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Supporting Information for *Angew. Chem. Int. Ed. Z 17576*

## **Design And Synthesis Of a Peptide that Binds Specific DNA Sequences Through Simultaneous Interaction In The Major and the Minor Groove**

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## Supporting Information

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### General

Solvents used were purified using procedures described by Perrin,<sup>1</sup> and were dried under Ar by distillation over the corresponding drying agent immediately before their use. Reactions were performed under Ar unless specifically stated.

Peptide synthesis was made using standard Fmoc solid phase synthesis protocols. Using mixtures of HBTU/HOBt as coupling agents and DIEA as base in DMF. Cleavage/deprotection of peptides was performed by treatment of the peptide bound to the solid support for 2.5 h with the following mixture: 830  $\mu$ L TFA, 25  $\mu$ L EDT, 50 mg PhOH, 50  $\mu$ L tianisole and 50  $\mu$ L H<sub>2</sub>O (300  $\mu$ L of this mixture for each 10 mg of resin)

NMR spectrums were registered with *Brucker WM-250* and *Brucker AMX-300* spectrometers. HPLC analysis and purification were made using a *Hewlett-Packard Serial 1100* with a diode array detector. Column packing used was *Merck LiChropher WP 300 RP-18* (5  $\mu$ M) for analytical columns and *Nucleosil 120-10 C-18* for semipreparative columns. Mixtures of CH<sub>3</sub>CN and water with 0.1% TFA were used as mobile phase with linear gradients. Detection was made at 220, 260 and 304 nm simultaneously.

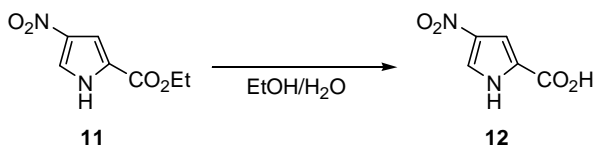
Qualitative molecular modelling was made using the *Builder* module of *InsightII 98.0* from MSI Molecular Simulations Inc. using a Silicon Graphics Octane with two parallel R10000 processors. Partial energy minimizations of the linkers were made using the *Discover* module of *Insight 98.0* using AMBER forcefield.<sup>2</sup> Graphical representations were made using program WebLabViewer Lite 3.7 from Molecular Simulations Inc. X-Ray structures used were directly downloaded from the Protein Data Bank server (<http://www.rcsb.org>).

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<sup>1</sup> Perrin, D.; Armarego, W. I. F. *Purification of Laboratory Chemicals*, 1998 Pergamon Press.

<sup>2</sup> Weiner, S. J.; Kollman, P. A.; Nguyen, D. T.; Case, D. A. *J. Comput. Chem.* **1986**, 7, 230.

### 4-nitro-1 H-pyrrole-2-carboxylic acid (**12**)



Pyrrole **11** (500 mg, 2.72 mmol) was treated with a 1M solution of NaOH in 70% aqueous EtOH (30 mL) at room temperature for 6 h. After, the resulting solution was carefully acidified with citric acid to pH=2 and was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried with MgSO<sub>4</sub>. Removal of the solvent under reduced pressure gave product **12** as a white solid. (360 mg, 85%). <sup>1</sup>H-NMR was consistent with previously reported data.<sup>3</sup>

**<sup>1</sup>H-NMR** d: 3.2 (s 1H), 7.2 (d, *J* = 1.7 Hz, 1H), 7.7 (d, *J* = 1.7 Hz, 1H).

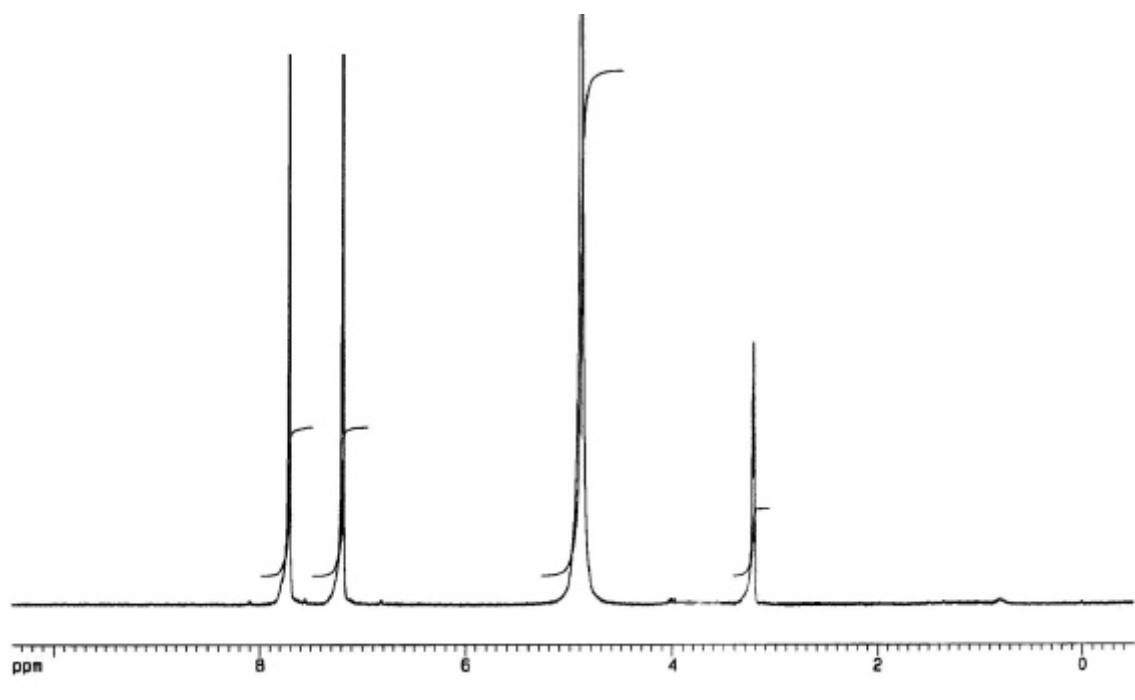
**MS:** m/z 156 (M<sup>+</sup>, 63), 138 (13), 122 (12), 108 (9), 92 (6), 65, (100).

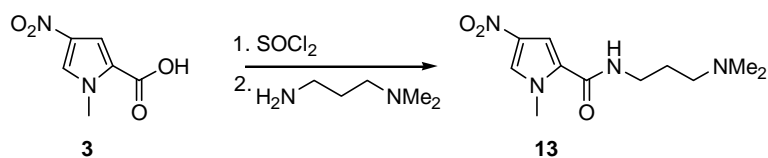
**HRMS:** calcd. for C<sub>5</sub>H<sub>4</sub>N<sub>2</sub>O<sub>4</sub> 156.01711, found 156.01718.

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<sup>3</sup> Lee, M.; Coulter, D. M.; Lown, J. W. *J. Org. Chem.* **1988**, *53*, 1855

## Supporting Information



***N*-[3-(dimethylamino)propyl]-1-methyl-4-nitro-1 *H*-pyrrole-2-carboxamide (**13**)**

1-methyl-4-nitro-1 *H*-pyrrole-2-carboxylic acid **3** (1.5 g, 8.8 mmol) was dissolved in SOCl<sub>2</sub> (6 mL) and the resulting suspension was refluxed under argon for 6 h. Solvent was removed under reduced pressure and the orange residue was redissolved in CH<sub>2</sub>Cl<sub>2</sub> which was again removed to ensure complete elimination of any residue of SOCl<sub>2</sub>. The resulting solid residue was dissolved in DMF (7 mL) and cooled to 0 °C. To this solution 3-dimethylaminopropylamine was added and the reaction mixture was allowed to stir at room temperature for 16 h. Water (20 mL) was added and the mixture was extracted with a mixture of EtOAc/hexane (3 x 20 mL). The combined organic layers were washed with aqueous NaHCO<sub>3</sub> (20 mL, 10%) and dried with MgSO<sub>4</sub>. Solvent was removed under reduced pressure. The product was purified by flash column chromatography to give **13**. (1.47 g, 67%) Spectroscopic data match reported data.<sup>4</sup>

**<sup>1</sup>H-NMR** d: 1.68-1.73 (m, 2H), 2.27 (s, 6H), 2.49 (t, *J* = 6.9, 2H), 3.46 (m, 2H), 4.01 (s, 3H), 6.9, 7.51 (2 s, 2H), 8.75 (s, 1H).

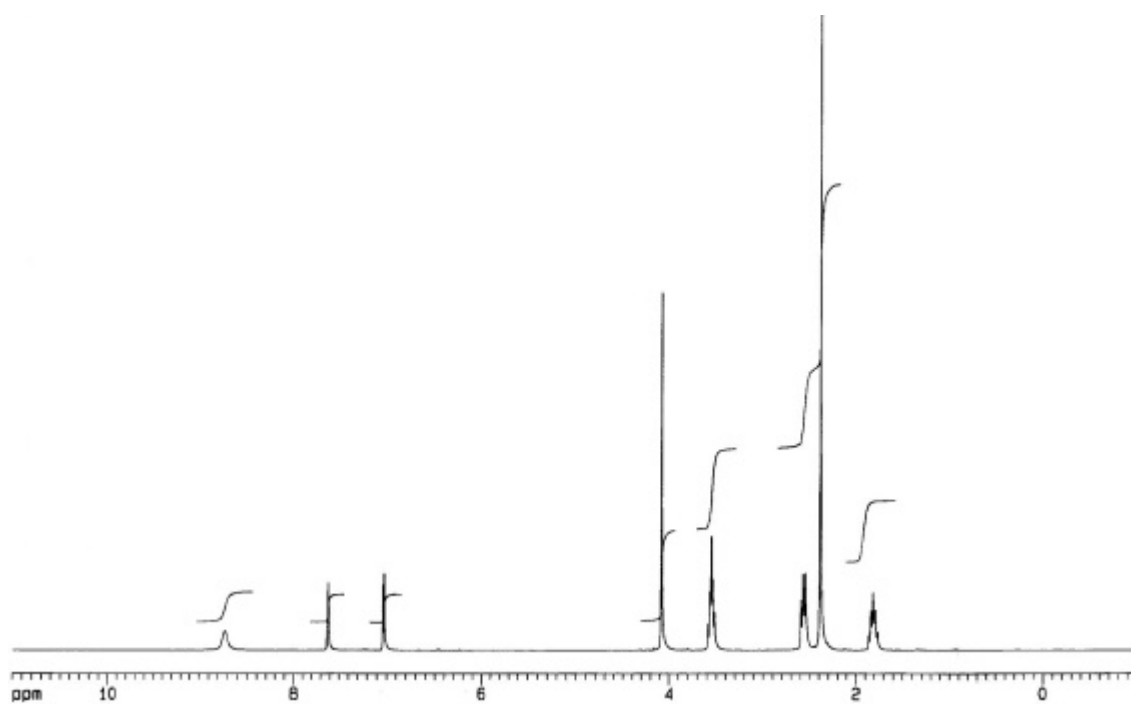
**MS:** *m/z* 254 (M<sup>+</sup>, 10), 181 (14), 153 (11), 131 (17), 58 (100).

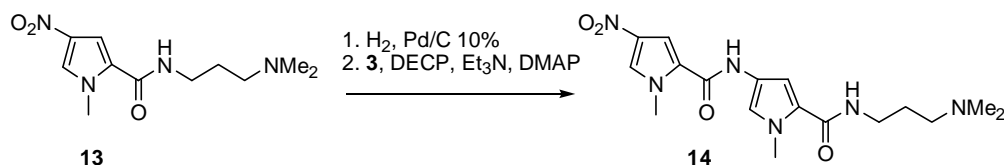
**HRMS:** calcd. for C<sub>11</sub>H<sub>18</sub>N<sub>4</sub>O<sub>3</sub> 254.13789, found 254.13997.

<sup>4</sup> Bruice, T. C.; Mei, H.; He, G.; Lopez, V. *Proc. Natl. Acad. Sci. USA* **1992**, *89*, 1700.

## Supporting Information

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***N*-[(dimethylamino)propyl]-1-methyl-4-[[[(1-methyl-4-nitro-1 *H*-pyrrole-2-yl)carbonyl]amino]-1 *H*-pyrrole-2-carboxamide (**14**)**

A solution of the pyrrole **13** (2.24 g, 8.84 mmol) in MeOH (100 mL) was hydrogenated over 10% palladium on charcoal (1.5 g) at room temperature for 1 h (balloon pressure). The catalyst was removed by filtration through celite, and the filtrate was concentrated under reduced pressure. The residue was dissolved in DMF (8 mL) and added over another solution previously prepared of 1-methyl-4-nitro-1 *H*-pyrrole-2-carboxylic acid **3** (1.66 g, 9.72 mmol), DECP (2 mL, 13.26 mmol), Et<sub>3</sub>N (6.16 mL, 44.2 mmol) and DMAP (10 mg) in DMF (12 mL) cooled at 0 °C. The resulting mixture was allowed to stir at room temperature for 12 h. MeOH (5 mL) was added and solvents were removed under reduced pressure, and the product was purified by flash column chromatography (1% Et<sub>3</sub>N, 10% MeOH, CH<sub>2</sub>Cl<sub>2</sub>) to afford **14** (1.4 g, 80%) as a yellowish solid whose spectroscopic data match previously reported.<sup>5</sup>

<sup>1</sup>H-NMR d: 1.6-1.7 (m, 2H), 2.27 (s, 6H), 2.42 (m, 2H), 3.46 (m, 2H), 3.82 (s, 3H), 4.01 (s, 4H), 6.53 (s, 1H), 7.17 (s, 1H), 7.39 (s, 1H), 7.54 (s, 1H), 9.06 (s, 1H).

