



Supporting Information

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Unprecedented Enantioselective Copper Catalyzed Conjugate Addition to Trisubstituted Cyclohexenones. Construction of Chiral Quaternary Centers.

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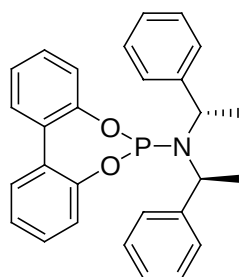
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General Procedures. ^1H (500 or 400 MHz) and ^{13}C (125 or 100 MHz) NMR spectra were recorded in CDCl_3 , and chemical shift (δ) are given in ppm relative to residual CHCl_3 . Evolution of reaction was followed by GC-MS Hewlett Packard (EI mode) HP6890-5973. Optical rotations were measured at 20°C in a 10 cm cell in the stated solvent; $[\alpha]_D$ values are given in $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$ (concentration c given as g/100 mL). Enantiomeric excesses were determined by chiral-GC (capillary column, 10 psi H_2). Temperature programs are described as follows: initial temperature ($^\circ\text{C}$) – initial time (min) – temperature gradient ($^\circ\text{C}/\text{min}$) – final temperature ($^\circ\text{C}$); retention time (R_T) are given in min. Flash chromatography were performed using silica gel 32-63 μm , 60 $\text{\AA}$ and a pentane-diethylether mixture as eluant.

All reactions were conducted under argon atmosphere. THF and diethylether were distilled from sodium-benzophenone ketyl under nitrogen. Substituted cyclohexenones (**3**, **4**, **7**, **9** and **11**) were synthesized by the reaction of corresponding Grignard reagent with 3-ethoxycyclohex-2-en-1-one (commercially available) in THF. All Grignard reagents were synthesized in THF by addition of the corresponding bromide onto magnesium. 3-methylcyclohex-2-en-1-one (**1**) (Aldrich), trimethylaluminium 2.0M in heptane (Fluka or Aldrich), triethylaluminium 0.9M in hexane (Fluka) and Copperthiophene carboxylate (FrontierScientific) were purchased and used without any further purification.

Typical procedure for ligand (L1 to L11) synthesis:

To a stirred mixture of Et₃N (111.1 mmol, 15.5 mL) and PCl₃ (22.2 mmol, 1.9 mL) at 0°C, a solution of amine (22.2 mmol) in THF (10 mL) was added and the reaction mixture was stirred for 3h at room temperature. Biphenol or binaphthol (22.2 mmol) in a solution of THF (5 mL) was slowly added to the reaction mixture at 0°C and then the suspension was stirred at RT overnight. The suspension was diluted in toluene (8 mL) and filtered on neutral alumina, the solution was concentrated and purified by flash chromatography through neutral alumina using dry toluene as eluent, to give the pure ligand as a white solid or a colorless oil.



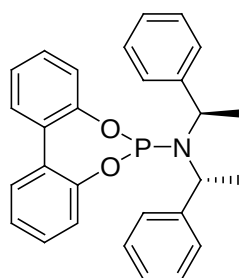
Ligand **L1** : see ref. [1].

¹H NMR (400 MHz, CDCl₃) : 7.49-7.45 (m, 2H), 7.38-7.10 (m, 16H), 4.60 (q, 1H, J = 7.0 Hz), 4.58 (q, 1H, J = 7.0 Hz), 1.73 (d, 6H, J = 7 Hz).

¹³C NMR (100 MHz, CDCl₃) : 152.0, 151.9, 151.1, 151.0, 143.0, 131.2, 130.0, 129.8, 129.1, 129.0, 128.9, 128.2, 127.9, 127.7, 126.6, 125.3, 124.0, 122.5, 122.0, 52.7, 52.6, 22.2.

³¹P (162 MHz, CDCl₃) : 147.0.

[α]_D : - 236 (c = 3.0 in toluene).



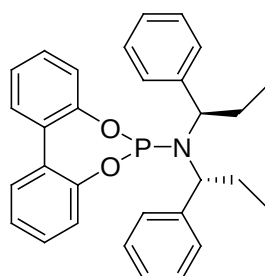
Ligand **L2** : see ref. [1].

¹H NMR (400 MHz, CDCl₃) : 7.49-7.45 (m, 2H), 7.38-7.10 (m, 16H), 4.60 (q, 1H, J = 7.0 Hz), 4.58 (q, 1H, J = 7.0 Hz), 1.73 (d, 6H, J = 7 Hz).

¹³C NMR (100 MHz, CDCl₃) : 152.0, 151.9, 151.1, 151.0, 143.0, 131.2, 130.0, 129.8, 129.1, 129.0, 128.9, 128.2, 127.9, 127.7, 126.6, 125.3, 124.0, 122.5, 122.0, 52.7, 52.6, 22.2.

³¹P (162 MHz, CDCl₃) : 147.0.

[α]_D : + 236 (c = 3.0, toluene).



Ligand **L3** : see ref. [1]

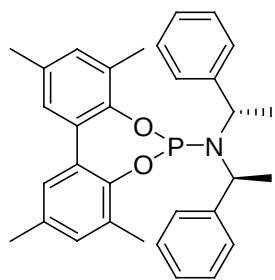
¹H NMR (400 MHz, CDCl₃) : 7.51-7.54 (m, 2H), 7.42-7.36 (m, 2H), 7.32-7.19 (m, 4H), 7.07-7.00 (m, 10H), 4.28 (t, 1H, J = 10.3 Hz), 4.27 (t, 1H, J = 10.3 Hz), 2.32-2.24 (m, 2H), 2.17-2.10 (m, 2H), 0.83 (t, 6H, J = 7.3 Hz).

¹³C NMR (100 MHz, CDCl₃) : 151.9, 151.8, 150.9, 141.0, 131.4, 129.9, 129.7, 129.1, 129.0, 128.3, 127.5, 126.2, 124.6, 123.7, 122.5,

121.7, 59.8, 59.7, 29.6, 28.3, 11.6.

³¹P (162 MHz, CDCl₃) : 144.4.

[α]_D : + 231 (c = 3.5, toluene).



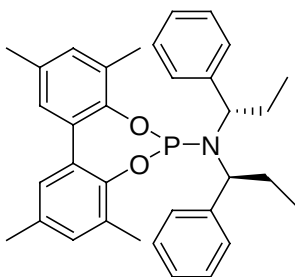
Ligand **L4** : see ref. [1]

^1H NMR (400 MHz, CDCl_3) : 7.30-6.98 (m, 14H), 4.78-4.58 (m, 2H), 2.47 (s, 3H), 2.35 (s, 3H), 2.33 (s, 3H), 2.09 (s, 3H), 1.71 (d, 6H, $J = 7\text{Hz}$).

^{13}C NMR (100 MHz, CDCl_3) : 148.0, 147.9, 147.1, 143.5, 137.5, 133.3, 132.6, 131.0, 130.9, 130.2, 129.3, 129.0, 128.2, 128.1, 127.9, 127.8, 127.6, 126.5, 125.3, 109.6, 52.5, 20.8, 17.3, 16.3.

^{31}P (162 MHz, CDCl_3) : 144.4.

$[\alpha]_{\text{D}}$: - 221 ($c = 2.2$, toluene).



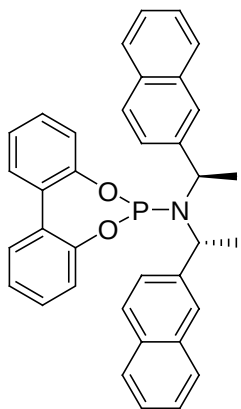
Ligand **L5** :

^1H NMR (400 MHz, CDCl_3) : 7.10-7.03 (m, 14H), 4.32 (s, 2H), 2.49 (s, 3H), 2.36 (s, 6H), 2.28 (m, 2H), 2.07 (s, 2H), 1.99 (s, 3H), 0.78 (s, 6H).

^{13}C NMR (100 MHz, CDCl_3) : 147.9, 147.3, 133.2, 132.7, 131.2, 131.1, 130.9, 130.4, 130.3, 129.3, 128.7, 128.2, 127.6, 126.4, 60.2, 23.2, 20.9, 20.8, 17.2, 16.4, 11.9.

^{31}P (162 MHz, CDCl_3) : 144.4.

$[\alpha]_{\text{D}}$: - 215 ($c = 1.06$, CHCl_3).



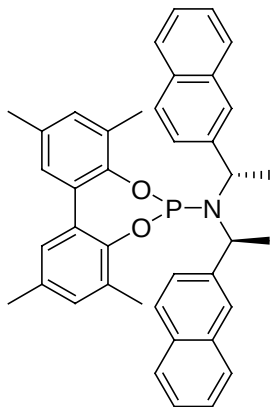
Ligand **L6** : see ref. [2]

^1H NMR (500 MHz, CDCl_3) : 7.53–7.17 (m, 22H), 4.82–4.78 (m, 2H), 1.90 (d, 6H, $J = 7.0\text{ Hz}$).

^{13}C NMR (125MHz, CDCl_3) : 151.9, 151.8, 151.0, 140.4, 132.9, 132.4, 131.2, 130.0, 129.8, 129.2, 129.0, 128.2, 127.9, 127.3, 127.0, 126.1, 125.7, 125.6, 125.3, 124.7, 124.1, 122.5, 122.1, 52.8, 52.7, 22.1.

^{31}P NMR (203 MHz; CDCl_3) : 146.3.

$[\alpha]_{\text{D}}$: + 405 ($c = 1.12$, CHCl_3).



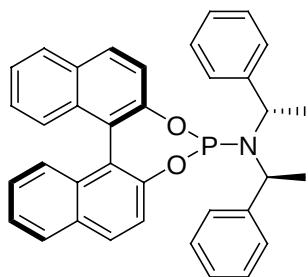
Ligand **L7** : see ref. [2]

^1H NMR (400 MHz, CDCl_3) : 7.67 (d, 2H, $J = 6,8\text{ Hz}$), 7.52–7.02 (m, 16H), 4.89 (m, 2H), 2.55 (s, 3H), 2.38 (s, 3H), 2.36 (s, 3H), 2.11 (s, 3H), 1.85 (d, 6H, $J = 4,0\text{ Hz}$).

^{13}C NMR (100 MHz, CDCl_3) : 147.5, 141.2, 133.8, 133.3, 133.1, 132.7, 131.4, 130.6, 130.5, 129.7, 128.6, 128.2, 127.7, 127.6, 127.2, 126.6, 126.0, 125.8, 53.1, 53.0, 21.2, 17.8, 16.9.

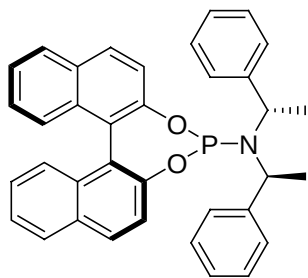
^{31}P NMR (162MHz, CDCl_3) : 142.0.

$[\alpha]_{\text{D}}$: - 435 ($c = 1.0$, CHCl_3).



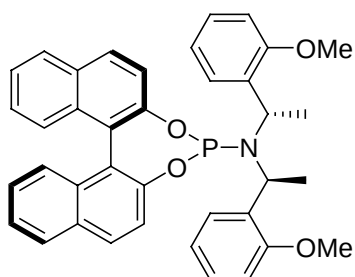
Ligand **L8** : see ref. [3]

All analysis are in accordance with reported data ^[3].



Ligand **L9** : see ref. [3]

All analysis are in accordance with reported data ^[3].



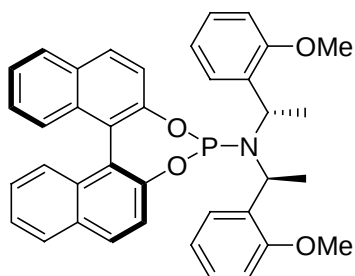
Ligand **L10** : see ref. [4]

¹H NMR (400 MHz, CDCl₃) : 8.05-5.03 (m, 20H), 5.07 (dq, 2H, J₁ = 1.04 Hz, J₂ = 7.08 Hz), 3.52 (s, 6H), 1.61 (d, 6H, J = 7.08 Hz).

¹³C NMR (100 MHz, CDCl₃) : 155.9, 150.7, 150.16, 133.0, 132.3, 131.4, 130.5, 129.5, 128.4, 128.2, 127.5, 127.4, 127.3, 127.2, 126.0, 125.8, 124.7, 124.4, 124.4, 124.3, 122.8, 122.5, 121.8, 119.4, 109.3, 65.9, 54.6, 48.3, 48.2, 22.2, 15.3.

³¹P NMR (162 MHz, CDCl₃) : 152.15.

[α]_D = - 272.2 (c 1.0, CHCl₃).



Ligand **L11** : see ref. [4]

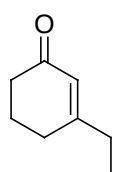
¹H NMR (400 MHz, CDCl₃) : 8.04-6.52 (m, 20H), 4.99 (dq, 2H, J₁ = 1.24 Hz, J₂ = 7.08 Hz), 3.58 (s, 6H), 1.55 (d, 6H, J = 7.08 Hz).

¹³C NMR (100 MHz, CDCl₃) : 156.1, 149.9, 132.8, 132.5, 131.4, 130.3, 129.2, 128.3, 128.0, 127.7, 127.6, 127.4, 127.3, 125.9, 125.6, 124.7, 124.5, 124.2, 122.7, 1212.5, 119.6, 109.2, 54.6, 50.3, 50.2, 22.6, 22.5.

³¹P NMR (162 MHz, CDCl₃) : 155.25.

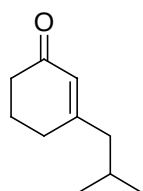
[α]_D = + 144.3 (c 1.1, CHCl₃).

Typical procedure for 3-substituted cyclohex-2-en-1-ones synthesis. A flame-dried flask was charged with Grignard reagent (2.0 eq.) and cooled to 0°C. The ethoxycyclohex-2-en-1-one (50 mmol) in THF (40mL) was added dropwise. Once the addition was complete the reaction mixture was left at room temperature until complete disappearance of the starting material. The reaction was hydrolyzed by addition of aqueous sulfuric acid (5% w/w). Diethyl ether (50 mL) was added and the aqueous phase was separated and extracted further with diethyl ether (3 x 20mL). The combined organic fractions were washed with NaHCO₃, brine and water, dried over sodium sulfate, filtered and concentrated *in vacuo*. The oily residue was purified by Kugelrohr distillation under reduced pressure.



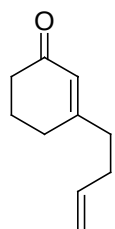
3-ethylcyclohex-2-enone (3) ^[5]:

¹H NMR (500 MHz, CDCl₃) : 5.87 (t, 1H, J = 1.4 Hz), 2.36 (t, 2H, J = 3.3 Hz), 2.29 (t, 2H, J = 5.7 Hz), 2.27-2.22 (m, 2H), 1.99 (m, 2H), 1.1 (t, 3H, J = 7.4 Hz).
¹³C NMR (125 MHz, CDCl₃) : 200, 167.8, 124.5, 37.4, 30.8, 29.7, 22.7, 11.2.



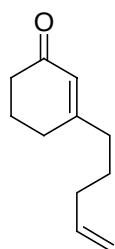
3-isobutylcyclohex-2-enone (4) ^[6]:

¹H NMR (400 MHz, CDCl₃) : 5.84 (s, 1H), 2.35 (t, 2H, J = 6.7 Hz), 2.25 (t, 2H, J = 5.9 Hz), 2.07 (d, 2H, J = 7.3 Hz), 1.97 (quint., 2H, J = 6.4 Hz), 1.87 (m, 1H), 0.9 (d, 6H, J = 6.6 Hz).
¹³C NMR (100 MHz, CDCl₃) : 199.9, 165.7, 126.9, 47.7, 37.3, 29.7, 26.4, 22.8, 22.5.



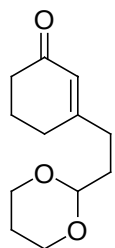
3-(3-butenyl)cyclohex-2-enone (7) ^[7]:

¹H NMR (500 MHz, CDCl₃) : 5.80 (s, 1H), 5.75-5.67 (m, 1H), 4.98 (dd, 1H, J₁ = 17.1 Hz, J₂ = 1.5 Hz), 4.92 (dd, 1H, J₁ = 9.4 Hz, J₂ = 1.6 Hz), 2.29-2.19 (m, 8H), 1.94-1.89 (m, 2H).
¹³C NMR (125 MHz, CDCl₃) : 199.9, 165.6, 137.1, 126.1, 115.7, 37.5, 37.4, 31.1, 29.9, 22.8.



3-(3-pentenyl)cyclohex-2-enone (9) ^[8]:

¹H NMR (400 MHz, CDCl₃) : 5.82 (s, 1H), 5.78-5.68 (m, 1H), 4.99-4.91 (m, 2H), 2.31-1.90 (m, 10H), 1.59-1.51 (m, 2H).
¹³C NMR (100 MHz, CDCl₃) : 200.0, 166.5, 138.0, 125.9, 115.5, 37.5, 37.5, 33.4, 29.9, 26.2, 22.9.

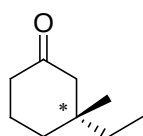


3-[2-(1,3-dioxan-2-yl)ethyl]cyclohex-2-enone (11) ^[8]:

¹H NMR (400 MHz, CDCl₃) : 5.70 (s, 1H), 4.39 (t, 1H, J = 5.0Hz), 3.93 (dd, 2H, J₁ = 10.6Hz, J₂ = 5.0Hz), 3.60 (td, 2H, J₁ = 12.4Hz, J₂ = 2.3Hz), 2.21-2.14 (m, 6H), 1.86-1.80 (m, 2H), 1.65-1.60 (m, 2H), 1.20 (dt, 2H, J₁ = 13.4Hz, J₂ = 1.2Hz).

