



Supporting Information

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An Efficient CuO Nanoparticle Catalyzed C-S Cross Coupling of Thiols with Iodobenzene

L. Rout, T. K. Sen and T. Punniyamurthy*

Materials and Methods

Thiols, aryl halides and CuO nanoparticle (particle size 33 nm, surface area 29 m²/g and purity 99.99%) were purchased from Aldrich. NMR spectra (400 MHz for ¹H and 100 MHz for ¹³C) were recorded using DRX-400 Varian spectrometer using CDCl₃ as solvent and Me₄Si as internal standard. Flash column chromatography was performed on silica gel (230-400 mesh) using ethyl acetate and hexane as eluent. Melting points were determined on Buchi B-540 and uncorrected. Centrifugation was carried out using SIGMA 3K30 Sartorius for 1 h with 10000 rpm.

General Procedure for C-S Coupling Reaction

Thiol (1 mmol), aryl halide (1.1 mmol), CuO nanoparticle (1.26 mol%) and KOH (1.5 mmol) were stirred in DMSO (1 mL) at 80 °C under N₂ atmosphere for the appropriate time. The progress of the reaction was monitored by TLC using ethyl acetate and hexane as eluent. After completion, the reaction mixture was treated with ethyl acetate (10 mL) and water (3 mL). The organic layer was separated and the aqueous layer was extracted with ethyl acetate (3 x 5 mL). The combined organic solution was washed with brine (3 x 5 mL) and water (1 x 5 mL). Drying (Na₂SO₄) and evaporation of the solvent provided a residue which was purified on short pad of silica gel using ethyl acetate and hexane.

4-Tolylphenyl sulfide (Table 4, entry 2).^[1]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.30-7.25 (m, 6H), 7.20-7.12 (m, 3H), 2.31 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.80, 137.31, 132.48, 131.43, 130.26, 129.94, 129.24, 126.59, 21.34.

p-Anisyl phenyl sulfide (Table 4, entry 3).^[1]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.39-7.37 (dd, *J* = 8.8, 2.8 Hz, 2H), 7.23-7.12 (m, 5H), 6.88-6.85 (dd, *J* = 8.8, 2.4 Hz, 2H), 3.78 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 159.99, 138.79, 135.56, 129.10, 128.32, 125.91, 124.41, 115.15, 55.52.

4-Nitrophenyl phenyl sulfide (Table 4, entry 4).^[2]

Yellow solid; m.p. 54 °C; ¹H NMR (CDCl₃, 400 MHz) δ 8.00 (d, *J* = 8.8 Hz, 2H), 7.31-7.23 (m, 3H), 7.06 (d, *J* = 8.8 Hz, 2H), 6.72 (d, *J* = 8.4 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 158.94, 142.89, 136.24, 131.33, 125.91, 124.84, 115.15.

4-Bromophenyl phenyl sulfide (Table 4, entry 5).^[3]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.41 -7.39 (dd, *J* = 6.8, 4.8 Hz, 2H), 7.36 -7.25 (m, 5H), 7.18 -7.16 (dd, *J* = 6.8, 4.8 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 135.99, 132.74, 132.59, 132.06, 129.89, 128.08.

2-Bromophenyl phenyl sulfide (Table 4, entry 6).^[4]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.57 -7.55 (d, *J* = 8 Hz, 1H), 7.49 -7.44 (m, 2H), 7.41-7.36 (m, 3H), 7.24 -7.23 (m, 1H), 7.16-7.12 (m, 1H), 7.05-7.01 (m, 1H), 6.92-6.90 (d, *J* = 8 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ 133.74, 133.23, 129.91, 129.86, 128.95, 128.70, 128.02, 127.89, 127.44.

2-Naphthylphenyl sulfide (Table 4, entry 7).^[3]

Colorless solid, m.p. 49 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.83-7.76 (m, 2H), 7.74-7.70 (m, 2H), 7.48 -7.44 (m, 2H), 7.41-7.36 (m, 2H), 7.32 -7.23 (m, 4H); ¹³C NMR (CDCl₃, 100 MHz) δ 131.14, 130.08, 129.44, 129.07, 128.95, 127.94, 127.62, 127.27, 126.80, 126.42.

Benzylphenyl sulfide (Table 4, entry 8).^[5]

White solid, m.p. 40°C; ¹H NMR (CDCl₃, 400 MHz) δ 7.31 -7.23 (m, 10H), 4.12 (s, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ 130.08, 129.05, 128.70, 127.38, 126.56, 39.28.

(*n*-Ethylthio)benzene (Table 5, entry 1).^[6]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.33 -7.25 (m, 4H), 7.17-7.15 (m, 1H), 2.94 (m, 2H), 1.30 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 136.79, 129.16, 129.0, 127.66, 127.32, 125.92, 27.78, 14.54.

(*n*-Butylthio)benzene (Table 5, entry 2).^[7]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.32 -7.24 (m, 4H), 7.16-7.14 (m, 1H), 2.88 (t, *J* = 7.6 Hz, 2H), 1.64-1.59 (m, 2H), 1.47-1.41 (m, 2H), 0.90 (t, *J* = 2.7 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.55, 137.18, 130.32, 128.90, 127.53, 125.67, 33.28, 31.31, 22.08, 13.78.

(*n*-Hexylthio)benzene (Table 5, entry 3).^[8]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.31 -7.24 (m, 4H), 7.14-7.13 (m, 1H), 2.89 (t, *J* = 7.6 Hz, 2H); 1.65-1.59 (m, 2H), 1.42-1.39 (m, 2H), 1.37-1.25 (m, 2H), 0.85 (t, *J* = 6.8 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.24, 128.93, 125.72, 33.68, 31.53, 29.26, 28.69, 22.70, 14.19.

(*n*-Octylthio)benzene (Table 5, entry 4).^[9]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.32 -7.25 (m, 4H), 7.17-7.15 (m, 1H), 2.91 (t, *J* = 7.6 Hz, 2H), 1.66-1.56 (m, 2H), 1.43-1.26 (m, 10H), 0.87 (t, *J* = 6.4 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ 137.63, 137.24, 130.39, 128.96, 125.74, 33.69, 31.98, 29.35, 29.31, 29.03, 22.83, 14.29.

(*n*-Decylthio)benzene (Table 5, entry 5).^[10]

Colorless oil, ¹H NMR (CDCl₃, 400 MHz) δ 7.32 -7.59 (m, 4H), 7.17-7.15 (m, 1H), 2.91 (t, *J* = 7.6 Hz, 2H), 1.66-1.60 (m, 4H), 1.42-1.37 (m, 12H), 0.88 (t, *J* = 6 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz) 137.43, 129.01, 125.81, 33.76, 32.12, 29.84, 29.55, 29.37, 29.06, 22.90, 14.33.

(*n*-Dodecylthio)benzene (Table 5, entry 6).^[11]

Colorless oil; ¹H NMR (CDCl₃, 400 MHz) δ 7.33 -7.26 (m, 4H), 7.25-7.13 (m, 1H), 2.91(t, *J* = 7.6 Hz, 2H), 1.66-1.56 (m, 4H), 1.43-1.25 (m, 12H), 0.88-0.86 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 137.43, 129.01, 125.80, 33.76, 32.12, 29.83, 29.55, 29.45, 29.37, 29.05, 28.74, 26.62, 23.73, 22.90, 14.32.

1,4-Diphenyl butanethiol (Scheme 2).^[12]

Colorless solid, m.p. 84 °C; ¹H NMR (CDCl₃, 400 MHz) δ 7.29 -7.23 (m, 8H), 7.16 -7.14 (m, 2H), 2.90-2.87 (m, 4H), 1.76-1.73 (m, 4H); ¹³C NMR (CDCl₃, 100 MHz) 137.63, 129.44, 129.10, 33.47, 28.32.