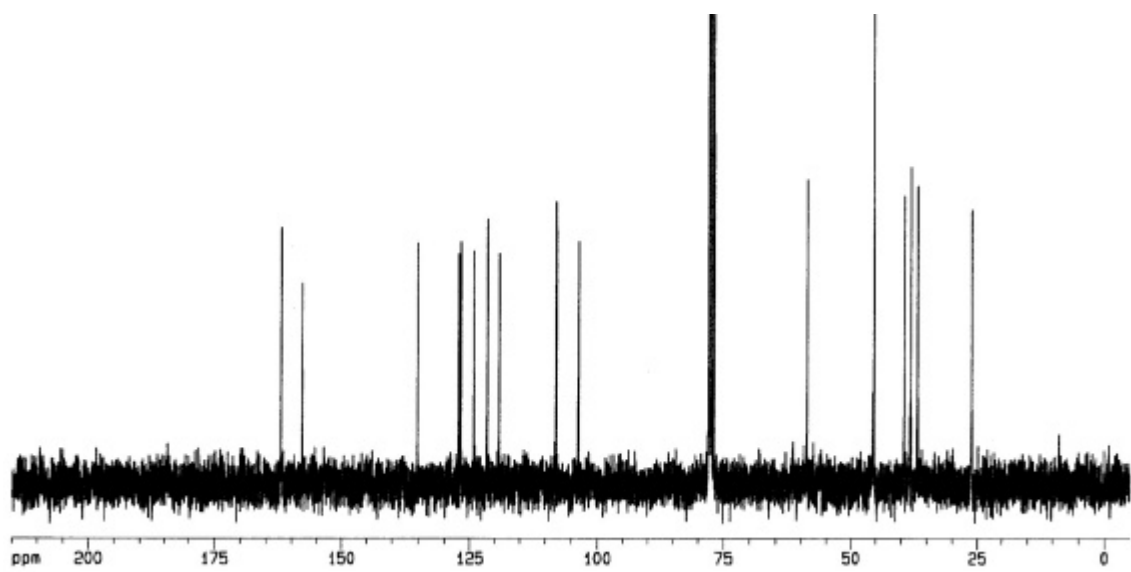
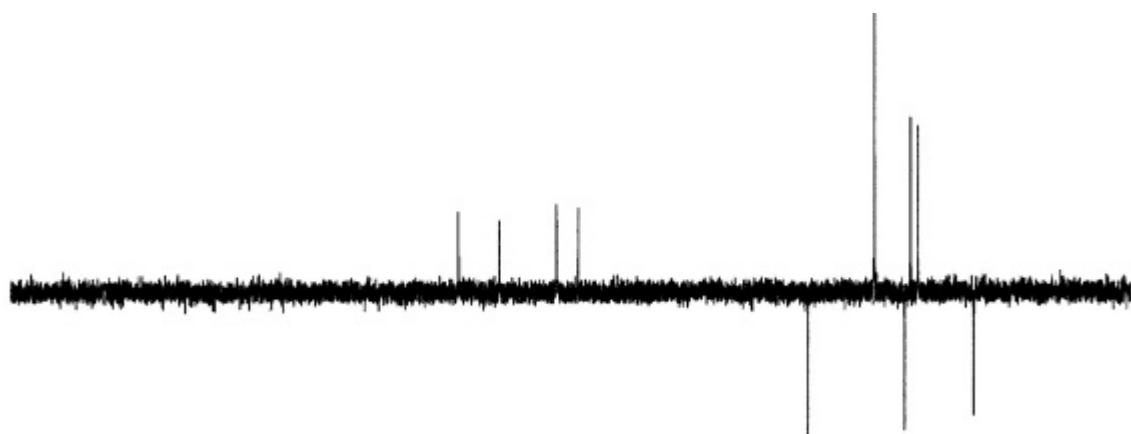
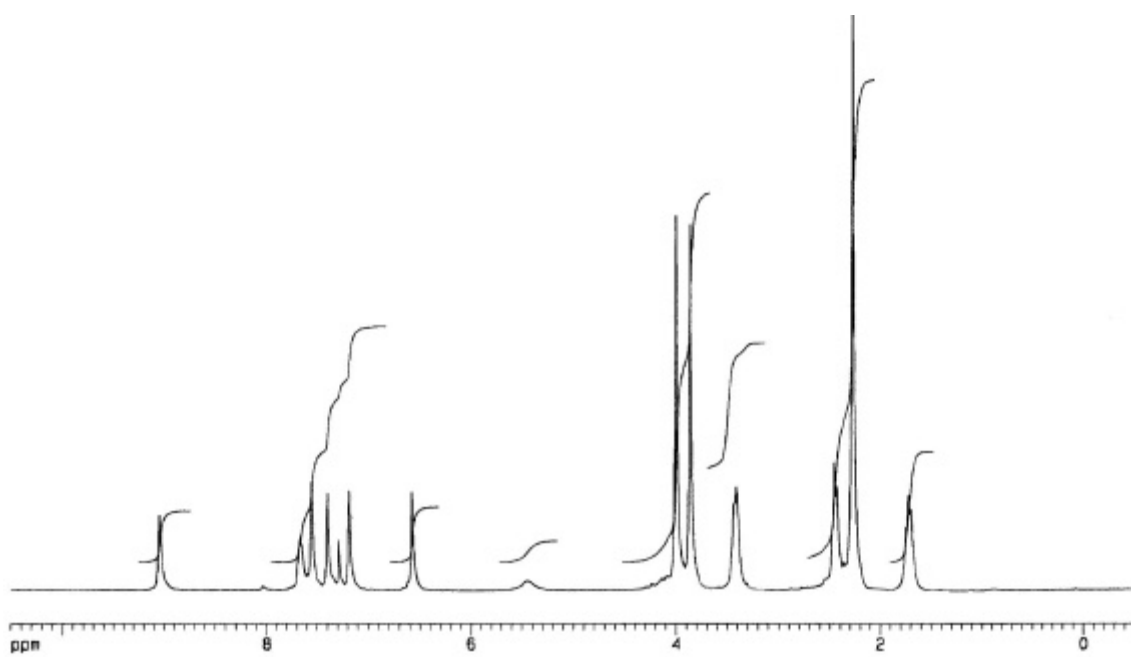
<sup>13</sup>C-NMR d: 26.1 (CH<sub>2</sub>), 37.5 (CH), 38.6 (CH), 40.0 (CH<sub>2</sub>), 45.1 (CH), 58.9 (CH<sub>2</sub>), 103.6 (CH), 107.5 (CH), 119.1 (CH), 121.8 (C), 124.3 (C), 126.8 (CH), 127.2 (C), 135.0 (C), 157.7 (C), 161.2 (C).

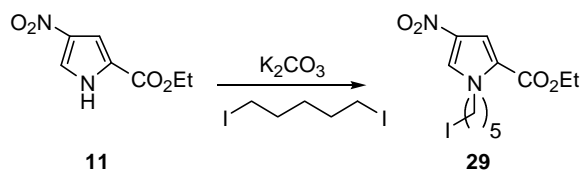
MS: m/z 376 (M<sup>+</sup>, 4), 307 (6), 155 (7), 154 (100).

HRMS: calcd. for C<sub>17</sub>H<sub>24</sub>N<sub>6</sub>O<sub>4</sub> 376.1859, found 376.19469.

<sup>5</sup> Nishiwaki, E. T.; Shigeaki, L.; Hideaki, S.; Masayuki. *Heterocycles*, **1988**, *27*, 1945.

## Supporting Information



1-(5-iodopentyl)-4-nitro-1 *H*-pyrrole-2-ethyl carboxylate (**29**)

To a solution of pyrrole **11** (1.7 g, 9.23 mmol) in dry acetone (60 mL) was added dry potassium carbonate (3.5 g) and 1,5-diiodopentane (11.96 g, 5.5 mL, 36.9 mmol). The reaction mixture was refluxed for 16 h. The suspension was filtered through celite and the filtrate was concentrated. The product was purified by flash column chromatography (EtOAc/hexane) to afford **29** as a pale-yellow oil that solidify on standing. (86%).

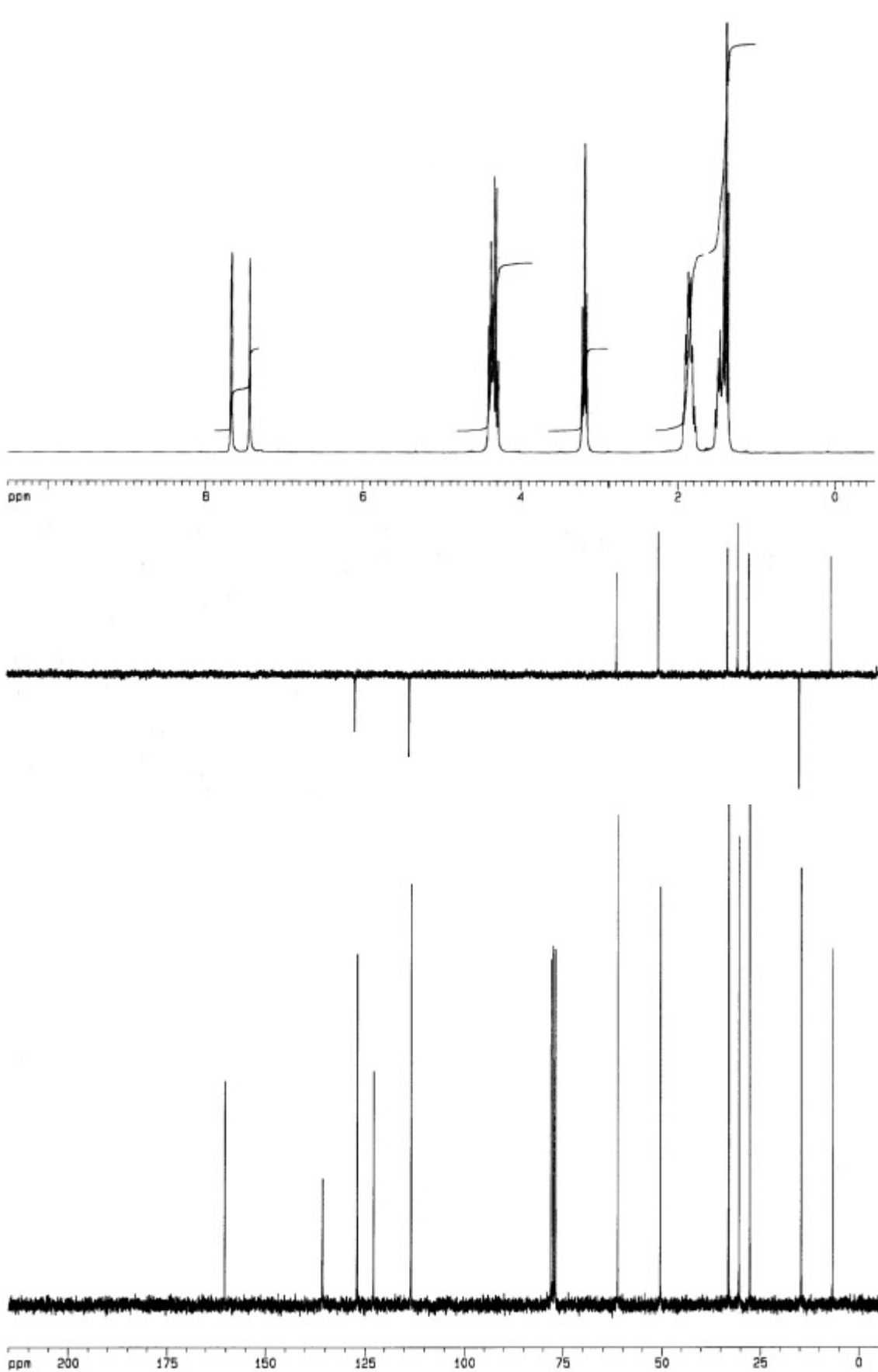
**<sup>1</sup>H-NMR** d: 1.3-1.6 (m, 5H), 1.83 (m, 4H), 3.16 (t,  $J = 6.9$  Hz, 2H), 4.22-4.37 (m, 4H), 7.42 (s, 1H), 7.68 (s, 1H).

**<sup>13</sup>C-NMR** d: 6.7 (CH<sub>2</sub>), 14.6 (CH), 27.6 (CH<sub>2</sub>), 30.4 (CH<sub>2</sub>), 33.0 (CH<sub>2</sub>), 50.5 (CH<sub>2</sub>), 61.3 (CH<sub>2</sub>), 113.4 (CH), 122.8 (C), 127.1 (CH), 135.7 (C), 160.3 (C).

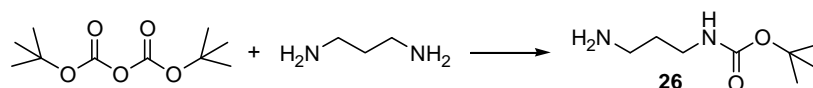
**MS:**  $m/z$  381 (MH<sup>+</sup>, 100), 364 (6), 307 (30), 289 (11), 253 (9).

**HRMS:** calcd. for C<sub>12</sub>H<sub>17</sub>IN<sub>2</sub>O<sub>4</sub> 381.02331, found 381.02963.

## Supporting Information



### **tert-butyl 3-aminopropylcarbamate (26)**



A solution of 1,3-diaminopropane (20.4 g, 23.5 mL, 0.275 mmol) in dioxane (95 mL) was cooled to 0 °C, and another solution of BOC<sub>2</sub>O (7.46 g, 0.034 mmol) in dioxane (50 mL) was added dropwise. The reaction mixture was allowed to stir for 2 days. The solvent was removed under reduced pressure and the residue was redissolved in water (150 mL) and partitioned with CH<sub>2</sub>Cl<sub>2</sub> (5 x 20 mL). Combined organic layers were dried with MgSO<sub>4</sub> and concentrated under reduced pressure to afford **26** as a clear oil. Spectroscopic data match previously reported data.<sup>6</sup>

**<sup>1</sup>H-NMR** d (CDCl<sub>3</sub>): 1.14 y 1.28 (s, 9H), 1.50 (t, 2H), 2.62 (t, 2H), 3.11 (c, 2H), 5.22 (s broad, 1H).

**MS:** m/z 175 (MH<sup>+</sup>, 13), 145 (52), 117 (40), 101 (68), 89 (16), 57 (100).

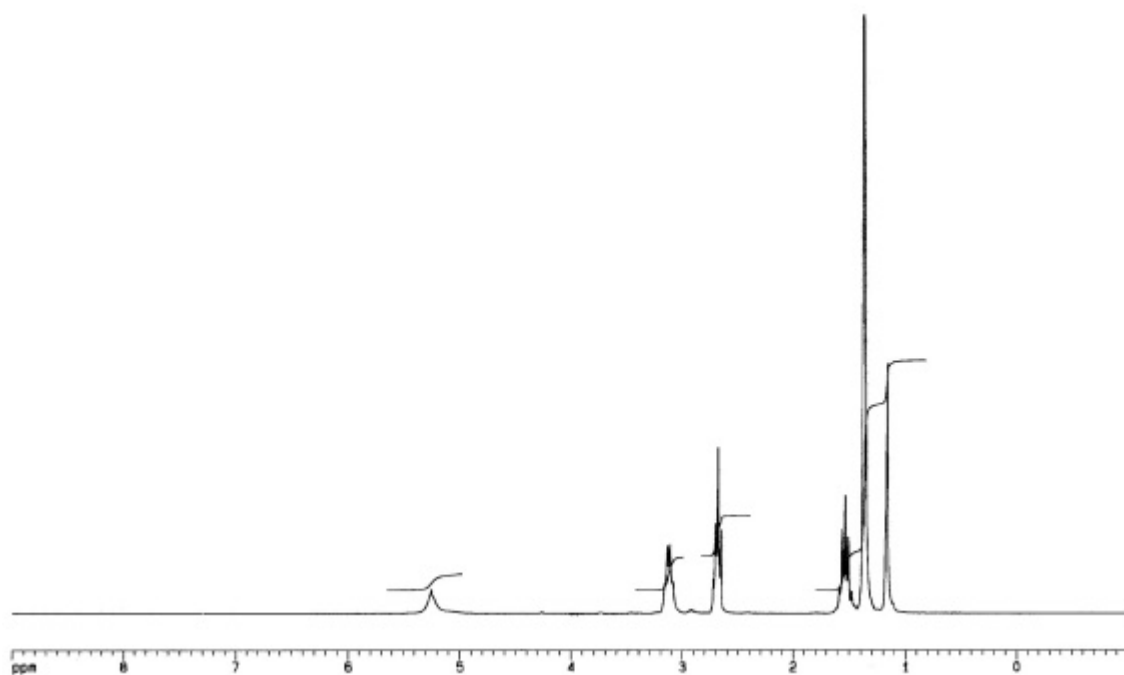
**HRMS:** calcd. for C<sub>8</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub> 174.13683, found 174.13639.

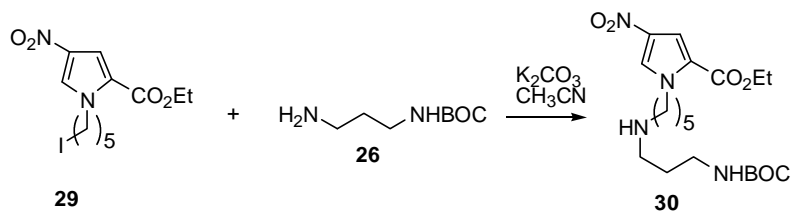
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<sup>6</sup> Lochner, M.; Geneste, H.; Hesse, M. *Helv. Chim. Acta*, **1998**, *81*, 2270.

## Supporting Information

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1-[5-({3-[(*tert*-butoxycarbonyl)amino]propyl}amino)pentyl]-4-nitro-1 *H*-pyrrole-2-ethyl carboxylate (**30**)

To a solution of iodide **29** (1 g, 2.63 mmol) in dry CH<sub>3</sub>CN (50 mL) was added dry K<sub>2</sub>CO<sub>3</sub> (5 g) and a solution of the amine **26** (1.22 g, 6.97 mmol) in CH<sub>3</sub>CN (50 mL). The resulting reaction mixture was refluxed for 12 h and the suspension was filtered through celite. The filtrate was concentrated under reduced pressure. The product was purified by flash column chromatography (5% MeOH/CH<sub>2</sub>Cl<sub>2</sub> → 1% Et<sub>3</sub>N, 10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>) to afford product **30** as a pale yellow oil (762 mg, 68%).

