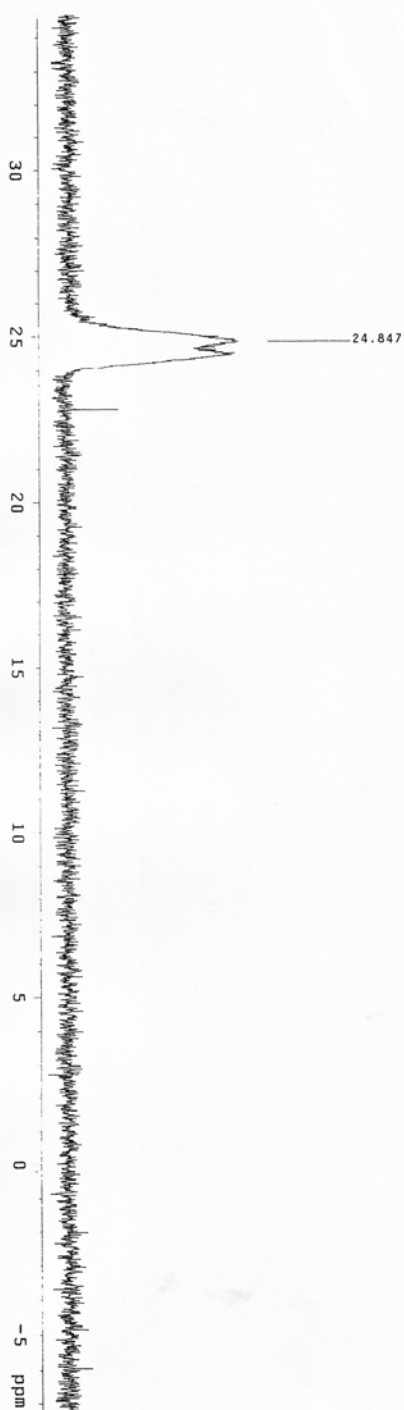
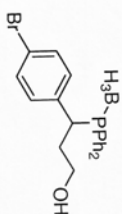


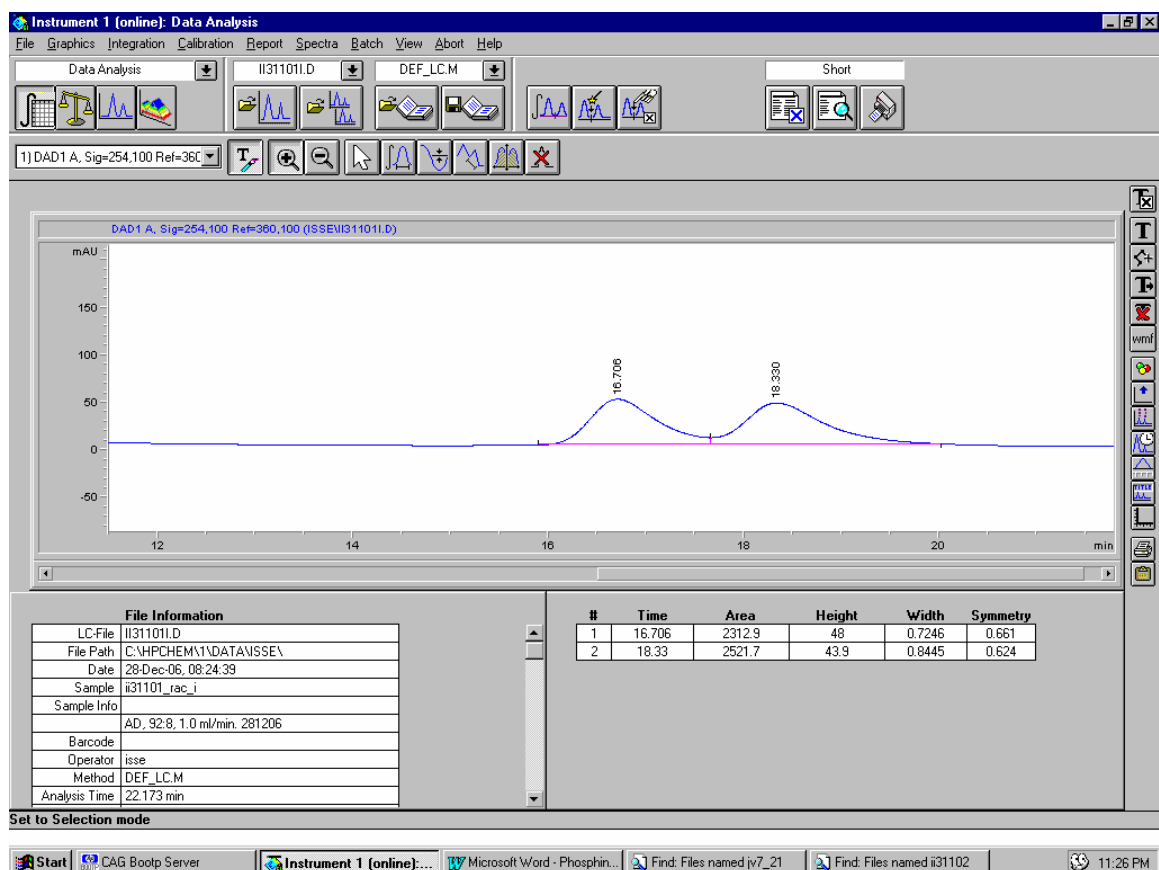
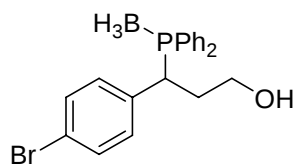


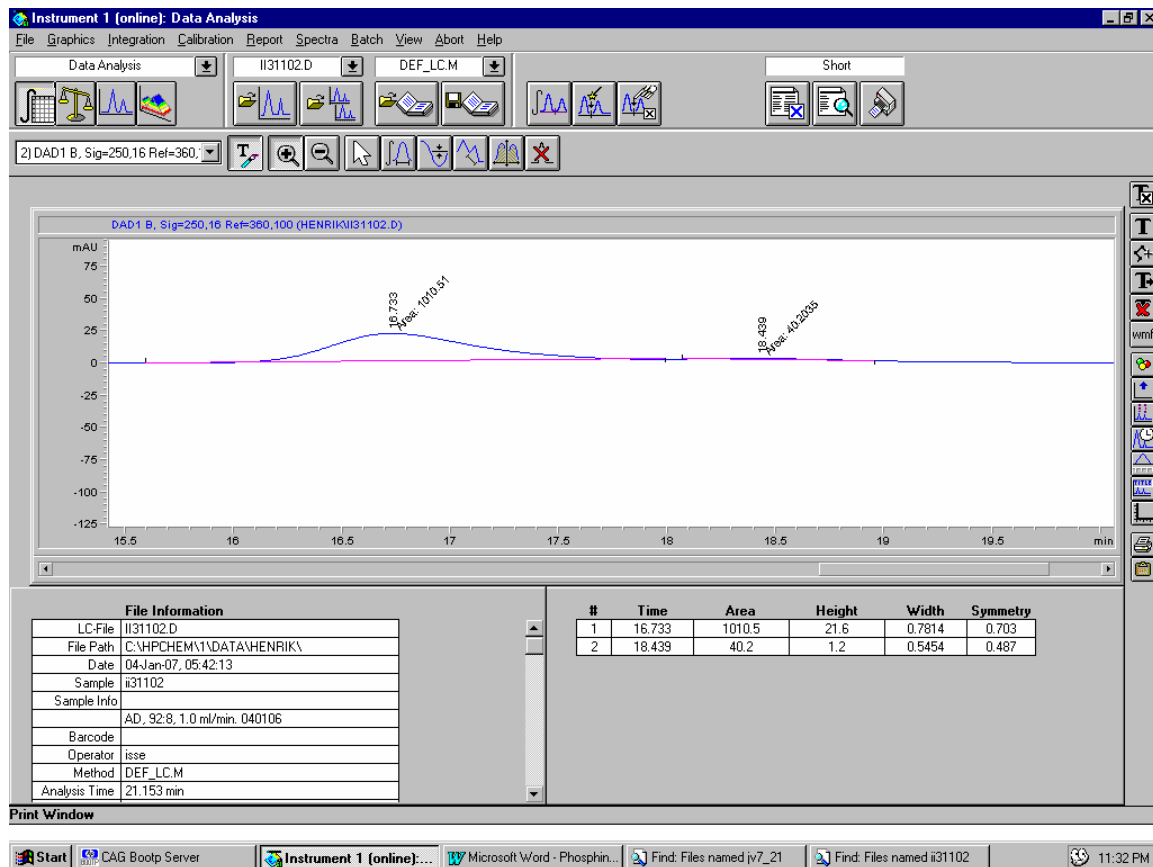
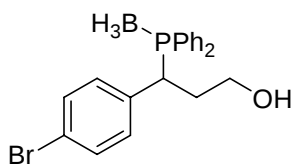


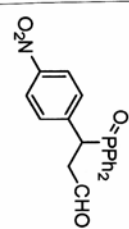
Pulse Sequence: zgpg30
Solvent: CDCl3
Temp: 24.0 C / 297.1 K
Mercury-40088 "nmr4"

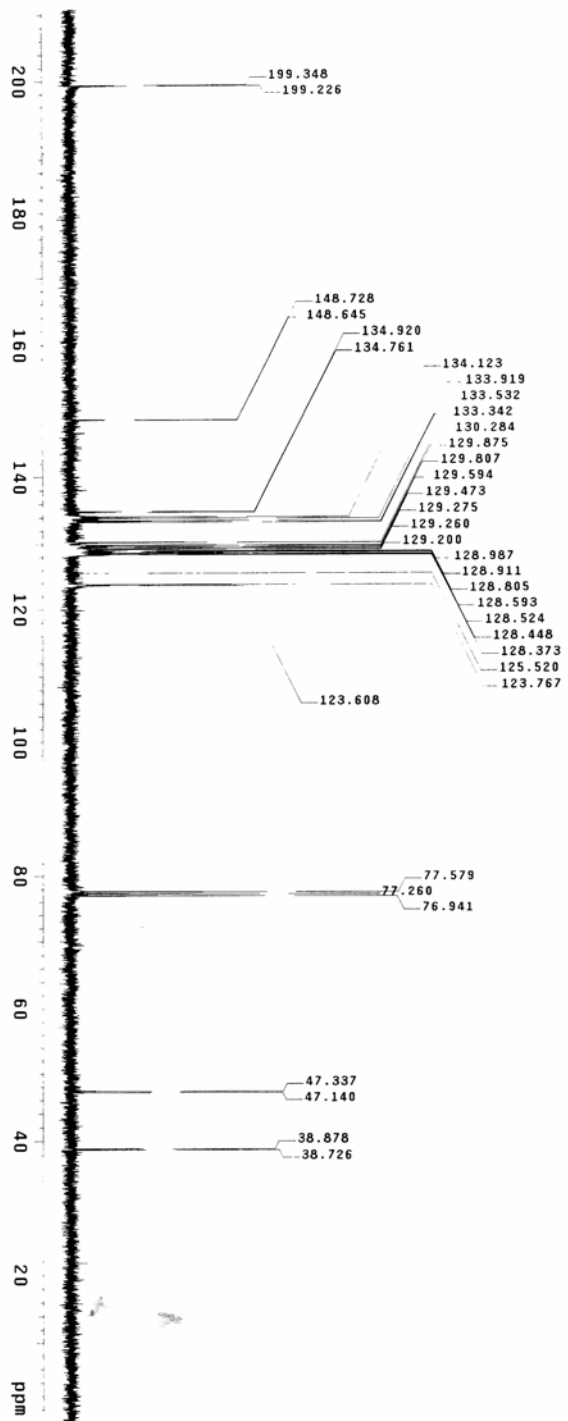
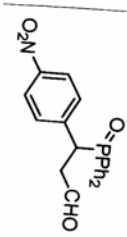
Pulse 27.6 degrees
Acq. time 0.800 sec
Width 20000.0 Hz
30 repetitions
Obscure P1, 181.883378 MHz
Obscure H1, 393.3075154 MHz
Power 42 dB
continuously on
VATZ-16 modulated
DATA PROCESSING 1.0 Hz
F1 size 32768
Total time 0 min, 27 sec