References

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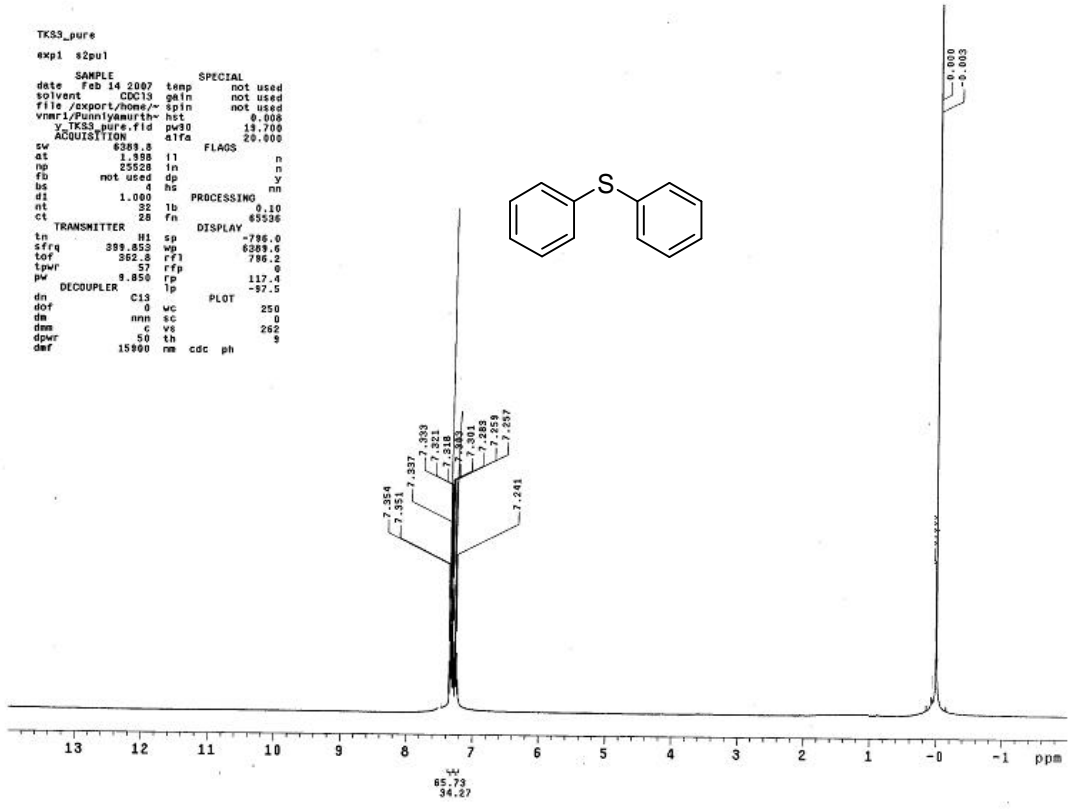
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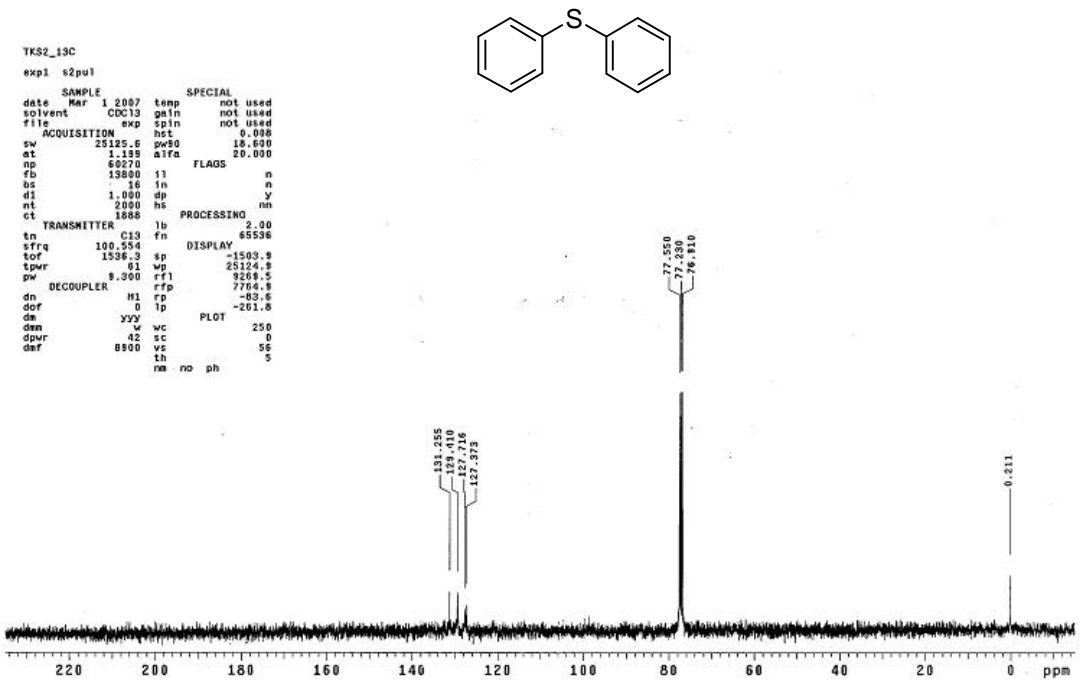
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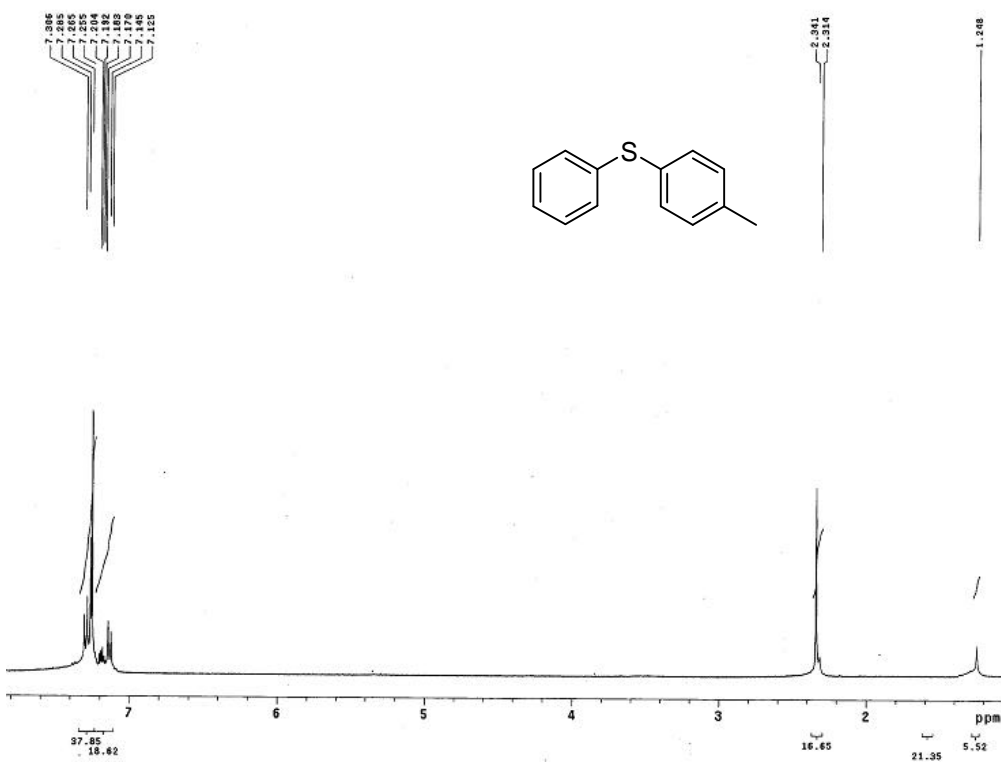
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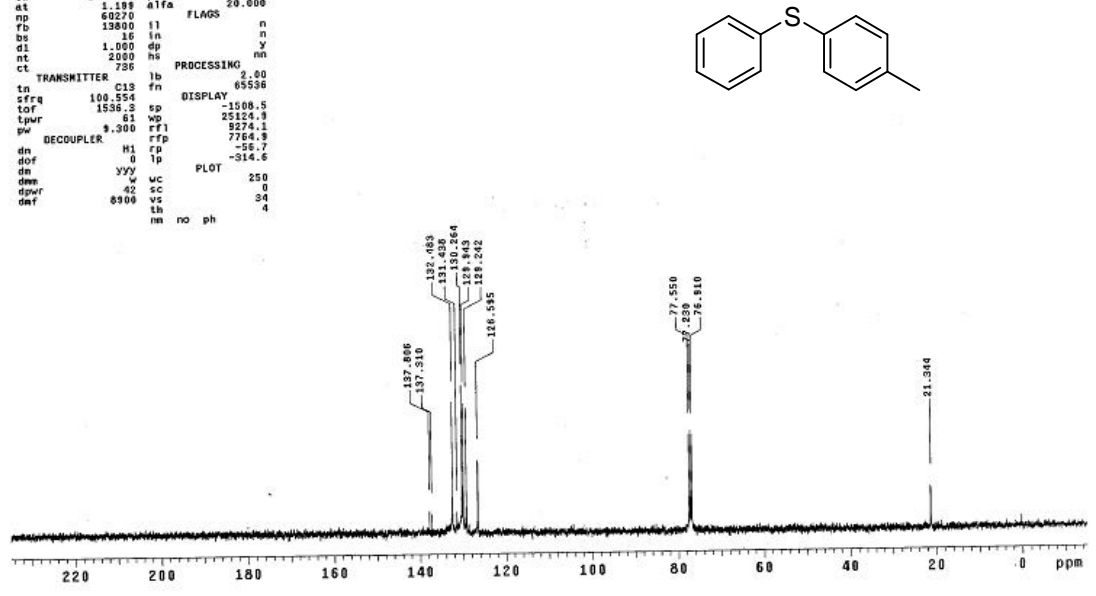
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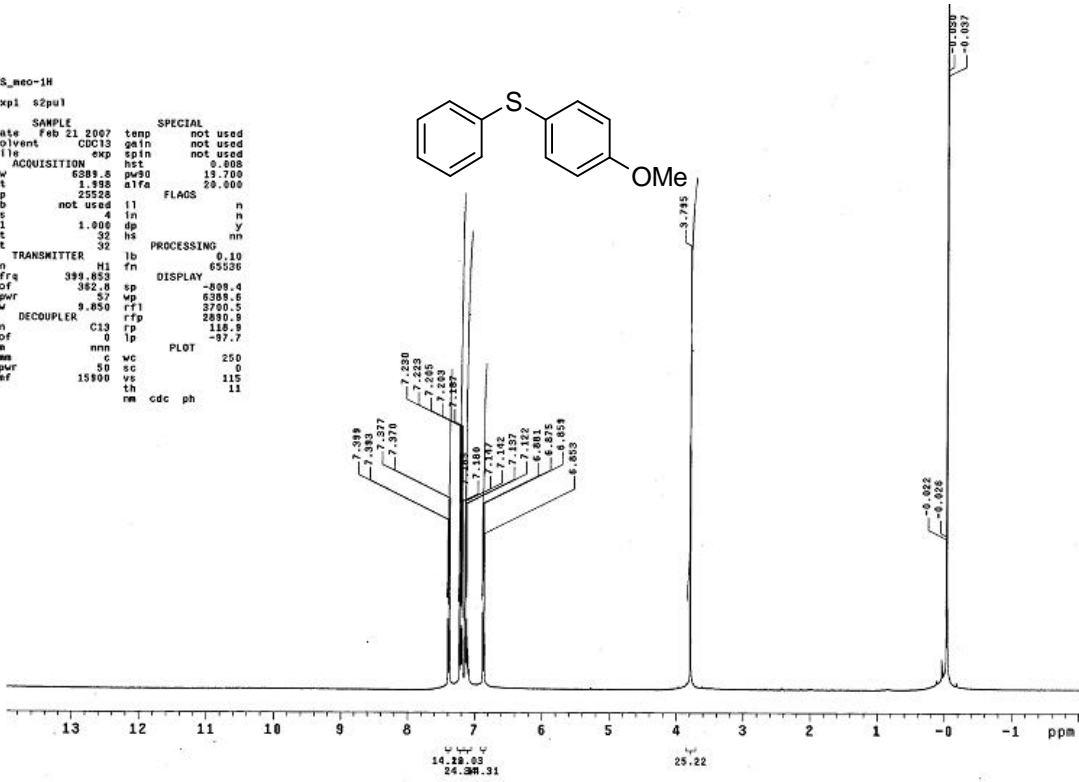
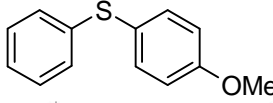