¹³C NMR (100 MHz, CDCl₃) : 199.6, 165.8, 125.4, 101.0, 66.7, 37.2, 32.1, 32.0, 29.6, 25.6, 22.6.

Typical procedure for enantioselective copper catalyzed conjugate addition with trialkylaluminium reagent. A flame-dried Schlenk tube was charged with copper salt (2.0 mol%) and the chiral ligand (4.0 mol%). Solvent (2 mL) was added and the mixture was stirred at room temperature for 30 min before being cooled to -30°C in an ethanol cold bath. The trialkylaluminium (2.0 eq. for Me_3Al , 1.0 mL of a 2M solution in heptane, or 1.4 eq. for Et_3Al , 1.6 mL of a 0.9M in hexane) was introduced dropwise at such rate that the temperature did not rise above -30°C , and the reaction mixture was stirred at -30°C for a further 5 min. Then the Michael acceptor (1.0 eq., 1.0 mmol) in diethylether or THF (0.5 mL) was added dropwise. Once the addition was complete the reaction mixture was left at -30°C overnight. The reaction was hydrolyzed by addition of MeOH at -30°C , followed by NH_4Cl sat. or HCl 2N (3mL) at room temperature. Diethyl ether (10 mL) was added and the aqueous layer was separated and extracted further with diethyl ether (3 x 3mL). The combined organic fractions were washed with brine (5 mL), dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The oily residue was purified by flash column chromatography (eluent = pentane : diethylether) to yield the 1,4-adduct. Gas Chromatography on a chiral stationary phase indicated the enantiomeric excess of 1-4adduct.

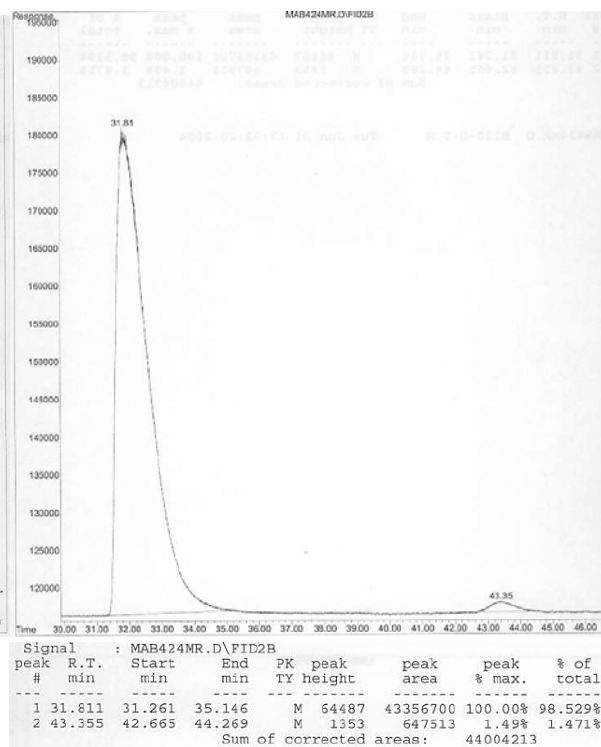
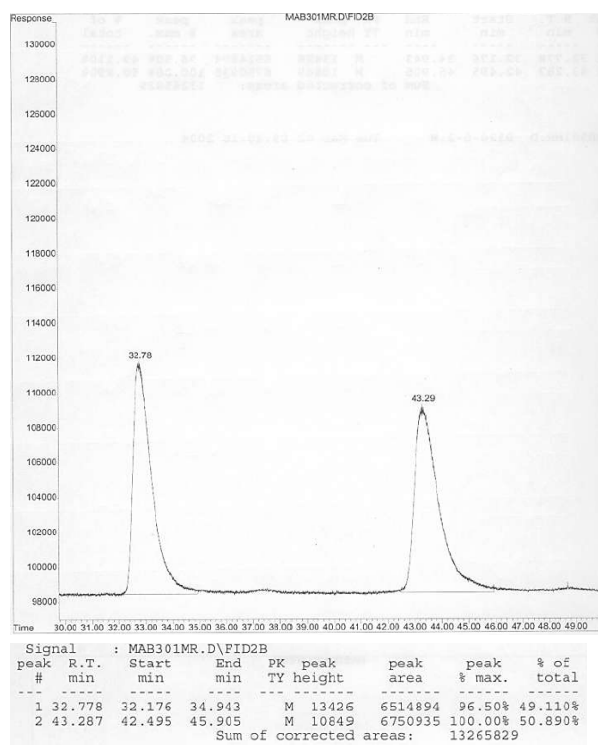


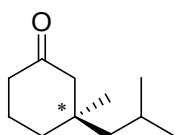
3-ethyl-3-methylcyclohexanone (2) ^[9]:

^1H NMR (400 MHz, CDCl_3) : 2.26 (t, 2H, $J = 6.6$ Hz), 2.16 (d of AB sys., 1H, $J = 13.6$ Hz), 2.08 (d of AB sys., 1H, $J = 13.6$ Hz), 1.88-1.81 (m, 2H), 1.64-1.48 (2m, 2H), 1.33 (q, 2H, $J = 7.3$ Hz), 0.88 (s, 3H), 0.83 (t, 3H, $J = 7.6$ Hz).

^{13}C NMR (100 MHz, CDCl_3) : 212.8, 53.7, 41.4, 39.0, 35.7, 34.3, 24.7, 22.5, 8.1.

$[\alpha]_{\text{D}} : +5.45$ (CHCl_3 , $c = 1.64$, $ee = 92\%$ *R*). Absolute configuration was assigned in analogy with **13**. Enantiomeric excess was measured by chiral GC (lipodex E, isotherm 60°C , $R_{\text{t}1} = 32.8$ (*R*), $R_{\text{t}2} = 43.3$ (*S*)).





3-ethyl-3-methylcyclohexanone (5):

The crude mixture was purified by flash chromatography (pentane : Et₂O, 80:20, R_f = 0.53, anisaldehyde as revelator) to afford the title compound as a colorless oil.

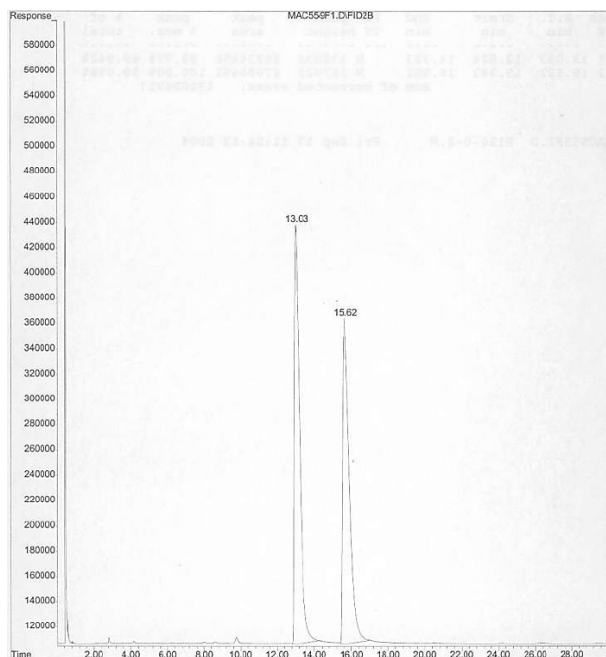
¹H NMR (400 MHz, CDCl₃) : 2.29-2.23 (m, 2H), 2.19 (d of AB sys., 1H, J = 13.4 Hz), 2.09 (d of AB sys., 1H, J = 13.4 Hz), 1.93-1.51 (3m, 5H), 1.19 (t, 2H, J = 5.0 Hz), 0.93 (s, 3H), 0.91 (dd, 6H, J₁ = 6.6 Hz, J₂ = 1.0 Hz).

¹³C NMR (100 MHz, CDCl₃) : 212.7, 54.6, 51.2, 41.3, 39.7, 36.8, 25.8, 25.7, 25.6, 24.2, 22.5.
IR (neat, cm⁻¹) : 2956, 1716, 1467.

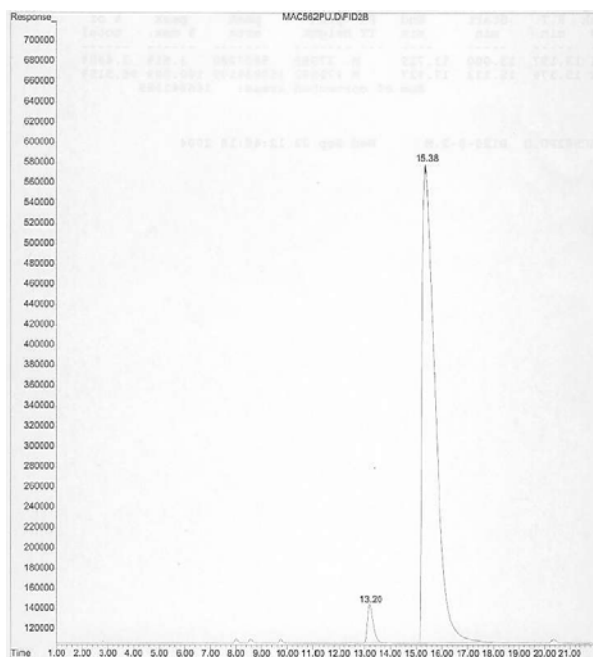
MS (IE) : 153(2), 125 (9), 112 (9), 111 (100), 110 (3), 107 (2), 98 (2), 97 (9), 95 (7), 93 (2), 85 (1), 84 (2), 83 (22), 82 (4), 81 (2), 79 (2), 77 (1), 71 (2), 70 (6), 69 (25), 68 (3), 67 (6), 65 (1), 58 (1), 57 (5), 56 (15), 55 (98), 54 (2), 53 (5).

HRMS (ESI-MS) : [M+Na]⁺ found 191.1408390, calcd. for C₁₁H₂₀ONa : 191.1406364.

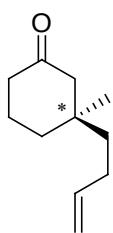
[α]_D : -5.25 (CHCl₃, c = 1.73, ee = 93% R. Absolute configuration was assigned in analogy with **13**). Enantiomeric excess was measured by chiral GC (lipodex E, isotherm 80°C, Rt₁ = 13.0 (S), Rt₂ = 15.6 (R)).



Signal : MAC559F1.D\FID2B								
peak #	R.T. min	Start min	End min	PK TY	peak height	peak area	peak % max.	% of total
1	13.033	12.824	14.323	M	332036	66934456	99.77%	49.942%
2	15.622	15.383	16.952	M	257923	67090461	100.00%	50.058%
Sum of corrected areas: 134024917								



Signal : MAC562PU.D\FID2B								
peak #	R.T. min	Start min	End min	PK TY	peak height	peak area	peak % max.	% of total
1	13.197	13.000	13.719	M	37989	5807280	3.61%	3.485%
2	15.376	15.133	17.927	M	472582	160834109	100.00%	96.515%
Sum of corrected areas: 166641389								

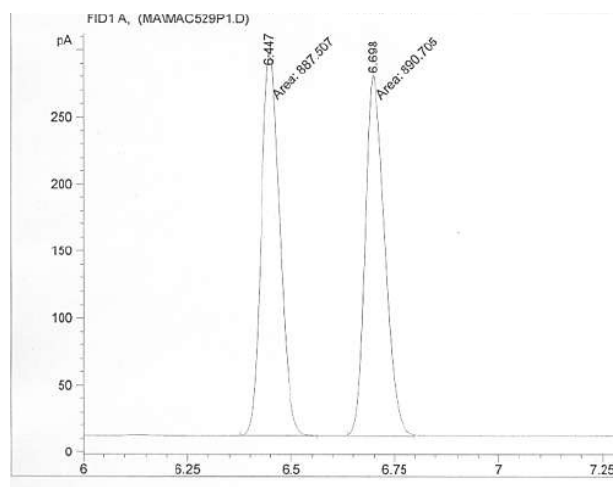


3-(3-butenyl)-3-methylcyclohexanone (8) [7]:

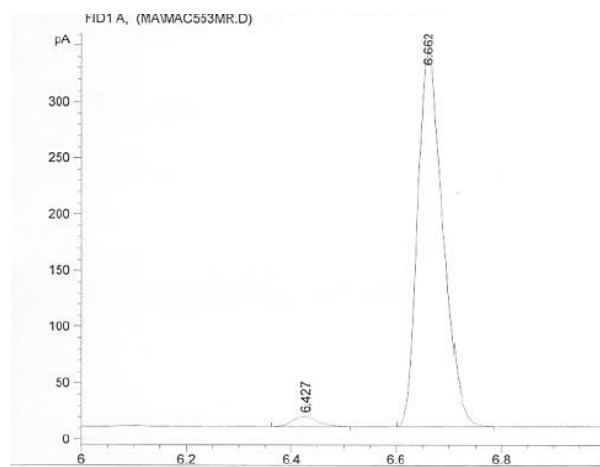
¹H NMR (500 MHz, CDCl₃) : 5.83-5.75 (m, 1H), 5.00 (dd, 1H, J₁ = 17.0 Hz, J₂ = 1.7 Hz), 4.93 (dd, 1H, J₁ = 10.3 Hz, J₂ = 1.3 Hz), 2.32-2.24 (m, 2H), 2.20 (d of AB sys., 1H, J = 13.4 Hz), 2.12 (d of AB sys., 1H, J = 13.4 Hz), 2.05-1.98 (m, 2H), 1.90-1.83 (m, 2H), 1.66-1.53 (m, 2H), 1.39-1.34 (m, 2H), 0.93 (s, 3H).

¹³C NMR (125 MHz, CDCl₃) : 212.5, 139.1, 114.7, 54.0, 41.3, 41.2, 38.9, 36.2, 28.2, 25.2, 22.4.

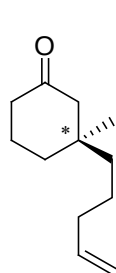
$[\alpha]_D$: +0.90 (CHCl₃, c = 1.71, ee = 93% *R*. Absolute configuration was assigned in analogy with **13**). Enantiomeric excess was measured by chiral GC (Hydrodex B-3P, isotherm 130°C, *R*_{t1} = 6.4 (*S*), *R*_{t2} = 6.7 (*R*)).



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.447	MM	0.0517	887.50702	285.96204	49.91007
2	6.698	MM	0.0553	890.70526	268.21765	50.08993
Totals :				1778.21228	554.17969	



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.427	BB	0.0477	29.00474	8.70781	2.52924
2	6.662	BB	0.0496	1117.77417	332.44604	97.47076
Totals :				1146.77891	341.15385	

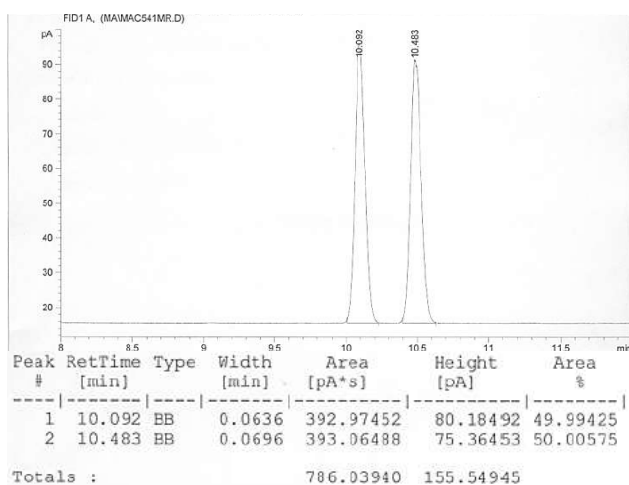


3-(3-pentenyl)-3-methylcyclohexanone (10) ^[10]:

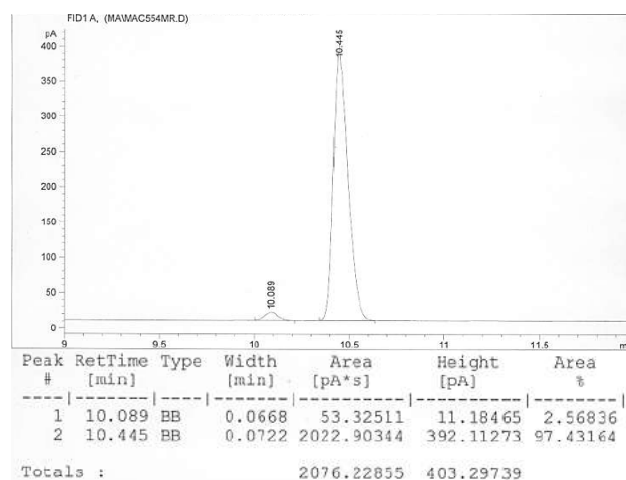
¹H NMR (400 MHz, CDCl₃) : 5.82-5.72 (m, 1H), 5.00-4.91 (m, 2H), 2.25 (t, 2H, J = 6.8 Hz), 2.18 (d of AB sys., 1H, J = 13.4 Hz), 2.09 (d of AB sys., 1H, J = 13.4 Hz), 2.03-1.98 (m, 2H), 1.87-1.81 (m, 2H), 1.65-1.49 (2m, 2H), 1.36-1.23 (m, 4H), 0.90 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) : 212.6, 138.9, 115.0, 54.1, 41.3, 38.9, 36.1, 34.6, 25.4, 23.0, 22.4.

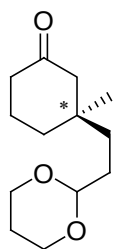
$[\alpha]_D$: -1.67 (CHCl₃, c = 1.70, ee = 93% *S*. Absolute configuration was assigned in analogy with **13**). Enantiomeric excess was measured by chiral GC (Hydrodex B-3P, isotherm 130°C, *R*_{t1} = 10.1 (*R*), *R*_{t2} = 10.5 (*S*)).