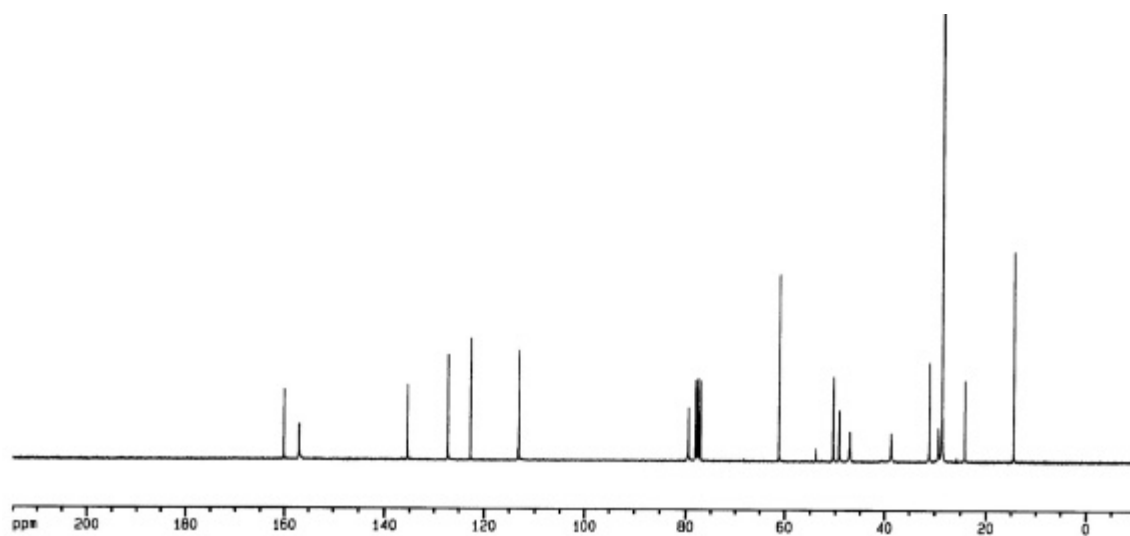
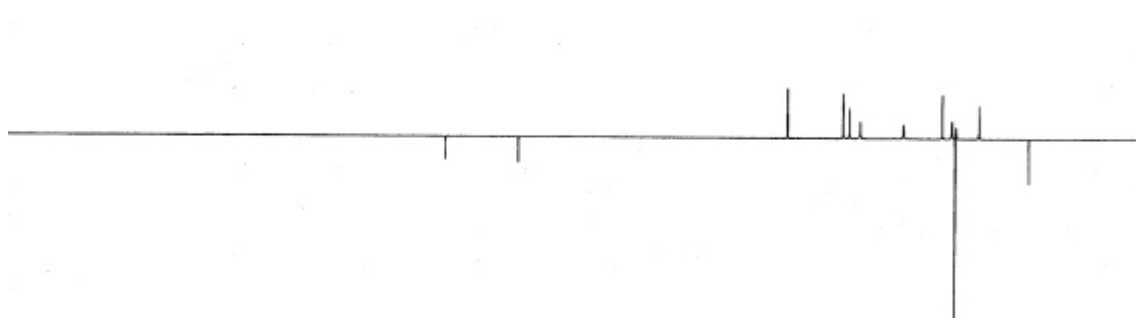
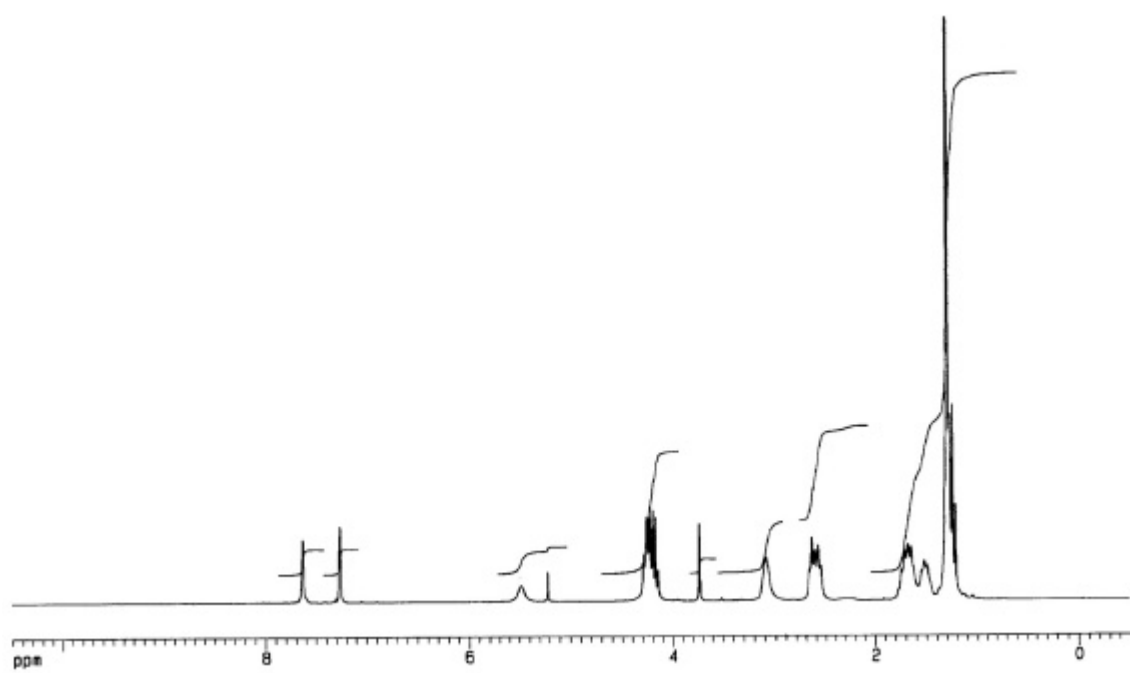
<sup>1</sup>H-NMR d: 1.12-2.38 (m, 12H) 1.45 (m, 2H) 1.52-1.77 (m, 4H), 3.05-3.26 (m, 6H), 2.48-2.63 (m, 4H), 3.09 (m, 2H), 4.15 (m, 4H), 5.42 (s broad, 1H) 7.22 (s, 1H), 7.61 (d broad, 1H).

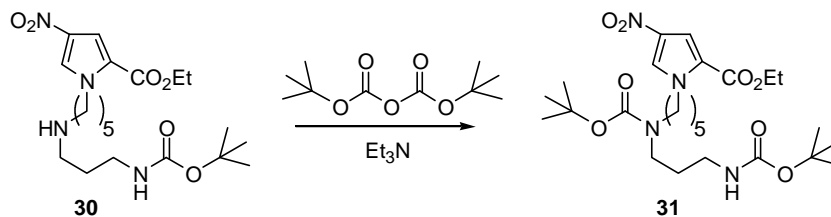
<sup>13</sup>C-NMR d: 14.5 (CH), 24.2 (CH<sub>2</sub>), 28.5 (CH<sub>2</sub>), 28.6 (CH), 28.9 (CH<sub>2</sub>), 31.1 (CH<sub>2</sub>), 37.1 (CH<sub>2</sub>), 47.1 (CH<sub>2</sub>), 49.3 (CH<sub>2</sub>), 50.5 (CH<sub>2</sub>), 54.8 (CH<sub>2</sub>), 61.2 (CH<sub>2</sub>), 79.6 (C), 113.2 (CH), 122.7 (C), 127.3 (CH), 135.5 (C), 152.3 (C), 160.3 (C).

MS: m/z 427 (MH<sup>+</sup>, 100), 413 (13), 371 (10), 307 (6).

HRMS: calcd. for C<sub>20</sub>H<sub>34</sub>N<sub>4</sub>O<sub>6</sub> 426.24784, found 426.25674.

## Supporting Information



**1-[5-((*tert*-butoxycarbonyl){3-[(*tert*-butoxycarbonyl)amino]propyl)amino)penthyl]-4-nitro-1 *H*-pyrrole-2-ethyl carboxylate (**31**)**

To a solution of amine **30** (570 mg, 1.4 mmol) in  $\text{CH}_2\text{Cl}_2$  (25 mL) was added  $\text{Et}_3\text{N}$  (0.97 mL, 7 mmol) and the solution was cooled to 0 °C. A solution of  $\text{BOC}_2\text{O}$  (436 mg, 2.0 mmol) in  $\text{CH}_2\text{Cl}_2$  (10 mL) was added and the resulting mixture was allowed to stir for 4 h at room temperature and was washed un with saturated aqueous solution of  $\text{NaHCO}_3$ . The combined organic layers were dried with  $\text{MgSO}_4$  and solvent evaporated *in vacuo*. The crude residue was purified by flash column chromatography (20% EtOAc/hexane) affording product **31** as a oily solid (707 mg, 96%).

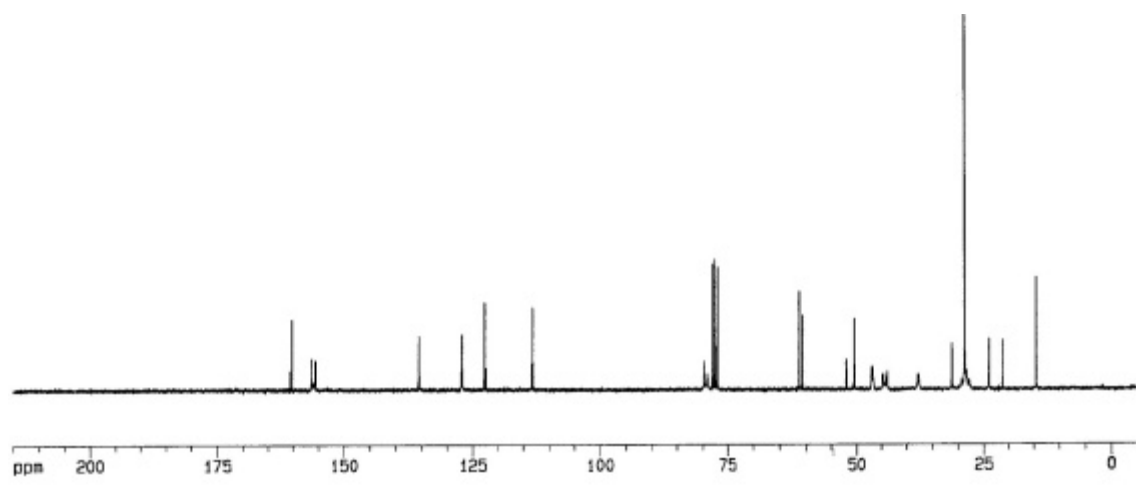
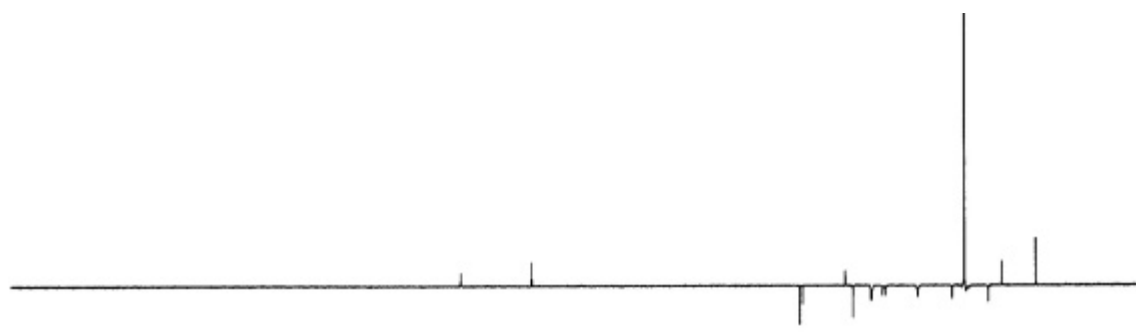
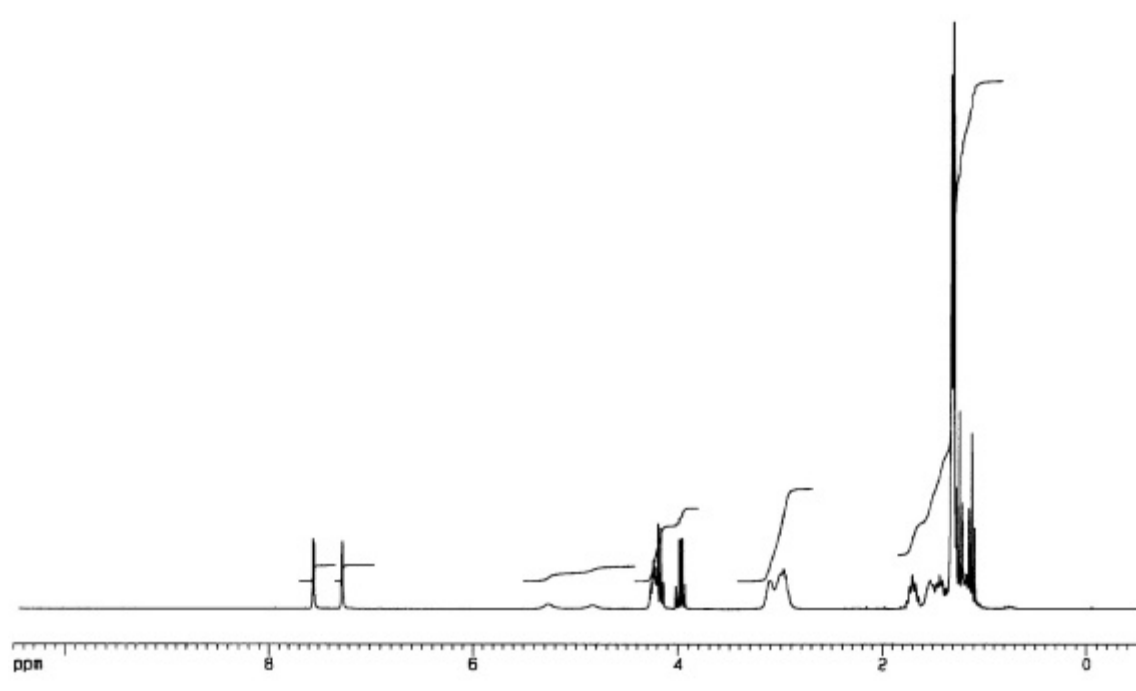
**$^1\text{H-NMR}$  d:** 1.20-1.65 (m, 25H) 2.93-3.22 (m, 6H), 4.15-4.23 (m, 4H), 7.28 (s, 1H), 7.56 (s, 1H).

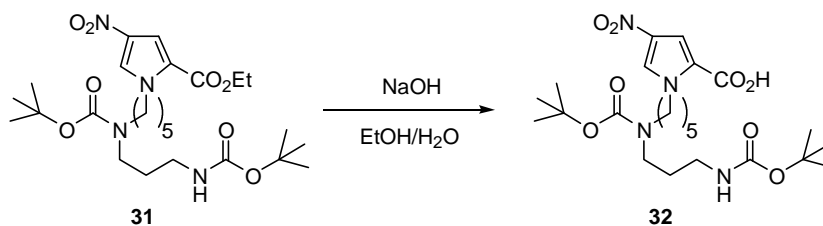
**$^{13}\text{C-NMR}$  d:** 14.4 (CH), 21.2 (CH), 23.9 ( $\text{CH}_2$ ), 28.7 (CH), 31.2 ( $\text{CH}_2$ ), 37.7 ( $\text{CH}_2$ ), 44.0 ( $\text{CH}_2$ ), 44.1 ( $\text{CH}_2$ ), 46.9 ( $\text{CH}_2$ ), 50.6 ( $\text{CH}_2$ ), 52.1 (CH), 60.6 ( $\text{CH}_2$ ), 61.2 ( $\text{CH}_2$ ), 77.6 (C), 79.8 (C), 113.3 (CH), 122.7 (C), 127.1 (CH), 135.6 (C), 156.3 (C), 156.7 (C), 160.2 (C).

**MS:**  $m/z$  452 (0.1), 113 (12), 308 (3), 57 (100).

**HRMS:** calcd. for  $\text{C}_{25}\text{H}_{42}\text{N}_4\text{O}_8$  526.30027, found 526.30169.

## Supporting Information



**1-[5-((*tert*-butoxycarbonyl){3-[(*tert*-butoxycarbonyl)amino]propyl)amino)penthyl]-4-nitro-1-*H*-pyrrole-2-carboxylic acid (**32**)**

Pyrrole **31** (1.06g 2.03 mmol) was treated with a solution of NaOH (4 mL, 1 M en 70% EtOH/H<sub>2</sub>O) at room temperature for 2 h. The solution was acidified with a saturated solution of citric acid to pH 5. This was then extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic extracts were dried with MgSO<sub>4</sub> and solvent was evaporated to give product **32** as a white powder (800 mg, 79%).