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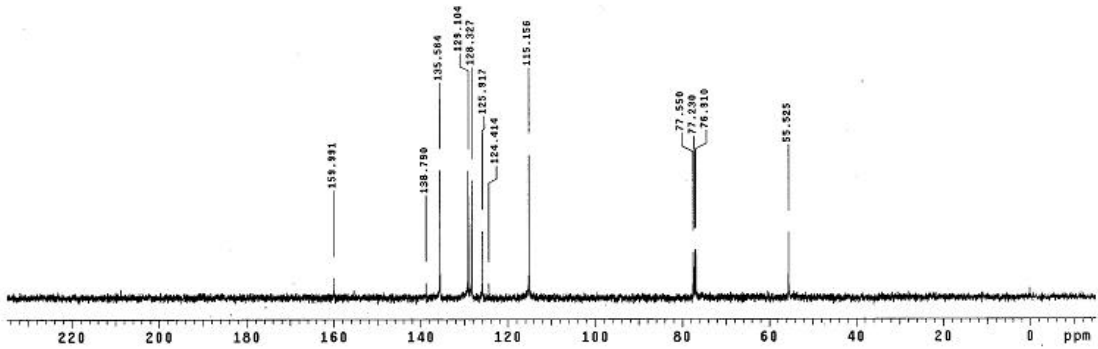
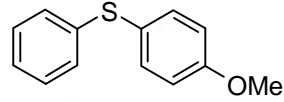


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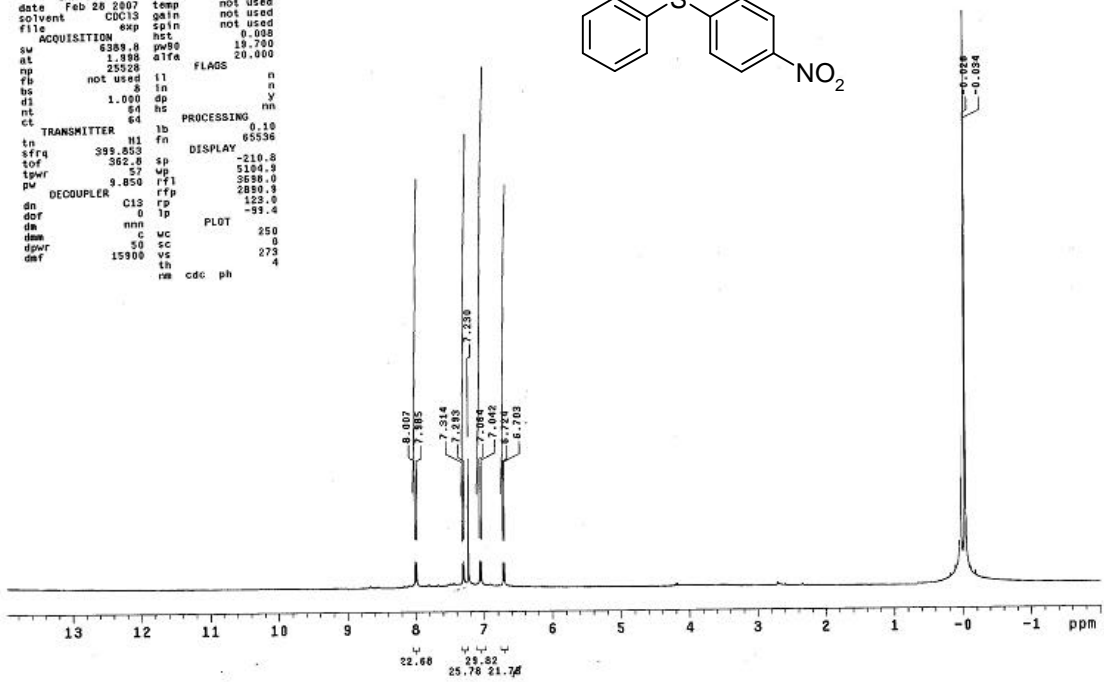
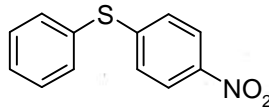
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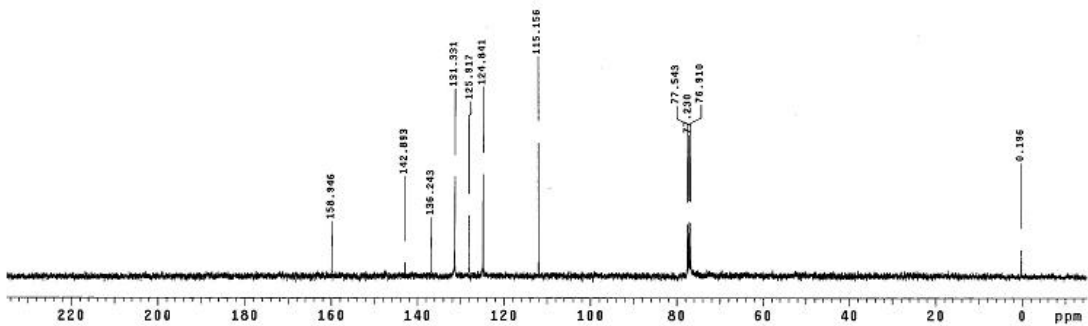
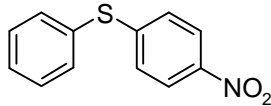
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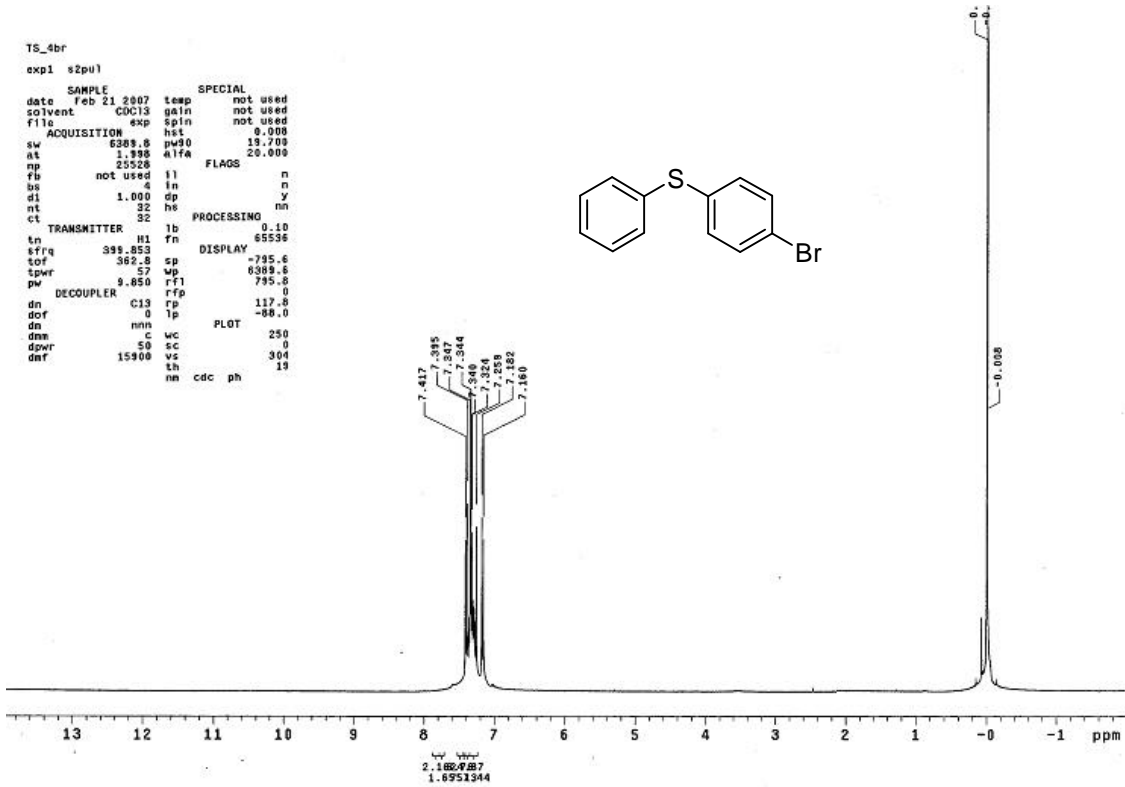
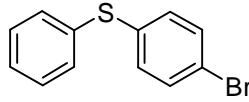