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	10.092	BB	0.0636	392.97452	80.18492	49.99425
2	10.483	BB	0.0696	393.06488	75.36453	50.00575
Totals :				786.03940	155.54945	



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	10.089	BB	0.0668	53.32511	11.18465	2.56836
2	10.445	BB	0.0722	2022.90344	392.11273	97.43164
Totals :				2076.22855	403.29739	

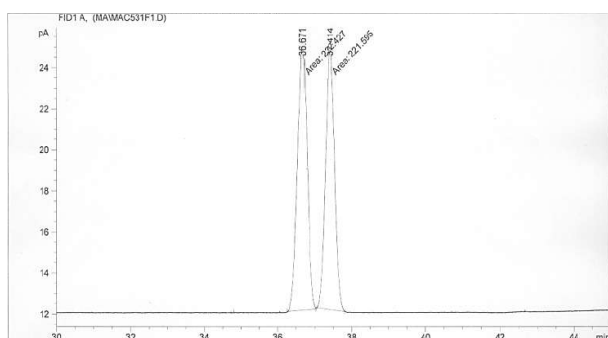


3-[2-(1,3-dioxan-2-yl)ethyl]-3-methylcyclohex-2-enone (12) ^[11, 12]:

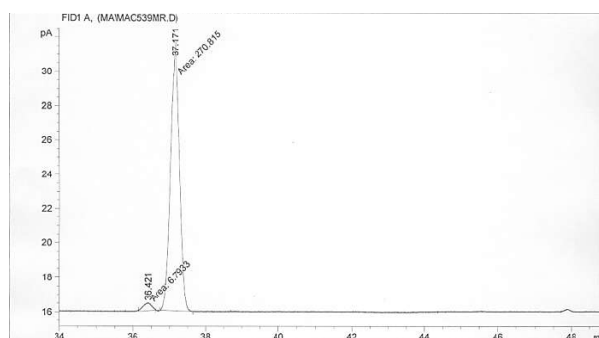
¹H NMR (500 MHz, CDCl₃) : 4.46 (t, 1H, J = 5.0 Hz), 4.08 (dd, 2H, J₁ = 10.6 Hz, J₂ = 4.9 Hz), 3.73 (tt, 2H, J₁ = 12.3 Hz, J₂ = 2.4 Hz), 2.29-2.22 (m, 2H), 2.17-2.00 (m, 3H), 1.89-1.78 (m, 2H), 1.63-1.49 (m, 4H), 1.38-1.30 (m, 3H), 0.89 (s, 3H).

¹³C NMR (125 MHz, CDCl₃) : 212.4, 102.9, 67.2, 54.1, 41.3, 38.4, 35.9, 35.8, 29.7, 26.1, 25.1, 22.4.

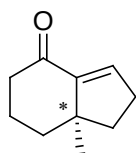
[α]_D : +0.08 (CHCl₃, c = 1.25, ee = 94% *R*. Absolute configuration was assigned in analogy with **13**). Enantiomeric excess was measured by chiral GC (Hydrodex B-3P, isotherm 130°C, Rt₁ = 36.7 (S), Rt₂ = 37.4 (R)).



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	36.671	MM	0.2894	222.42731	12.81153	50.09367
2	37.414	MM	0.2830	221.59549	13.05064	49.90633
Totals :				444.02280	25.86216	



Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	36.421	MM	0.2559	6.79330	4.42467e-1	2.44708
2	37.171	MM	0.2882	270.81500	15.66130	97.55292
Totals :				277.60830	16.10377	



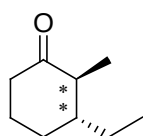
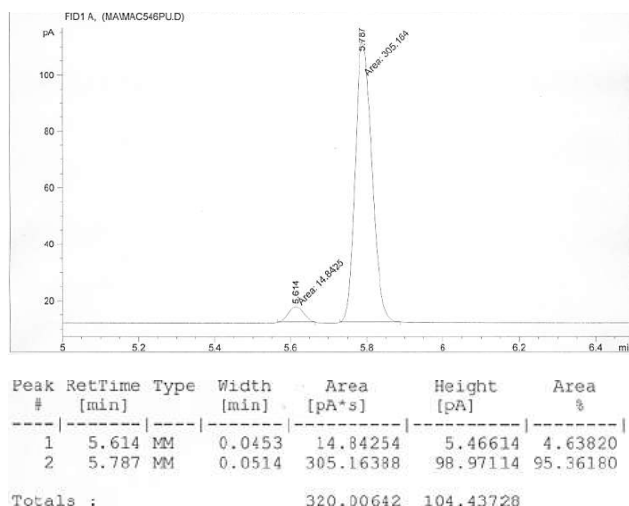
1,2,5,6,7,7a-hexahydro-7a-methylinden-4-one (13) ^[12, 13]:

According to literature references ^[12,13], the 1,4-adduct **12** (565.8 mg, 2.5 mmol) was dissolved in a 6N HCl solution in THF (10 mL) at room temperature. After being stirred at room temperature for 18 hours, the mixture was neutralized at 0°C with a sodium bicarbonate saturated solution, diluted with diethylether, washed with three portions of water, and one of brine, dried over anhydrous sodium sulfate, filtered and concentrated *in vacuo*. The crude mixture was purified by flash chromatography (pentane : diethylether) to afford 255.2 mg (68% yield) of a pale yellow oil.

¹H NMR (400 MHz, CDCl₃) : 6.43 (t, 1H, J = 3.0 Hz), 2.54-2.18 (3m, 5H), 2.03-1.79 (m, 6H), 1.57 (td, 1H, J₁ = 13.9 Hz, J₂ = 4.8 Hz), 1.07 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) : 200.8, 149.7, 136.4, 47.7, 42.5, 40.0, 38.5, 30.1, 24.2, 21.5.

[α]_D : -74.6 (CHCl₃, c = 1.53, ee = 90% *R*. Absolute configuration was attributed in comparison with literature data^[12]). Enantiomeric excess was measured by chiral GC (Hydrodex B-3P, isotherm 130°C, Rt₁ = 36.7 (S), Rt₂ = 37.4 (R)).



3-ethyl-2-methylcyclohexanone (15) ^[14]:

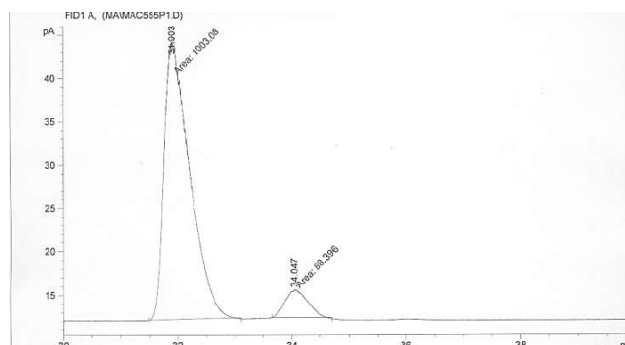
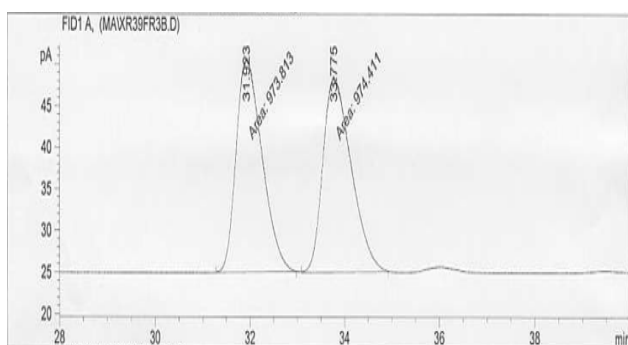
¹H NMR (500 MHz, CDCl₃) : 2.37 (m, 1H), 2.26 (m, 1H), 2.16 (m, 1H), 2.04 (m, 1H), 1.89 (m, 1H), 1.62 (m, 2H), 1.36 (m, 3H), 1.02 (d, 3H, J = 6.7 Hz), 0.90 (t, 3H, J = 7.4 Hz).

¹³C NMR (125 MHz, CDCl₃) *trans* diastereoisomer : 213.8, 49.5, 46.7, 41.5, 29.7, 26.2, 25.9, 11.8, 10.2.

¹³C NMR (125 MHz, CDCl₃) *cis* diastereoisomer : 214.8, 48.8, 43.9, 39.7, 26.5, 23.9, 21.9, 11.6, 11.3.

[α]_D : -12.7 (CHCl₃, c = 1.29, ee = 82% (2*S*, 3*R*)). Enantiomeric excess was measured by chiral GC (Hydrodex B-3P, isotherm 70°C, Rt₁ = 31.9 (2*S*, 3*R*), Rt₂ = 33.8 (2*R*, 3*S*)).

Trans-adduct after separation by flash-chromatography :



References :

- [¹] A. Alexakis, S. Rosset, J. Allamand, S. March, F. Guillen, C. Benhaim, *Synlett* **2001**, 9, 1375-1378.
- [²] A. Alexakis, D. Polet, C. Benhaim, S. Rosset, *Tetrahedron : Asymm.* **2004**, 15, 2199-2203.
- [³] (a) A.H.M. de Vries, A. Meetsma, B.L. Feringa, *Angew. Chem., Int. Ed. Engl.*, **1996**, 35, 2374-2376. (b) L.A. Arnold, R. Imbos, A. Mandoli, A.H.M. de Vries, R. Naasz, B.L. Feringa, *Tetrahedron* **2000**, 56, 2865-2878.
- [⁴] (a) K. Tissot-Kroset, D. Polet, A. Alexakis, *Angew. Chem., Int. Ed. Engl.*, **2004**, 43, 2426-2428. (b) K. Tissot-Croset, D. Polet, S. Gille, C. Hawner, A. Alexakis, *Synthesis* **2004**, 15, 2586-2590.
- [⁵] B.-D.Chong, Y. Ji, S.-S. Oh, J.D. Yang, W. Baik, S. Koo, *J. Org. Chem.* **1997**, 62, 9323-9325.
- [⁶] A.P. Kozikowski, S.H. Jung, *J. Org. Chem.* **1986**, 51, 3400-3402.
- [⁷] B. Chenera, C.P. Chuang, D.J. Hart, C.S. Lai, *J. Org. Chem.* **1992**, 57, 2018-2029.
- [⁸] J.P. Dittami, X.Y. Nie, H. Nie, H. Ramanathan, S. Breining, J. Bordner, D. Decosta, J. Kiplinger, P. Reiche, R. Ware, *J. Org. Chem.* **1991**, 56, 5572-5578.
- [⁹] J. Westermann, K. Nickisch, *Angew. Chem.* **1993**, 105, 1429-1431 (See also *Angew. Chem., Int. Ed. Engl.*, **1993**, 32, 1368-1370).
- [¹⁰] G.A. Molander, J.A. McKie, *J. Org. Chem.* **1994**, 59, 3186-3192.
- [¹¹] F. Leyendecker, F. Jesser, *Tetrahedron Lett.* **1980**, 21, 1311-1314.
- [¹²] T. Ohkubo, H. Akino, M. Asaoka, H. Takei, *Tetrahedron Lett.* **1995**, 36, 3365-3368.
- [¹³] A.S. Bal, A. Marfat, P. Helquist, *J. Org. Chem.* **1982**, 47, 5045-5050.
- [¹⁴] A. Alexakis, S. March, *J. Org. Chem.* **2002**, 67, 8753-8757.

PALAIS L PROTON CDCl3 u quest 1

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PROCNO : 1

*** Acquisition Parameters ***

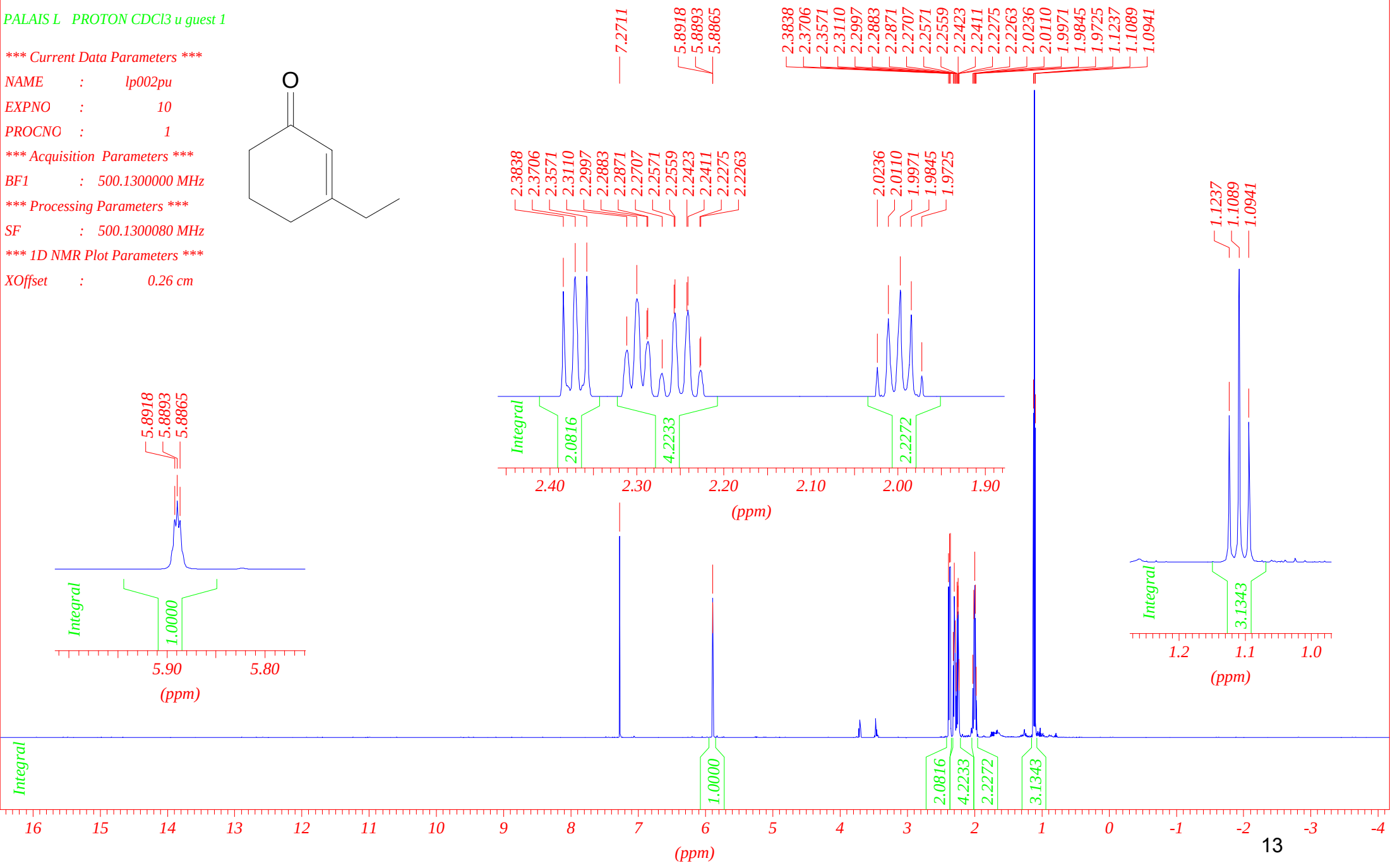
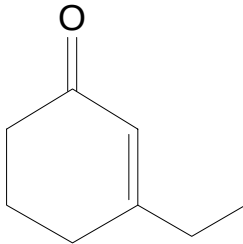
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PALAI5 L C13CPD CDCl3 u quest 1

*** Current Data Parameters ***

NAME : lp002pu

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PROCNO : 1

*** Acquisition Parameters ***

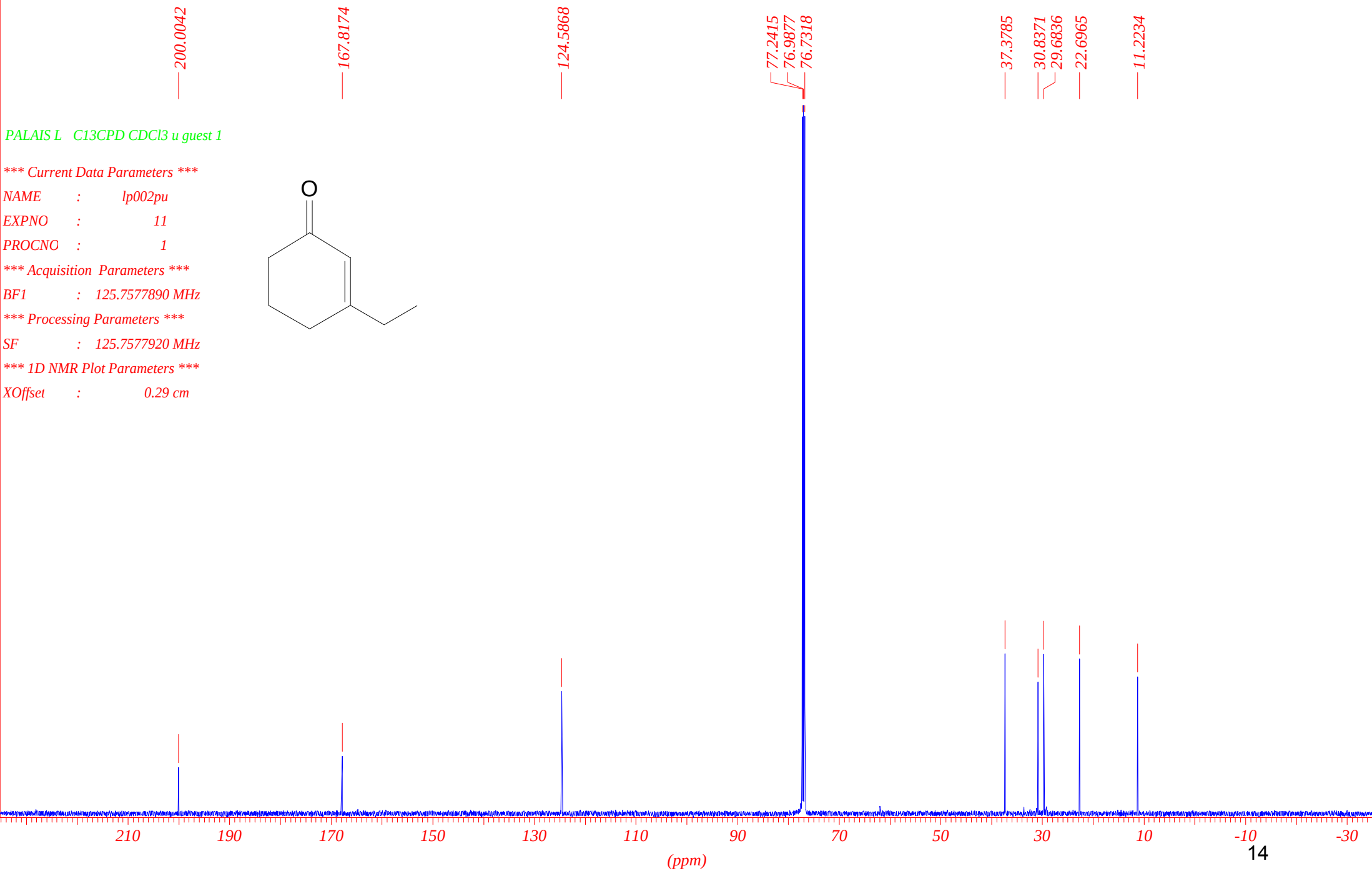
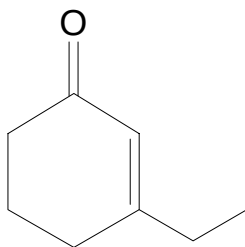
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*** Processing Parameters ***