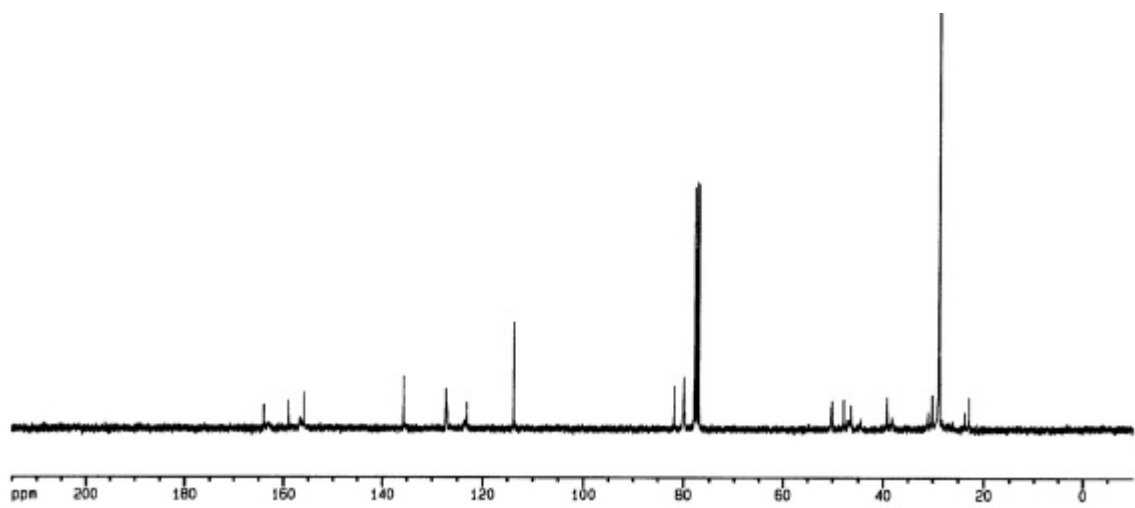
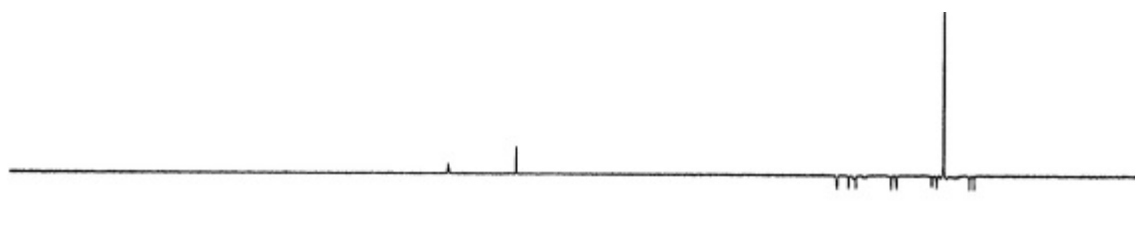
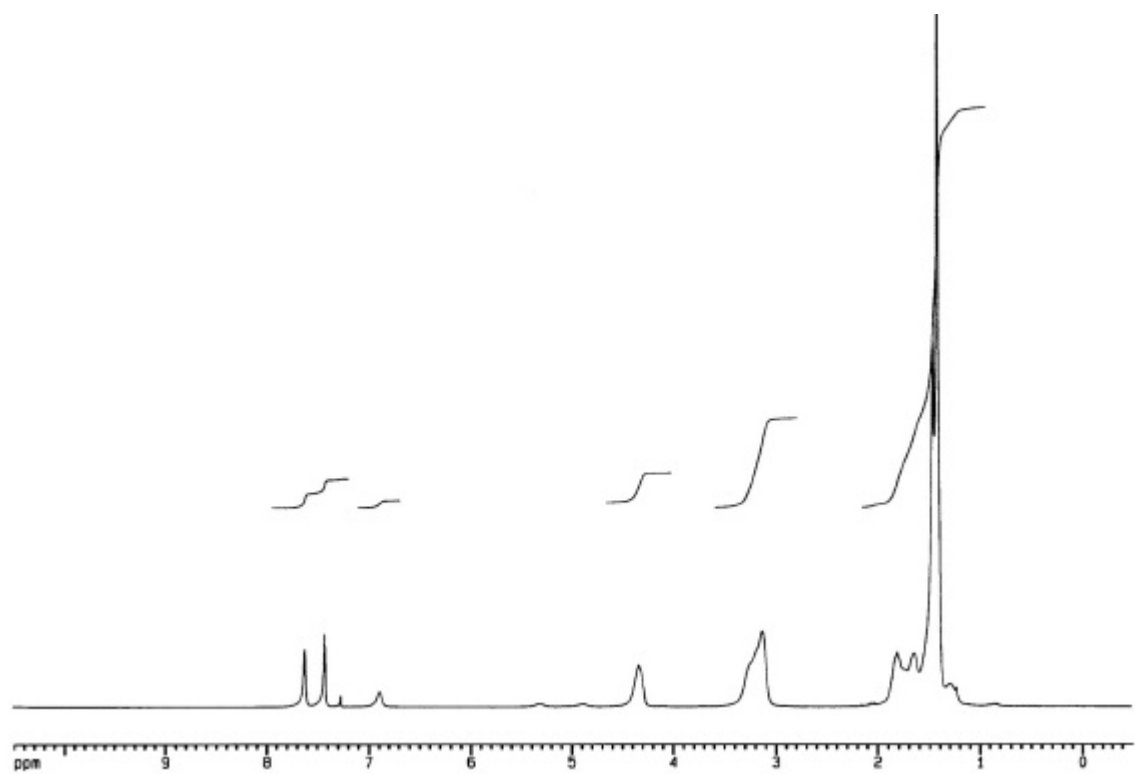
**<sup>1</sup>H-NMR** d (CD<sub>3</sub>OD): 1.20 (m, 18H), 1.5-1.9 (m, 6H) 3.1-3.4 (m, 6H), 4.2-4.4 (m, 2H), 7.26 (s, 1H), 7.62 (s, 1H).

**<sup>13</sup>C-NMR** d (CD<sub>3</sub>OD): 23.1 (CH<sub>2</sub>), 24.2 (CH<sub>2</sub>), 28.7 (CH), 30.2 (CH<sub>2</sub>), 38.1 (CH<sub>2</sub>), 44.1 (CH<sub>2</sub>), 46.9 (CH<sub>2</sub>), 47.5 (CH<sub>2</sub>), 51.2 (CH<sub>2</sub>), 79.5 (C), 81.8 (C), 114.1 (CH), 123.1, 126.9 (CH), 135.8 (C), 156.2 (C), 158.0 (C), 164.1 (C).

**MS:** m/z 499 (MH<sup>+</sup>, 61), 343 (12), 154 (100).

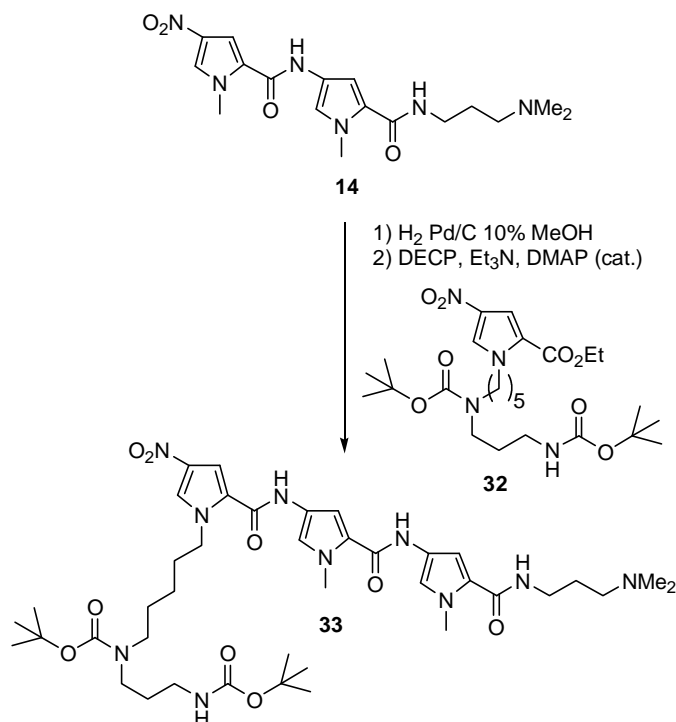
**HRMS:** calcd. for C<sub>23</sub>H<sub>38</sub>N<sub>4</sub>O<sub>8</sub> 498.26897, found 498.27919.

## Supporting Information



## Supporting Information

**N2-(5-[[3-(dimethylamino)propyl]carbamoyl]-1-methyl-1 H-3 pyrrolyl-4-[(1-{5-[(tert-butoxycarbonyl) {3-[(tert-butoxycarbonyl)amino]propyl]amino]pentyl}-4-nitro-1 H-2-pyrrolyl) carboxamido] -1-methyl-1 H-2-pyrrolicarboxamido (33)**



A solution of dipyrrole **14** (430 mg, 1.131 mmol) in MeOH (50 mL) was hydrogenated over 10% palladium on charcoal (200 mg) at room temperature (balloon pressure). The catalyst was removed by filtration through celite and the filtrates were concentrated *in vacuo*. The residue was dissolved in THF/DMF (10 mL 8:2) and added over another solution previously prepared of acid **32** (470 mg, 0.943 mmol), DECP (184 mg, 172  $\mu$ L, 1.131 mmol), Et<sub>3</sub>N (476 mg, 0.65 mL, 4.71 mmol) and DMAP (10 mg) in THF/DMF (12 mL 8:2) cooled at 0 °C. The resulting mixture was allowed to stir at room temperature for 10 h. MeOH (5 mL) was added and solvents were removed under reduced pressure, and the product was purified by flash column chromatography (1% Et<sub>3</sub>N, 10% MeOH, CH<sub>2</sub>Cl<sub>2</sub>) to afford **33** (766 mg, 82% as a yellowish solid whose

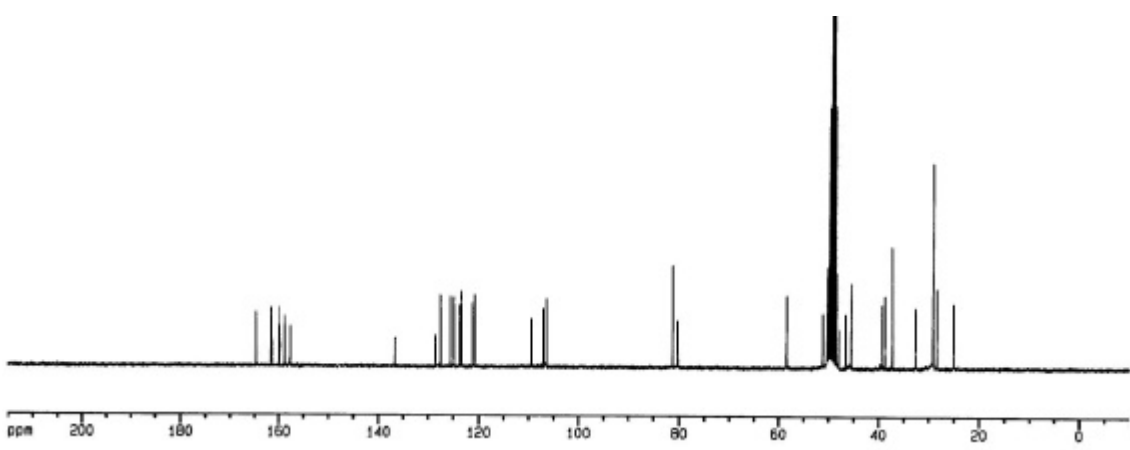
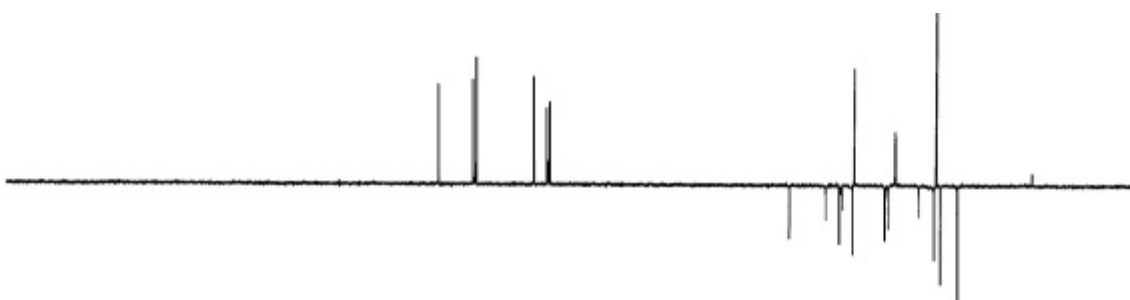
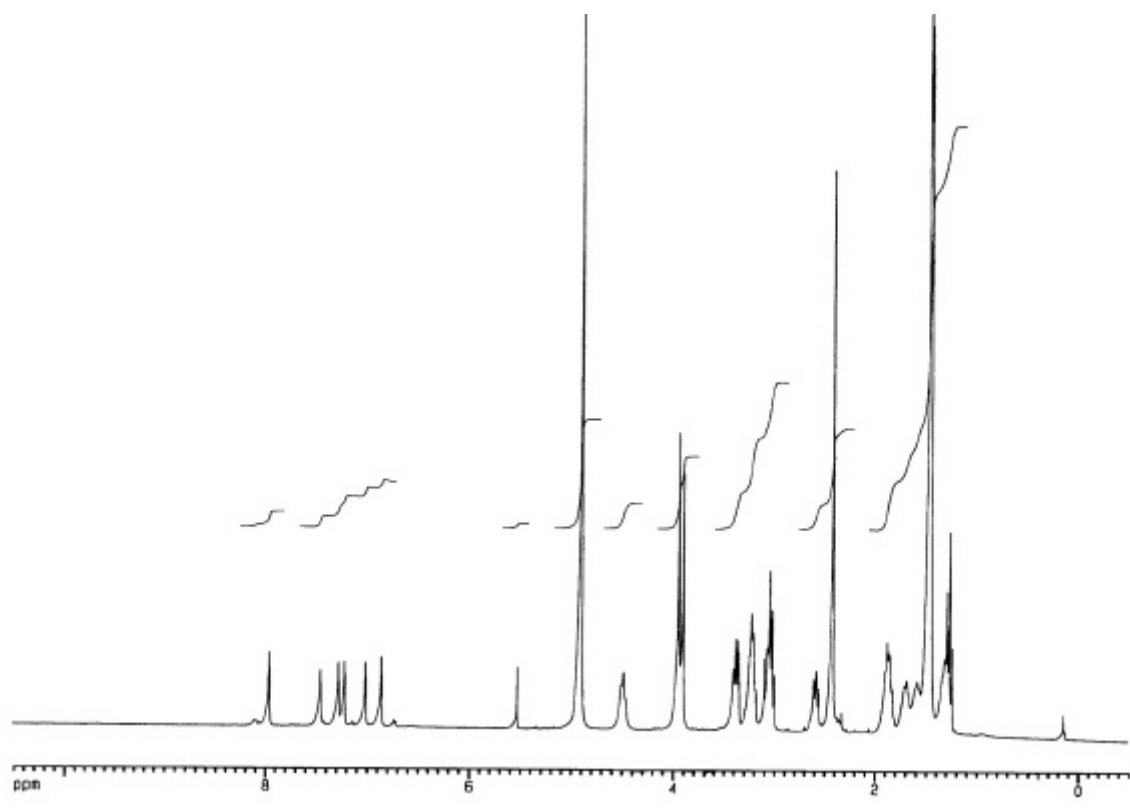
**<sup>1</sup>H-NMR** d (CD<sub>3</sub>OD): 1.2-1.4 (m, 4H), 1.5 (s, 18H), 1.6 (m, 2H), 1.7 (m, 2H), 1.8-2.0 (m, 4H), 2.38 (s, 6H), 2.56 (m, 2H), 3.05 (m, 4H), 3.21 (m, 4H), 3.30 (m, 2H), 3.79 (s, 3H), 3.81 (s, 3H), 4.42 (m, 2H), 6.81 (s, 1H), 7.0 (s, 1H), 7.19 (s, 1H), 7.26 (s, 1H), 7.41 (s, 1H), 7.91 (s, 1H).

**<sup>13</sup>C-NMR** d (CD<sub>3</sub>OD): 25.0 (CH<sub>2</sub>), 28.1 (CH<sub>2</sub>), 29.0 (CH), 30.0 (CH<sub>2</sub>), 32.6 (CH<sub>2</sub>), 37.5 (CH), 38.0 (CH<sub>2</sub>), 39.1 (CH<sub>2</sub>), 45.2 (CH), 46.1 (CH<sub>2</sub>), 50.0 (CH<sub>2</sub>), 50.1 (CH<sub>2</sub>), 51.1 (CH<sub>2</sub>), 58.3 (CH<sub>2</sub>), 80.0 (C), 81.1 (C), 106.4 (CH), 106.6 (CH), 109.7 (CH), 120.8 (CH), 121.1 (CH), 123.0 (C), 123.2 (C), 124.9 (C), 125.1 (C), 127.0 (CH), 128.1 (CH), 136.4 (C), 158.0 (C), 158.9 (C), 160.0 (C), 161.8 (C), 165.9 (C).