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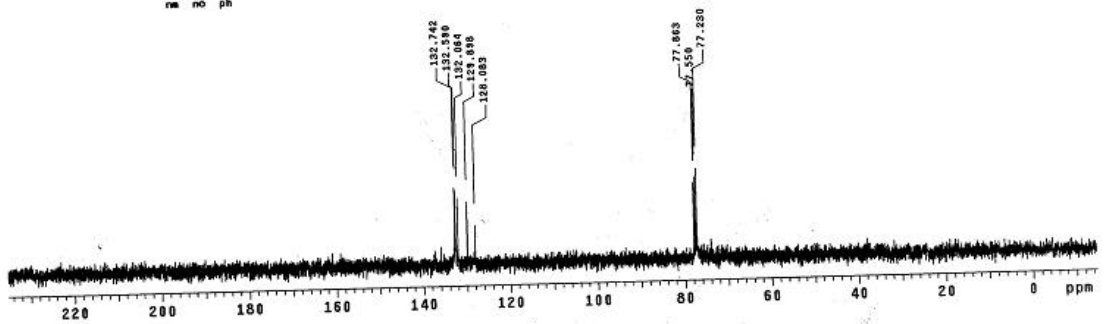
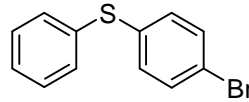


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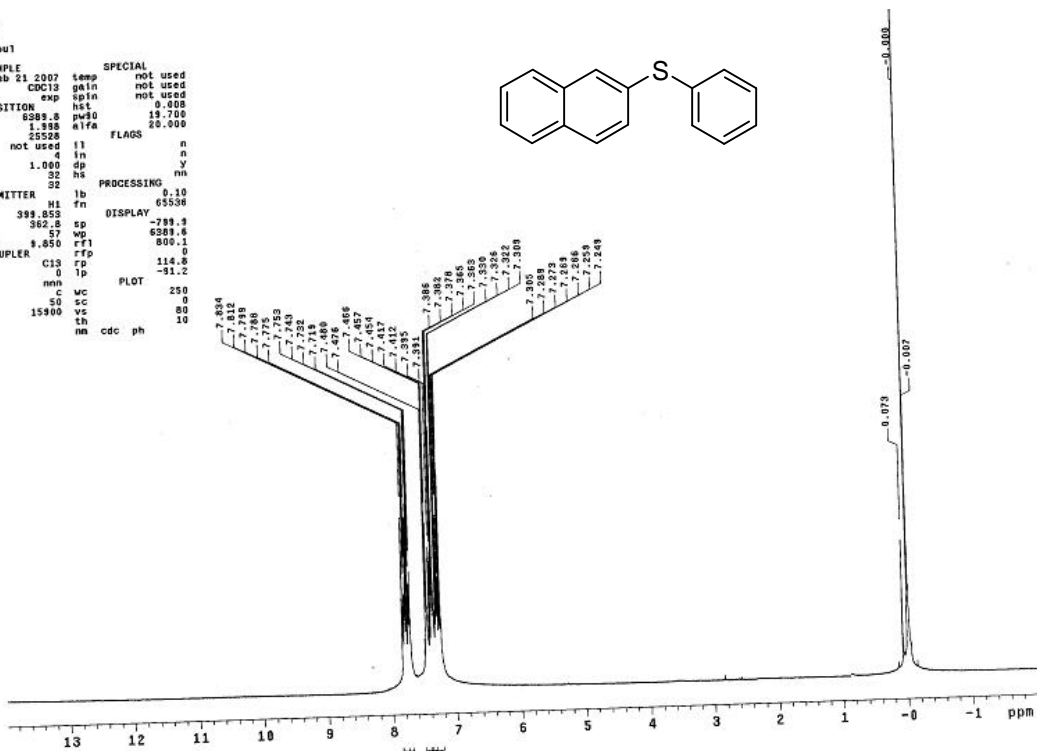
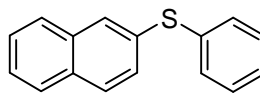
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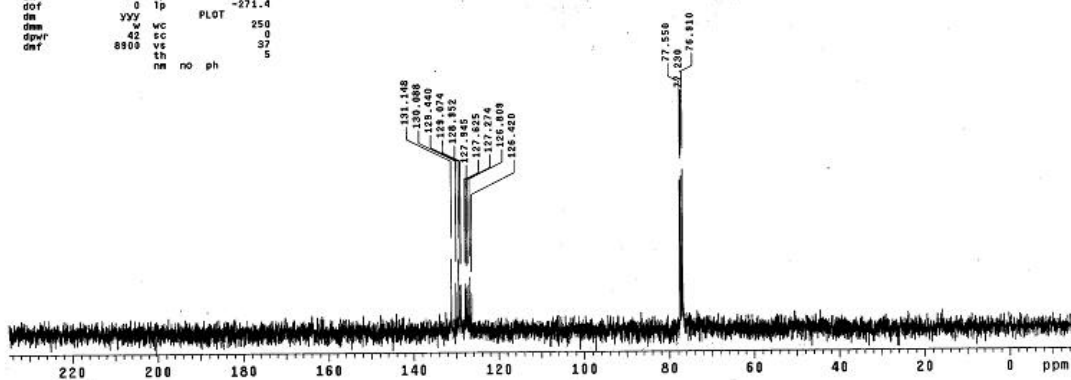
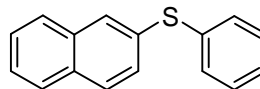
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 exp1 s2pu1