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*** 1D NMR Plot Parameters ***

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PALAI5 L C13DEPT135 CDCl3 u gues

*** Current Data Parameters ***

NAME : lp002pu

EXPNO : 12

PROCNO : 1

*** Acquisition Parameters ***

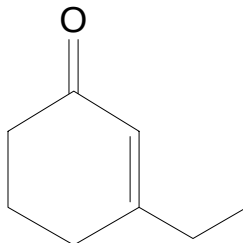
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*** Processing Parameters ***

SF : 125.7577920 MHz

*** 1D NMR Plot Parameters ***

XOffset : 0.29 cm



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37.3940

30.8506

29.6992

22.7101

11.2369

210

190

170

150

130

110

90

70

50

30

10

-10

-30

(ppm)

15

*** Current Data Parameters ***

NAME : lp020pu
EXPNO : 10
PROCNO : 1

*** Acquisition Parameters ***

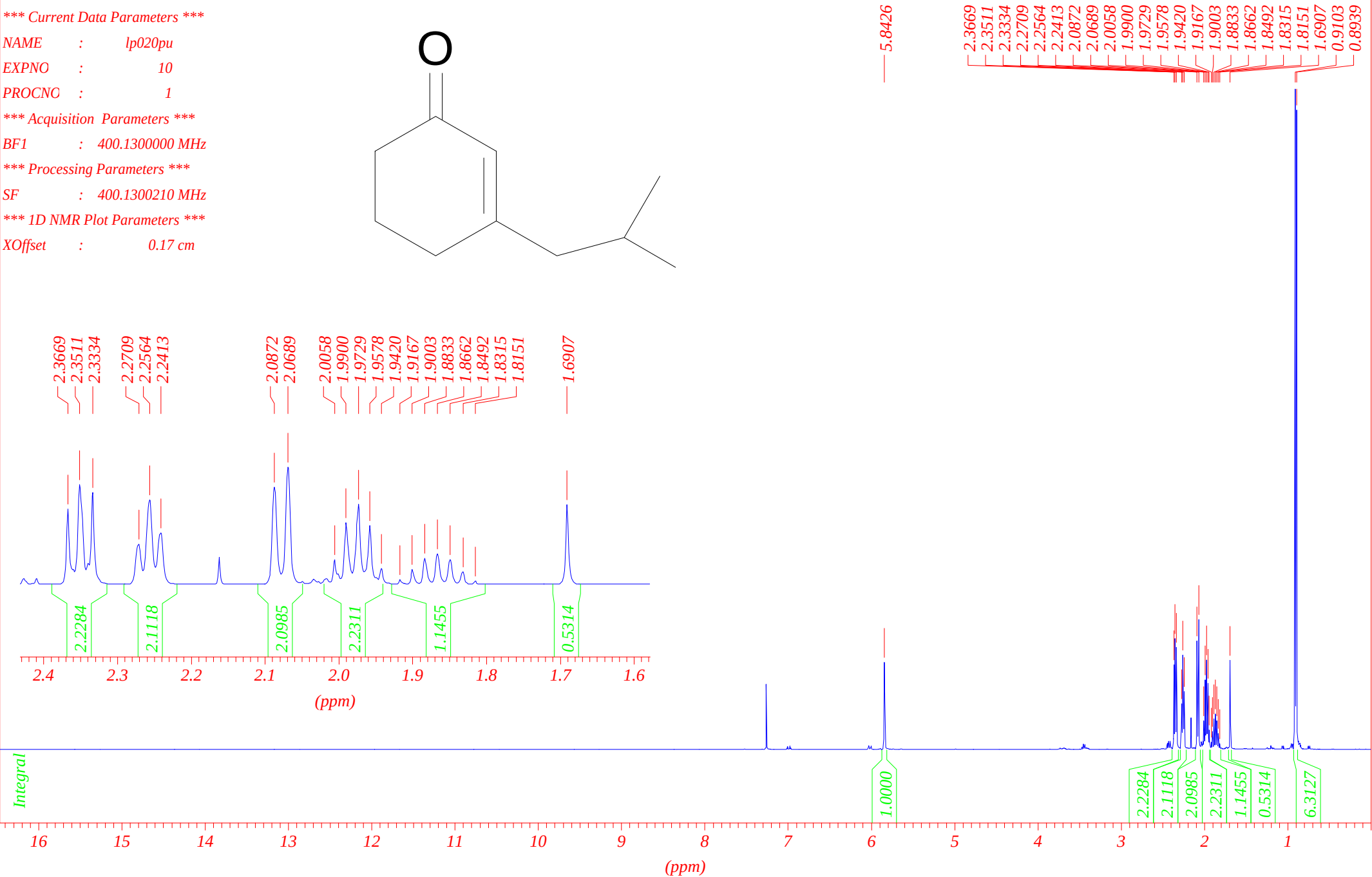
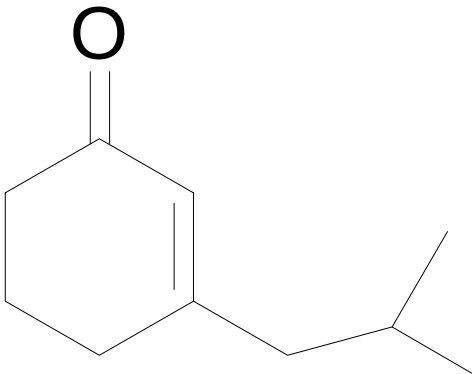
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Laetitia C13CPD CDCl3 u quest 6

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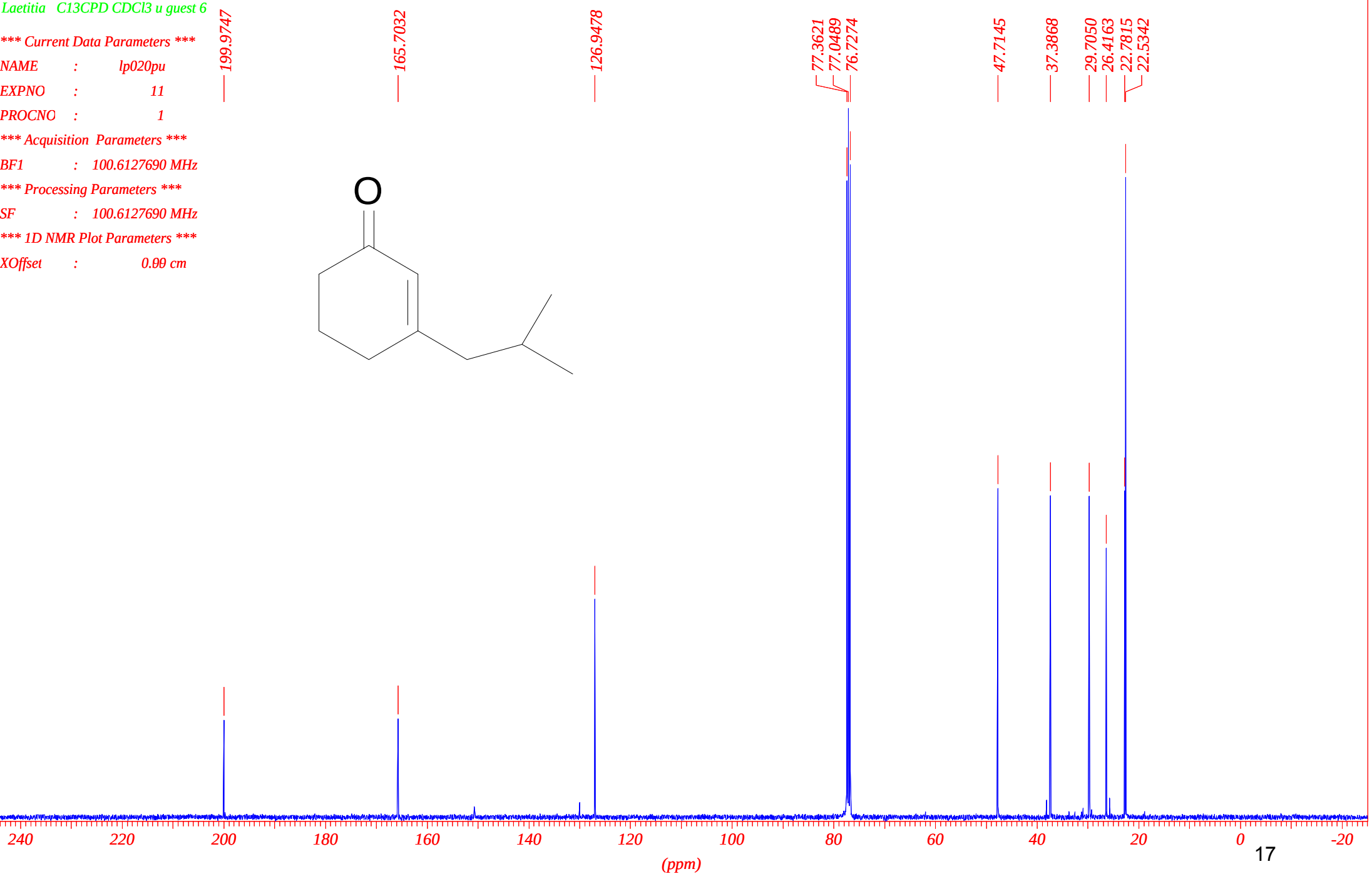
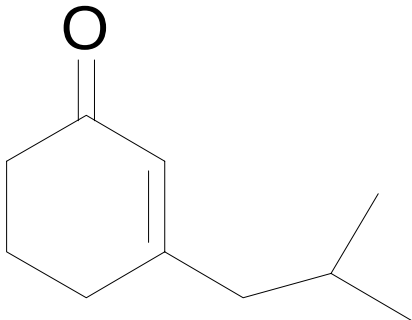
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*** Processing Parameters ***

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*** 1D NMR Plot Parameters ***

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Laetitia C13DEPT135 CDCl3 u guest t

*** Current Data Parameters ***

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*** Acquisition Parameters ***