i i31004_P

Pulse Sequence: s2pu1

Solvent: CDCl₃
Temp.: 25.0 C /

```
File: 1131004_P31
Message: 40066 "ner4"
```

Her. Cui y 40000

Pulse 27.6 degrees
Acq. time 0.800 sec

Width 20000.0 Hz

16 Repetitions
OBSERVE P31, 161.896

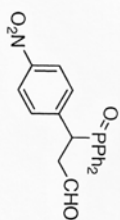
DECOUPLE H1, 399.93/
Power 42 dB

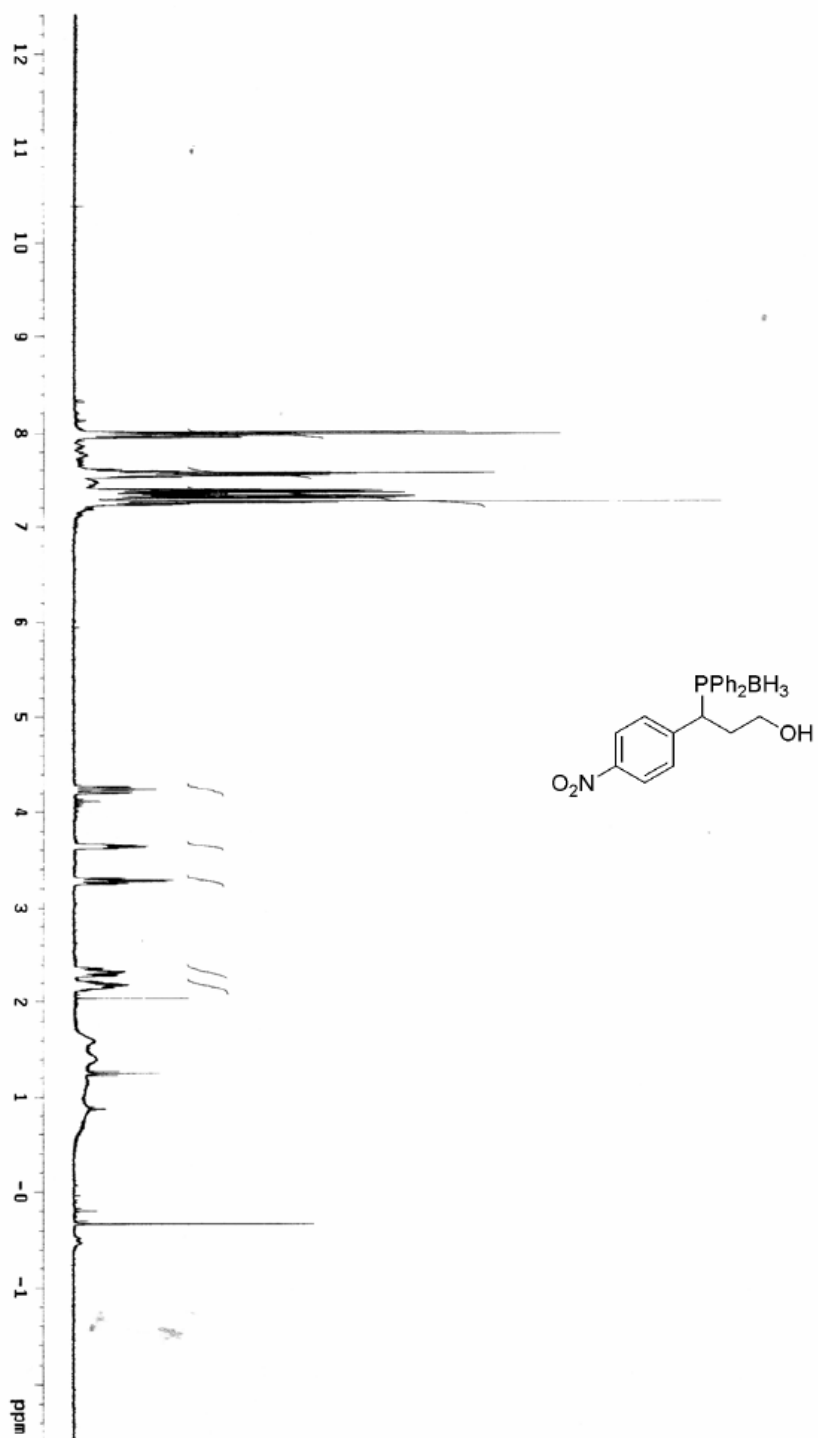
continuously on
WALTZ-16 modulated

DATA PROCESSING

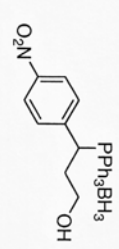
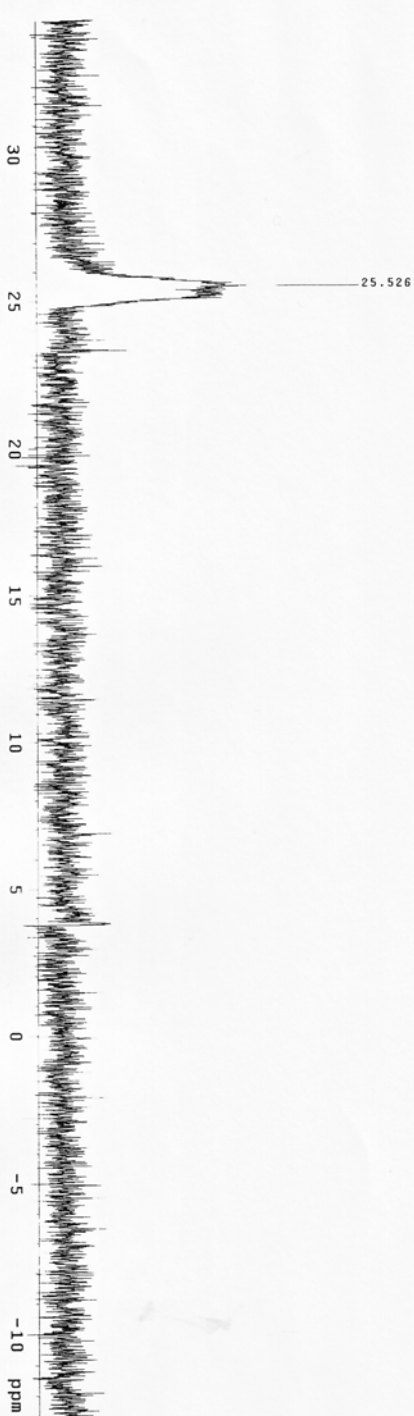
Line of accounting 1:0
FT size 32768

total time 0 min, 1s

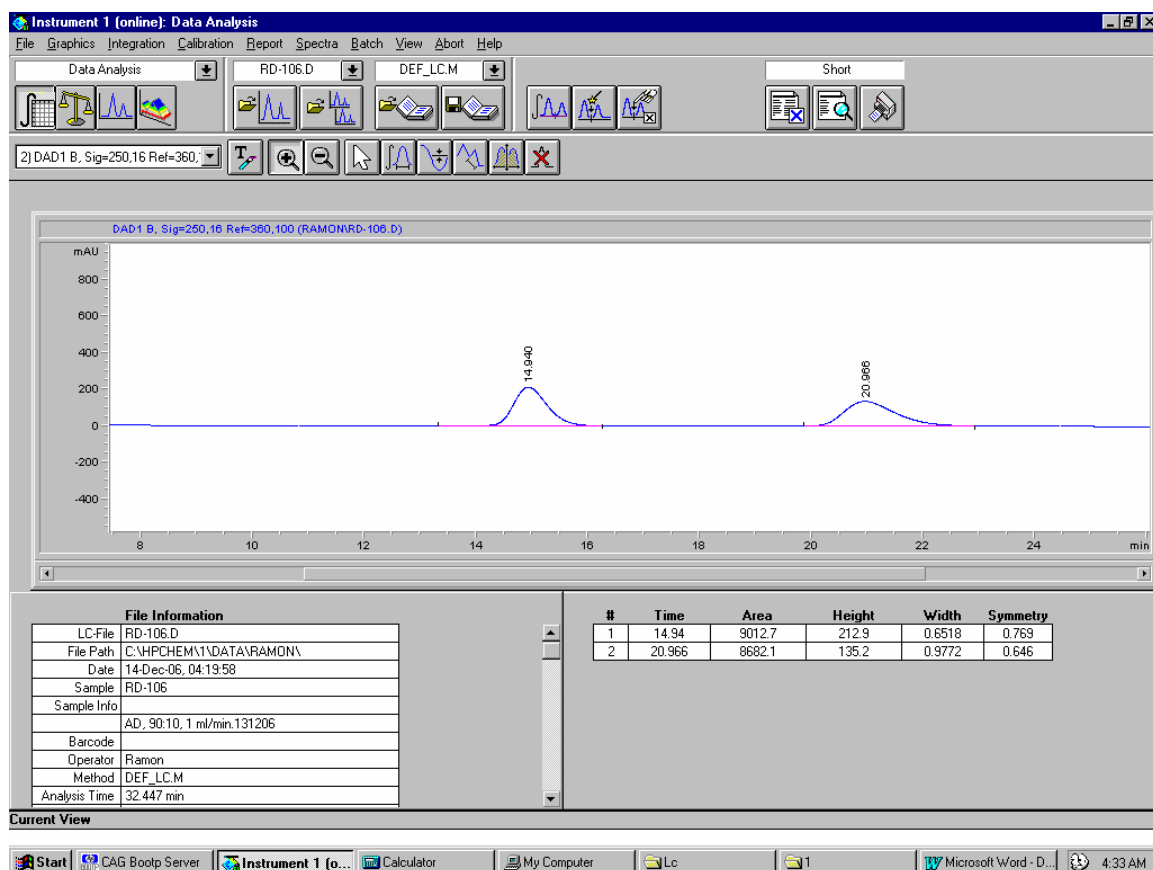
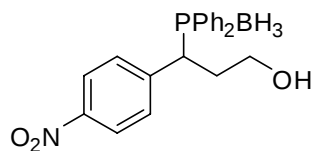


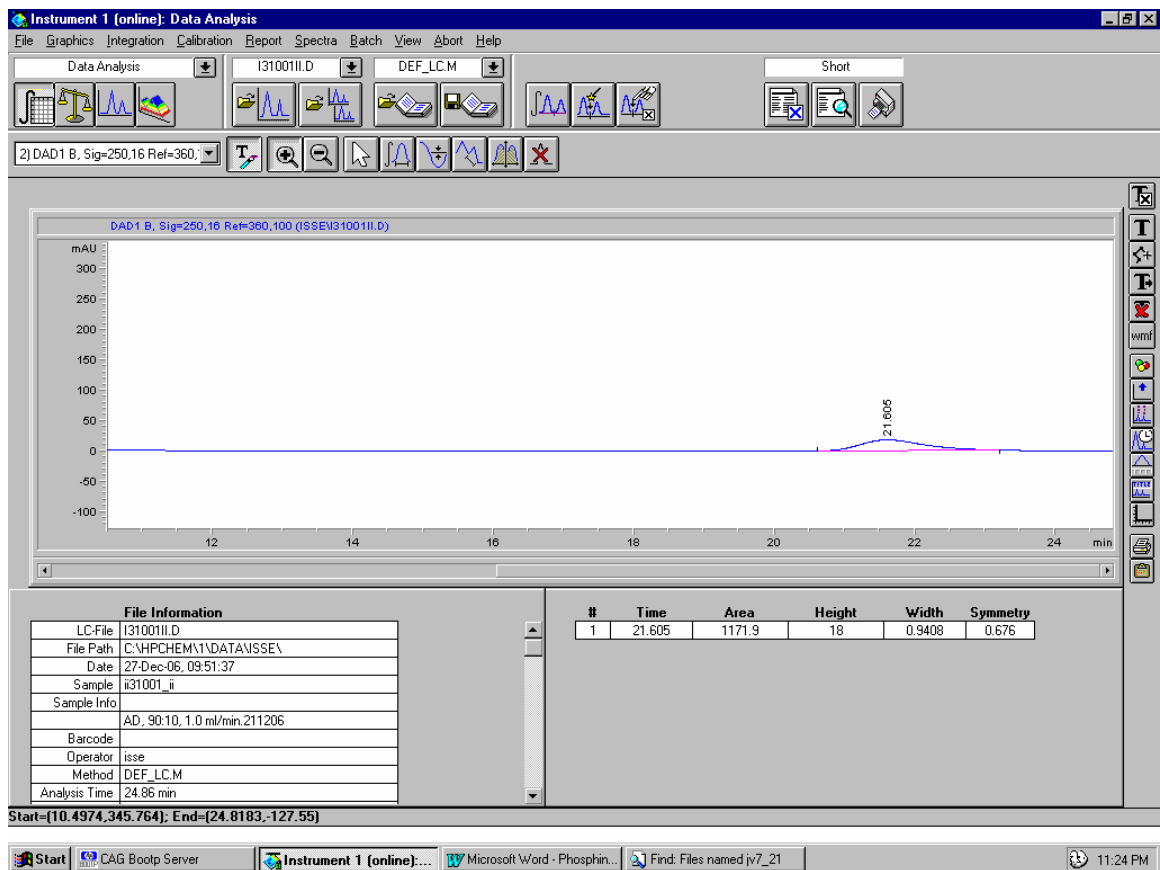
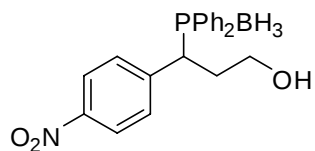


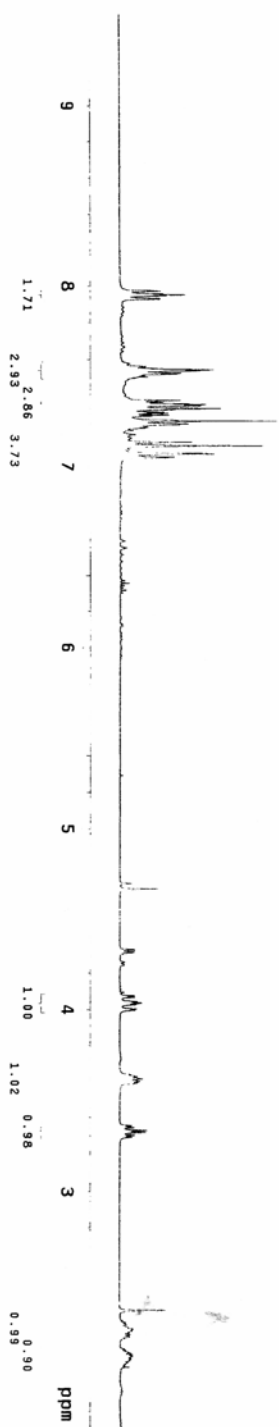
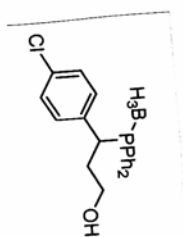
1131002_P31
Pulse Sequence: szpul
Solvent: CDCl3
Temp: 25.0 C / 299.1 K
File: 1131002_P31
Mercury-40086 "nmr4"
Pulse 27.6 degrees
Acq 3200.0 Hz
Acq 200.0 Hz
128 repetitions
OBSERVE P31, 161.8967379 MHz
DECOUPLE H1, 399.9376154 MHz
Power 42 dB
CONTINUOUSLY ON
SOLVENT DECOUPLED
DATA PROCESSING
Line broadening 1.0 Hz
FT size 32768
Total time 4 hr, 48 min, 3 sec