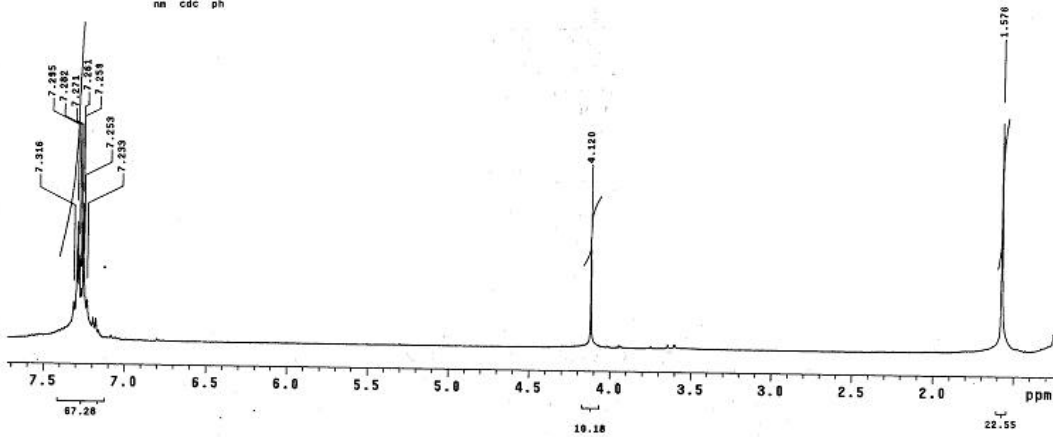
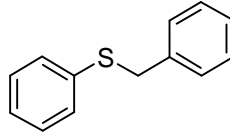
date	Feb 21 2007	temp	SPECIAL	not used
solvent	CDCl3	gain	not used	
file		spin	not used	
ACQUISITION	exp	hst	0.008	
sw	25125.6	pw90	18.800	
at	1.199	alfa	20.000	
nt	80270	FLAGS		
fb	13800	l1	n	
bs	32	ln	n	
d1	1.000	dp	y	
nt	2000	hs	nm	
ct		PROCESSING	2.00	
tn	TRANSMITTER	l1	fn	65536
sfrq	100.624	sp	DISPLAY	-1506.2
tof	1536.3	wp	25124.8	
tpwr	61	rf1	9271.8	
pw	8.300	rfo	7764.8	
dn	DECOUPLER	rp	-85.5	
dof	H1	lp	-271.4	
dm	yyy	WC	250	
dwm	42	SC	0	
dpwr	8900	VS	97	
dmf		TH	5	
		NM	no ph	



```

TS2_Pure
exp1 s2pu1
SAMPLE
date Feb 13 2007 temp not used
solvent CDCl3 gain not used
file
ACQUISITION exp spin not used
sv 6383.8 hst 0.008
et 1.998 pw90 19.700
ns 25520 alpha 20.000
fb not used 11 FLAGS n
bs 4 in n
dt 1.000 dp y
nt 32 hs nn
ct
TRANSMITTER 20 lb PROCESSING 0.10
tn H1 fn 65536
sfrq 399.853 DISPLAY -785.2
tof 382.8 sp -6389.6
tpwr 57 wp 785.4
pw 9.856 rfp 0
DECOUPLER C13 rp 119.1
dn 0 lp -78.5
dm nnn wc PLOT 250
dpr 50 sc 0
dnt 15300 vs 130
dmf nm cdc ph 20

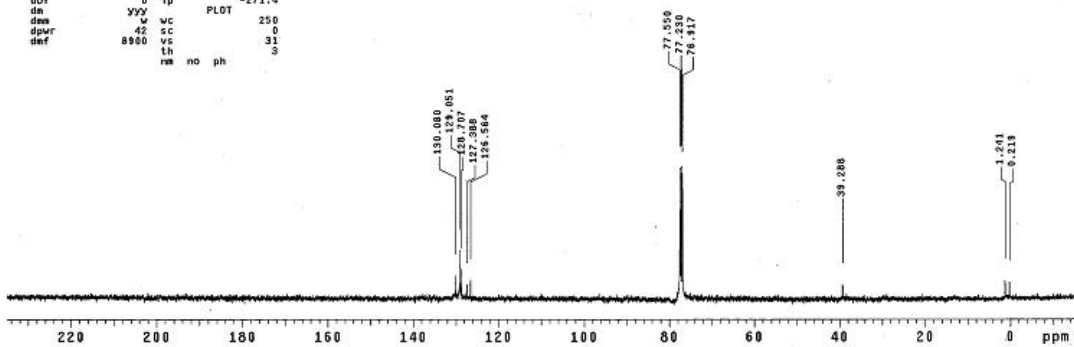
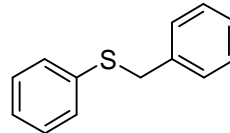
```



```

TS2_pure_13C
exp1 s2pu1
SAMPLE
date Feb 13 2007 temp not used
solvent CDCl3 gain not used
file
ACQUISITION exp spin not used
sv 25125.6 pw90 18.600
et 1.199 alpha 20.000
ns 68270 FLAGS n
fb 13800 11 n
bs 16 in n
dt 1.000 dp y
nt 3080 hs nn
ct
TRANSMITTER 3080 lb PROCESSING 2.00
tn C13 fn 65536
sfrq 100.536 DISPLAY -1502.4
tof 1536.3 sp -25124.9
tpwr 61 wp 1500.1
pw 9.300 rfp 0
DECOUPLER H1 rp -78.1
dn 0 lp -271.4
dm yyy wc PLOT 250
dpr 42 sc 0
dnt 8900 vs 31
dmf nm no ph 5

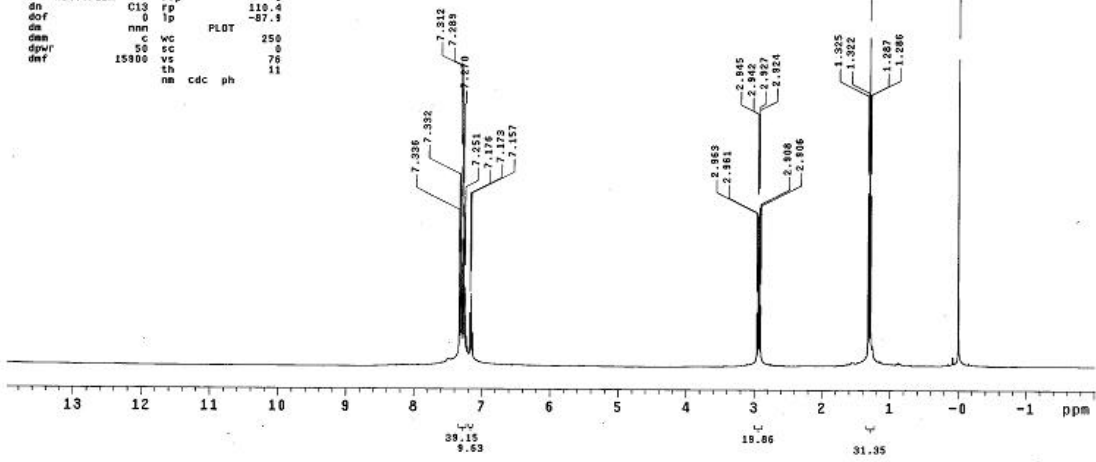
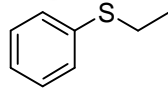
```



```

1t-phenylethyl
expl s2pu1
SAMPLE
date Feb 27 2007 temp not used
solvent CDC13 gain not used
file exp spin not used
ACQUISITION hst 0.008
sv 5389.8 pw90 15.700
at 1.988 a1fa 20.000
np 25528 FLAGS
fb not used i1 n
bs 4 in n
d1 1.000 dp y
nt 32 hs mn
ct 32
TRANSMITTER i1 b1 PROCESSING
tn HI F1 9.10
sfrq 399.853 DISPLAY 85538
tof 362.8 sp -805.2
tpwr 57 wp 8309.6
pw 9.850 rfl 805.4
DECOUPLER rfp 0
dn C13 rp 110.4
dof 0 lp -87.9
dm c wc PLOT 250
dpr 50 sc 8
dmr 15300 vc 76
th 11
nm cdc ph

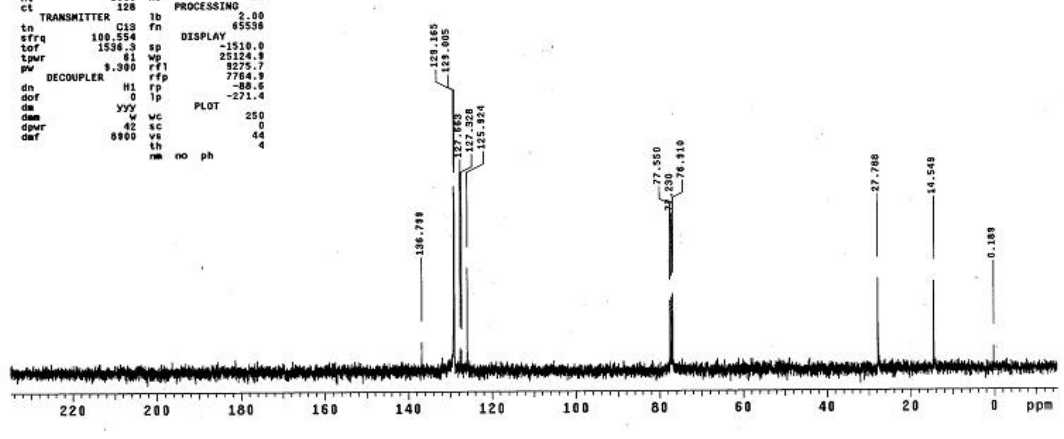
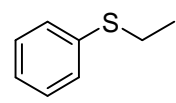
```



```

1t-phenylethyl
expl s2pu1
SAMPLE
date Feb 27 2007 temp not used
solvent CDC13 gain not used
file exp spin not used
ACQUISITION hst 0.008
sv 25125.6 pw90 18.600
at 1.199 a1fa 20.000
np 60279 FLAGS
fb 13000 i1 n
bs 18 in n
d1 1.000 dp y
nt 1000 hs mn
ct 128
TRANSMITTER i1 b1 PROCESSING
tn C13 F1 2.00
sfrq 100.554 DISPLAY 85538
tof 1536.3 sp -1510.0
tpwr 81 wp 28124.9
pw 9.300 rfl 9275.7
DECOUPLER H1 rfp 2764.9
dn v w wc PLOT 250
dof y y 0 lp -271.4
dm v wc 0
dpr 42 sc 44
dmr 8900 vc 76
th 4
nm no ph

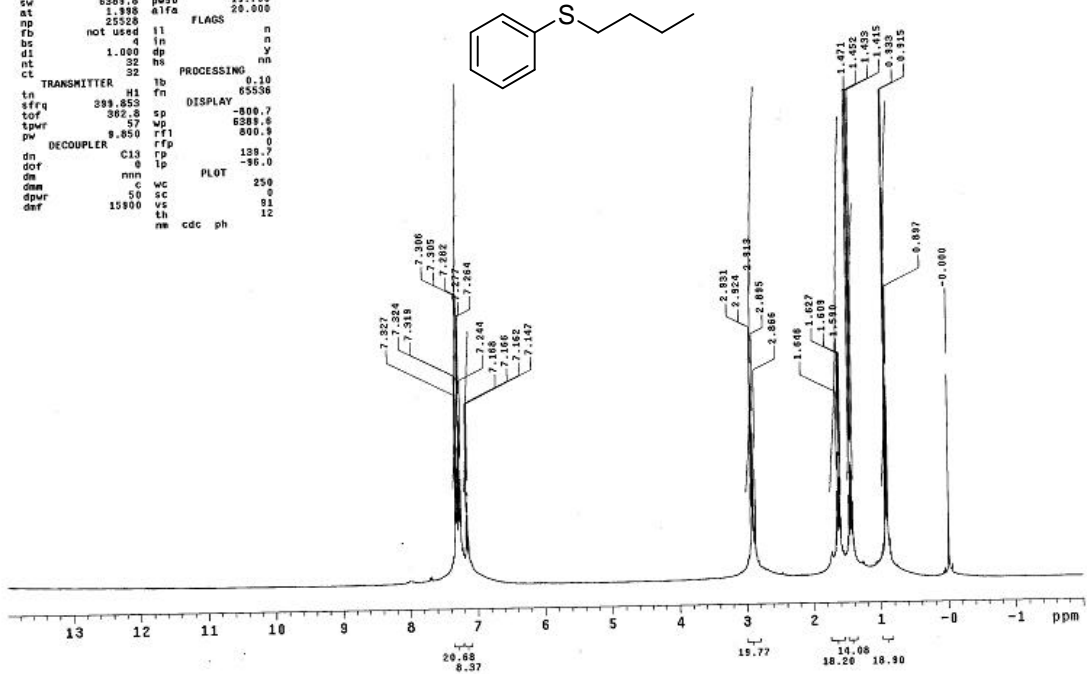
```