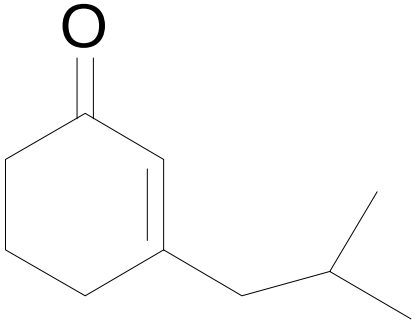
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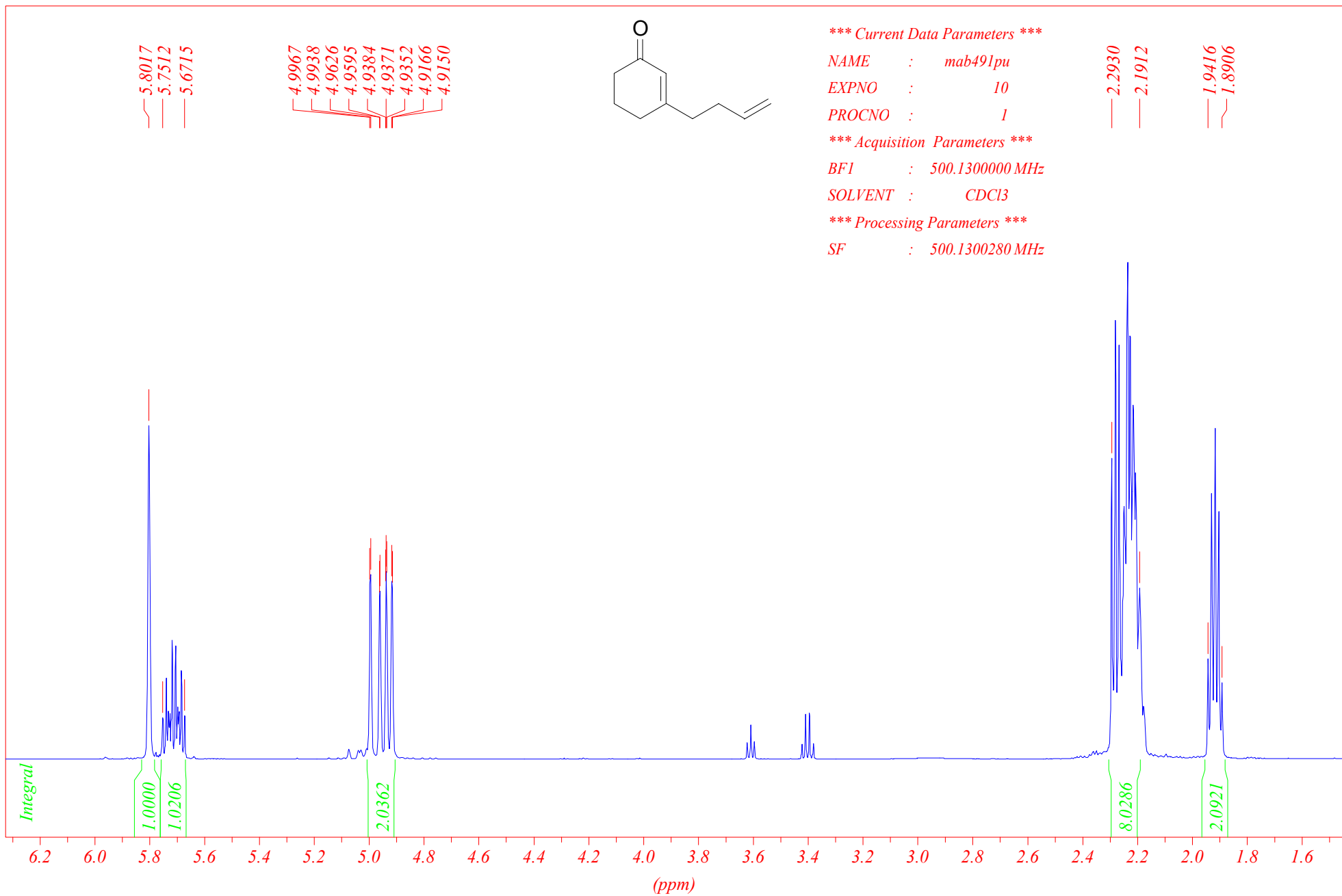
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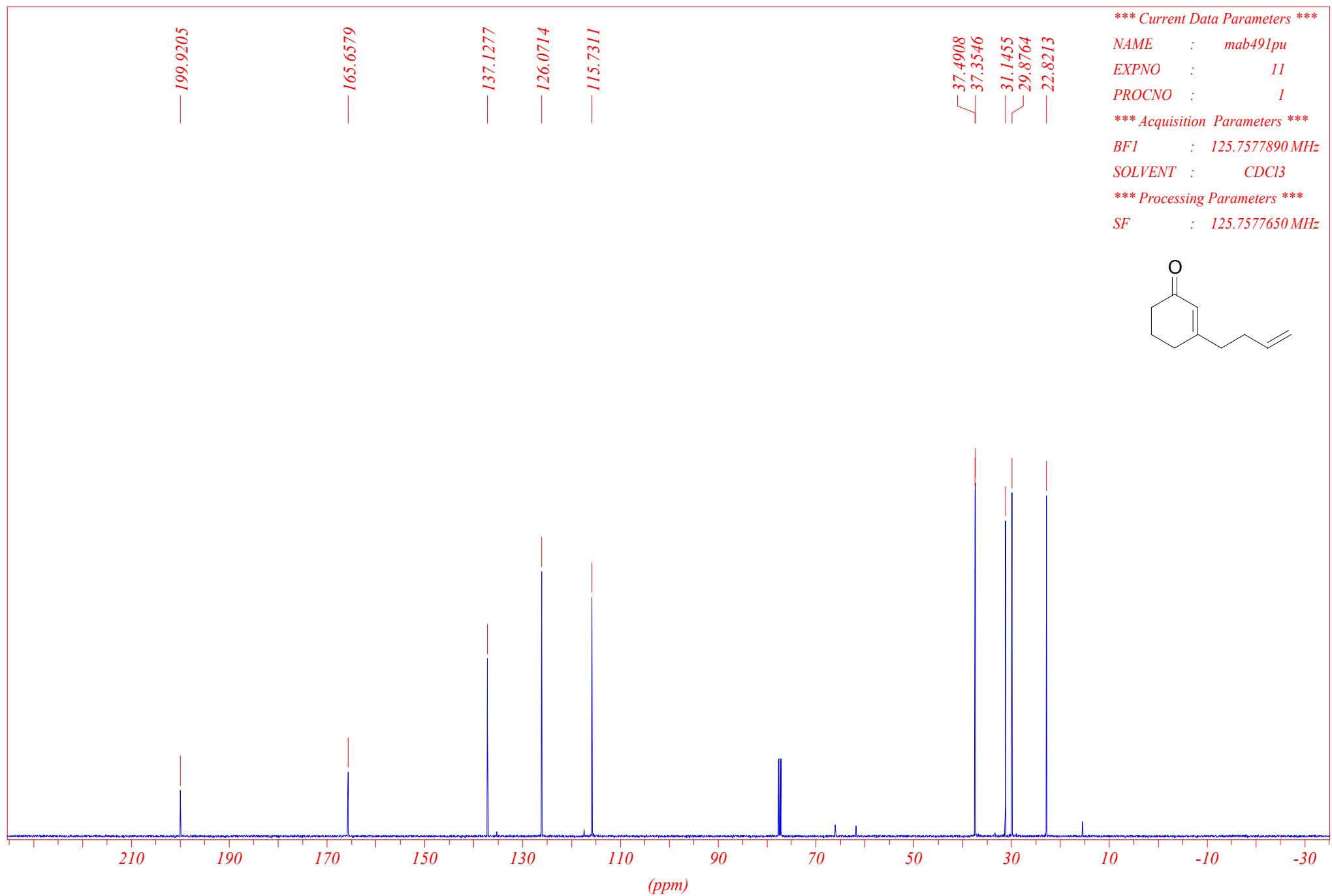
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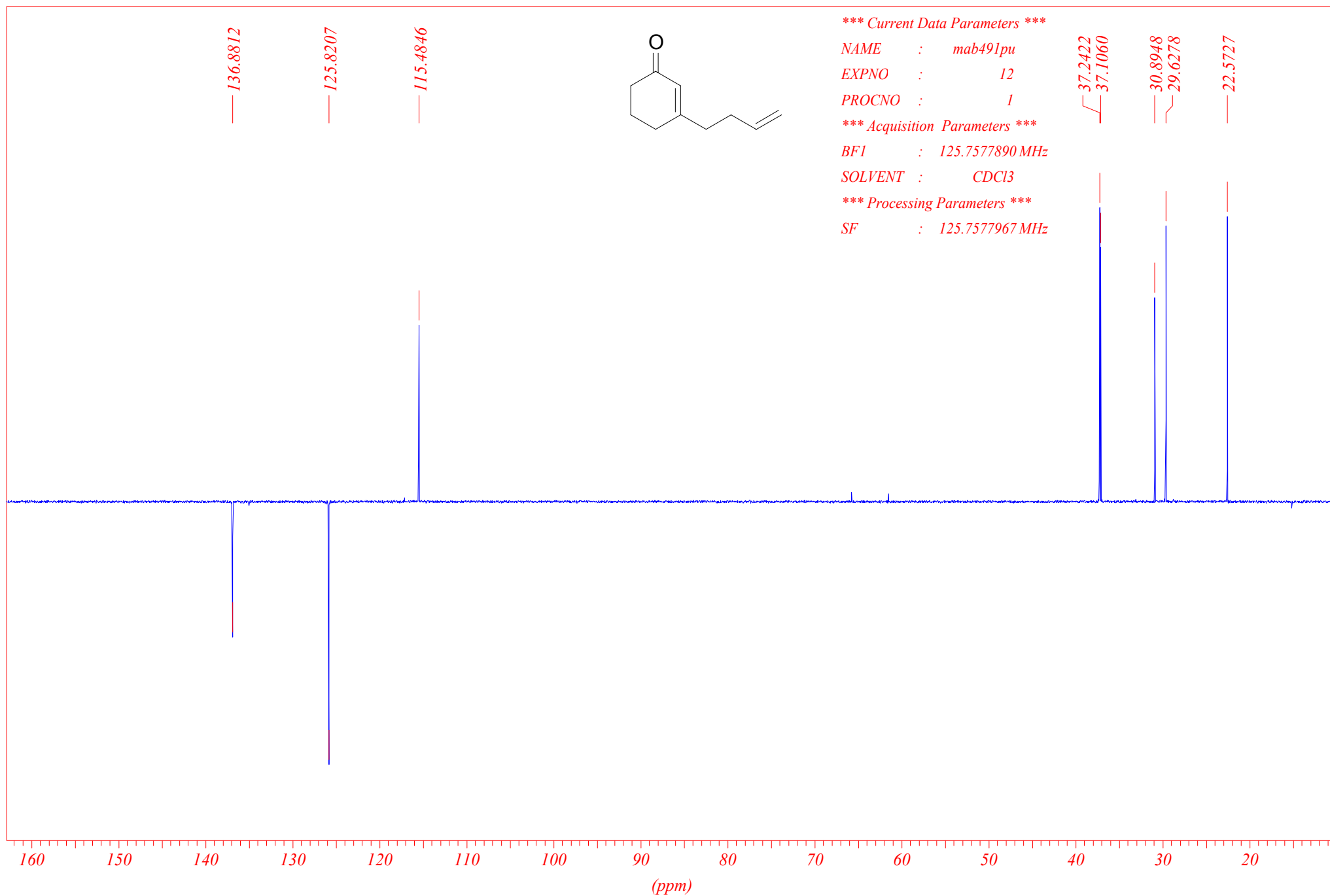
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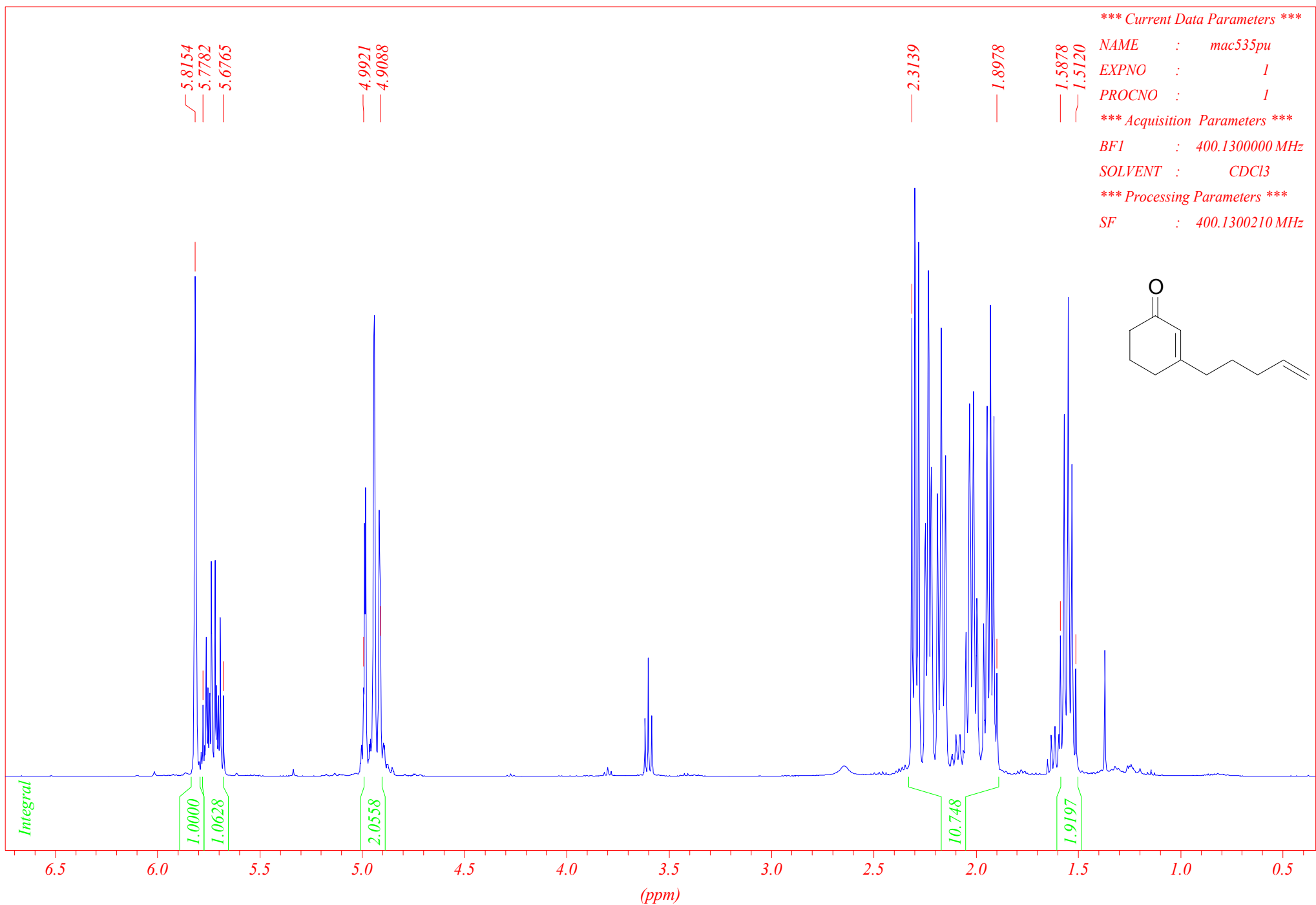
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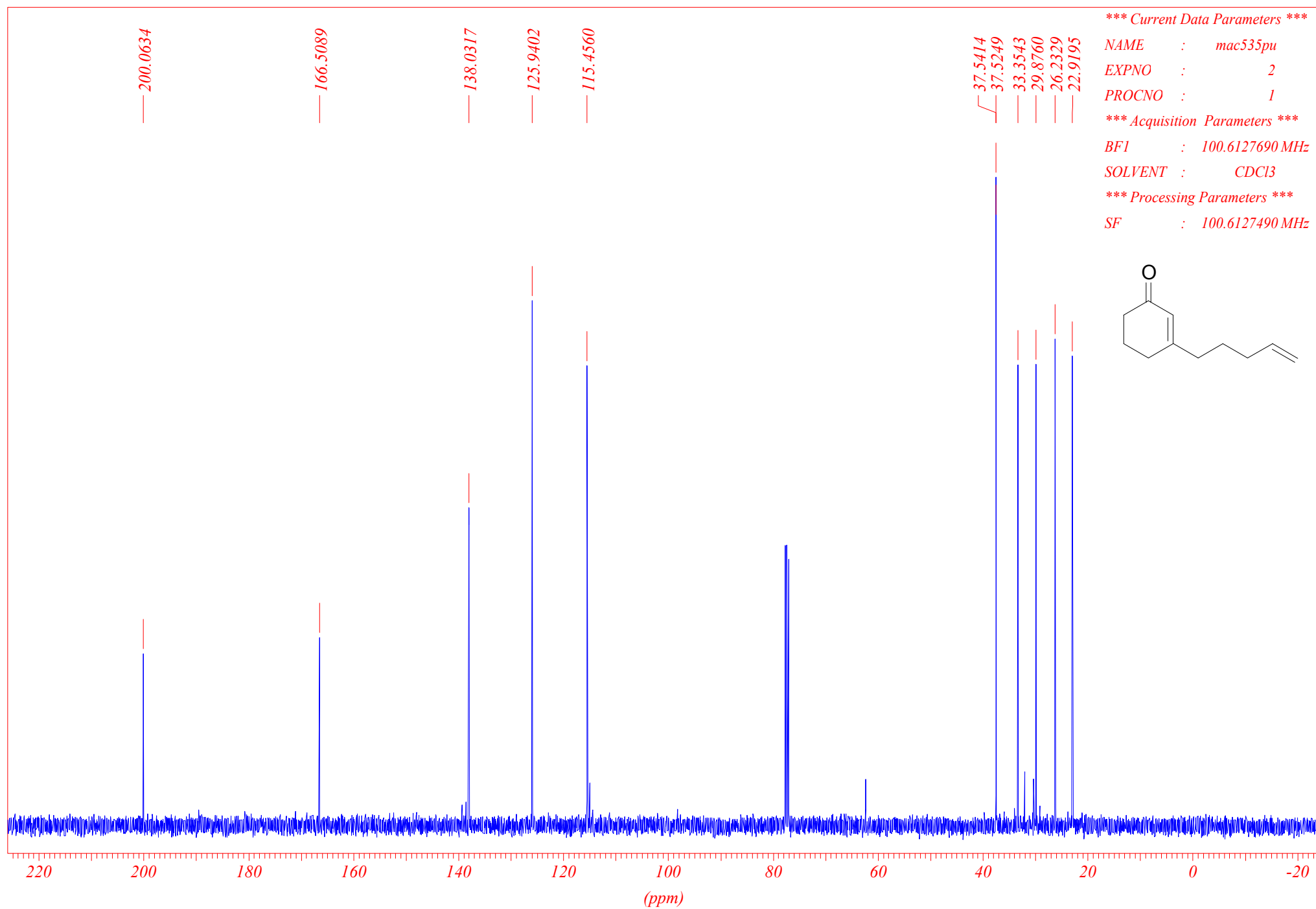