**MS:** *m/z* 827 (100), 627 (35), 307 (14), 179 (8).

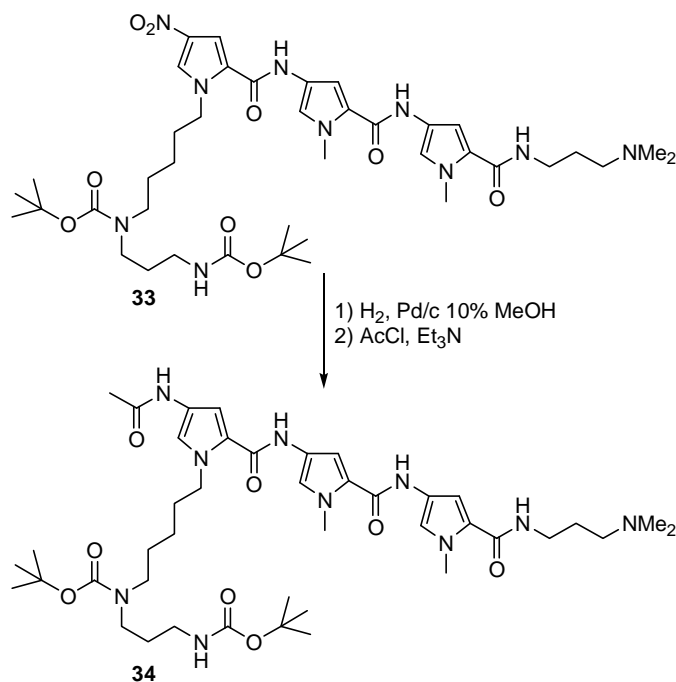
**HRMS:** calcd. for C<sub>40</sub>H<sub>62</sub>N<sub>10</sub>O<sub>9</sub> 826.46535, found 826.46597

## Supporting Information



## Supporting Information

**N2-(5-[[3-(dimethylamino)propyl]carbamoyl]-1-methyl-1*H*-pyrrolyl]-4-[[1-{5-((*tert*-butoxycarbonyl) {3-[[*tert*-butoxycarbonyl]amino]propyl]amino)pentyl]-4- (Methyl carboxamido) -1*H*-2-pyrrolyl] carboxamido} -1-methyl-1 *H*-2-pyrrole carboxamido (34)**



A solution of tripyrrole **33** (100 mg, 0.121 mmol) in MeOH (40 mL) was hydrogenated for 45 min over 10% palladium on charcoal (50 mg) at room temperature (balloon pressure). The catalyst was removed by filtration through celite and the filtrate was concentrated. The residue was immediately dissolved in DMF (10 mL) containing Et<sub>3</sub>N (0.17 mL, 1.209 mmol) and cooled to 0 °C. AcCl (9.5 μL, 0.1331 mmol in 200 μL of DMF) was added and the solution was stirred at 0 °C for 30 min and another at room temperature for another 10 h. Solvents were evaporated in vacuo and the resulting residue was purified by flash column chromatography (aluminium oxide, 5% MeOH, CH<sub>2</sub>Cl<sub>2</sub>) to give product **34** as a yellow solid (85 mg, 84%).

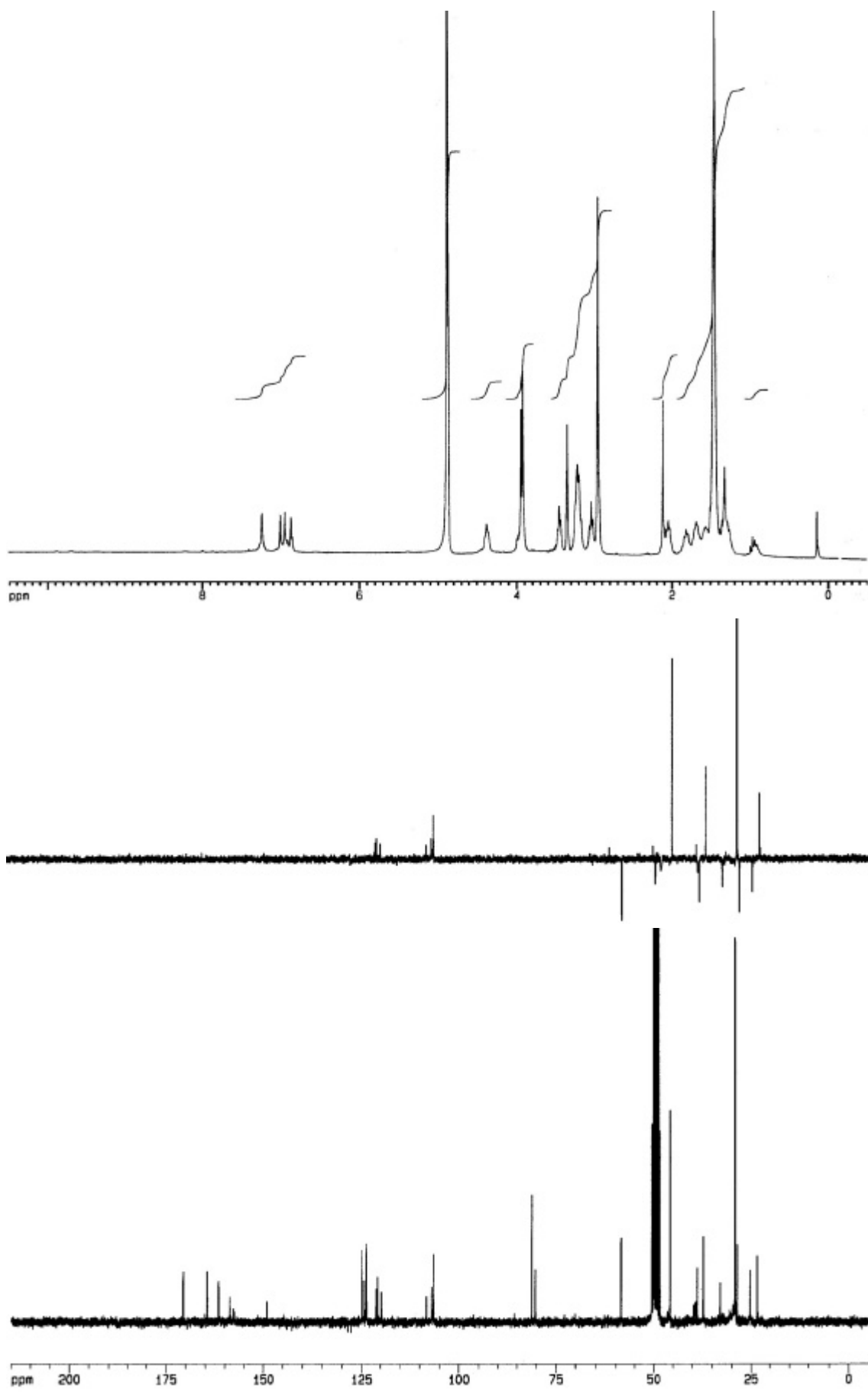
**<sup>1</sup>H-NMR** d (CD<sub>3</sub>OD): 1.2-1.4 (m, 4H), 1.4-1.6 (m, 20H), 1.6-1.7 (m, 4H), 1.96 (s, 3H), 2.2 (s, 6H), 2.35 (m, 2H), 2.89 (m, 2H), 3.01 (m, 4H), 3.20 (m, 2H), 3.71 (s, 3H), 3.79 (s, 3H), 4.21 (m, 2H), 6.67 (m, 2H), 6.82 (s, 1H), 7.16 (m, 3H).

**<sup>13</sup>C-NMR** d (CD<sub>3</sub>OD): 23.8 (CH), 25.1 (CH<sub>2</sub>), 28.8 (CH<sub>2</sub>), 29.2 (CH), 32.1 (CH<sub>2</sub>), 37.5 (CH), 38.9 (CH<sub>2</sub>), 39.2 (CH<sub>2</sub>), 45.1 (CH), 45.2 (CH<sub>2</sub>), 47.3 (CH<sub>2</sub>), 50.0 (CH<sub>2</sub>), 58.0 (CH<sub>2</sub>), 80.3 (C), 81.2 (C), 106.5 (CH), 106.9 (CH), 108.2 (CH), 119.9 (CH), 120.8 (CH), 121.2 (CH), 123.8, (C) 124.4 (C), 124.9 (C), 158.7 (C), 161.6 (C), 161.7 (C), 164.5 (C), 170.6 (C).

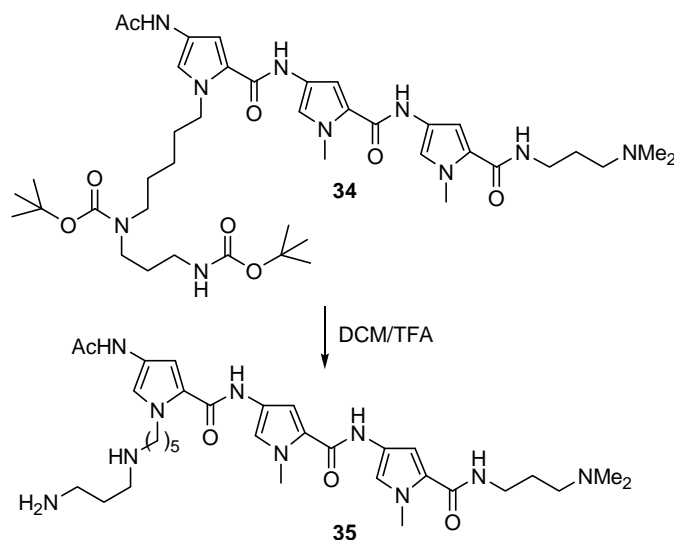
**MS:** m/z 839.5 (100), 639 (19), 309 (12), 231 (48).

**HRMS:** calcd. for C<sub>42</sub>H<sub>66</sub>N<sub>10</sub>O<sub>8</sub> 838.50651, found 838.46535.

## Supporting Information



***N*2-(5-([3-(dimetilamino)propil]carbamoil)-1-metil-1 *H*-pirrolil)-4-([1-(5-[(3-aminopropil) amino] pentil)-4-(metilcarboxamido)-1 *H*-2-pirrolil]carboxamido)-1-metil-1 *H*-2-pirrol carboxamido (35)**



A solution of tripyrrole **34** (84 mg, 0.1 mmol) in  $\text{CH}_2\text{Cl}_2$  (3 mL) was cooled to 0 °C and TFA (3 mL, 15 min) was added dropwise. The resulting orange solution was stirred at 0 °C for 1 h and at room temperature for another 2 h. Solvents were removed in vacuo at room temperature and residual TFA was removed by codistillation with  $\text{CH}_2\text{Cl}_2$ . The residue was identified as the desired product **35**, showing by HPLC to be highly pure. HPLC conditions: analytical column C-18 5 → 95 % B, 30 min.  $t_R = 9.315$ .

**$^1\text{H-NMR}$**  d ( $\text{CD}_3\text{OD}$ ): 1.14-1.35 (m, 4H), 1.5-1.9 (m, 4H), 2.0 (s, 3H), 2.79 (s, 6H), 2.8-3 (m, 2H), 3-3.1 (m, 2H), 3.3 (m, 4H), 3.70 (s, 3H), 3.72 (s, 3H), 4.2 (m, 2H), 6.6 (s, 1H), 6.65 (s, 1H), 6.7 (s, 1H), 7.09 (m, 3H).

**$^{13}\text{C-NMR}$**  d ( $\text{CD}_3\text{OD}$ ): 22.1 (CH), 24.0 ( $\text{CH}_2$ ), 24.9 ( $\text{CH}_2$ ), 25.2 ( $\text{CH}_2$ ), 29.8 (CH), 30.0 ( $\text{CH}_2$ ), 35.0 ( $\text{CH}_2$ ), 35.1 (CH), 35.2 ( $\text{CH}_2$ ), 44.3 (CH), 46.3 ( $\text{CH}_2$ ), 47.5 ( $\text{CH}_2$ ), 48.1 ( $\text{CH}_2$ ), 55.0 ( $\text{CH}_2$ ), 107.3 (CH), 111.9 (C), 116.0 (C), 121.5 (CH), 122.7 (CH), 124.1 (C), 124.2 (C), 124.4 (C), 160.0 (C), 165.3 (C), 171.2 (C).

**MS:**  $m/z$  639 ( $\text{MH}^+$ , 10), 422 (8), 391 (21), 309 (73), 278 (100).

**HRMS:** calcd. for  $\text{C}_{32}\text{H}_{50}\text{N}_{10}\text{O}_4$  638.40165, found 638.39561.

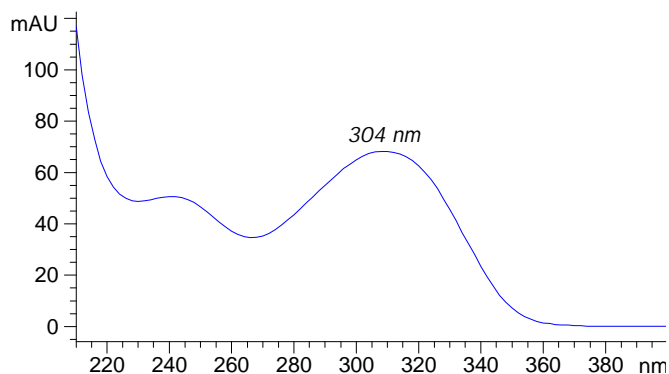
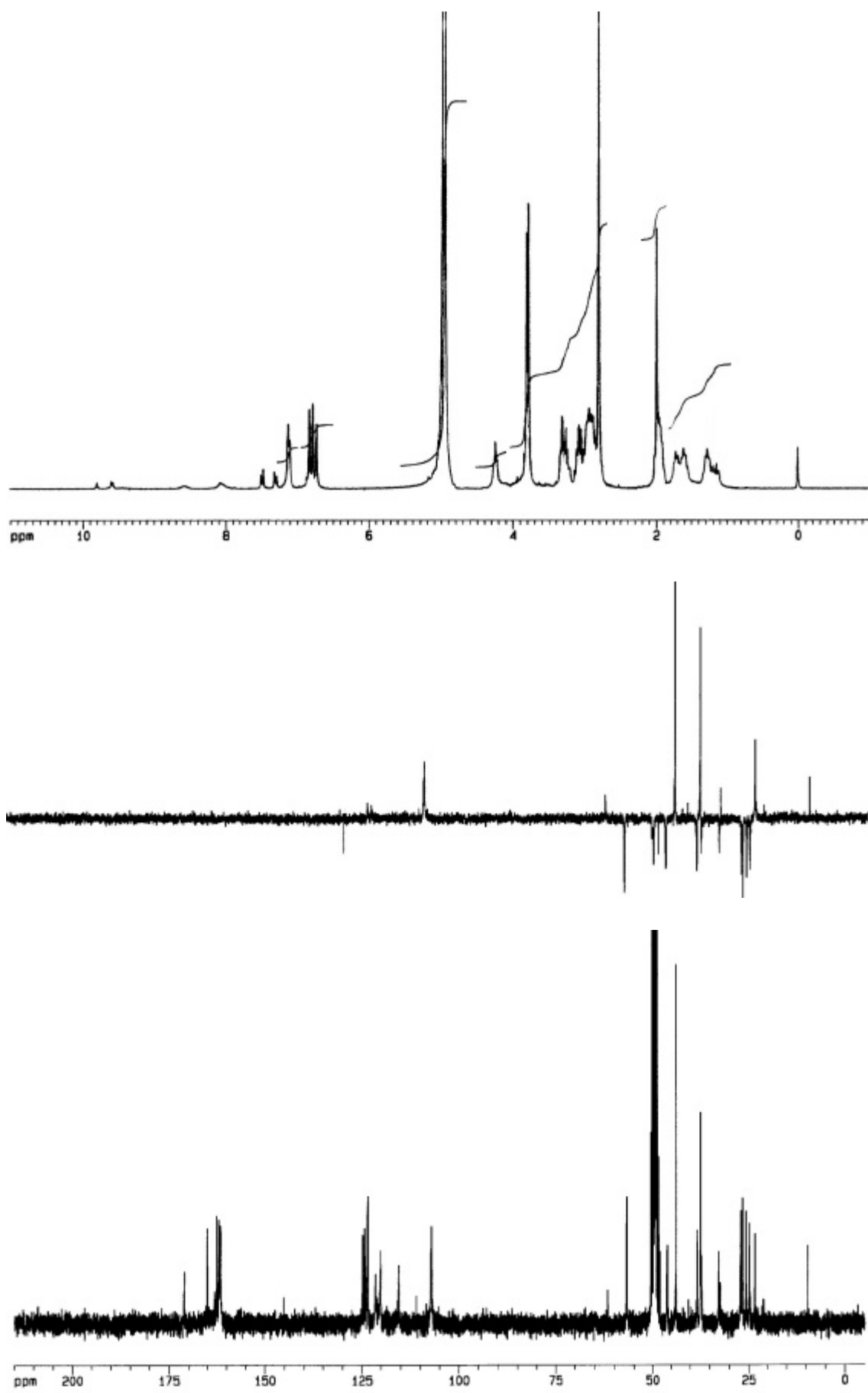
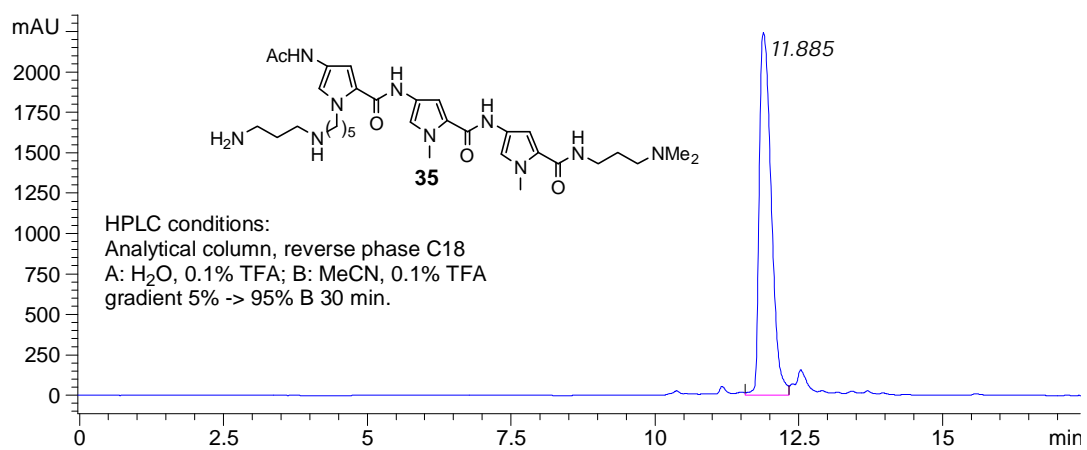


Figure 1. UV spectrum of **35**.