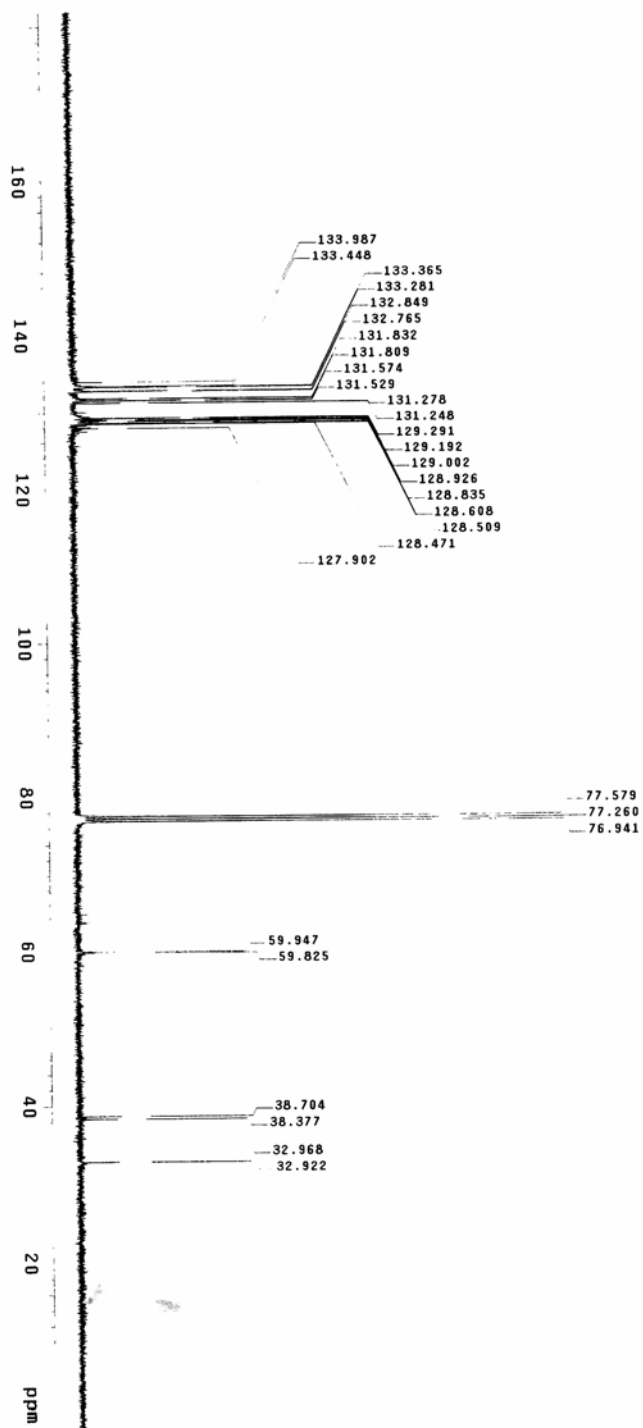
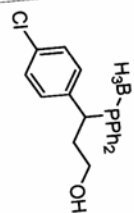




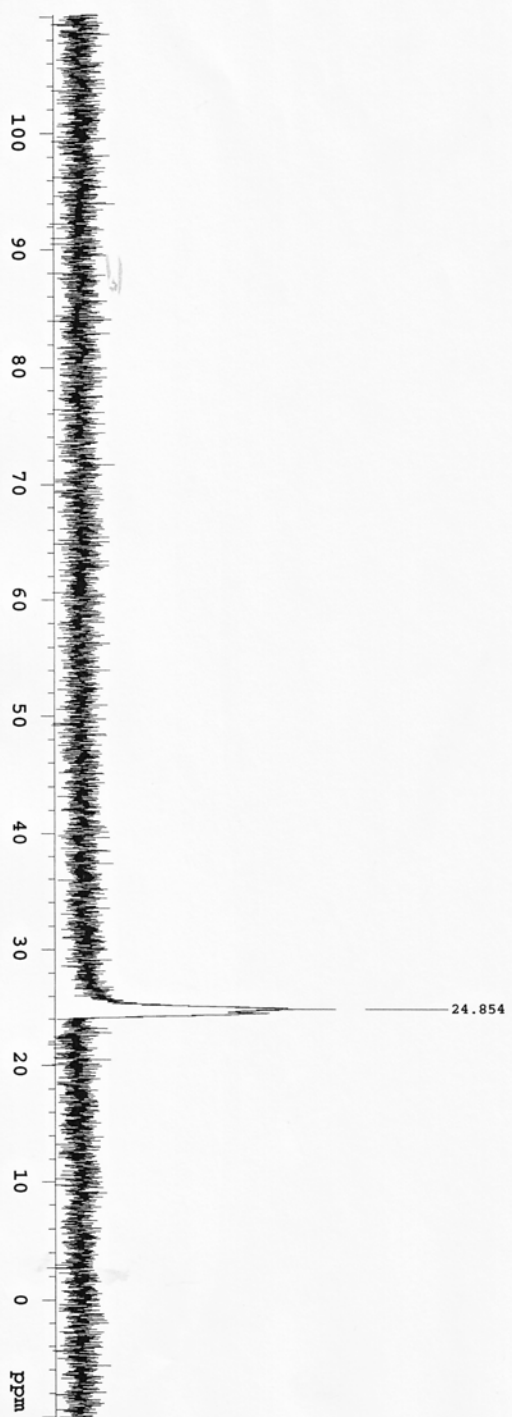
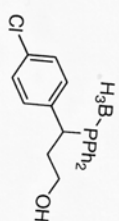


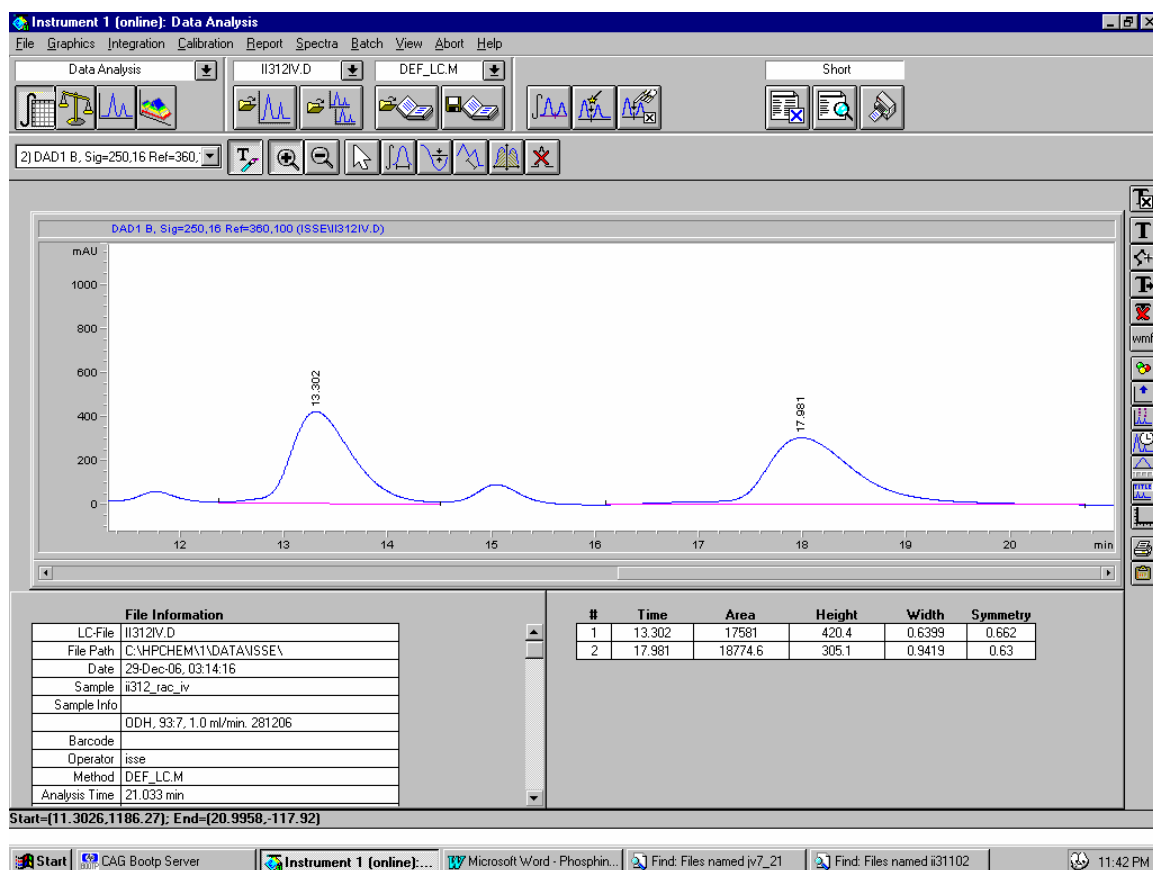
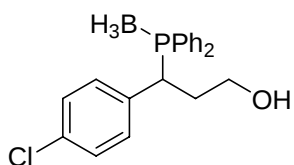


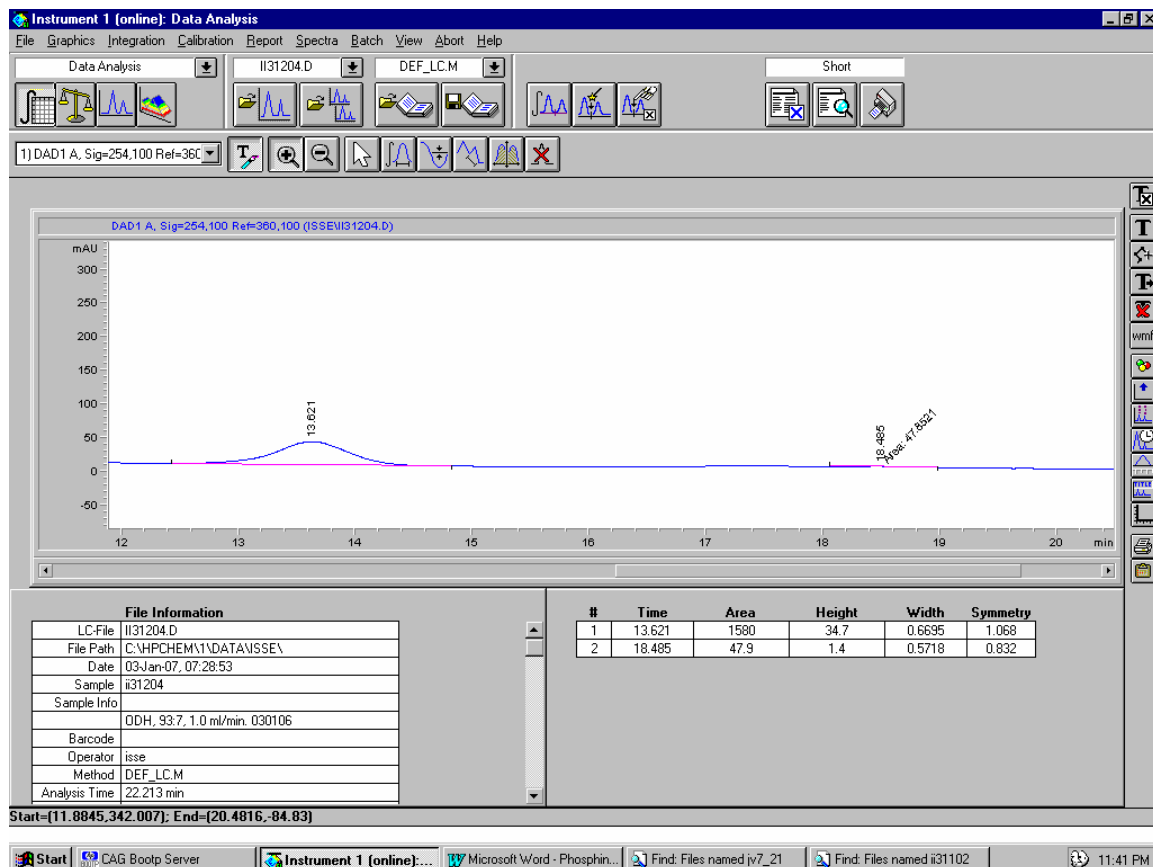
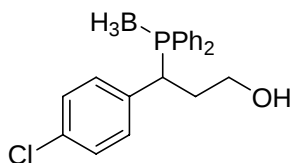


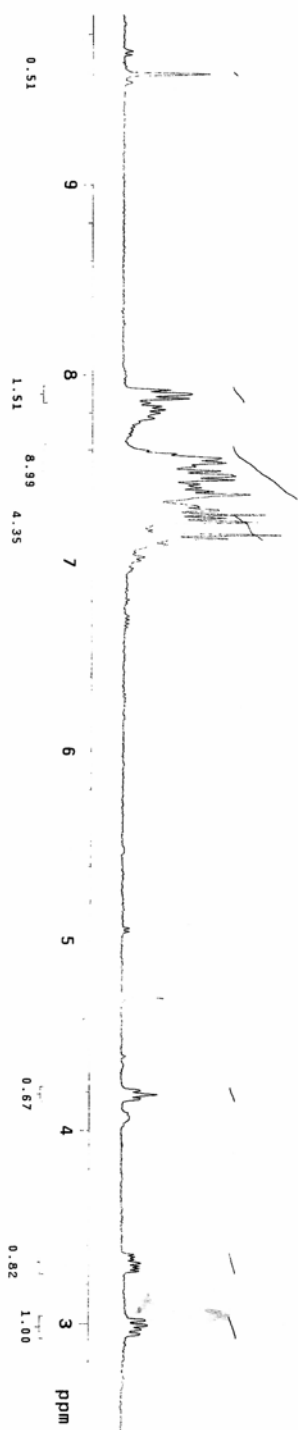
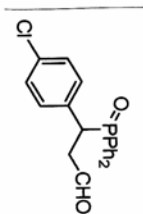


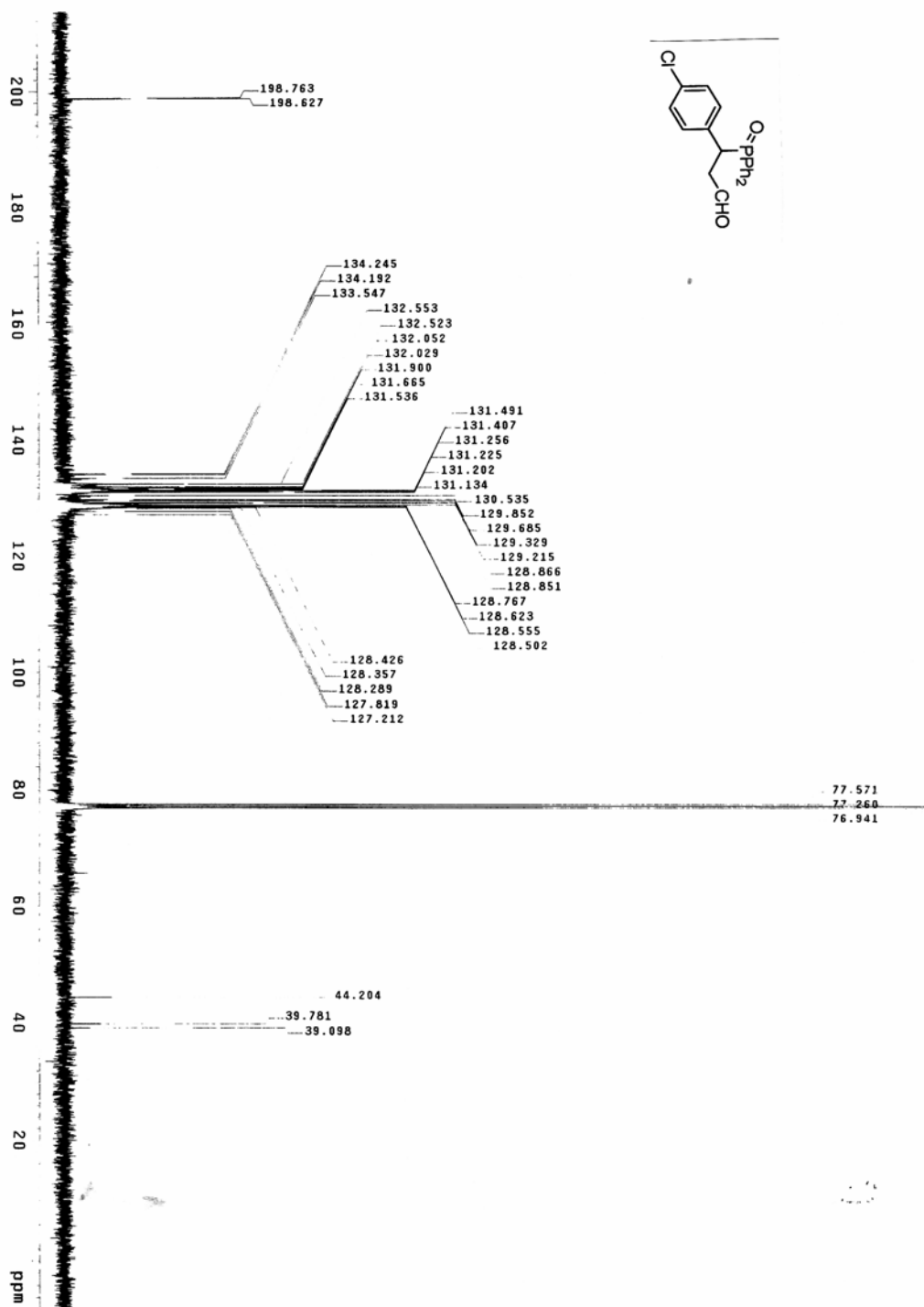
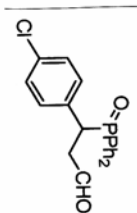
INDEX	FREQUENCY	PPM	HEIGHT
1	4023.730	24.854	45.4