TL-butyl-1M

expl s2pu1

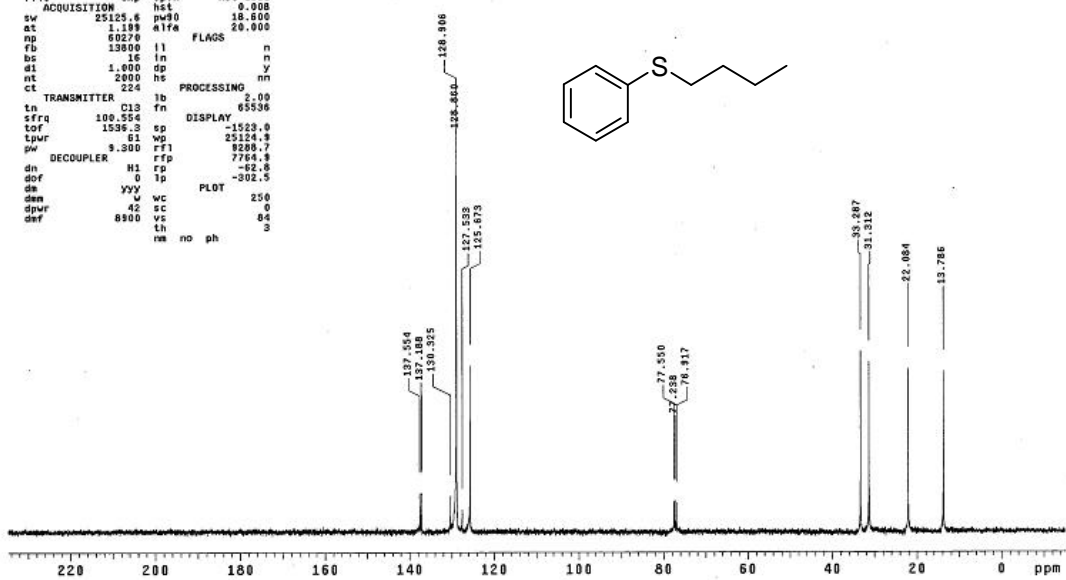
date	Feb 22 2007	temp	not used
solvent	CDCl3	gain	not used
file	exp	spin	not used
ACQUISITION	exp	hst	0.000
sw	6381.8	pu30	19.700
at	1.998	alfa	20.000
np	25528	FLAGS	
fb	not used	l1	n
bc	not used	l2	n
d1	1.000	dp	nn
nt	32	hs	
ct	32	PROCESSING	0.10
tn	TRANSMITTER	H1	fn
sfrq	399.853	sp	65536
tof	362.3	wp	-800.7
tpwr	5.7	rf1	6389.6
pw	9.850	rf2	800.9
DECOUPLER	C13	rp	138.7
dn	0	lp	-96.0
dof	0	nm	
dma	0	wc	250
dpwr	50	sc	0
def	15900	vs	91
		th	12
		ns	cdc ph



TS-butyl-13C

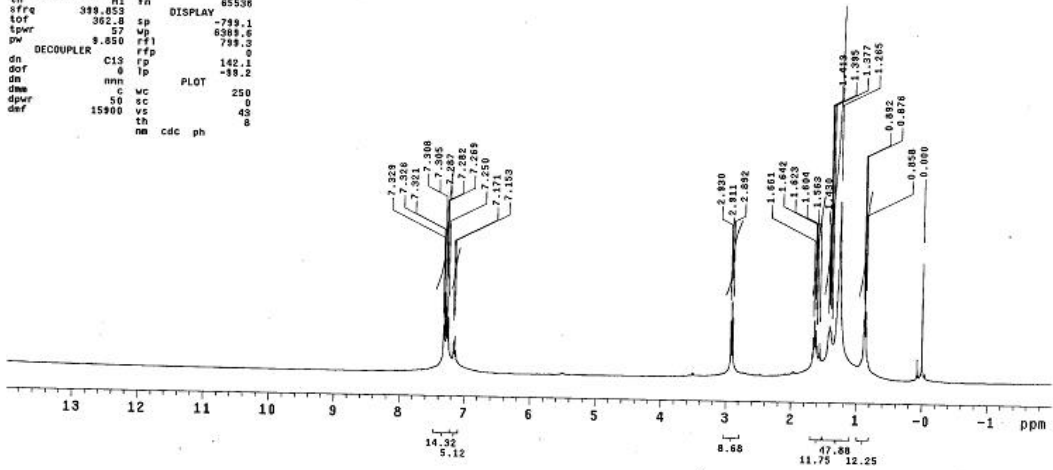
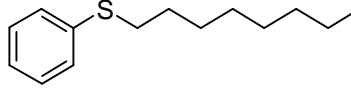
expl s2pu1

date	Feb 21 2007	temp	not used
solvent	CDCl3	gain	not used
file	exp	spin	not used
ACQUISITION	exp	hst	0.000
sw	25125.6	pu30	18.800
at	1.199	alfa	20.000
np	60270	FLAGS	
fb	13800	l1	n
bc	not used	l2	n
d1	1.000	dp	nn
nt	2000	hs	
ct	224	PROCESSING	2.00
tn	TRANSMITTER	H1	fn
sfrq	100.554	sp	65536
tof	1536.3	wp	-1523.0
tpwr	61	rf1	25124.3
pw	9.300	rf2	8288.7
DECOUPLER	C13	rp	7764.3
dn	0	lp	-92.8
dof	0	nm	-202.5
dma	yy	wc	250
dpwr	42	sc	0
def	8900	vs	84
		th	3
		ns	no ph