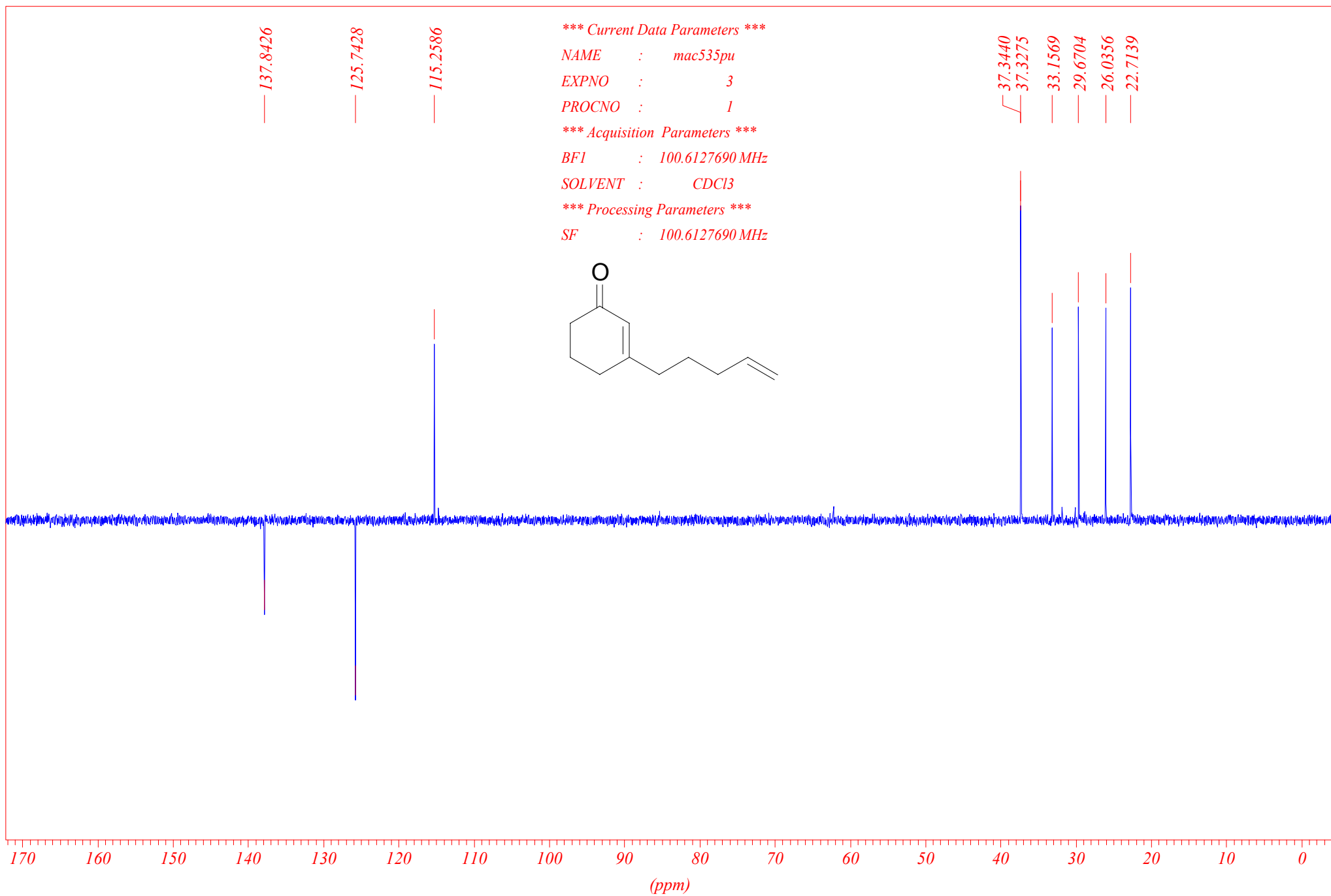


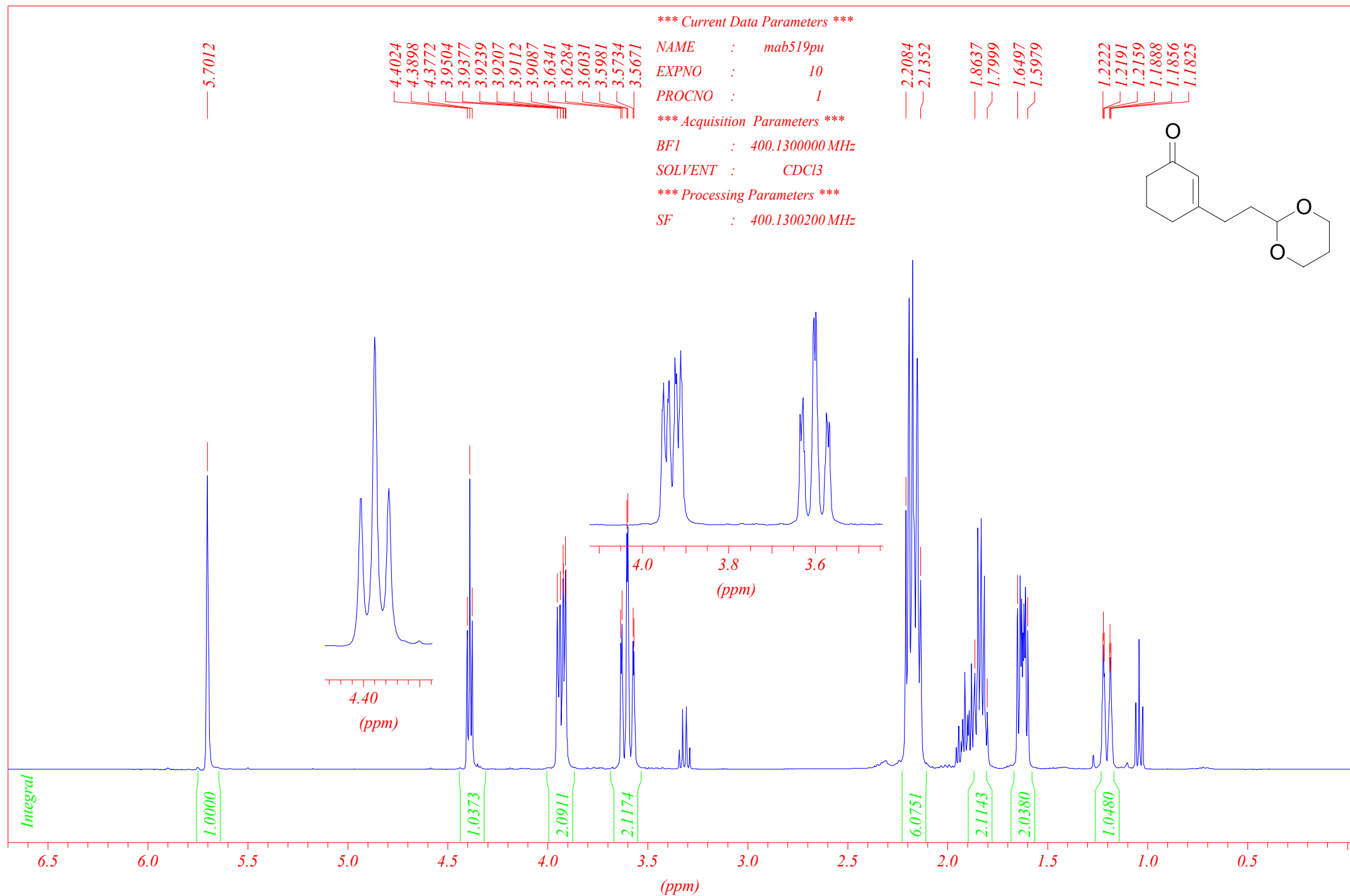


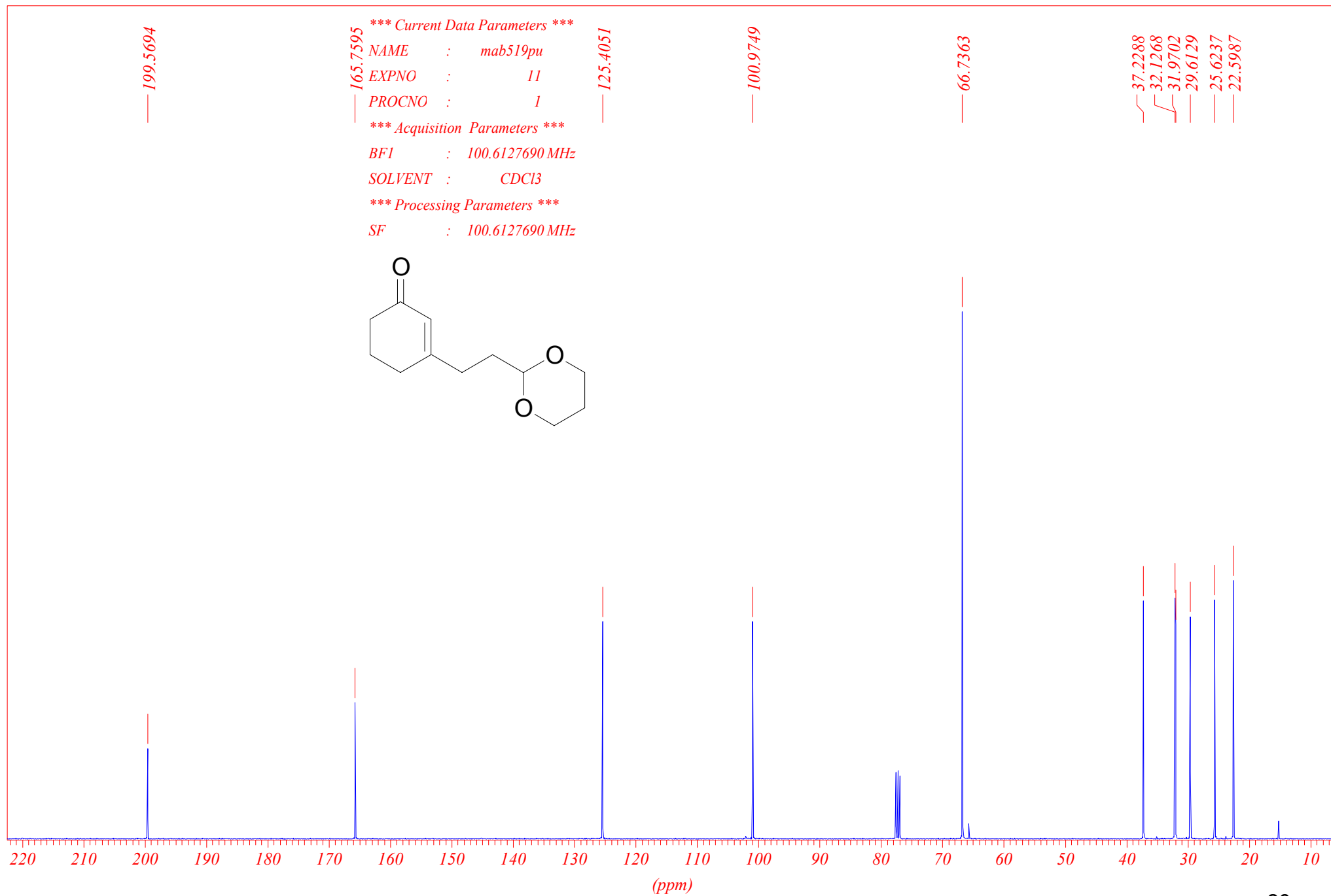


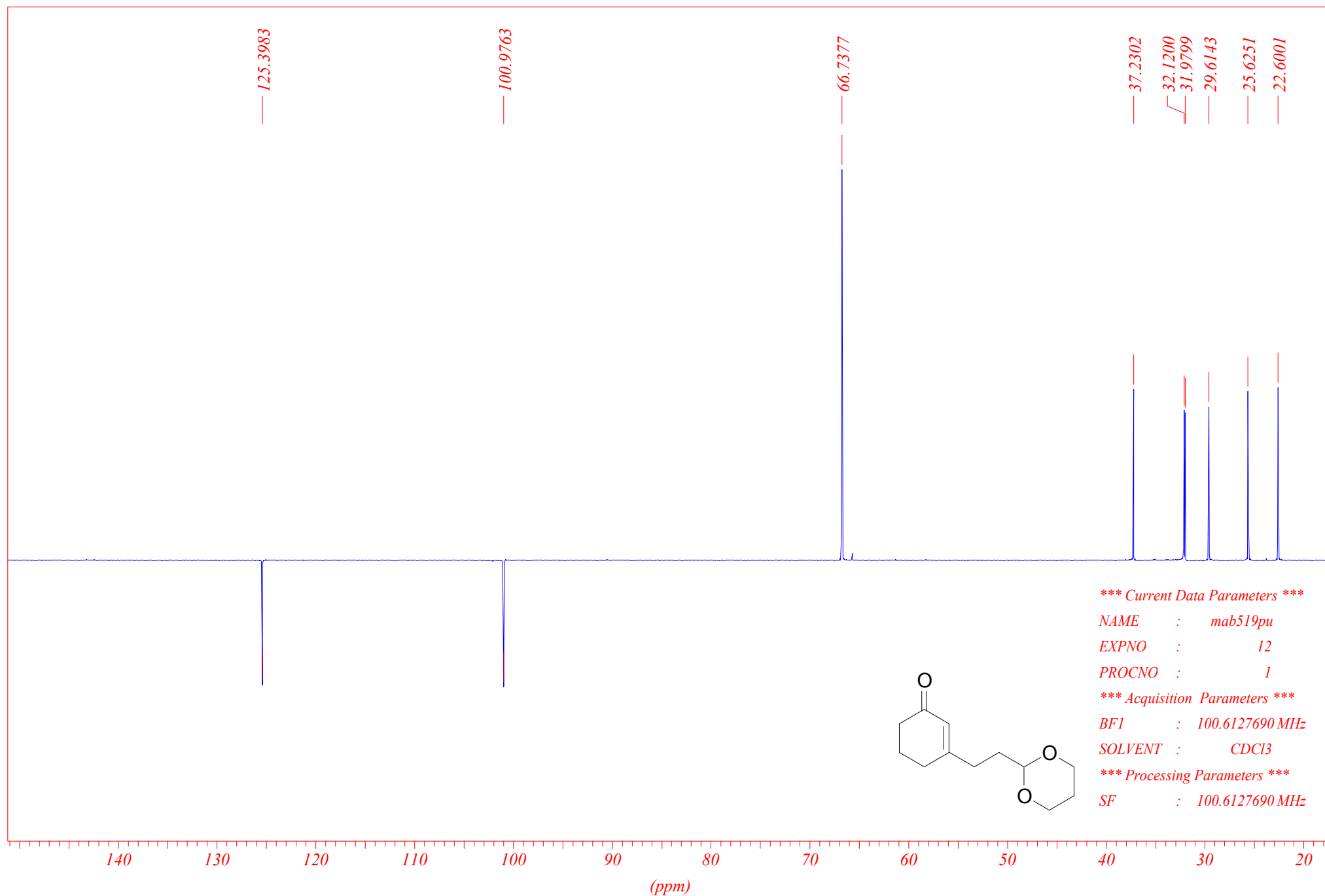


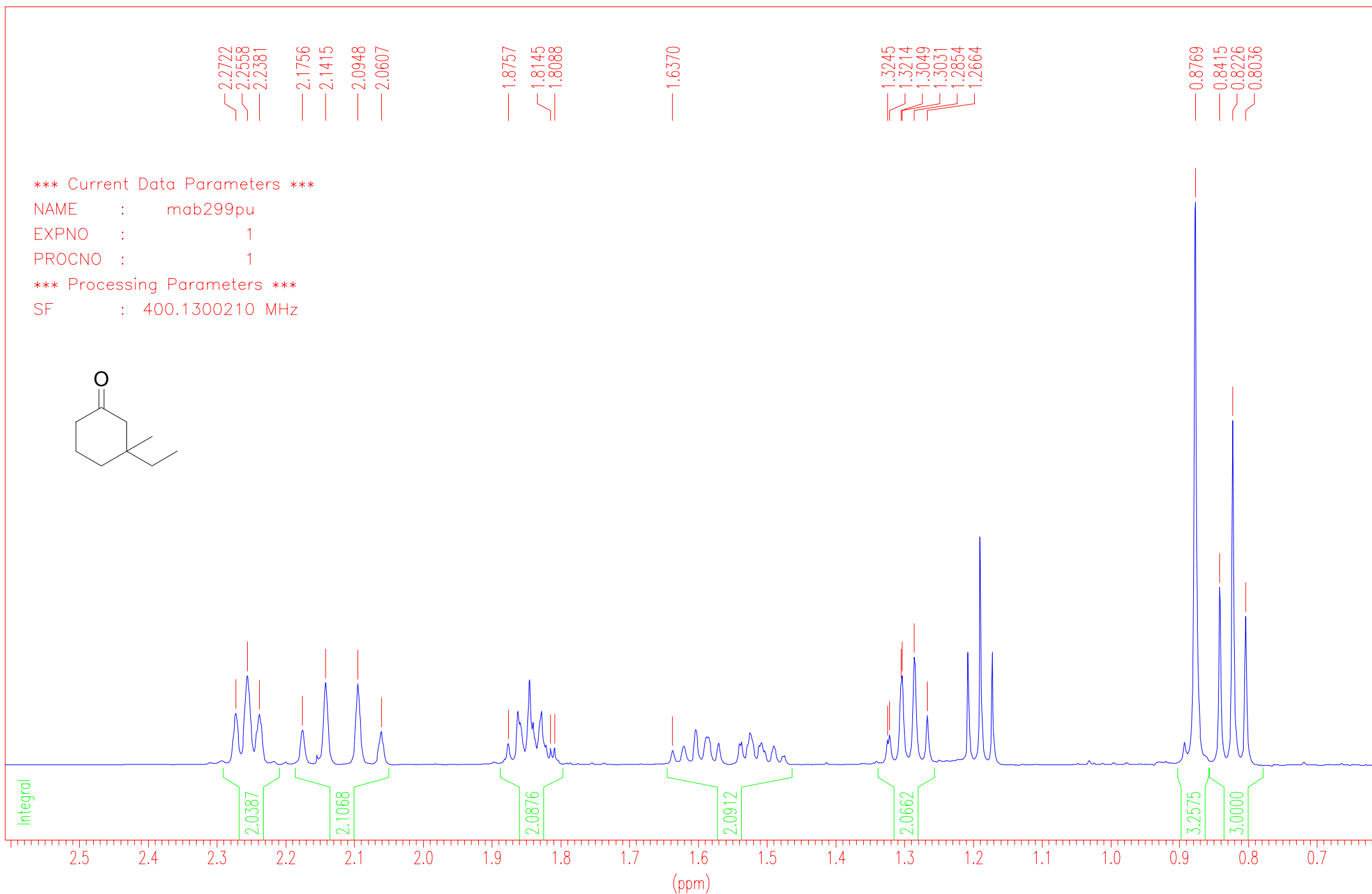


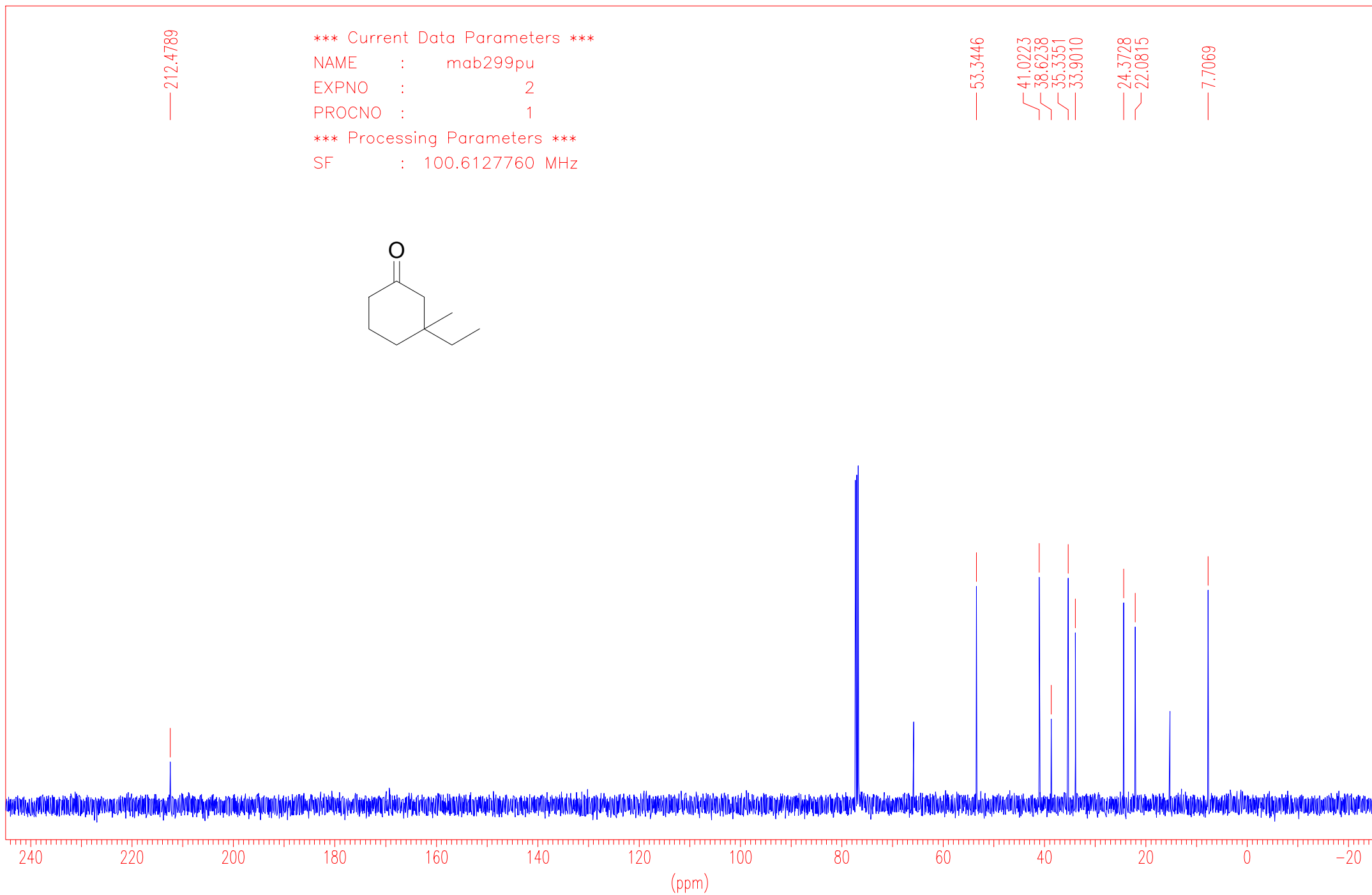












*** Current Data Parameters ***

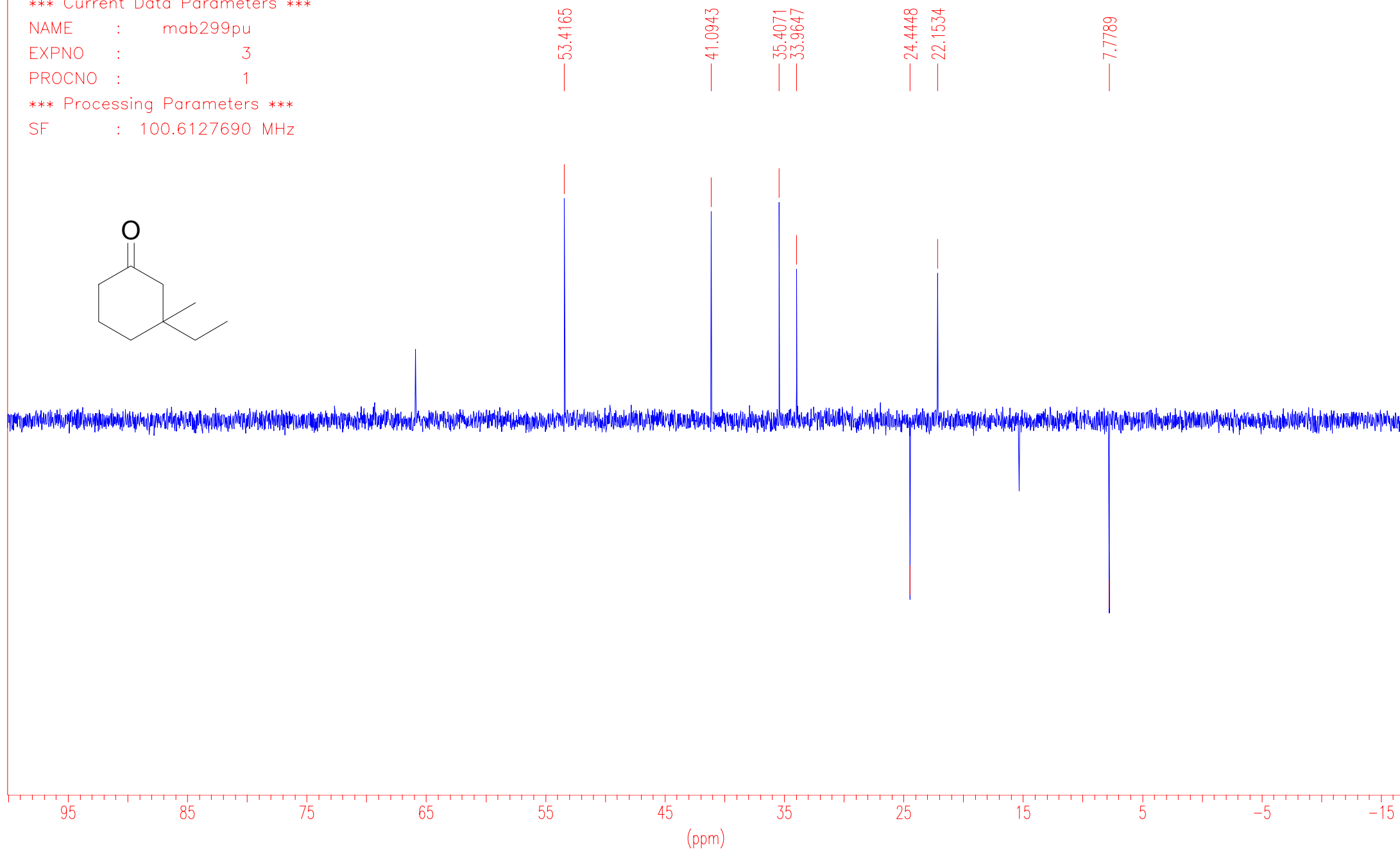
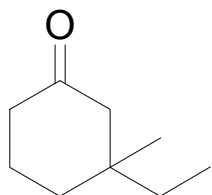
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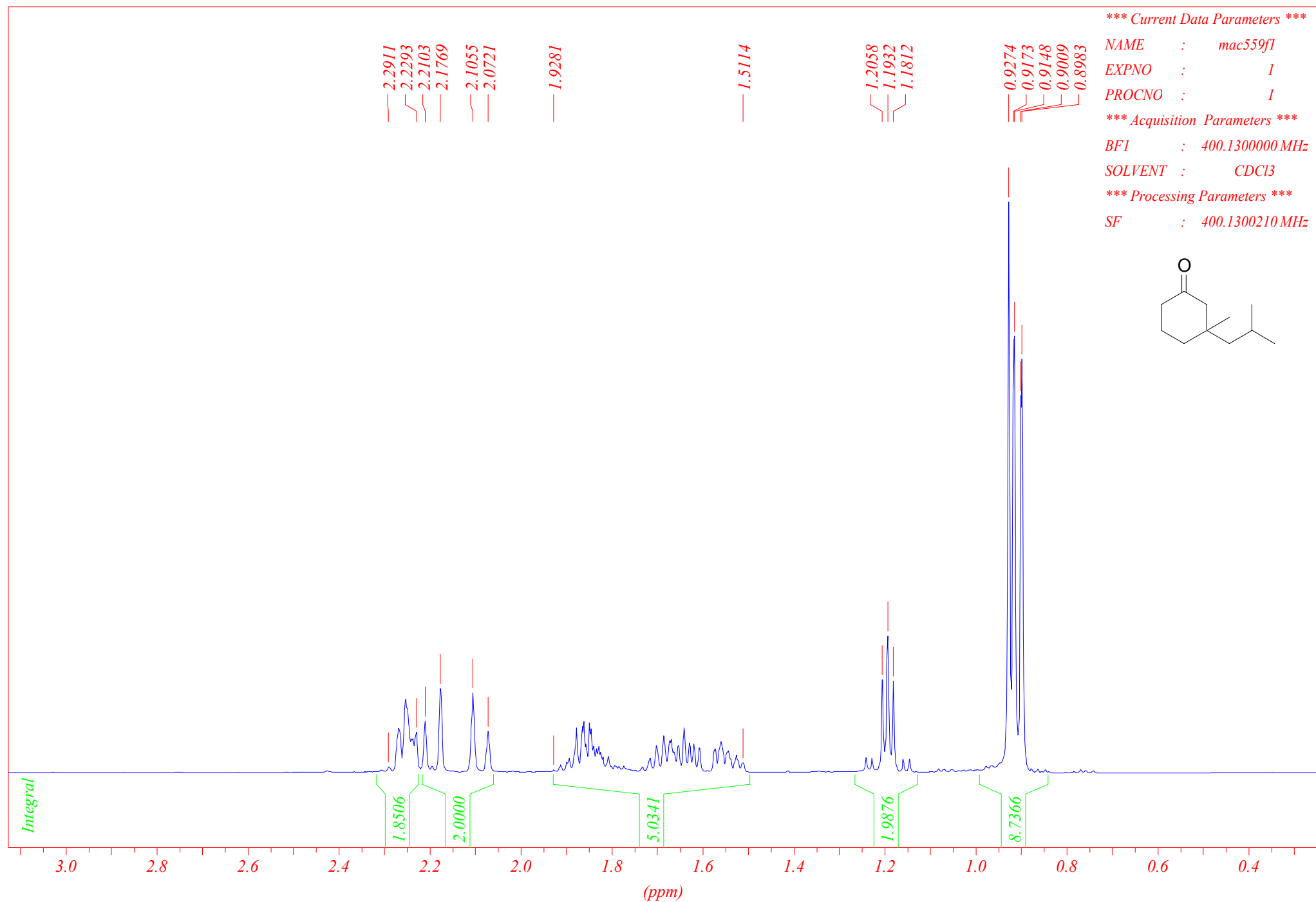