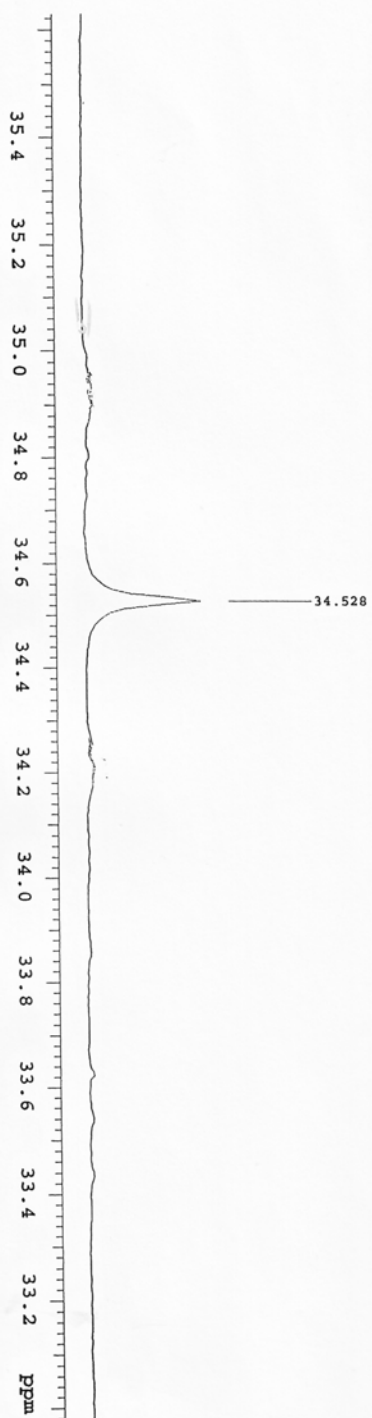
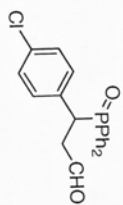


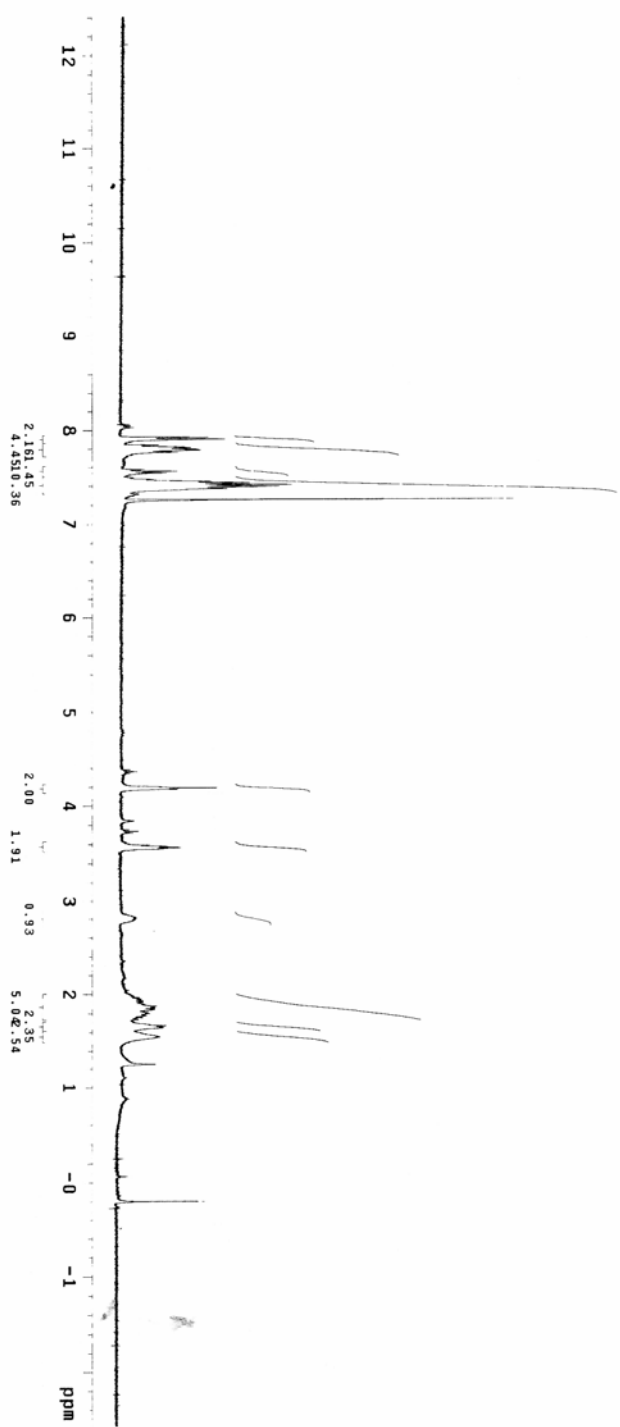
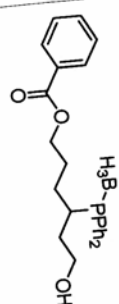


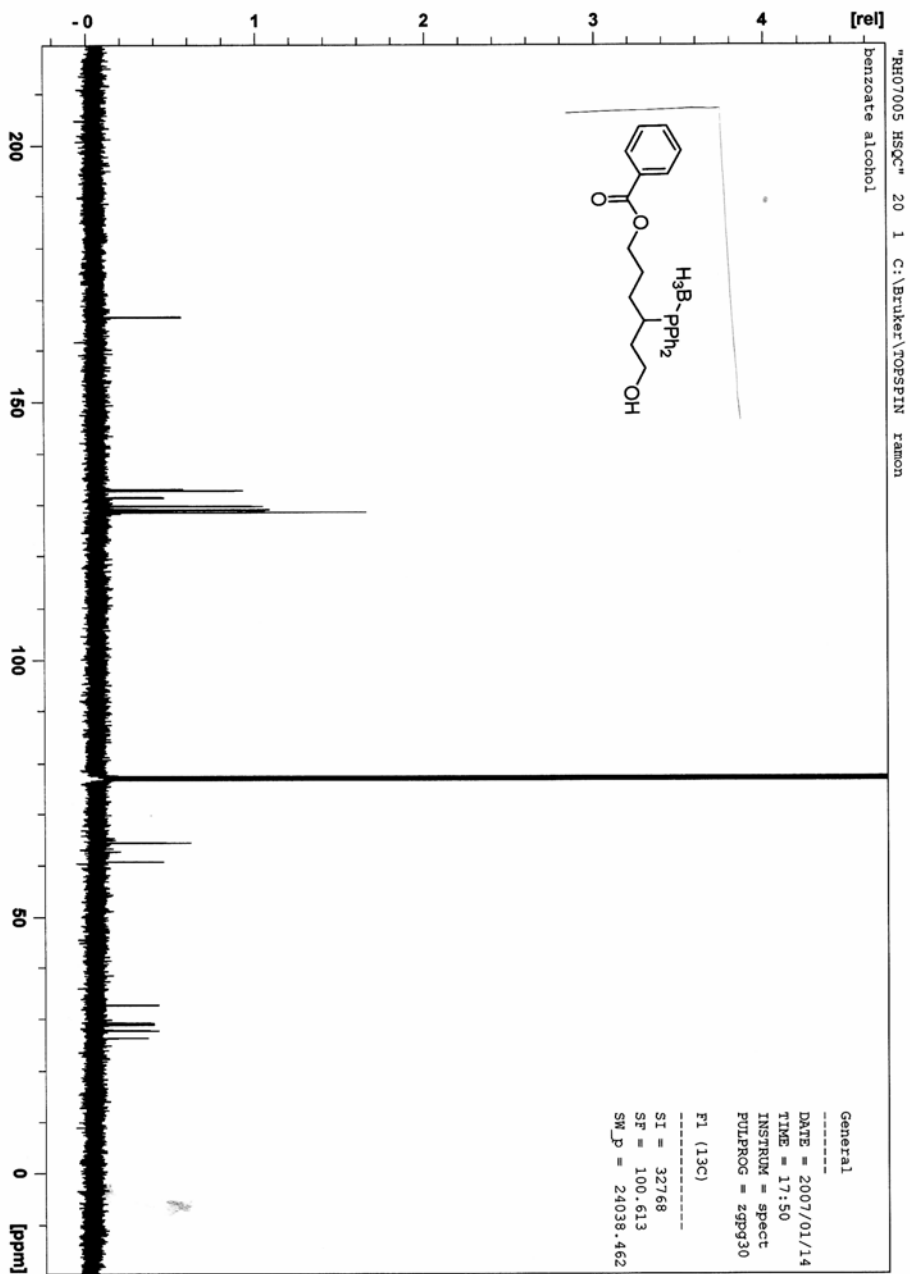


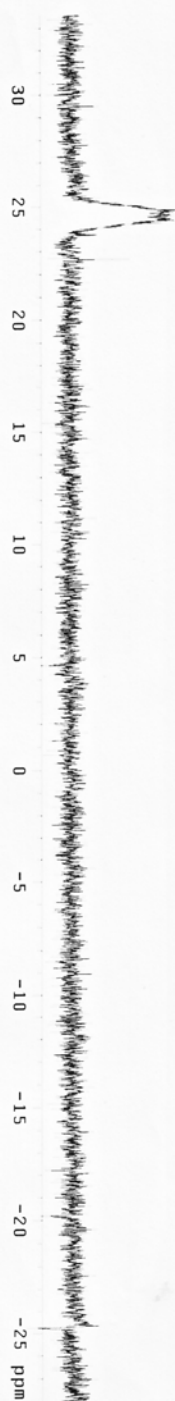
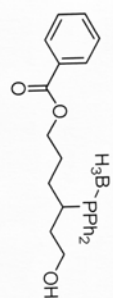


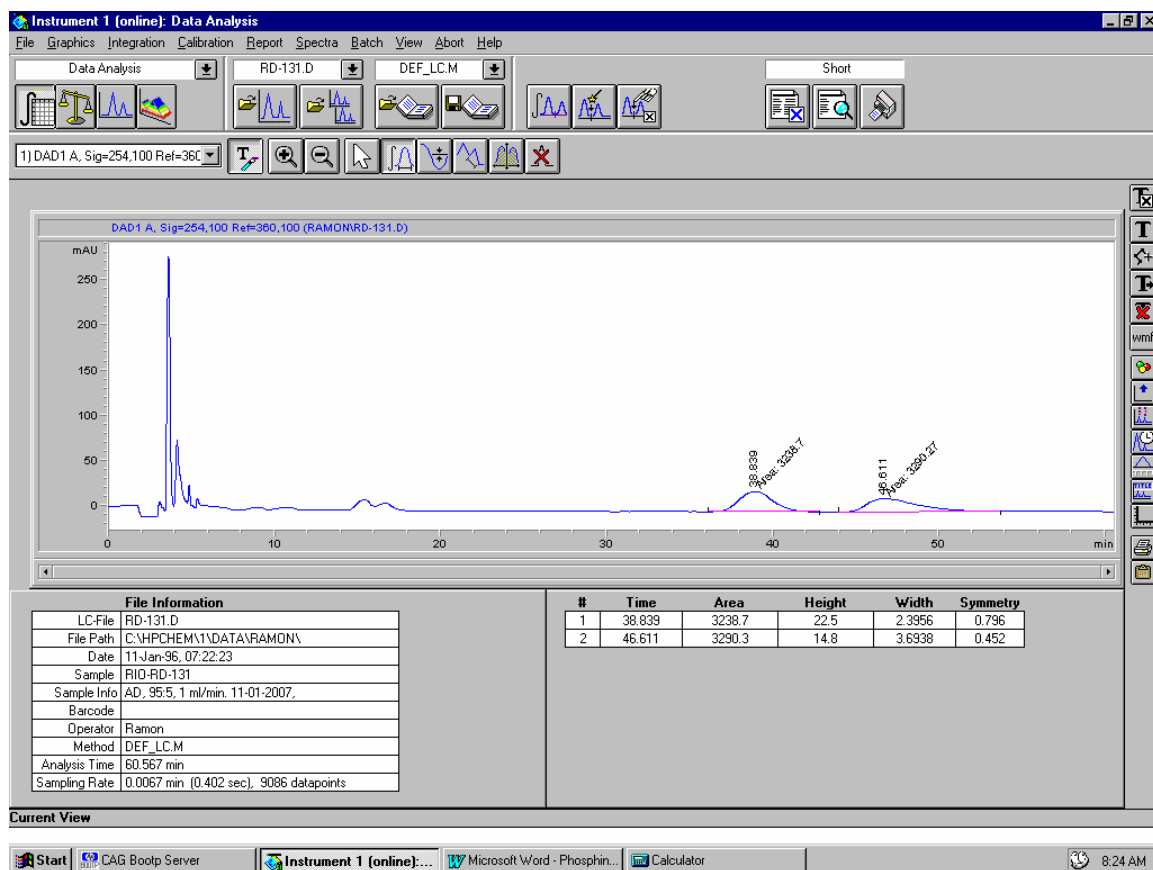
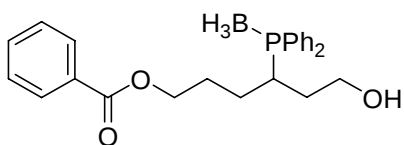