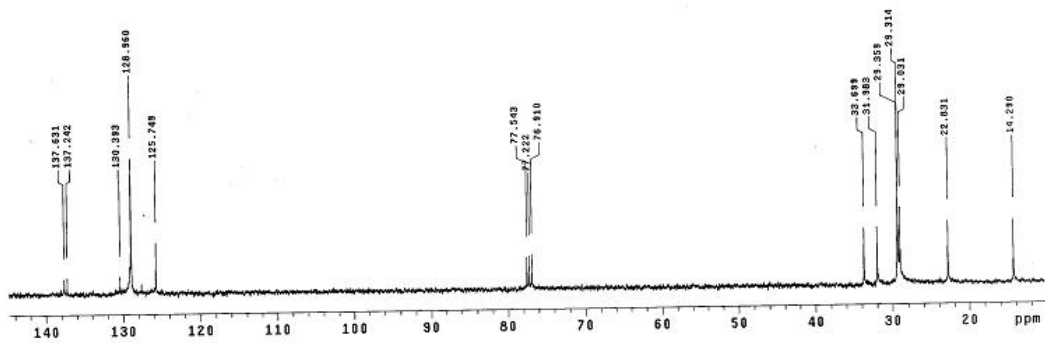
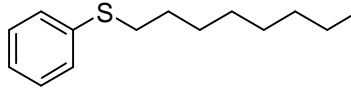
TL-octyl-1H
 exp1 s2pu1

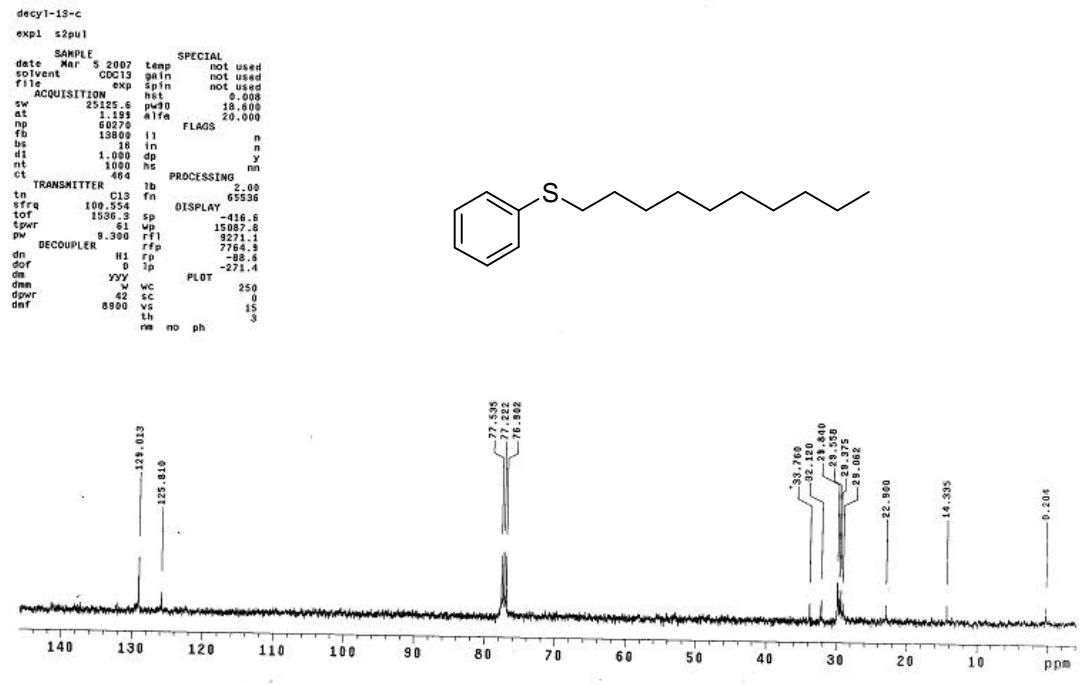
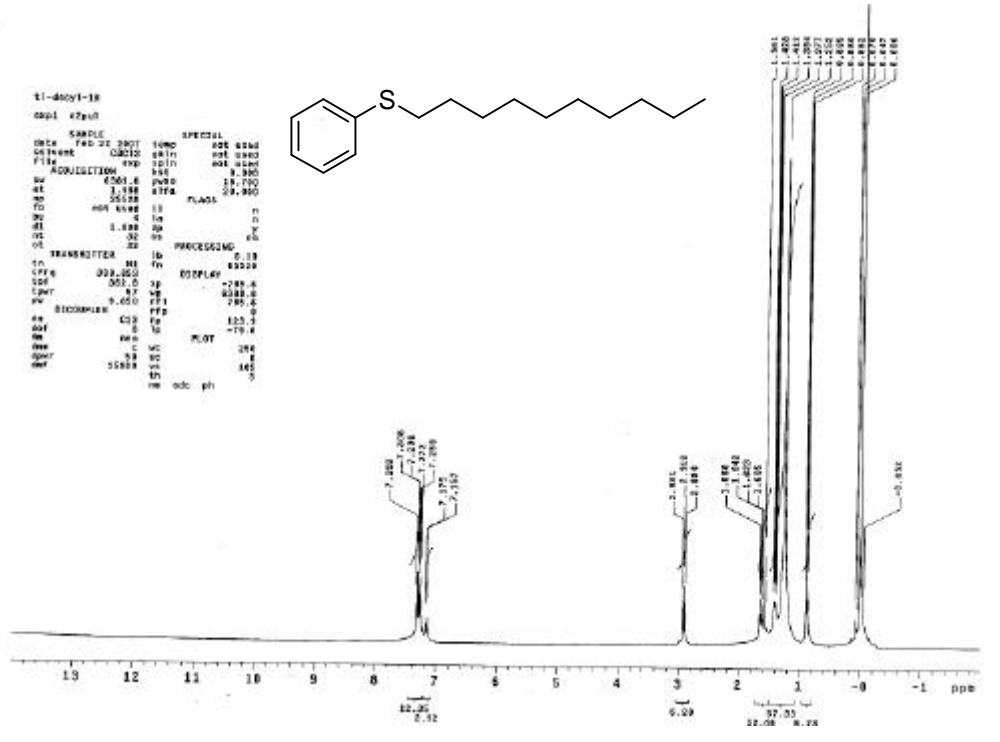
date	Feb 22 2007	temp	not used
solvent	CDCl3	gain	not used
file	exp	spin	not used
ACQUISITION	exp	hct	0.008
sw	6389.6	pu90	19.708
at	1.398	a1fa	20.000
np	25528	FLAGS	n
fb	not used	11	n
bc	4	in	n
d1	1.000	dp	y
nt	32	hs	n
ct	32	PROCESSING	na
tn		tb	0.10
sfrq	399.853	H1	fn
tof	362.0	sp	DISPLAY
tpwr	57	wp	-749.1
pw	9.850	rf1	6389.6
DECOUPLER	C13	rfp	739.3
dn	0	lp	142.1
dof	0	ip	-99.2
dm	nnh	wc	PLOT
dma	c	wc	250
dpwr	50	sc	0
def	15900	vs	43
		th	8
		nm	cdc ph



TS-octyl-13C
 exp1 s2pu1

date	Feb 22 2007	temp	not used
solvent	CDCl3	gain	not used
file	exp	spin	not used
ACQUISITION	exp	hct	0.008
sw	25125.6	pu90	18.000
at	1.199	a1fa	20.000
np	60270	FLAGS	n
fb	15800	11	n
bc	16	in	n
d1	1.000	dp	y
nt	2000	he	nn
ct	240	PROCESSING	2.00
tn		tb	fn
sfrq	100.554	DISPLAY	65538
tof	1536.3	sp	1026.5
tpwr	61	wp	13531.2
pw	9.300	rf1	3278.0
DECOUPLER	H1	rfp	7184.3
dn	0	lp	-75.8
dof	0	ip	-294.1
dm	vyy	wc	PLOT
dma	42	sc	250
dpwr	8900	vs	33
def		th	3
		nm	no ph