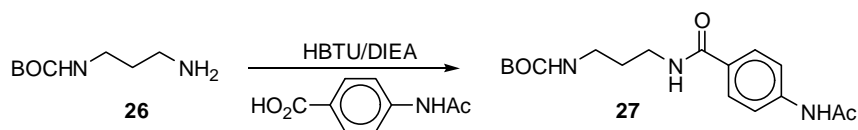
## Supporting Information



## Supporting Information



**Figure 2.** HPLC chromatogram of **35**.

**tert-butyl 3-[[4-(acetylamino)benzoyl]amino]propylcarbamate (27)**

To a solution of amine **26** (250 mg, 1.435 mmol) and DIEA (1.252 mL, 7.17 mmol) in DMF (5 mL) was added a mixture of acetamidobenzoic acid (334 mg, 1.86 mmol) and HBTU (707 mg, 1.86 mmol) prepared 5 min before. The resulting solution was stirred for 4 h and H<sub>2</sub>O (50 mL) was added and the mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The combined organic layers were dried with Na<sub>2</sub>SO<sub>4</sub> and the solvents removed *in vacuo* to give product **27** as a light-yellow oil that solidified on standing. (336 mg, 70%).

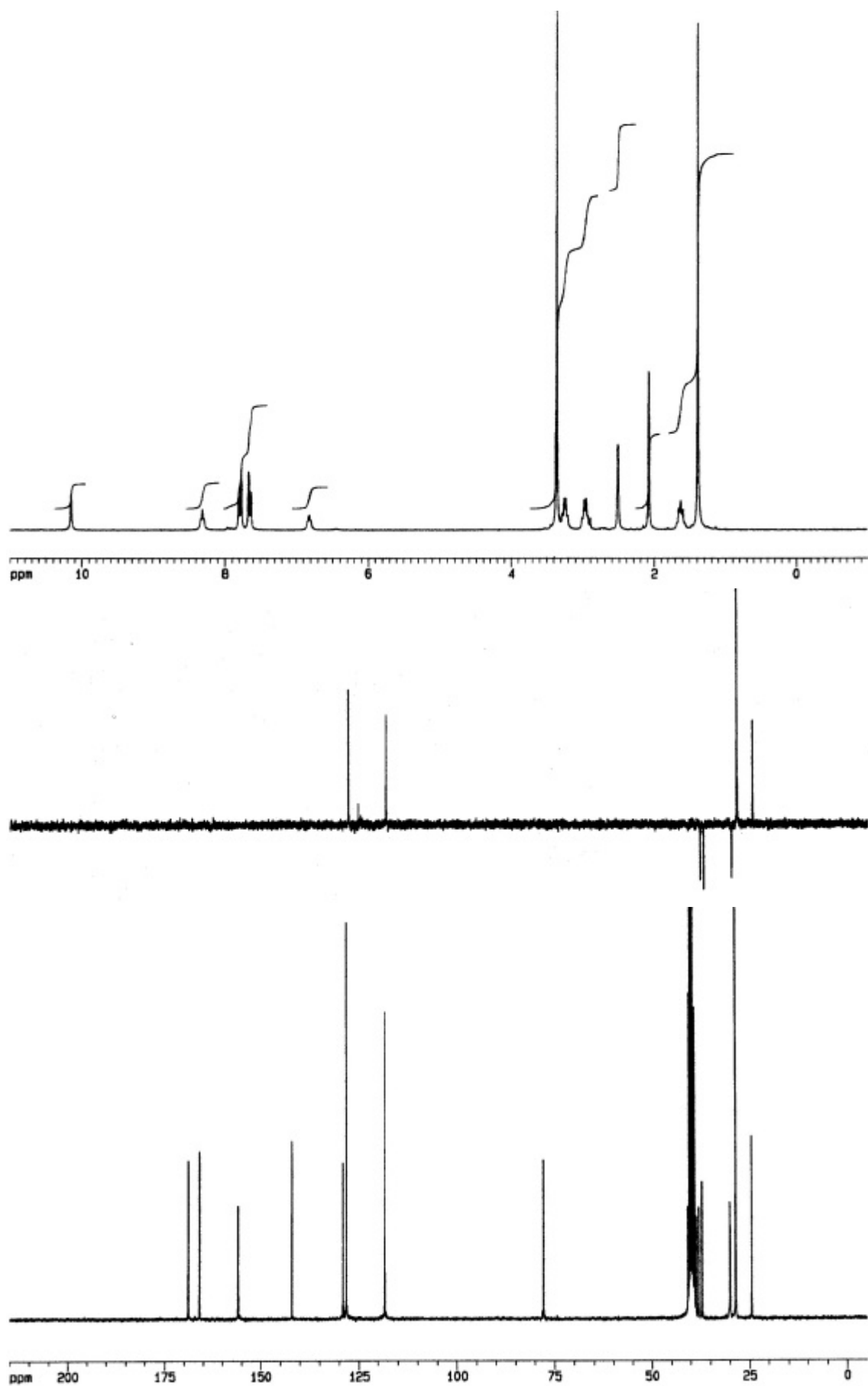
**<sup>1</sup>H-RMN d** (DMSO-D<sub>6</sub>): 1.38 (s, 9H), 1.60 (m, 2H), 2.84 (s, 3H), 2.96 (m, 2H), 3.24 (m, 2H), 6.78 (t, 1H), 7.67 (d, 2H), 7.78 (d, 2H), 8.28 (t, 1H), 10.13 (s, 1H).

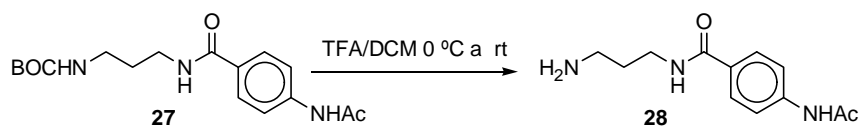
**<sup>13</sup>C-RMN d** (DMSO-D<sub>6</sub>): 24.43 (CH), 28.57 (CH), 29.99 (CH<sub>2</sub>), 37.21 (CH<sub>2</sub>), 38.05 (CH<sub>2</sub>), 77.83 (C), 118.37 (CH), 128.27 (CH), 129.17 (C), 142.13 (C), 155.94 (C), 169.01 (C), 169.21 (C).

**MS**: 335 (M<sup>+</sup>, 66), 201 (13), 101 (80), 58 (54), 57 (100).

**HRMS**: calcd. for C<sub>17</sub>H<sub>25</sub>N<sub>3</sub>O<sub>4</sub> 335.18451, found 335.12455.

## Supporting Information



4-(acetylamino)-*N*-(3-aminopropyl)benzamide (**28**)

To a suspension of **27** (297 mg, 0.886 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5 mL) cooled to 0 °C, TFA (5 mL) was added dropwise, observing the formation of a clear solution that was stirred for 1 h and then heated to room temperature and stirred for another hour. Solvents were removed *in vacuo* and the residue redissolved in CH<sub>2</sub>Cl<sub>2</sub> and removed again to ensure complete removal of TFA. HPLC of the white powder obtained showed only one product that was confirmed as product **28** by MS and NMR analysis.

<sup>1</sup>H-RMN-d (DMSO-D<sub>6</sub>): 1.84 (m, 2H), 2.12 (s, 3H), 2.85 (m, 2H), 3.37 (m, 2H), 7.56 (d, 2H), 7.69 (d, 2H), 7.88 (s, 1H).

MS: 235 (M<sup>+</sup>, 62), 177 (34), 162 (23), 134 (63), 43 (100).

HRMS: calcd. for C<sub>12</sub>H<sub>17</sub>N<sub>3</sub>O<sub>2</sub> 235.13208, found 235.13387.

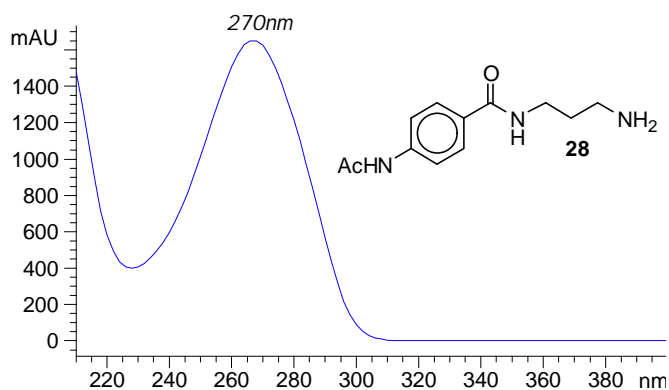
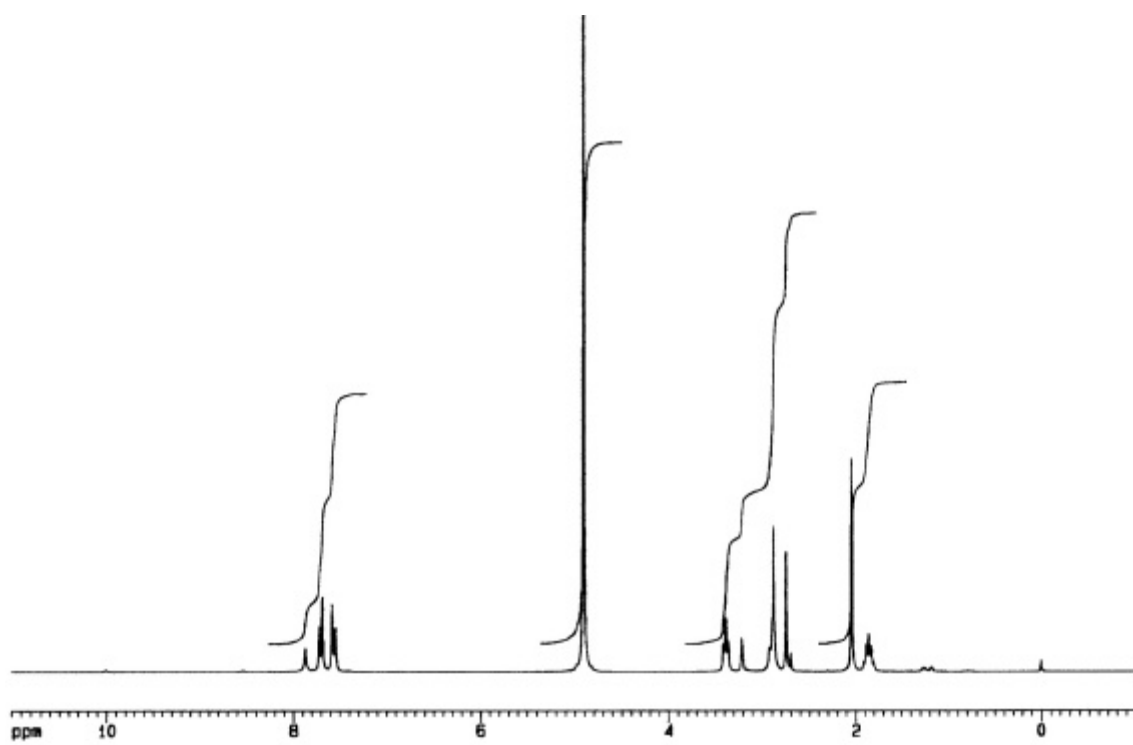


Figure 3. UV spectrum of amine **28**.

## Supporting Information



## Supporting Information

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### CD and UV spectra.

CD spectra were recorded using a *Jasco J-715* spectrometer coupled with a refrigerated water bath *Nestlab RTE-111*. All measurements were made using a 0.2 cm cell at 4 °C. Concentration of the peptide hybrids was determined by UV spectroscopy. All UV spectra were recorded using a *Varian Cary 100 Bio* with 1 cm cells.

All CD measurements were made using 300  $\mu$ L of solutions of ds-DNA of known concentration (5  $\mu$ M) at pH = 7.5 (phosphate buffer 10 mM) and NaCl (100 mM). Over those solutions a known volume of the mother solution of the molecule studied was added. Final concentration of peptide and DNA were always near 5  $\mu$ M. The resulting mixtures were incubated for 5 min before registering.

<b><i>CD spectra parameters:</i></b>	
<b>Cell Length:</b>	0.2 cm
<b>Concentration:</b>	5 x 10 <sup>-6</sup> M
<b>Solvent:</b>	H <sub>2</sub> O/buffer
<b>Temperature:</b>	4 °C
<b>Data mode:</b>	Mol. Ellip.
<b>Range:</b>	195-380 nm
<b>Resolution:</b>	0.2 nm
<b>Band width:</b>	2.0 nm
<b>Sensitivity:</b>	10 mdeg
<b>Response:</b>	0.25 sec
<b>Speed:</b>	100 nm/min
<b>Accumulation:</b>	10 scans

The spectra showed are the average of 10 scans and were slightly smoothed using the “smooth” macro implemented in the program *Kaleidagraph* (v 3.5 by Synergy Software). Spectra of the peptides in presence of DNA were calculated as the difference between the spectra of the peptide - DNA mixture and the spectrum of free DNA.

### Determination of the affinity constant for tripyrrole derivatives.

CD spectroscopy was used to determine the affinity constant of the tripyrrole derivatives for selected DNA sequences. Considering the equilibrium  $L + M \rightleftharpoons LM$ , where L is the ligand, M the macromolecule, and LM the complex formed by their association, the *affinity constant*  $K_{\text{aff}}$  is defined as:

$$K_{\text{aff}} = \frac{[ML]}{[M][L]}$$

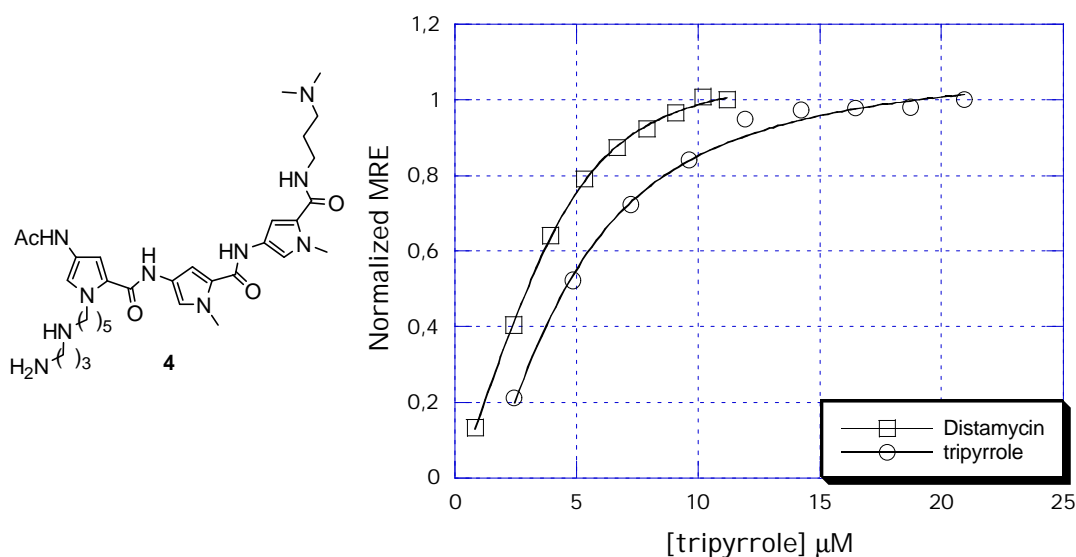
In the equilibrium, the exact concentration of free ligand [L] is given by the following expression:

$$[L] = 0.5 \left\{ ([M]_{\text{T}} + [L]_{\text{T}} + 1/K_{\text{aff}}) - \sqrt{K_{\text{aff}} ([M]_{\text{T}} + [L]_{\text{T}} + 1/K_{\text{aff}})^2 - 4 K_{\text{aff}} [M]_{\text{T}} [L]_{\text{T}}} \right\}$$