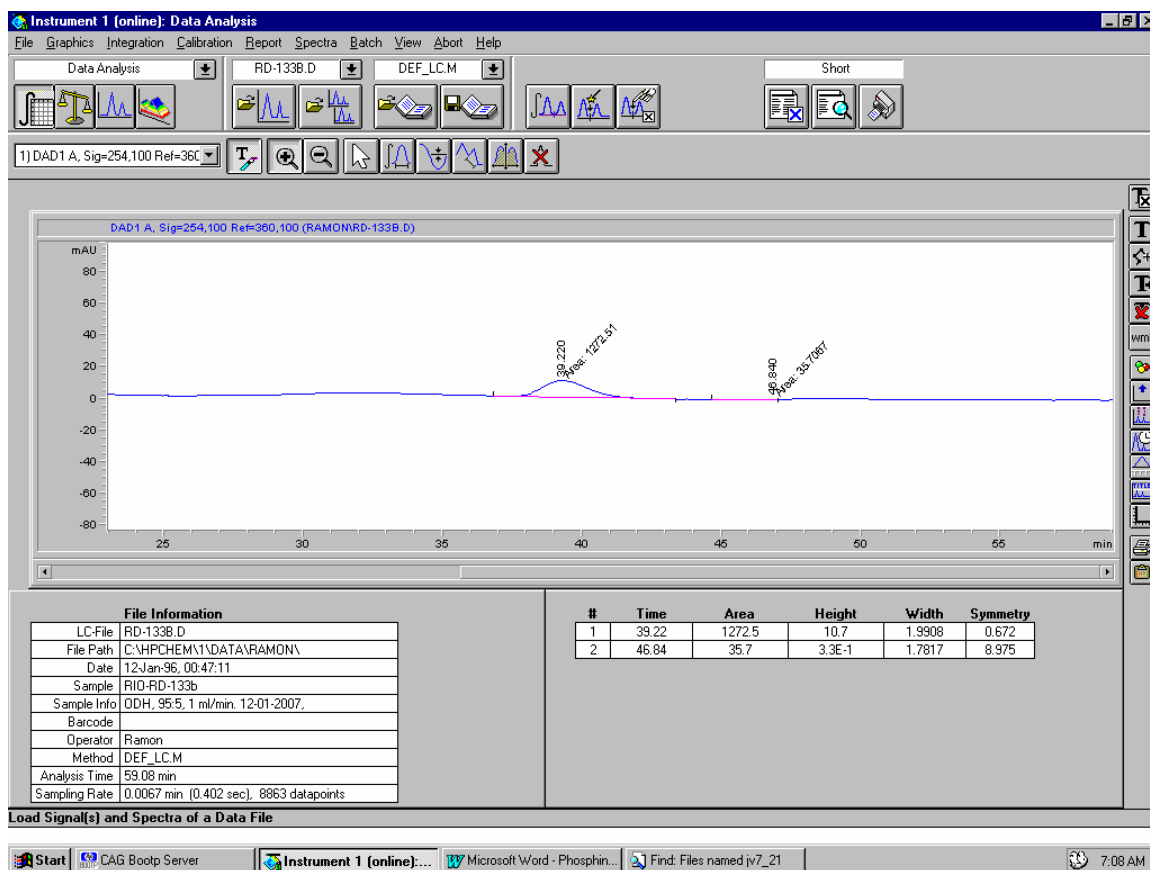
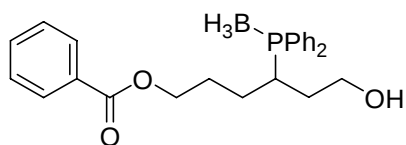
1

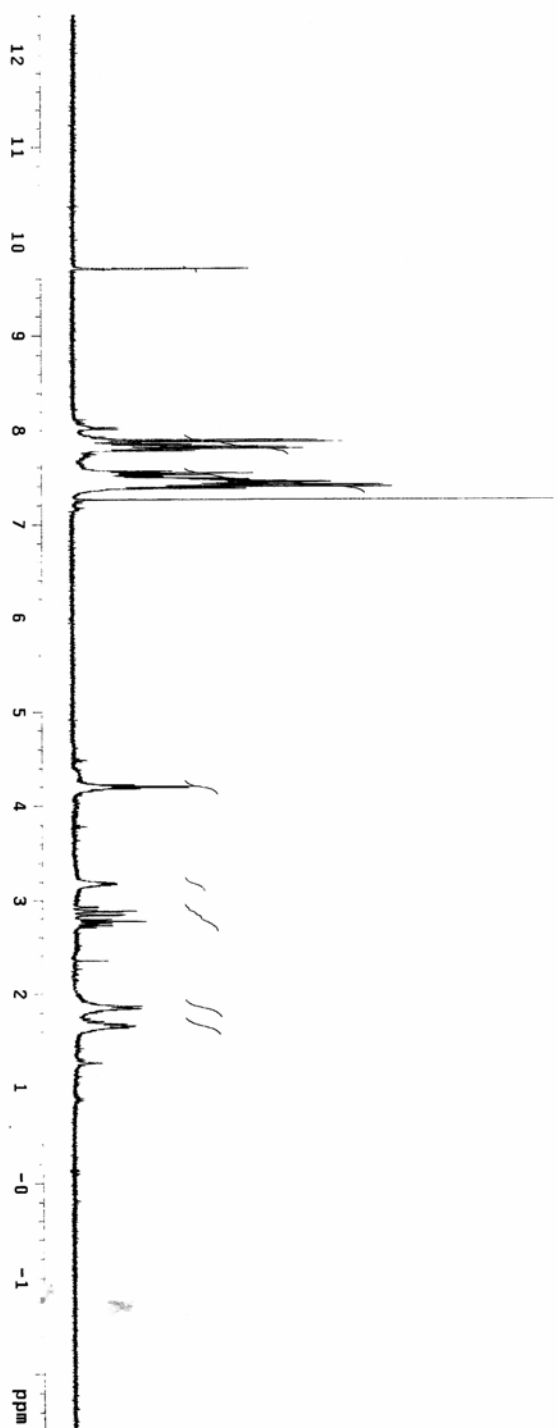
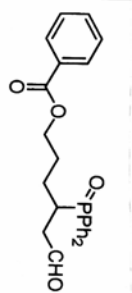


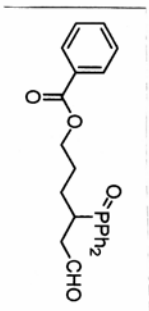


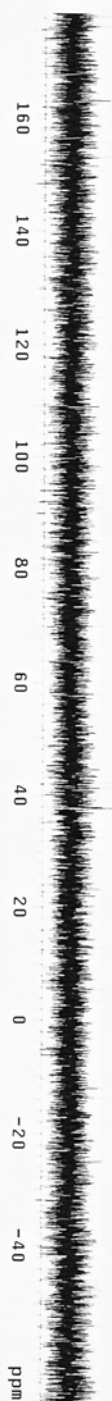
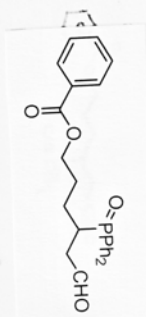




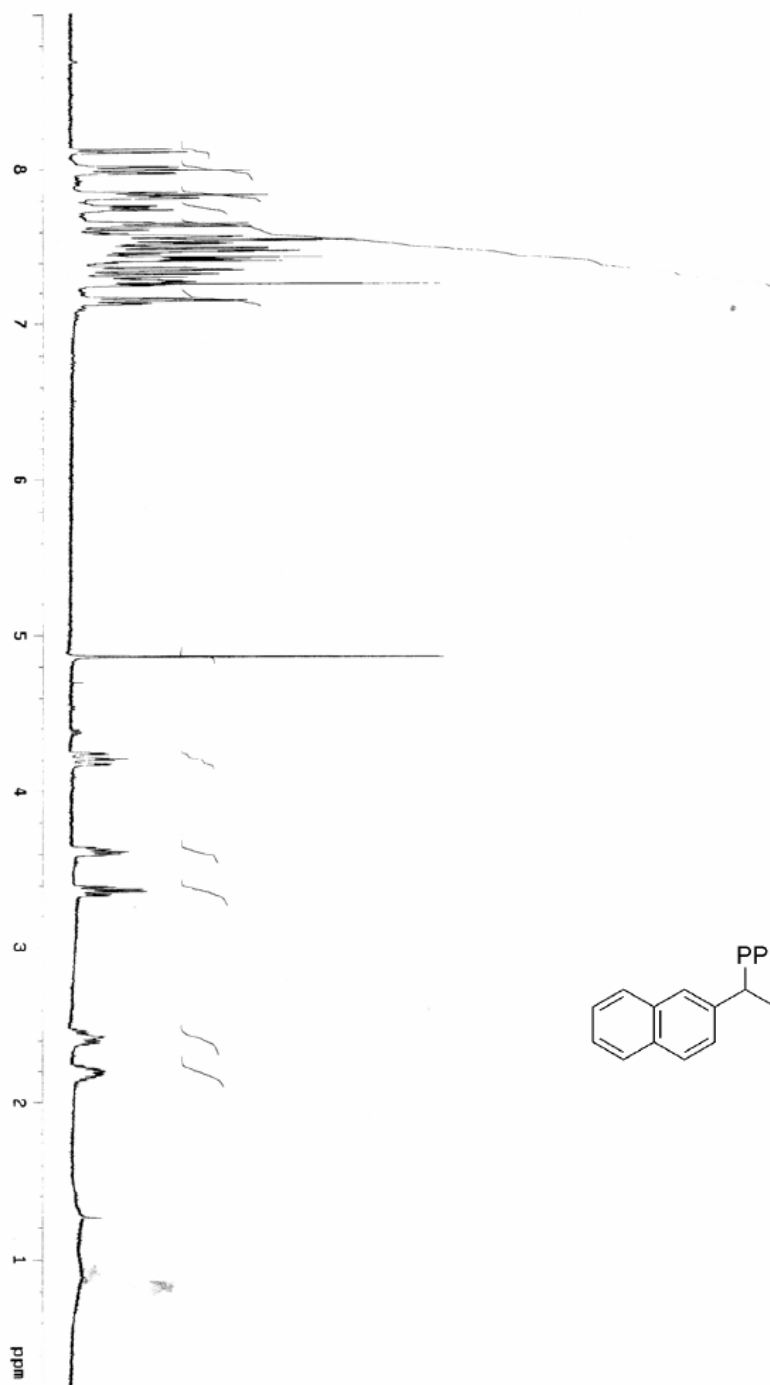
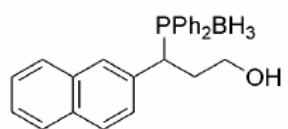


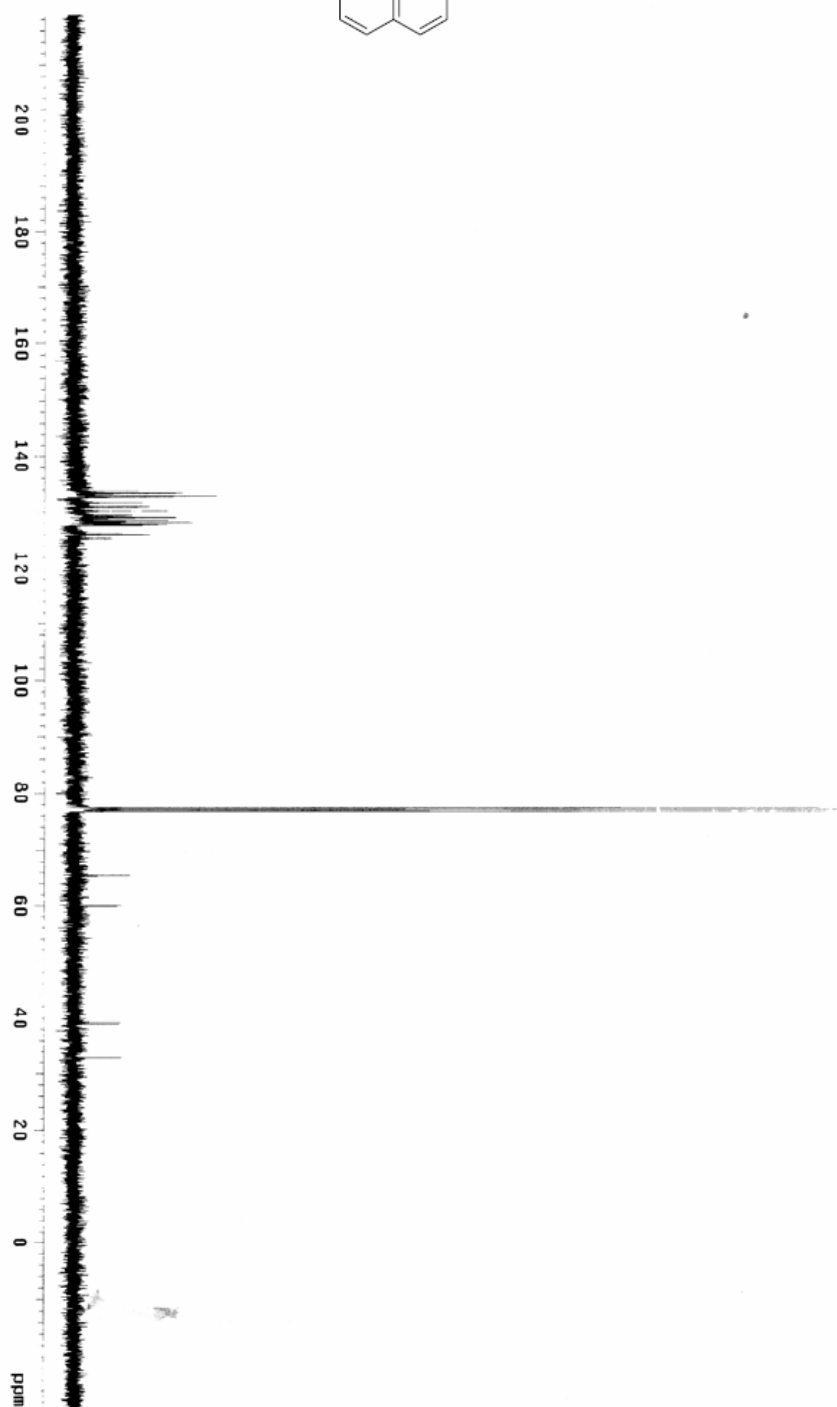
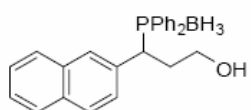