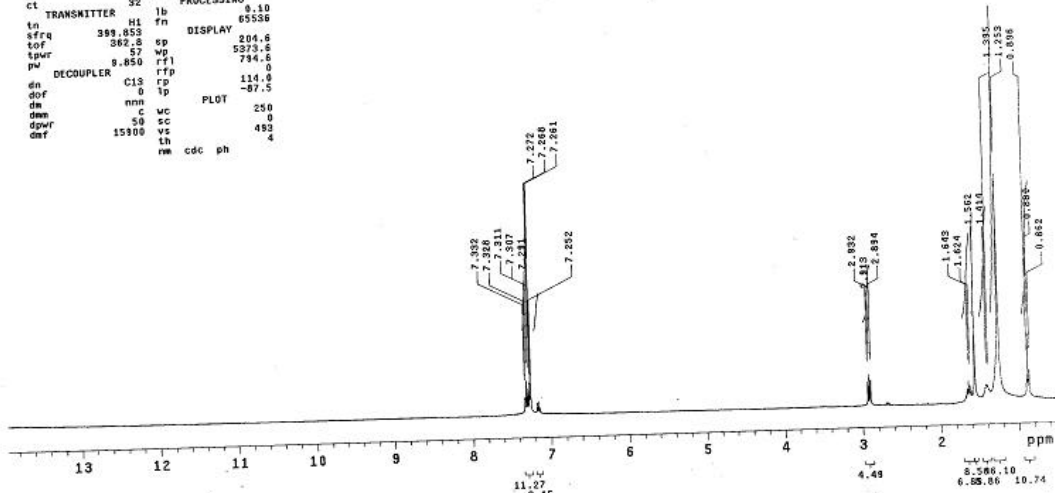
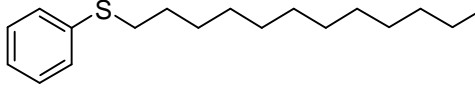




```

L_dodecyl
exp1 s2pul
SAMPLE
date Mar 5 2007 temp SPECIAL
solvent CDCl3 gain not used
file exp s2in not used
ACQUISITION hst 0.008
sw 6389.8 pu90 19.700
at 1.898 a1fa 20.000
np 25528 FLAGS
pb not used i1 n
bs 1.000 in n
dl 1.000 dp y
nt 32 hs PROCESSING
ct 32 mn
TRANSMITTER lb fn 0.10
tn H1 fn 65536
sfrq 399.853 DISPLAY
tof 362.8 sp 204.6
tpwr 57 wp 5370.6
pw 8.650 rfp 794.6
DECOUPLER C13 rfp 0
dn H1 rp 114.0
dof 0 tp -87.5
dm nmn c wc PLOT 250
dwm 50 sc 0
dpwr 15900 vs vs 483
dmf 15900 th th 4
nm cdc ph

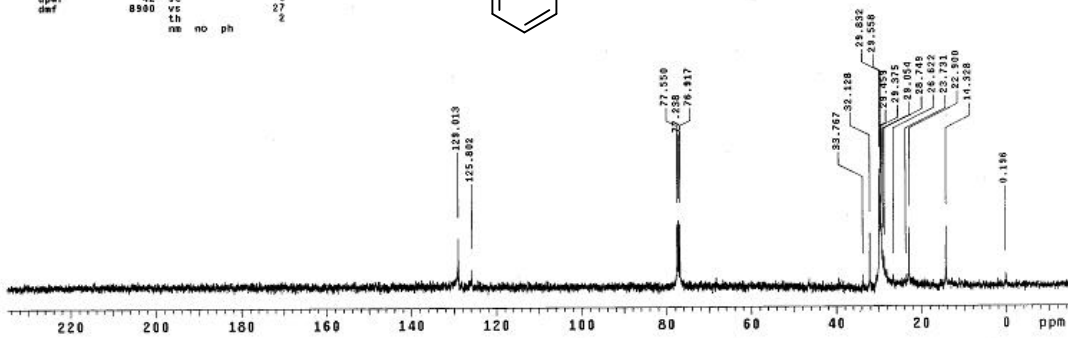
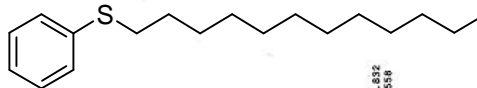
```



```

Dodecyl_TS
exp1 s2pul
SAMPLE
date Mar 7 2007 temp SPECIAL
solvent CDCl3 gain not used
file exp s2in not used
ACQUISITION hst 0.008
sw 25325.6 pu90 18.800
at 1.199 a1fa 20.000
np 80270 FLAGS
pb 13600 i1 n
bs 16 in n
dl 1.000 dp y
nt 1000 hs PROCESSING
ct 608 mn
TRANSMITTER lb fn 2.00
tn C13 fn 65536
sfrq 100.504 DISPLAY
tof 1536.3 sp -1505.4
tpwr 61 wp 25124.3
pw .300 rfp 8273.1
DECOUPLER H1 rfp 7764.3
dn H1 rp -84.8
dof 0 tp -271.4
dm yyy w wc PLOT 250
dwm 42 sc 0
dpwr 8900 vs vs 27
dmf 8900 th th 2
nm no ph

```

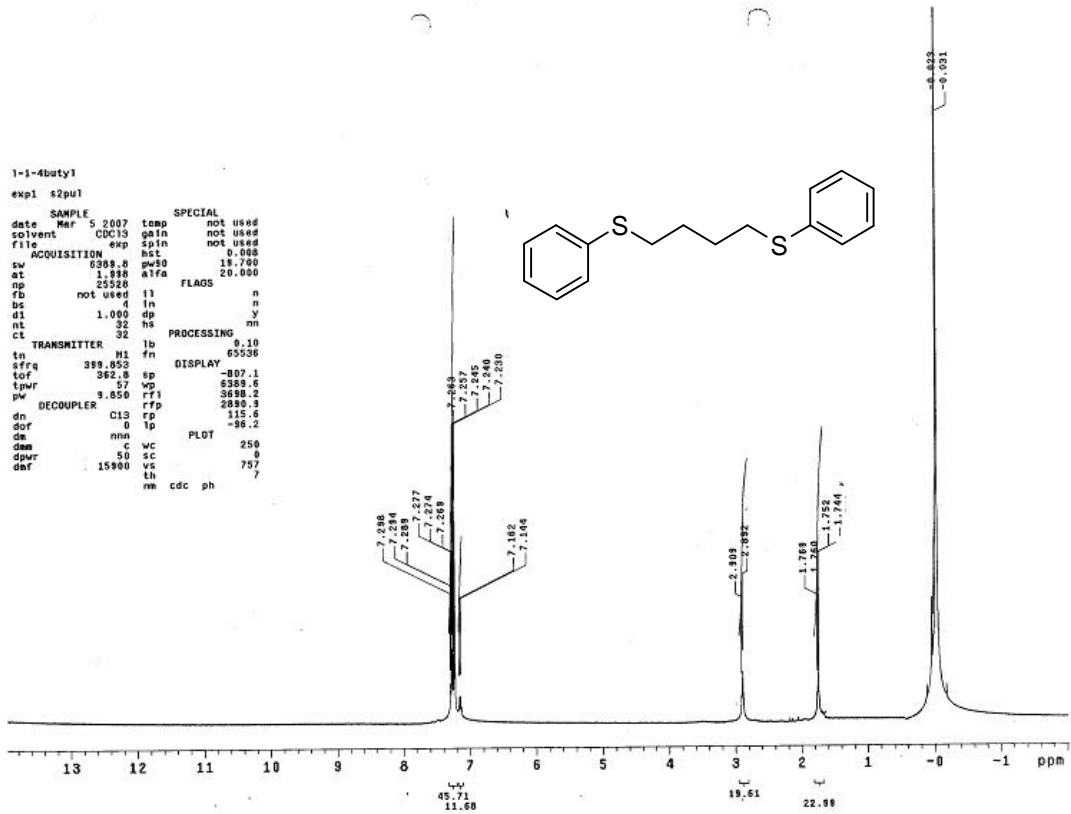


1-1-4butyl
exp1 s2pu1

```

SAMPLE          SPECIAL
date Mar 5 2007 temp not used
solvent CDC13 gain not used
file           exp spin not used
ACQUISITION    hst 0.000
sw 6389.8 pw50 18.700
at 1.898 atrs 20.000
np 25228
fd not used li n
ds 4 in n
d1 1.000 dp y
nt 32 hs nm
ct 32
TRANSMITTER    lb 0.10
tn n1 fn 65036
sfrs 389.853 DISPLAY
tof 362.8 sp -807.1
tpwr 57 wp 6389.6
pw 9.850 rfp 3698.2
DECOUPLER      C13 rp 2898.9
dr 0 lp -95.2
dof nna c wc PLOT 250
da 50 sc 0
dpmr 15900 vs 757
daf nm cdc ph 7

```



1-1-4butyl130
exp1 s2pu1

```

SAMPLE          SPECIAL
date Mar 5 2007 temp not used
solvent CDC13 gain not used
file           exp spin not used
ACQUISITION    hst 0.000
sw 6389.8 pw50 18.700
at 1.898 atrs 20.000
np 25228
fd not used li n
ds 4 in n
d1 1.000 dp y
nt 32 hs nm
ct 32
TRANSMITTER    lb 0.10
tn n1 fn 65036
sfrs 389.853 DISPLAY
tof 362.8 sp -807.1
tpwr 57 wp 6389.6
pw 9.850 rfp 3698.2
DECOUPLER      C13 rp 2898.9
dr 0 lp -95.2
dof nna c wc PLOT 250
da 50 sc 0
dpmr 15900 vs 757
daf nm cdc ph 7

```