The recorded signal (CD in our case) can be related with that concentration resulting in the equation used to fit the data.<sup>7</sup>

$$[q] = 0.5 [q]_{\infty} \left\{ ([M]_{\text{T}} + [L]_{\text{T}} + K_{\text{D}}) - \sqrt{([M]_{\text{T}} + [L]_{\text{T}} + K_{\text{D}})^2 - 4 [M]_{\text{T}} [L]_{\text{T}}} \right\}$$

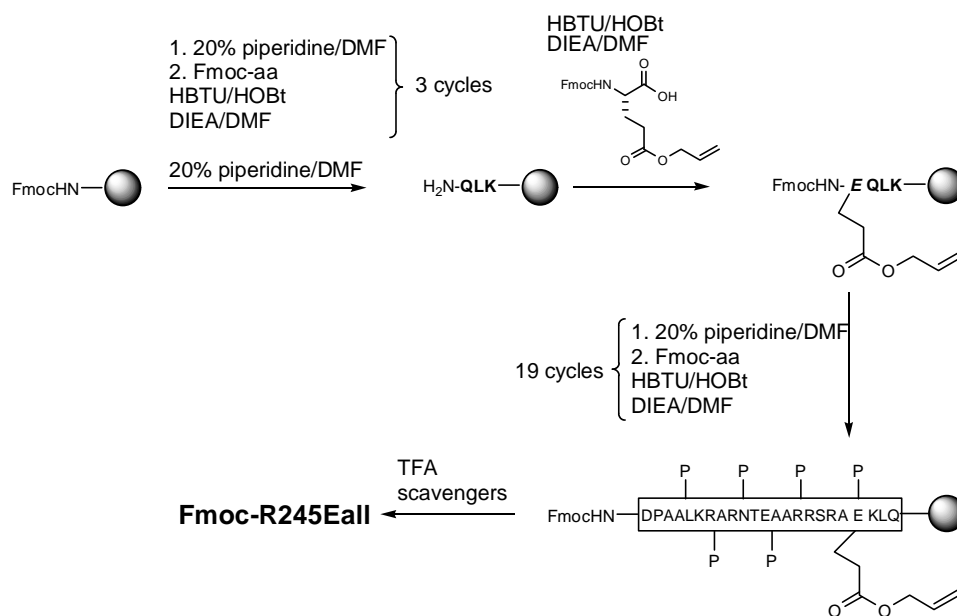
Where  $[q]_{\infty}$  represents the value of ellipticity at infinitum and  $[q]$  the measured value of ellipticity at each point.  $[M]_{\text{T}}$  is the total concentration of the macromolecule (DNA),  $[L]_{\text{T}}$  the total concentration of ligand (tripyrrrole),  $K_{\text{D}}$  is the dissociation constant. Data obtained for tripyrrole **4** are shown. Affinity constants are in both cases in the micromolar range ( $\sim 0.8 \mu\text{M}$  for tripyrrole **3** and  $\sim 1.0 \mu\text{M}$  for tripyrrole **4**). Curve fitting was carried out using nonlinear least-squares analysis.



**Figure 4.** CD titration of tripyrrole **4** with ds-DNA 5'-CGCGAAAAAGCGC-3' compared with Distamycin. Values of MRE obtained at 330 nm are normalized and given arbitrary the value of 1 on saturation for easier comparison.

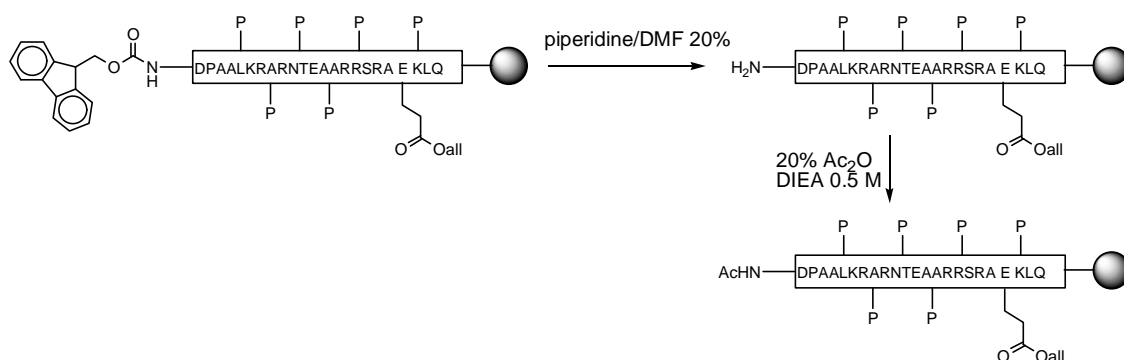
<sup>7</sup> Pilch, D. S.; Poklar, N.; Baird, E. E.; Dervan, P. B.; Breslauer, K. *Biochemistry*, **1999**, *38*, 2143.

### Solid Phase Synthesis of peptide Fmoc-R245Eall



Synthesis of peptide **Fmoc-R245Eall** was performed using standard Fmoc SPPS conditions, starting with Rink MBHA amide resin (217 mg, 0.1 mmol, load 0.46 mmol/g). After completion of the synthesis deprotection/cleavage of 50 mg resin afforded after RP-HPLC 8 mg of a peptide identified by MS to be the desired product. (approx. 44%)

### Preparation of peptide R245E: Acetylation of N terminal residue.



Over 165 mg of **Fmoc-R245E-all** peptide bound to the resin, a solution of piperidine (5 mL, 20% in DMF) was added and argon was passed through the mixture for 45 min. The resin was washed with DMF (3 x 5 mL x 5 min). Deprotection of Fmoc group can be confirmed by RP-HPLC analysis of the products by standard TFA/scavengers mixture treatment. Deprotection of Fmoc results in a shift on the  $t_R$ .

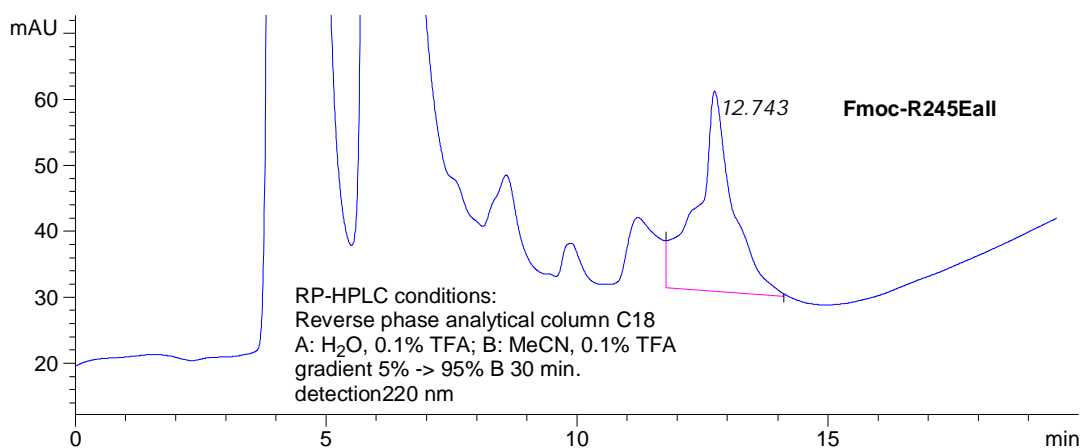


Figure 5. peptide **Fmoc-R245Eall**.

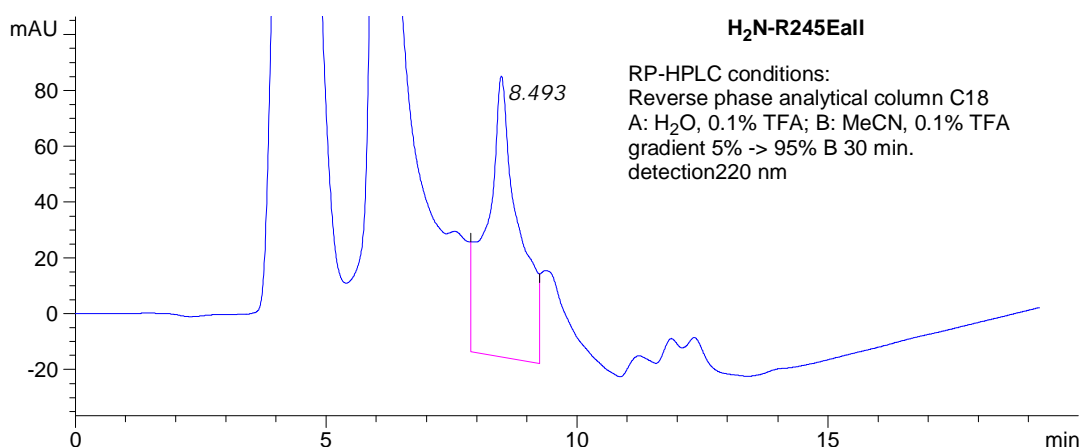


Figure 6. peptide **H<sub>2</sub>N-R245Eall**.

Acetylation was carried out immediately after deprotection of the N terminal Fmoc protecting group. After washing the resin with DMF (3 x 5 mL x 5 min), DIEA (2.5 mL, 0.2 M in DMF) and Ac<sub>2</sub>O (5 mL, 20% en DMF) were added. The resulting mixture was shaken for 1 h. After

## Supporting Information

filtration the resin was washed with DMF (3 x 5 mL x 5 min), *i*PrOH (5 mL x 5 min) and Et<sub>2</sub>O (5 mL x 5 min.). HPLC analysis of the residue obtained after deprotection/cleavage clearly showed remove of the product at 8.5 min and a new product with retention time of 9.3 min, corresponding to the acetylated product as shown by MS (electrospray).

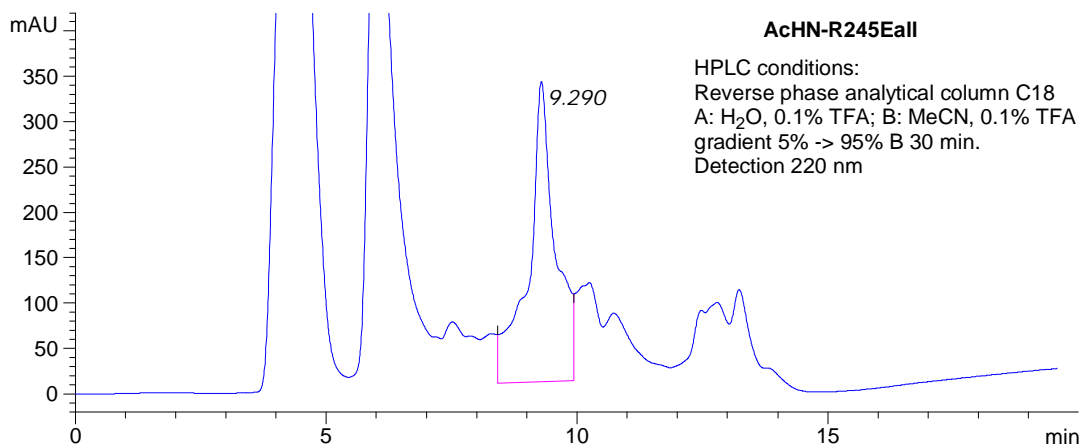


Figure 7. HPLC of AcHN-R245Eall

**R245Eall MS (ES):** [MH<sup>+</sup>] m/z calcd. for C<sub>112</sub>H<sub>198</sub>N<sub>43</sub>O<sub>34</sub> 2691.0, observed 2690.6.

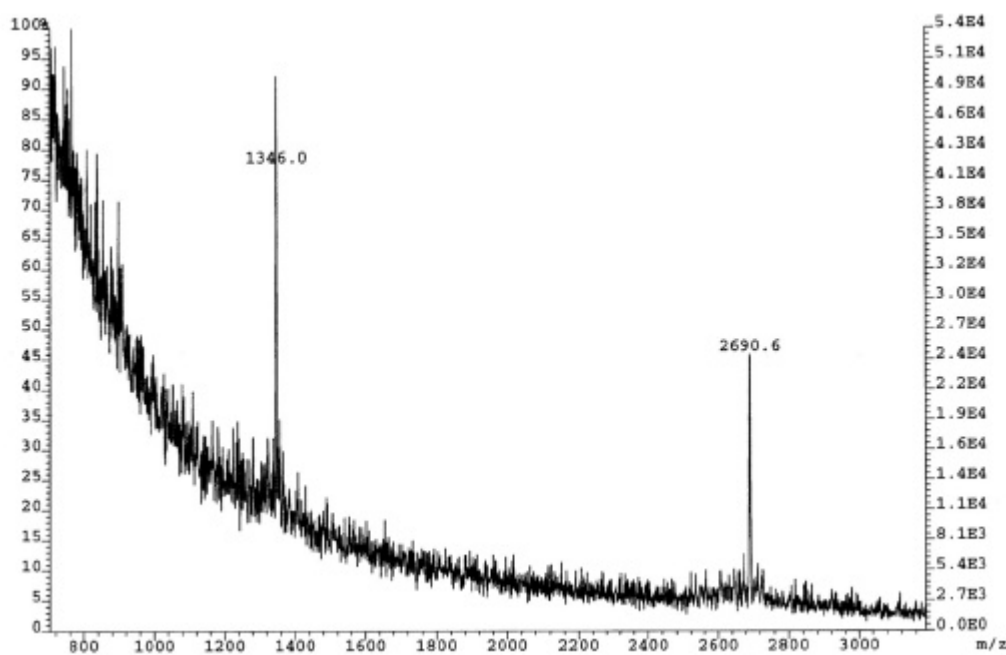
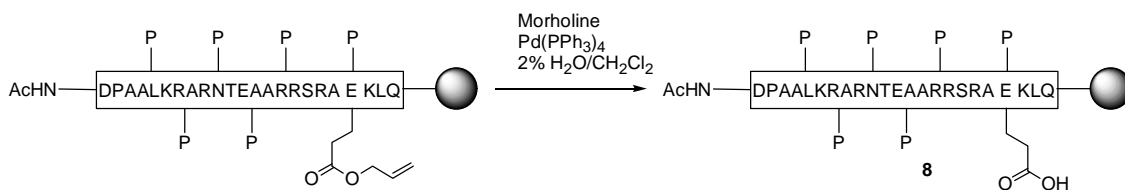


Figure 8. MS of peptide AcHN-R245Eall.

### Preparation of peptide R245E: deprotection of allyl protecting group



Over 161 mg of resin (approx. 0.025 mmol of peptide) DMF (10 mL) was added and the resin was shaken for 4 h. The resin was filtrated and 2% H<sub>2</sub>O/CH<sub>2</sub>Cl<sub>2</sub> (6mL), morpholine (0.42 mL, 4.84 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (29 mg, 0.025 mmol) were added. The mixture was shaken under argon for 12 h. The resin was filtered and washed with DMF (3 x 5 mL x 5 min), *i*PrOH (2 x 5 mL x 5 min) and CH<sub>2</sub>Cl<sub>2</sub> (5 mL x 5 min).

Cleavage and deprotection of the product with the standard mixture, and its HPLC analysis shows a new product with retention time of 16.9 min.

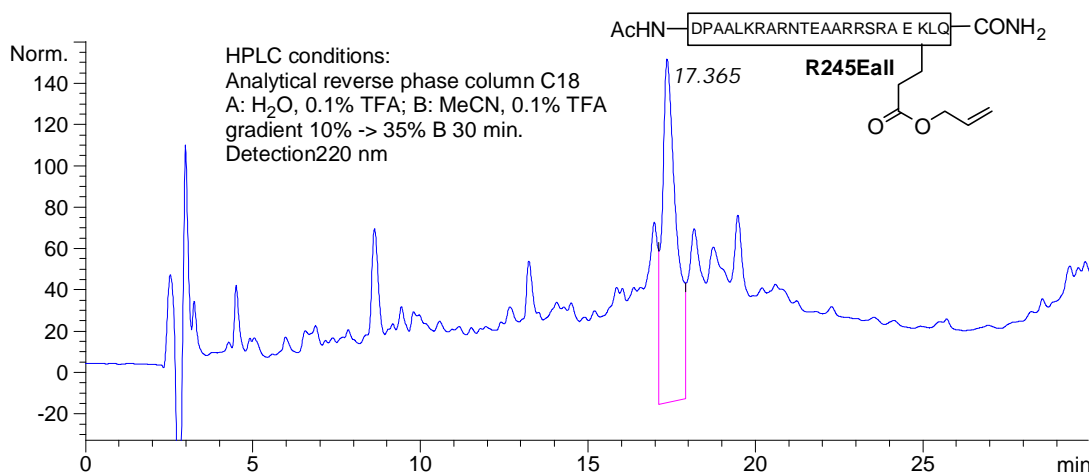


Figure 9. HPLC of peptide R245Eal.