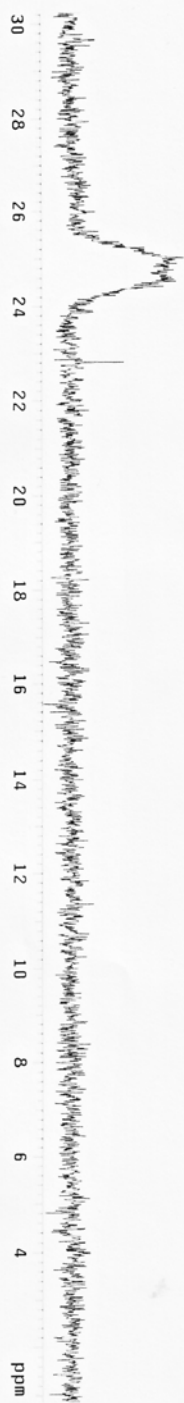
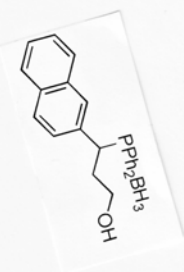


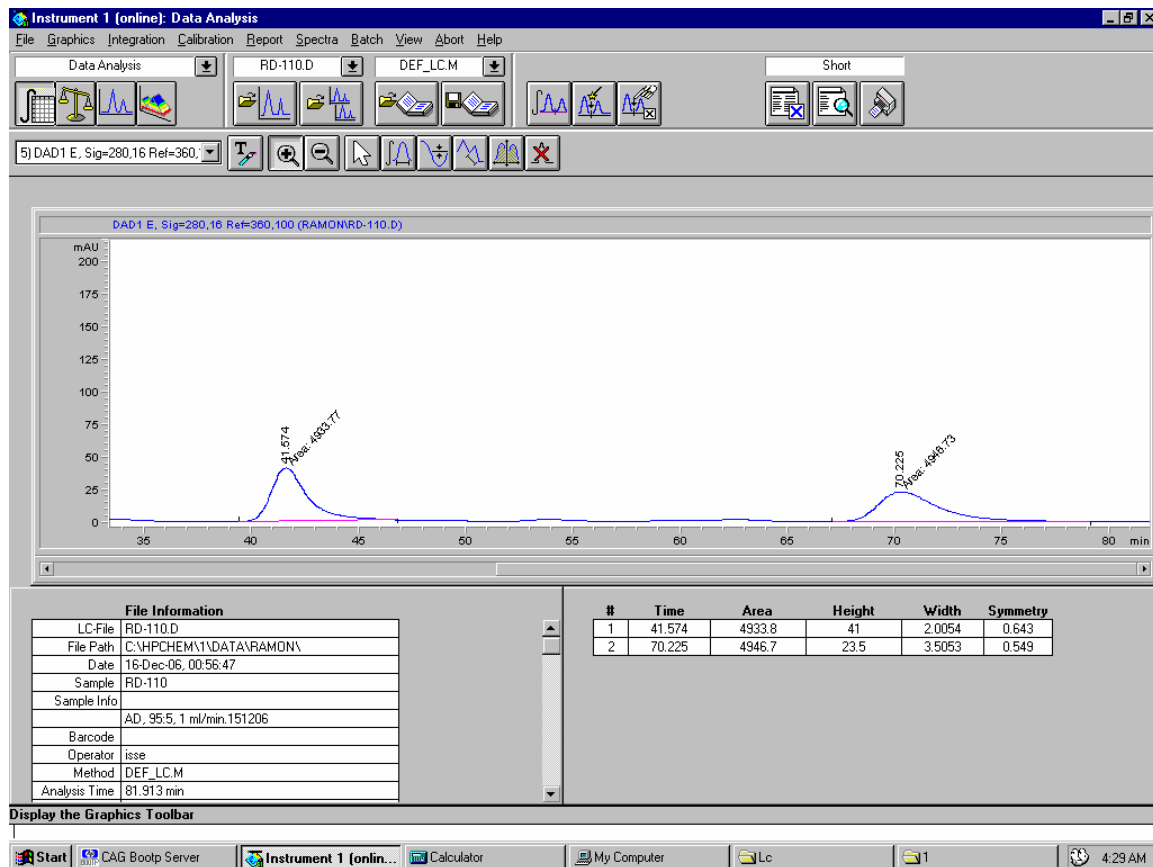
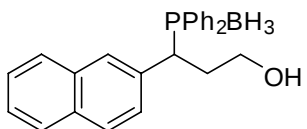


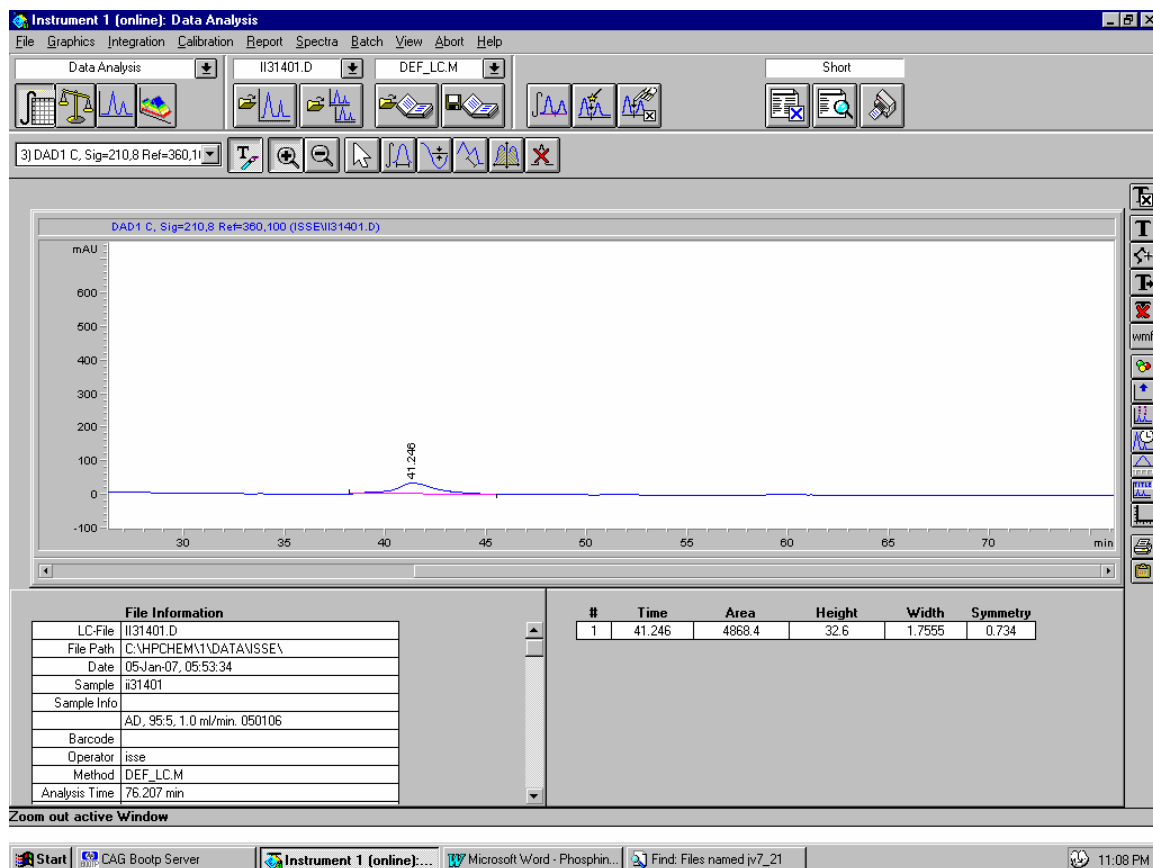
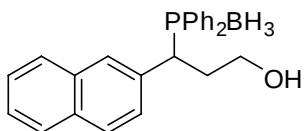
37.697

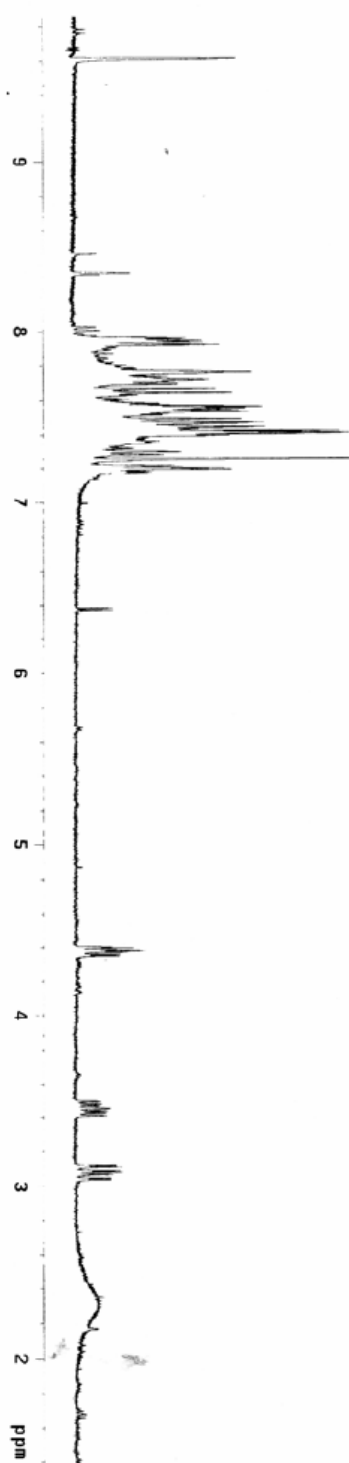
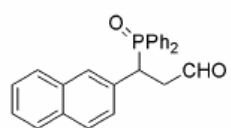


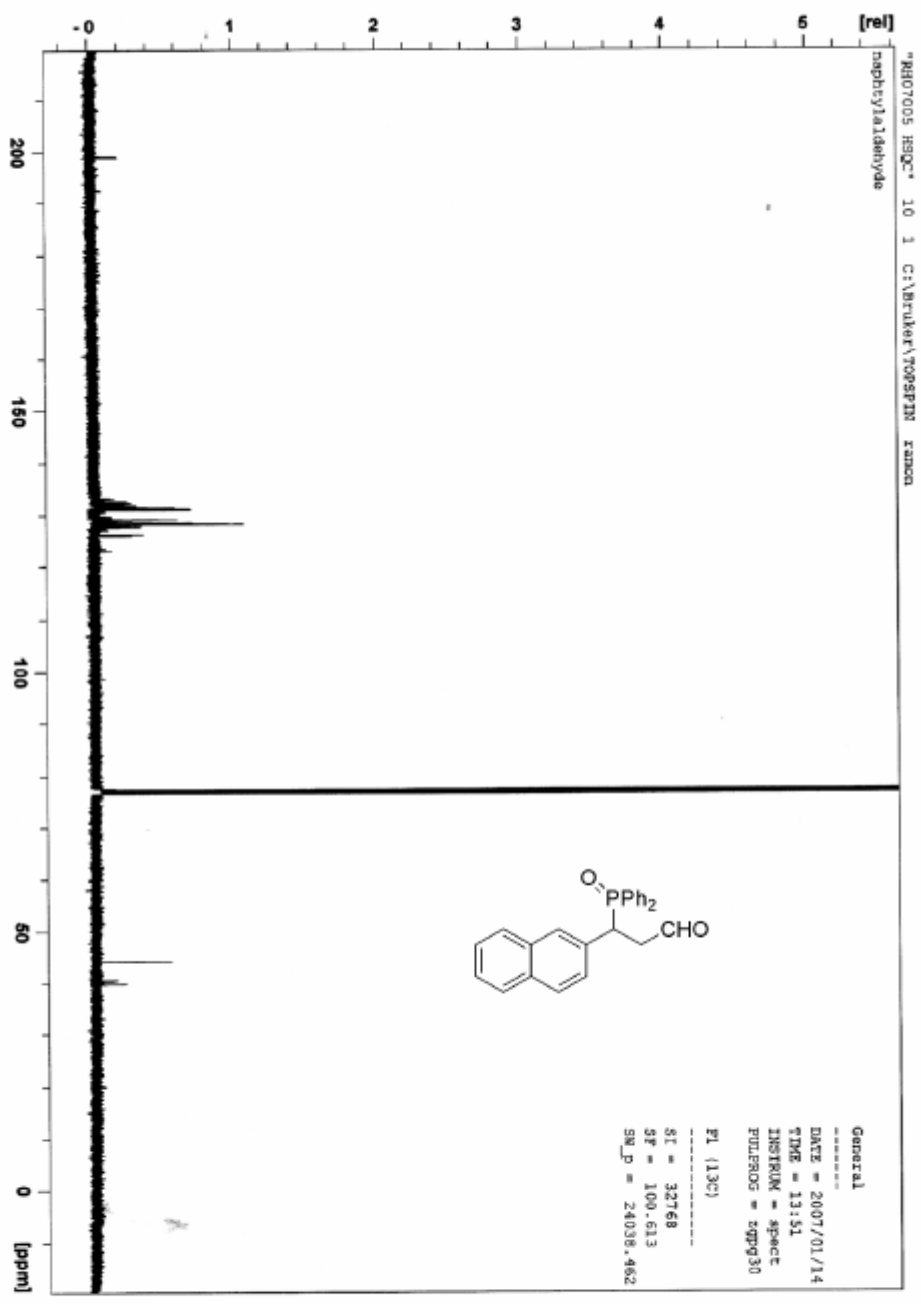












jv7_21_4R

Pulse Sequence: szpul

Solvent: CDCl3

Temp. 40.0 C / 313.1 K

Mercury-400DB "nmr4"