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*** Processing Parameters ***

SF : 100.6127690 MHz





*** Current Data Parameters ***

NAME : mac559f1

EXPNO : 2

PROCNO : 1

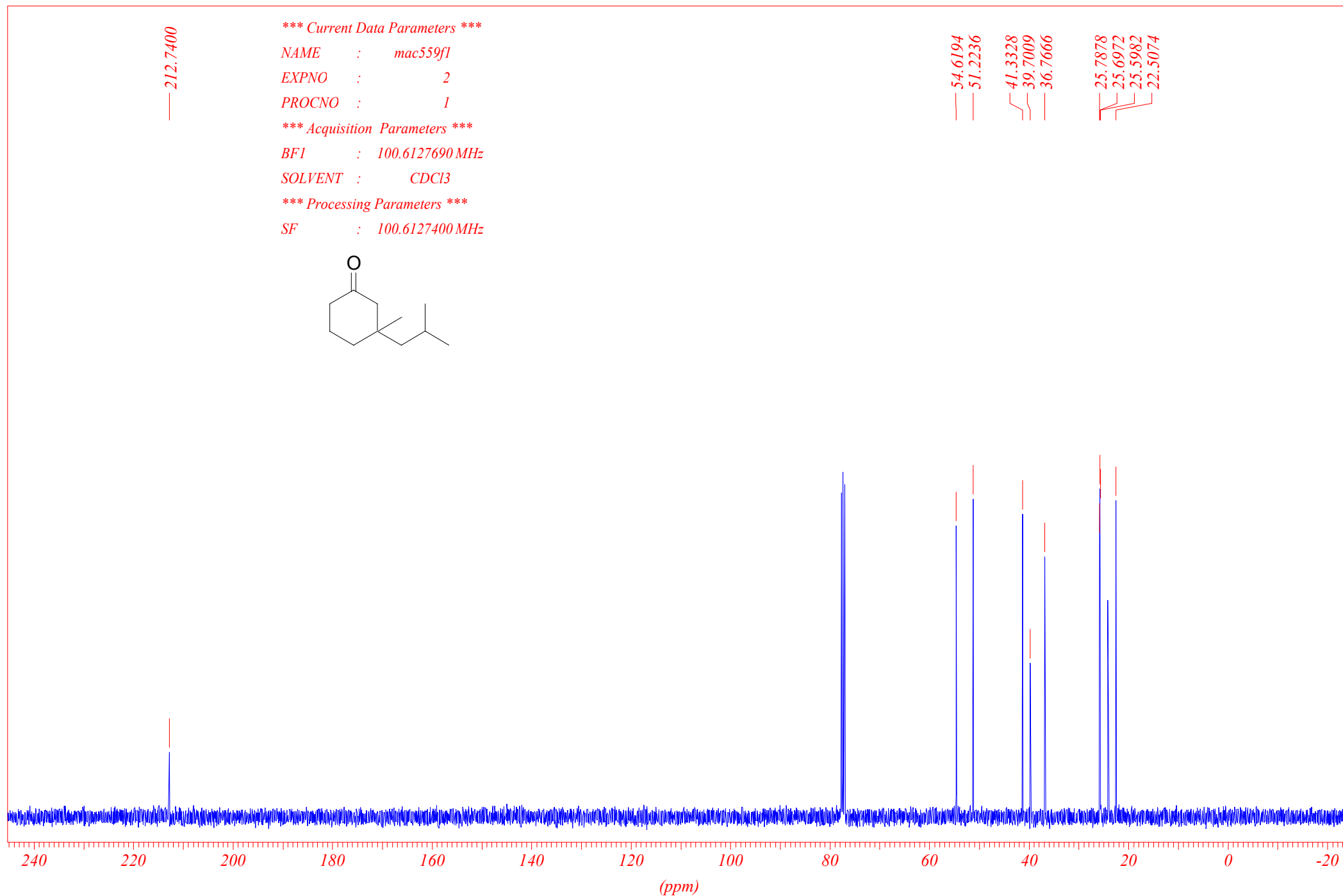
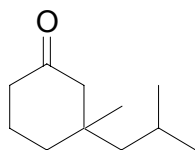
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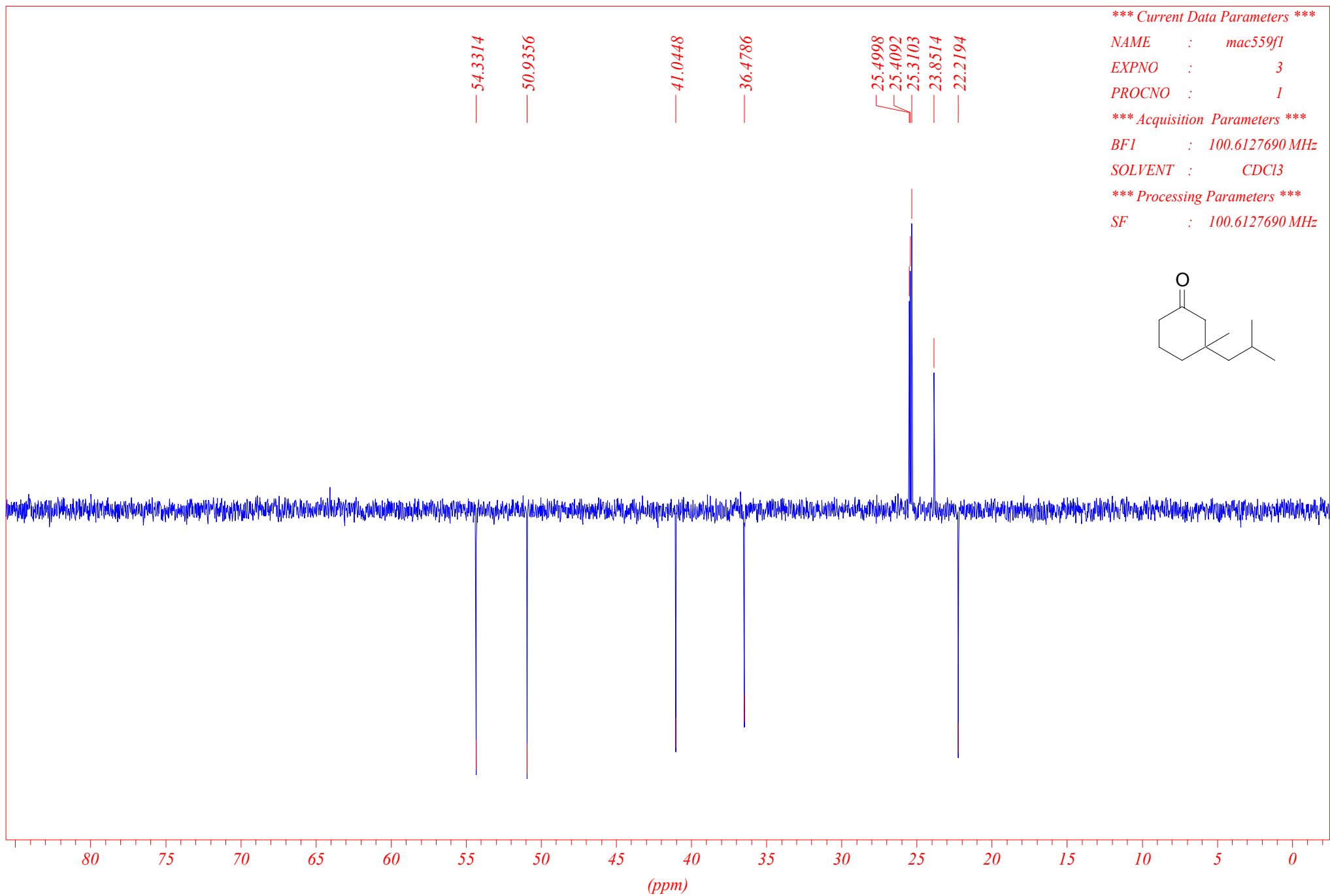
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SOLVENT : CDCl3