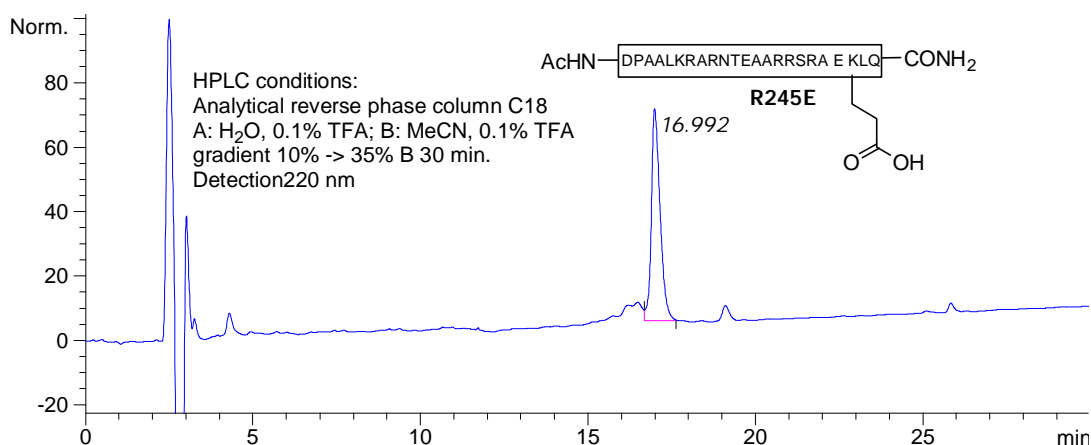
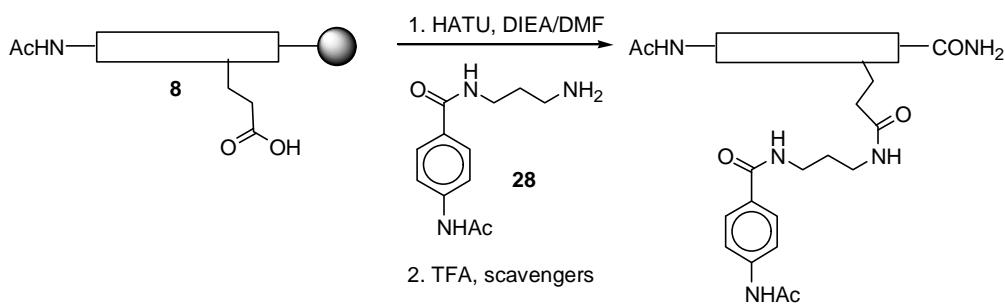


Figure 10. HPLC of peptide R245E.

**R245E MS:** [MH<sup>+</sup>] m/z calculated for C<sub>109</sub>H<sub>194</sub>N<sub>43</sub>O<sub>34</sub> 2649.4, observed 2650.0.

## Coupling of peptide R245E with 28



Resin with peptide **8** (15 mg) placed in an *Eppendorff* vial was treated with 1 mL of DMF during 1 h to ensure correct resin swelling. Resin was filtrated and DIEA was added (100  $\mu$ L, 0.5M in DMF). HATU (2.1 mg in 100  $\mu$ L of the same solution DIEA 0.5 M in DMF) was added and the mixture shaken for 5 minutes. Over the resulting activated acid, amine **28** was added (12 mg in 100  $\mu$ L of the same solution DIEA 0.5 M in DMF) and the mixture shaken for 3 h at room temperature. The resin was then filtrated and washed with DMF (3 x 0.6 mL x 5 min), *i*PrOH (1 x 0.6 mL x 5 min), finally with Et<sub>2</sub>O and dried under Ar.

The cleavage/deprotection of the obtained resin was performed using standard conditions previously described, The residue was purified by RP-HPLC to give a white powder identified as the desired product **R245EABA** by MS (ES).

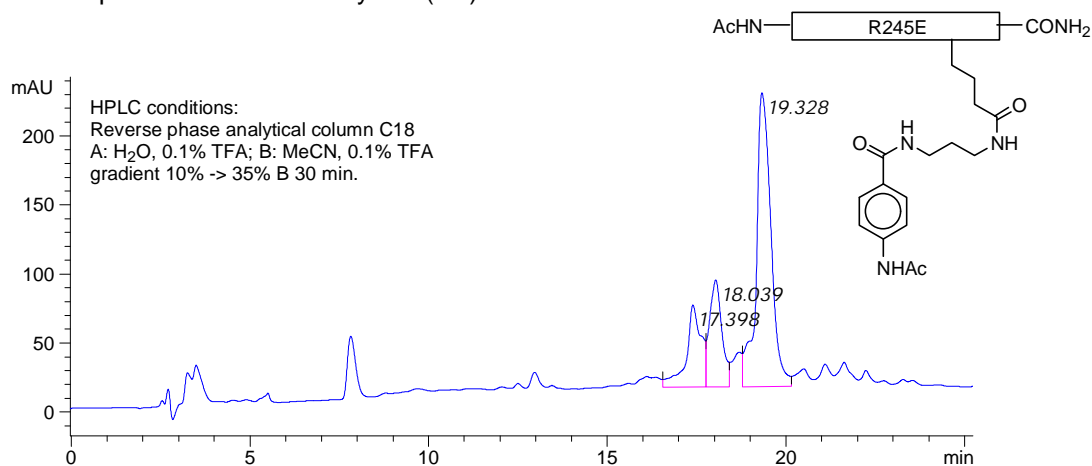


Figure 11. Coupling between **R245E** and amine **28**.

## Supporting Information

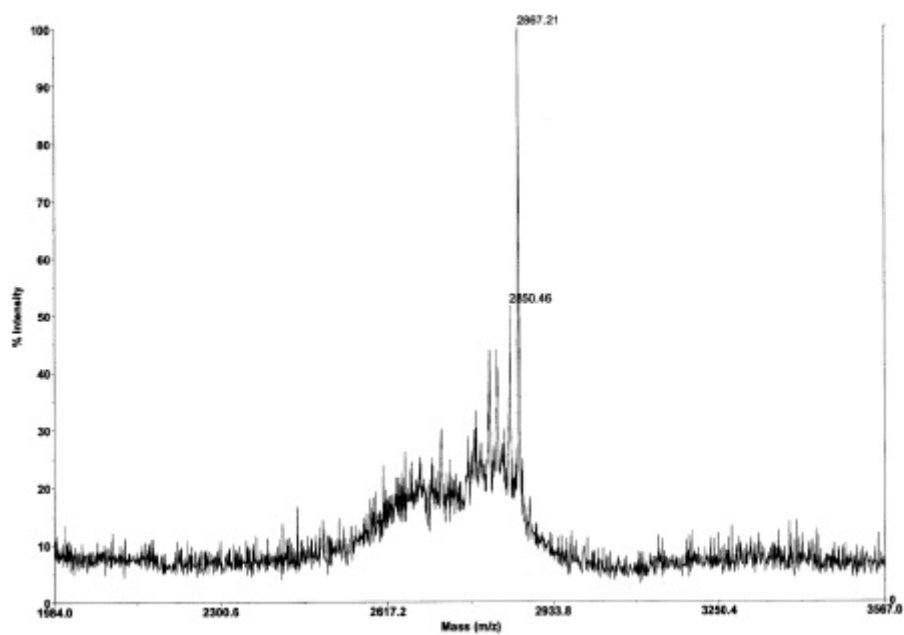
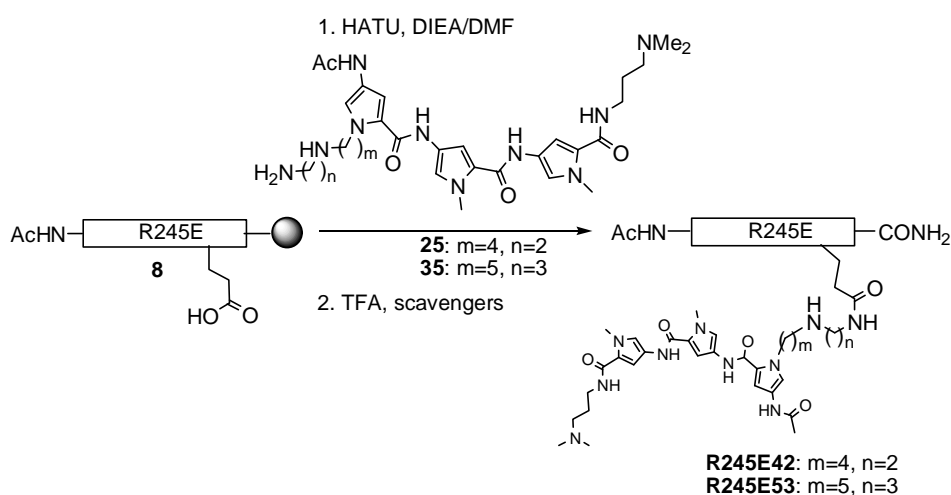


Figure 12. MS of peptide R245EABA.

**MS(ES):**  $[MH^+]$  m/z calcd. 2865.7, observed 2867.2.

Coupling of peptide R245E with tripyrrole **25** and **35**.

Coupling procedure of tripyrrole **25** and **35** with peptide **R245E** was identical in both cases, as example, we report the experimental procedure followed with tripyrrole **35**.

Over 20 mg of **R245E** bound to the resin placed in an *Eppendorf*, DMF was added (1 mL) and the mixture was shaken for 1 h to ensure resin swelling. DMF was removed and HATU (3.5 mg in 100  $\mu$ L DIEA 0.5 M in DMF) was added. The resulting mixture was shaken for 5 min. and tripyrrole **35** (25 mg in 200  $\mu$ L DIEA 0.5 M in DMF) was added over the resin. The reaction mixture was shaken for 2 hours and then washed with DMF (3 x 0.6 mL x 5 min), *i*PrOH (1 x 0.6 mL x 5 min) and finally Et<sub>2</sub>O. Cleavage/deprotection with standard conditions afforded a major product that was purified by RP-HPLC. MS analysis confirmed that product as the desired product **R245E53**. Yields were similar in both cases around 40-50%.

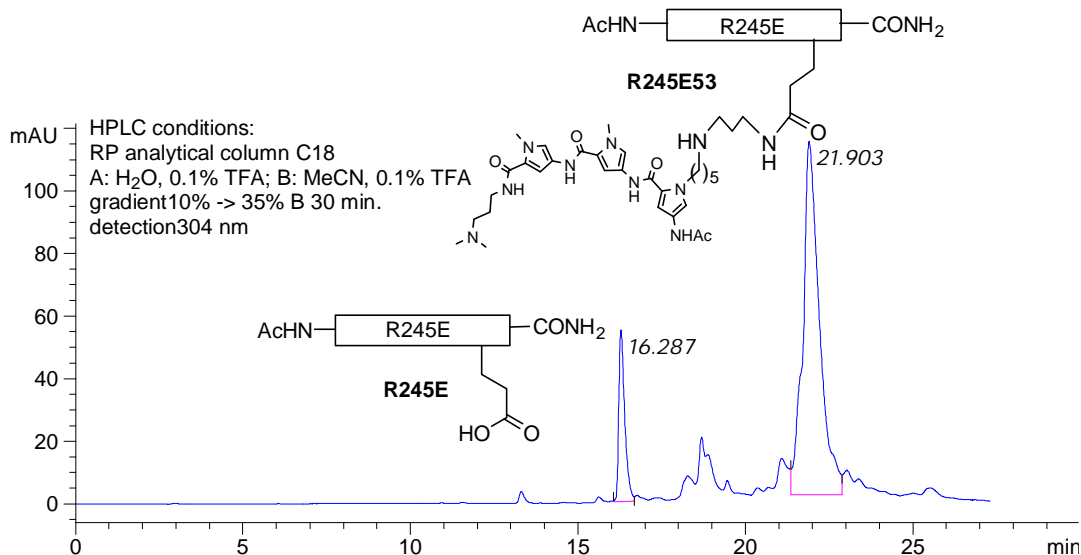


Figure 13. HPLC chromatogram of cleavage mixture.

**R245E42 MS(ES)**: [MH<sup>+</sup>] m/z calcd. for C<sub>139</sub>H<sub>238</sub>N<sub>53</sub>O<sub>37</sub> 3241.8, observed 3243.4.

**R245E53 MS(ES)**: [MH<sup>+</sup>] m/z calcd. for C<sub>141</sub>H<sub>242</sub>N<sub>53</sub>O<sub>37</sub> 3270.8, observed 3271.4.

## Supporting Information

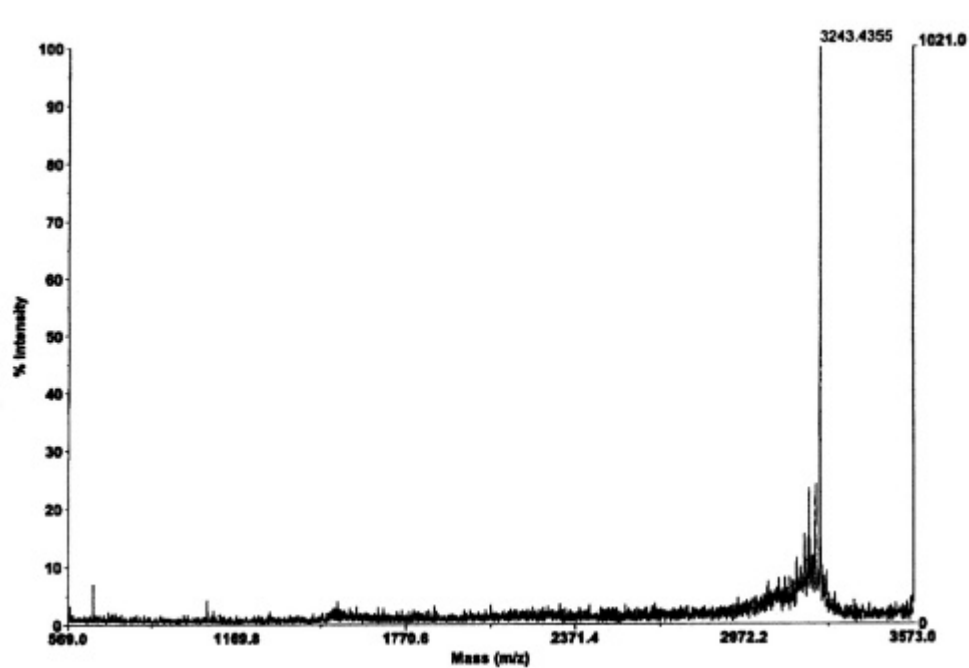


Figure 14. MS (ES) of peptide hybrid R245E42.

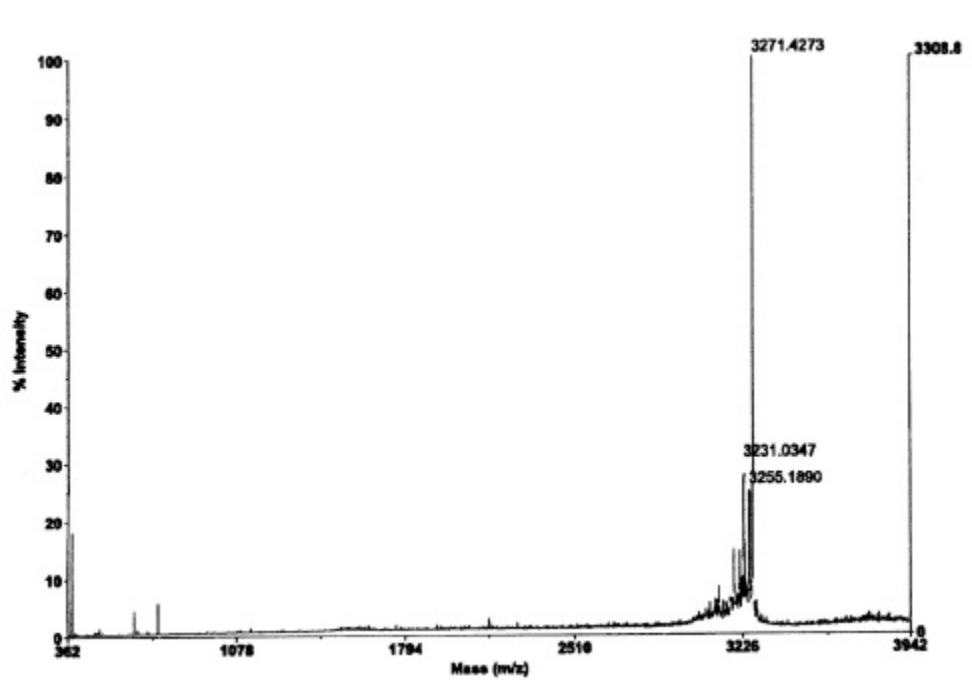


Figure 15. MS (ES) of peptide hybrid R245E53.