Relax. delay 1.000 sec

Pulse 41.6 degrees

Acq. time 2.733 sec

Width 5995.2 Hz

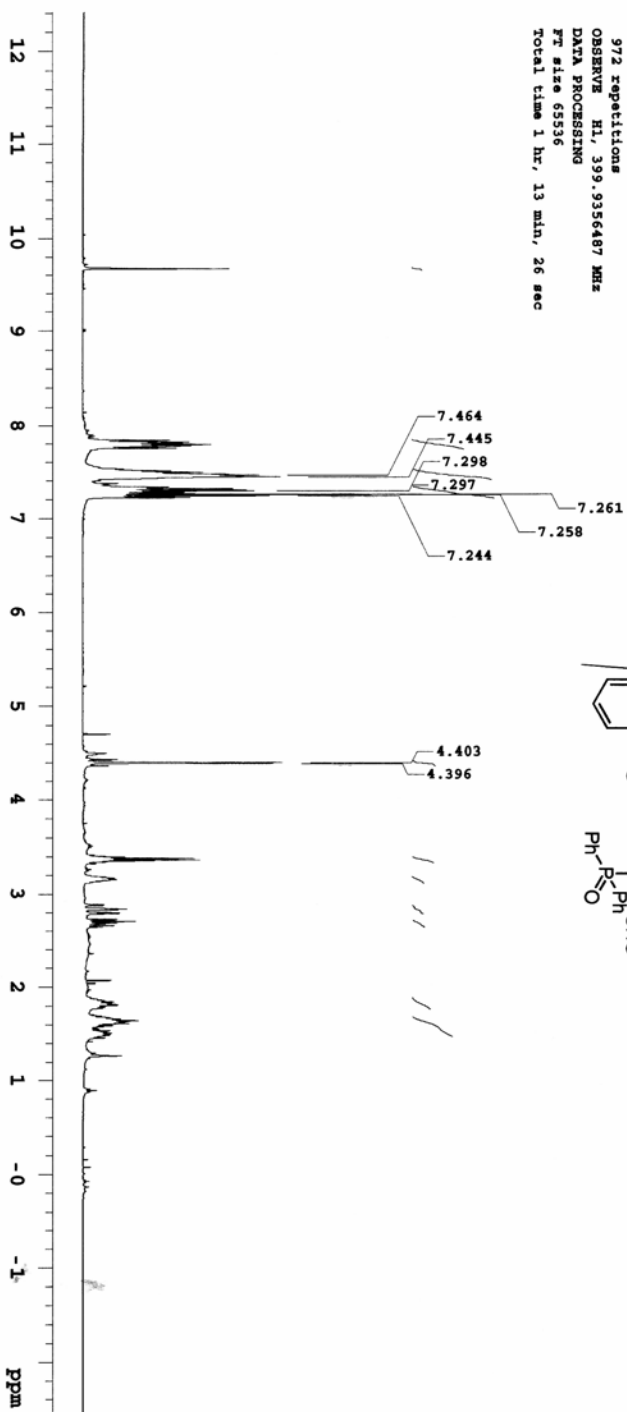
972 repetitions

OBSERVE H1, 399.9356487 MHz

DATA PROCESSING

FT size 65536

Total time 1 hr, 13 min, 26 sec



jr07_21_4

Pulse Sequence: szpul

Solvent: CDCl₃

Temp. 40.0 C / 313.1 K

Mercury-400DB "nmr4"

Relax. delay 1.000 sec

Pulse 48.9 degrees

Acq. time 1.199 sec

Width 25000.0 Hz

1792 repetitions

OBSERVE C13, 100.5638771 MHz

DECOUPLE H1, 399.9376154 MHz

Power 42 dB

continuously on

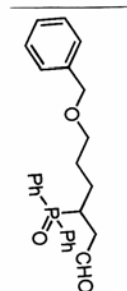
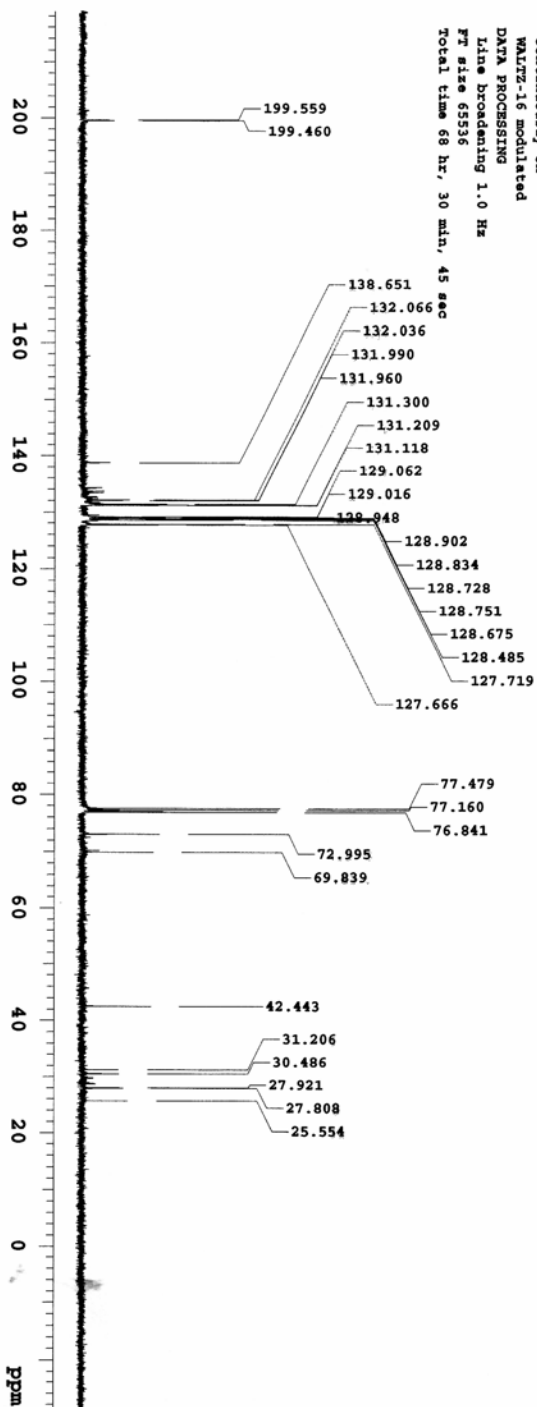
WALTZ-16 modulated

DATA PROCESSING

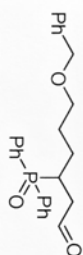
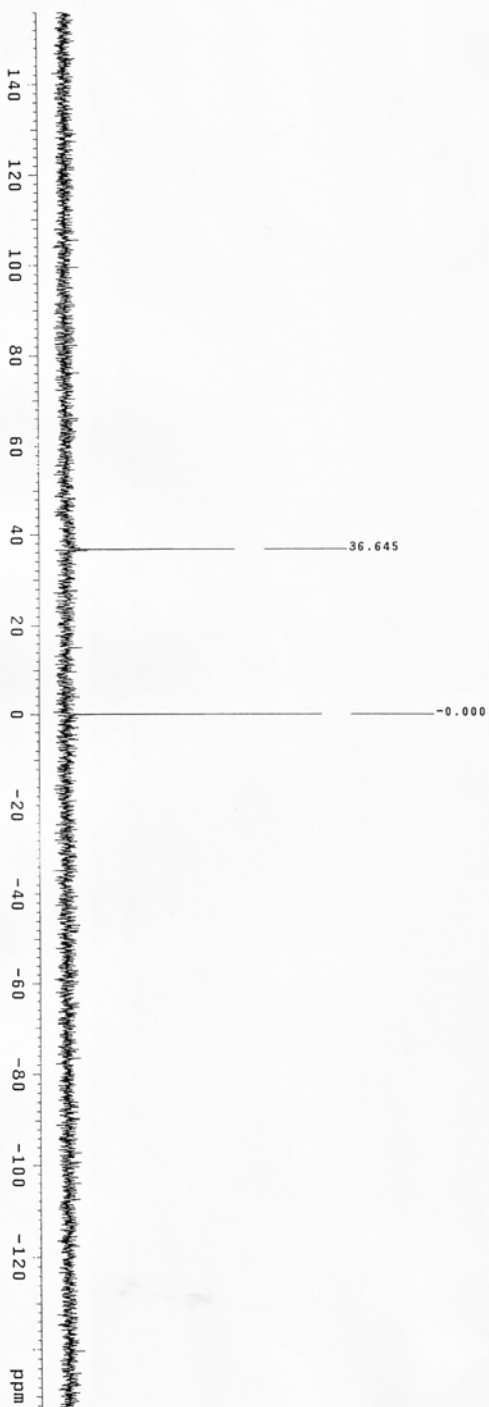
Line broadening 1.0 Hz

FT size 65536

Total time 68 hr, 30 min, 45 sec



Jv07_21_3P
 Pulse Sequence: szpul
 Solvent: CDCl3
 Temp: 25.0 C / 298.1 K
 File: Jv07_21_3P
 Mercury-400WB 7mm-4"
 Pulse 27.6 degrees
 Acq time 0.680 sec
 Width 5000.0 Hz
 98 repetitions
 OBSERVE P31, 161.8969256 MHz
 DECOUPLE H1, 399.3976154 MHz
 500.136480 MHz
 Continuously on
 WALTZ-16 modulated
 DATA PROCESSING
 Line broadening 5.0 Hz
 FT size 131072
 Total time 56 min, 25 sec



jv07_17_4

Pulse Sequence: szpul

Solvent: CDCl3

Temp. 25.0 C / 298.1 K

File: jv07_17_4

Mercury-400DB "nmr4"

Relax. delay 1.000 sec

Pulse 41.6 degrees

Acq. time 2.733 sec

Width 5995.2 Hz

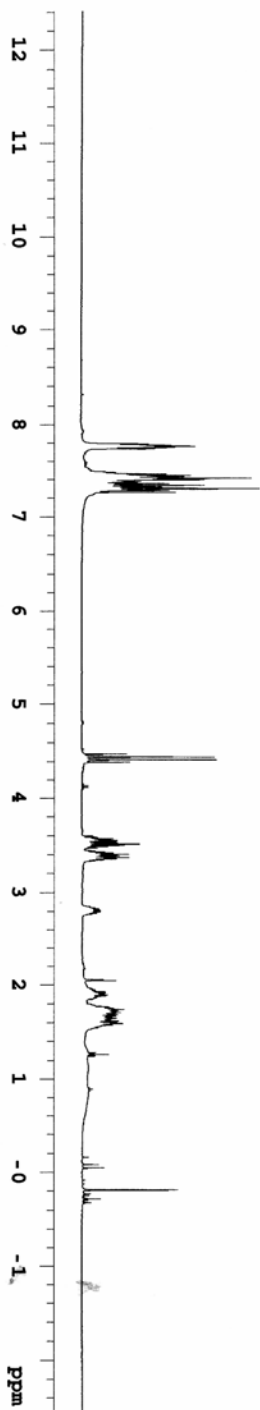
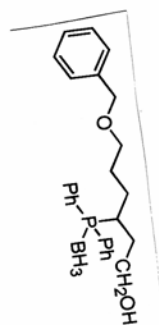
68 repetitions

OBSERVE H1, 399.9356487 MHz

DATA PROCESSING

FT size 65536

Total time 1 hr, 13 min, 26 sec



jv07_17_4

Pulse Sequence: szpul

Solvent: CDCl₃

Temp. 25.0 C / 298.1 K

File: jv07_17_4c

Mercury-400BB "nmr4"

Relax. delay 1.000 sec

Pulse 48.9 degrees

Acq. time 1.199 sec

Width 25000.0 Hz

352 repetitions

OBSERVE C13, 100.563885 MHz

DECOUPLE H1, 399.9376154 MHz

Power 42 dB

continuously on

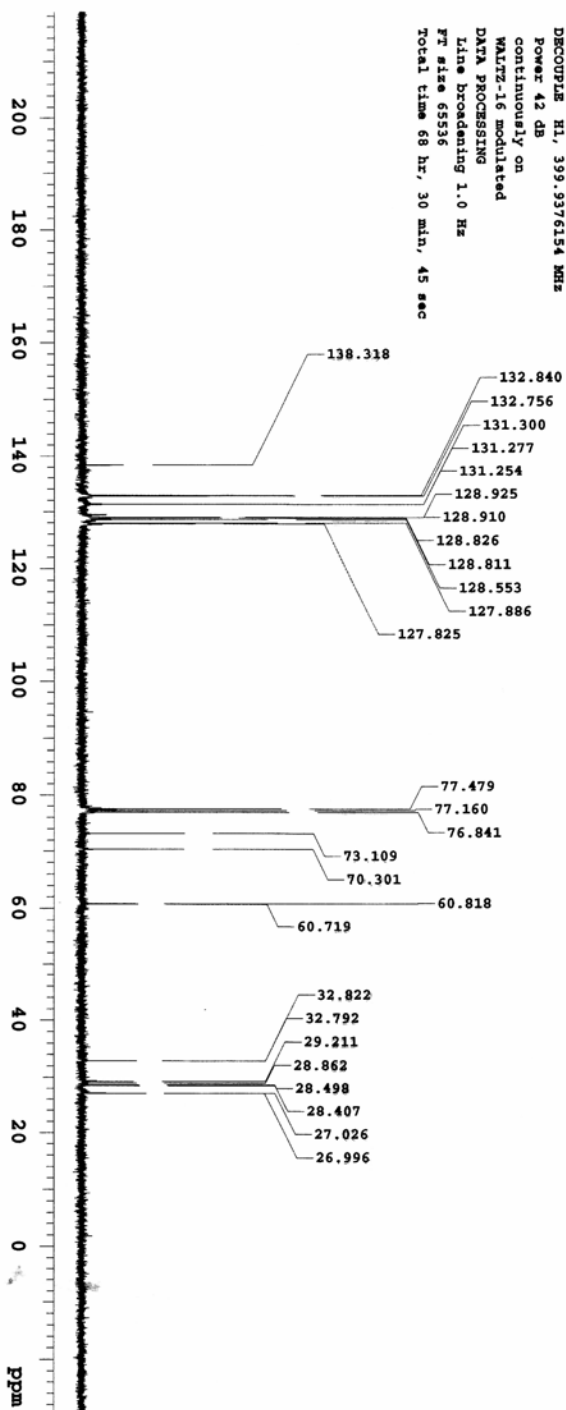
WALTZ-16 modulated

DATA PROCESSING

Line broadening 1.0 Hz

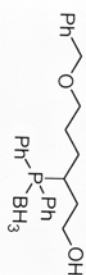
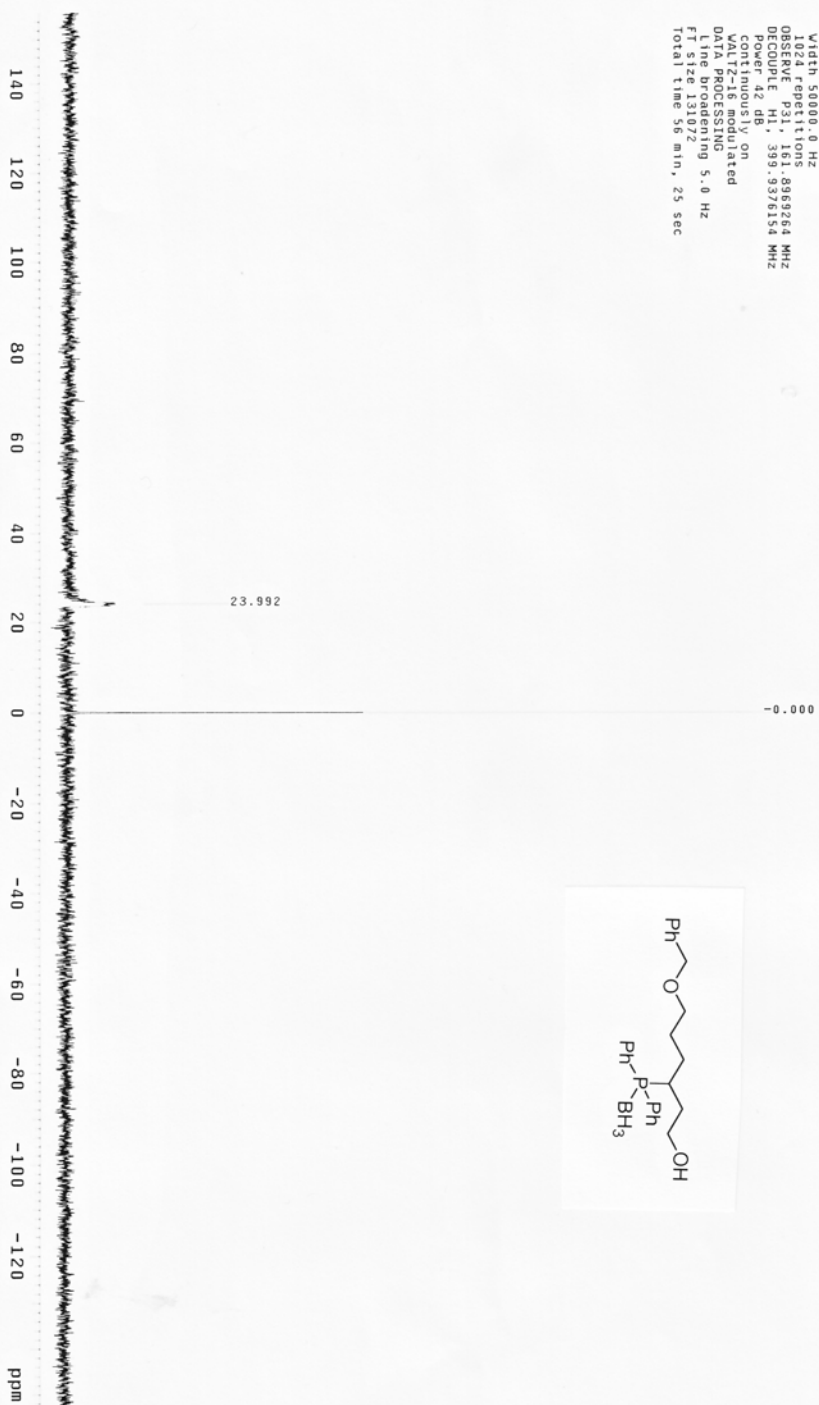
FT size 65536

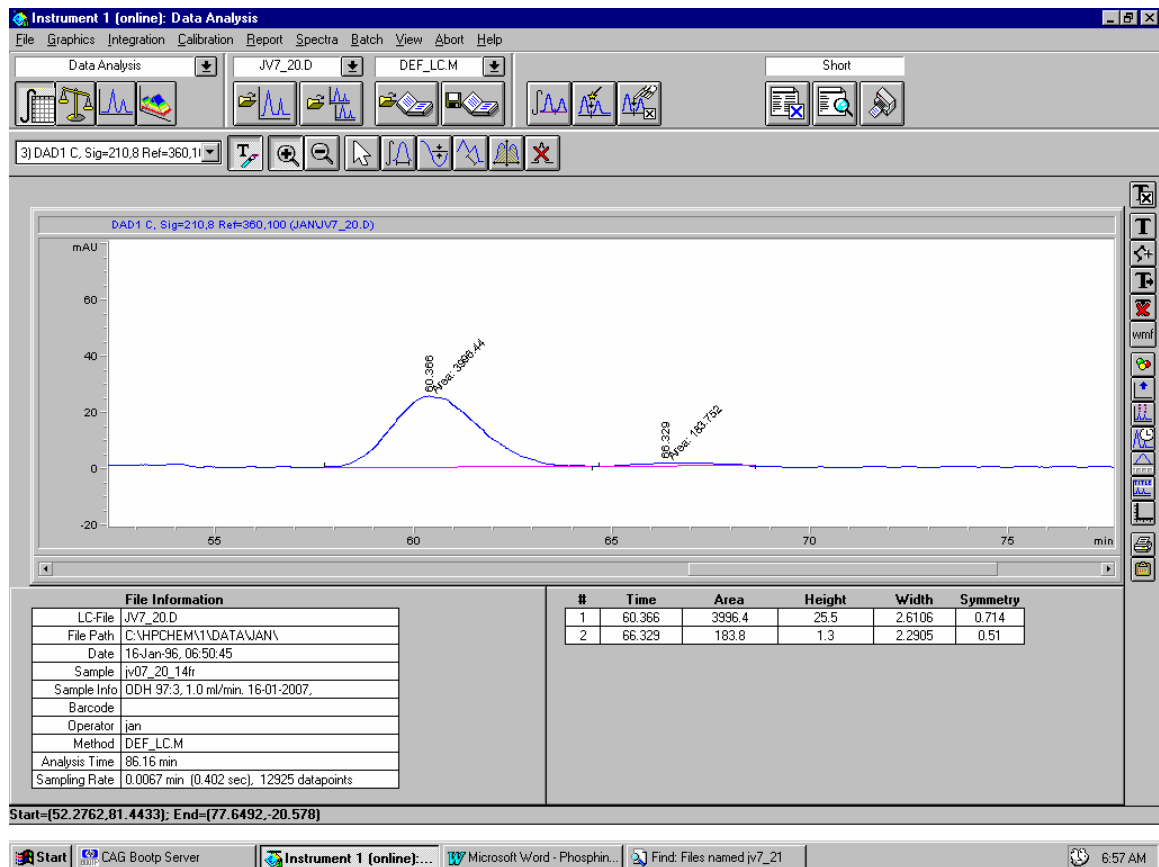
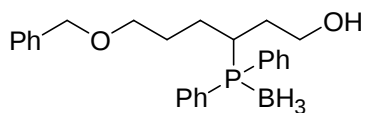
Total time 68 hr, 30 min, 45 sec

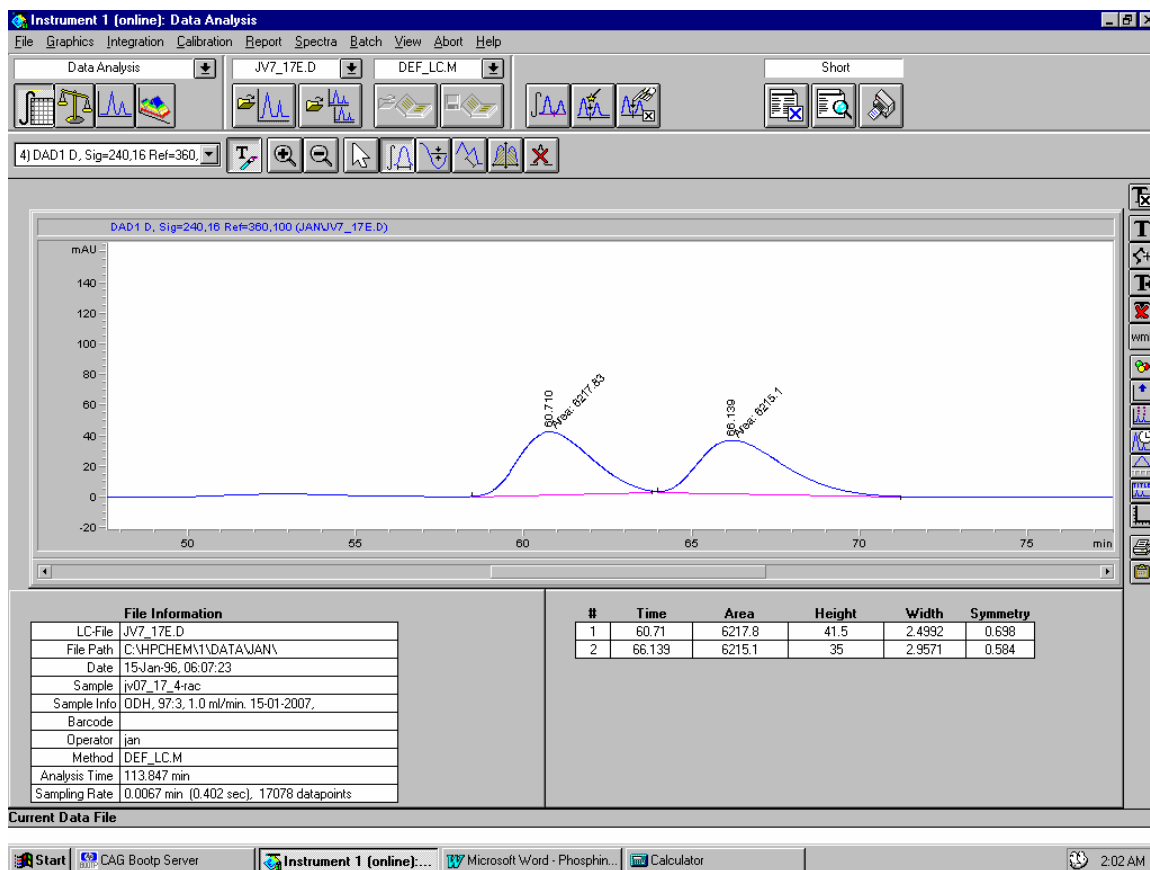
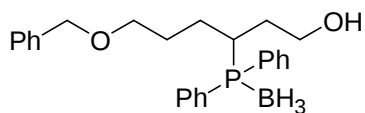


PHOSPHORUS OBSERVE
STANDARD PARAMETERS
PHOSPHATE REGION

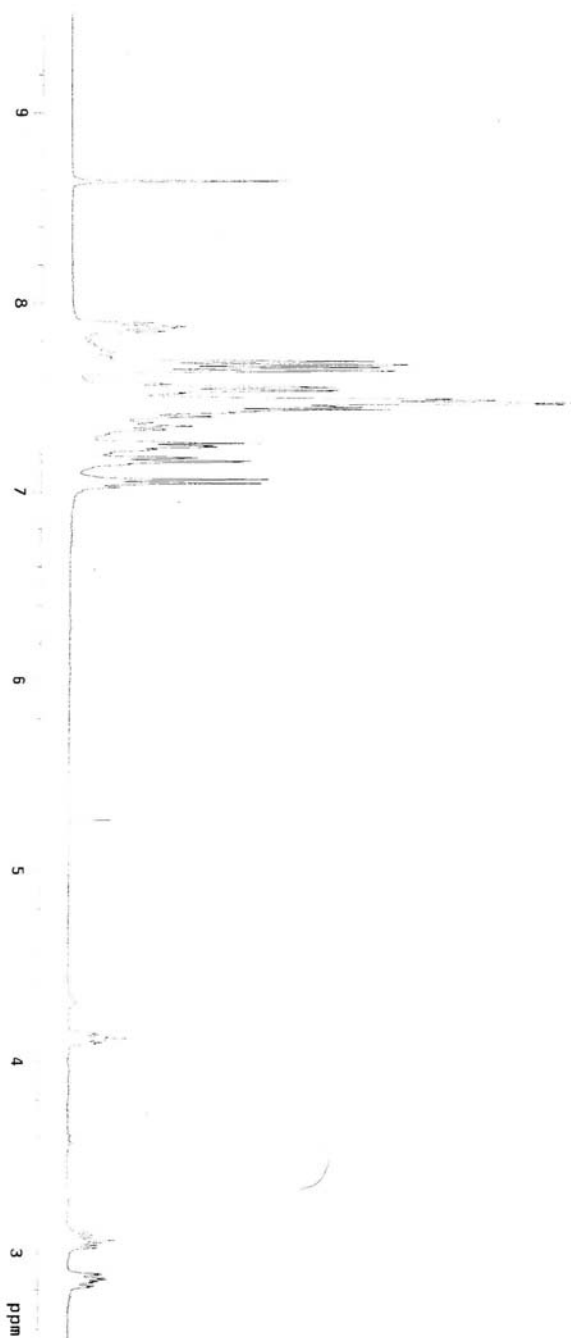
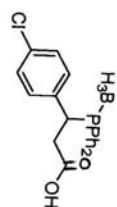
Pulse Sequence: szpu1
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
File: jv07_20_3p
Mercury-40058 "nmr4"
Pulse 27.6 degrees
Acq. time 0.600 sec
Sweep 130.0 kHz
1024 repetitions
OBSERVE P31, 161.8969264 MHz
DECOUPLE H1, 399.9376154 MHz
Power 42 dB
continuously on
SOLVENT DECOUPLED
DATA ACQUISITION
Line broadening 5.0 Hz
FT size 131072
Total time 56 min, 25 sec







1131209_flash
Pulse Sequence: zgpg30
Solvent: CDCl3
Temp: 25.0 C / 298.1 K
File: 1131209_flash_EtOAc
Mercury-400BB "nmr4"
Relax, delay 1.000 sec
Pulse 41.6 degrees
Pulse program: zgpg30
Width 5995.2 Hz
16 repetitions
OBSERVE H1, 399.9356500 MHz
DATA PROCESSING
F2 514.855000
Total time 1 min, 2 sec

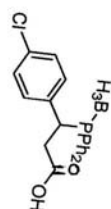


1131209_flash_13C

Pulse Sequence: zgpg30

Solvent: CDCl3
Temp: 298.1 K
File: 1131209_flash_13C
Mercury-4000B "nmr1"

Relax. delay 1.000 sec
Pulse 48.9 degrees
Acq. time 1.159 sec
Sweep 135.1 kHz
672 repetitions
OBSERVE C13, 100.626785 MHz
DECOUPLE H1, 399.9376154 MHz
Power 42 dB
CONTINUOUS ON
WALTZ16 MODULATED
DATA PROCESSING
Line broadening 1.0 Hz
FT size 65536
Total time 13 hr, 42 min, 9 sec



200 180 160 140 120 100 80 60 40 20 ppm