*** Processing Parameters ***

SF : 100.6127400 MHz





*** Current Data Parameters ***

NAME : mac529p1

EXPNO : 10

PROCNO : 1

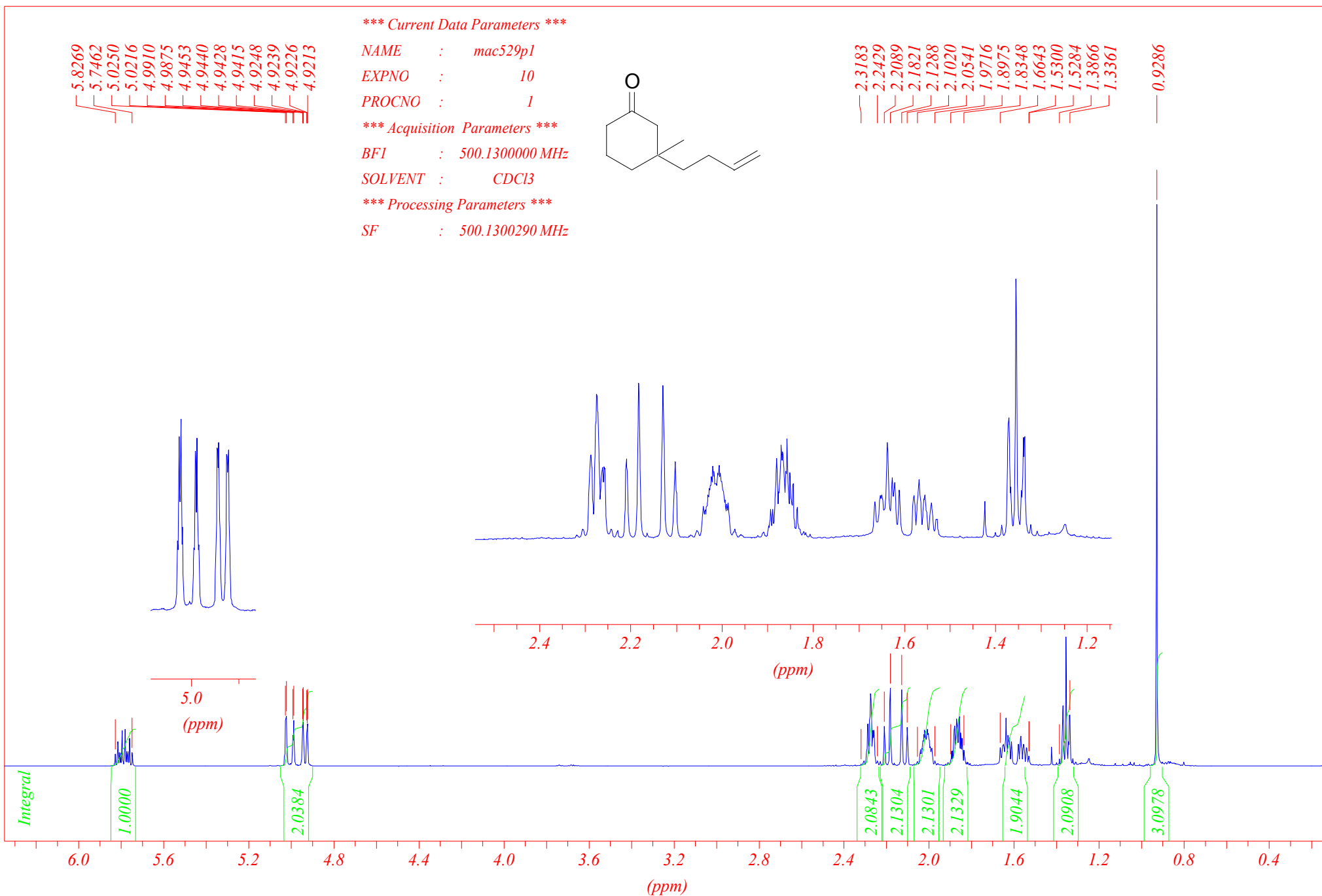
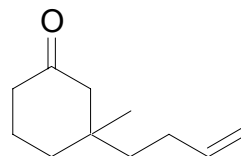
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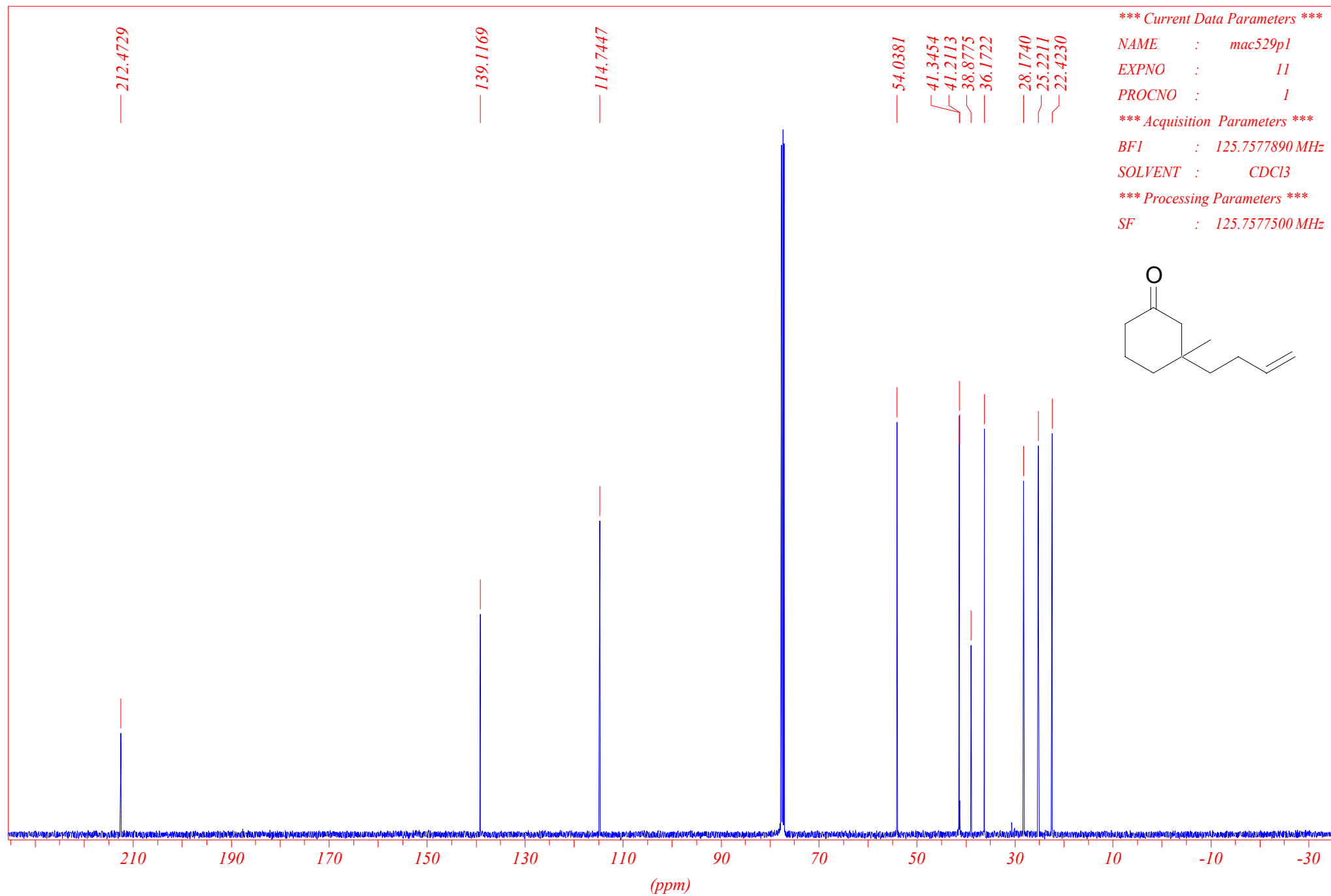
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SOLVENT : CDCl3

*** Processing Parameters ***

SF : 500.1300290 MHz





*** Current Data Parameters ***

NAME : mac529p1

EXPNO : 12

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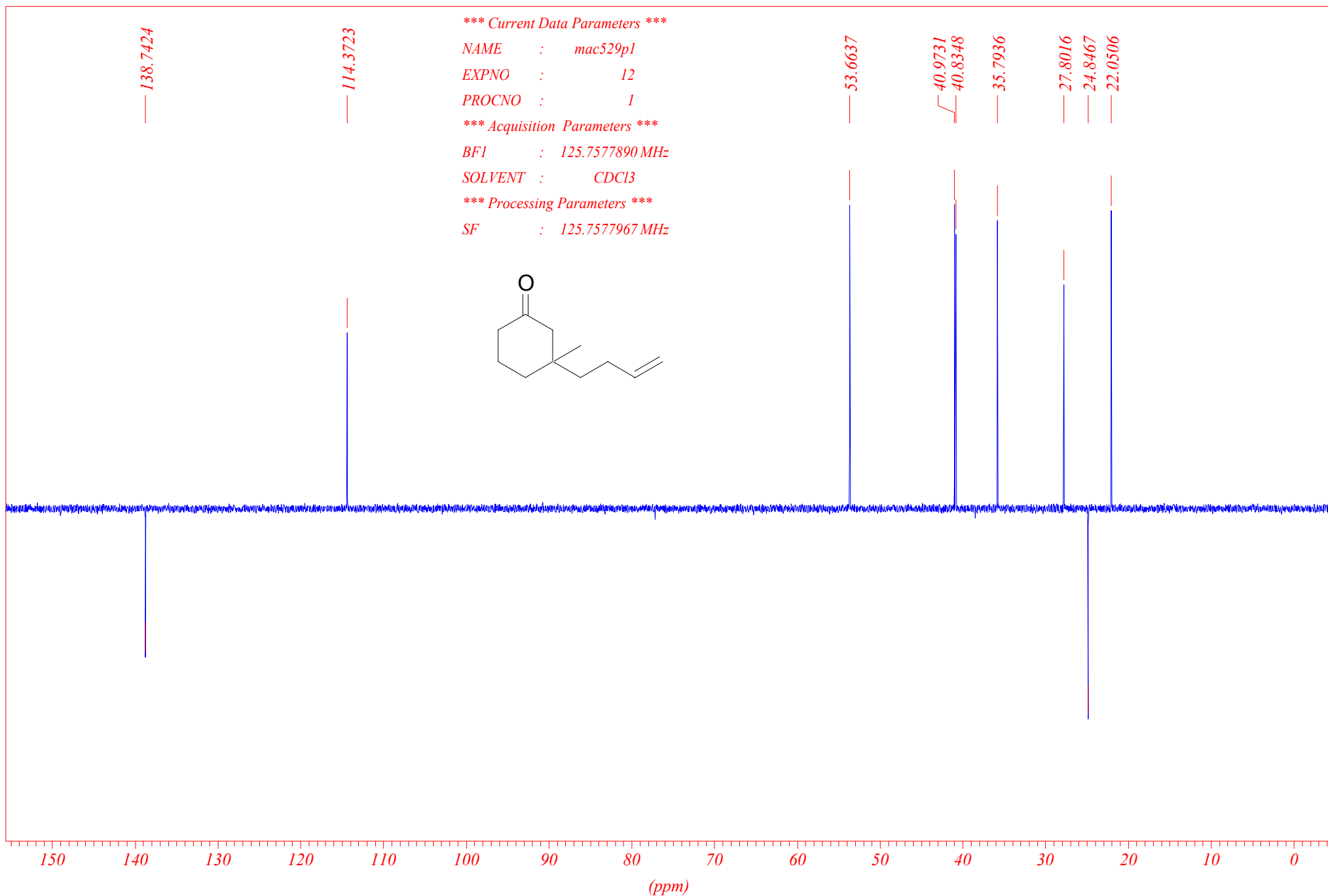
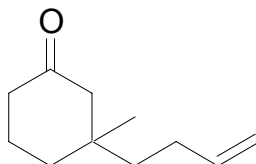
*** Acquisition Parameters ***

BF1 : 125.7577890 MHz

SOLVENT : CDCl₃

*** Processing Parameters ***

SF : 125.7577967 MHz



*** Current Data Parameters ***

NAME : mac541mr
EXPNO : 1
PROCNO : 1

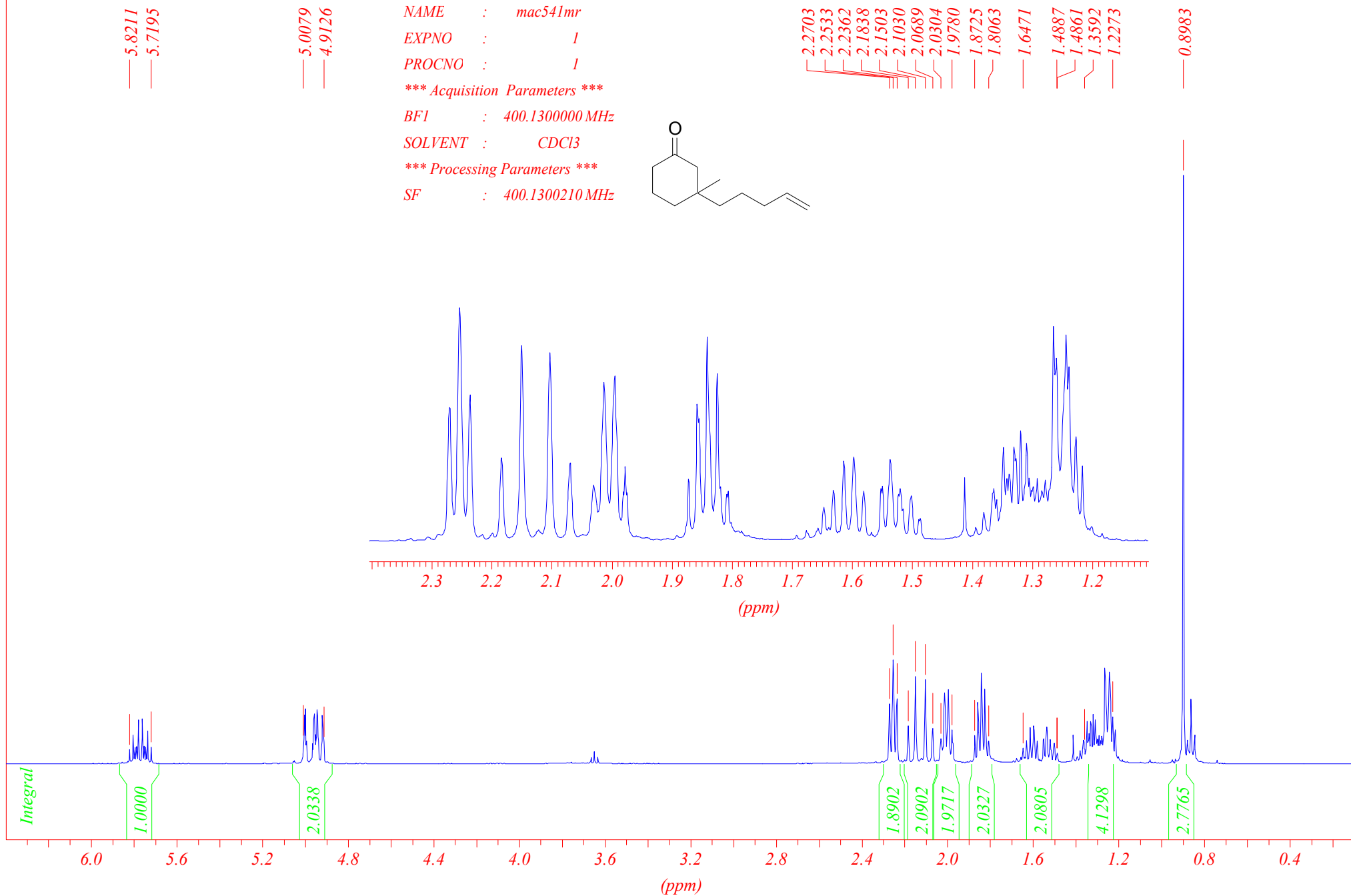
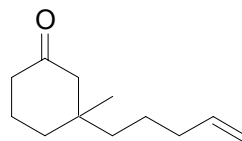
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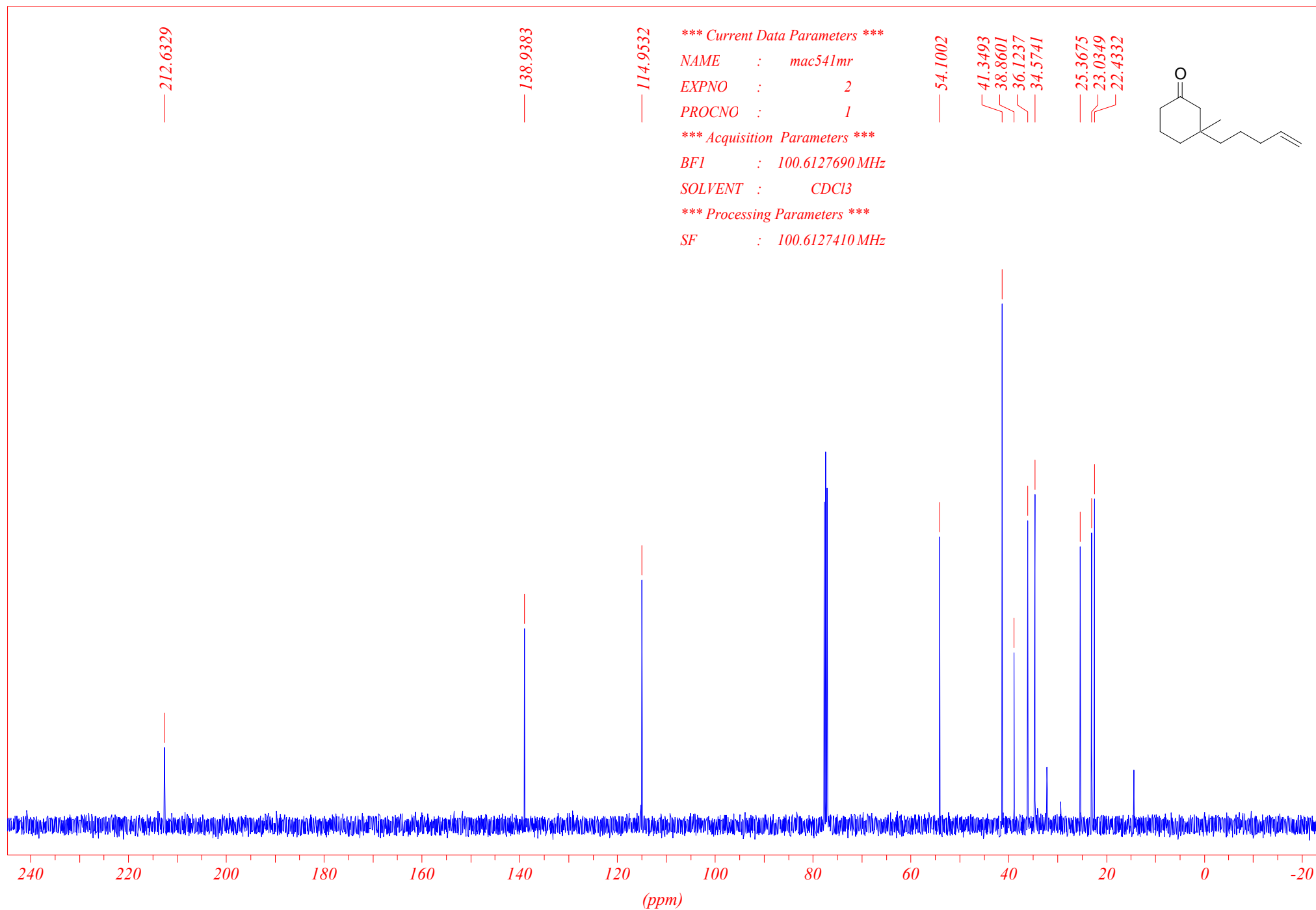
BF1 : 400.130000 MHz

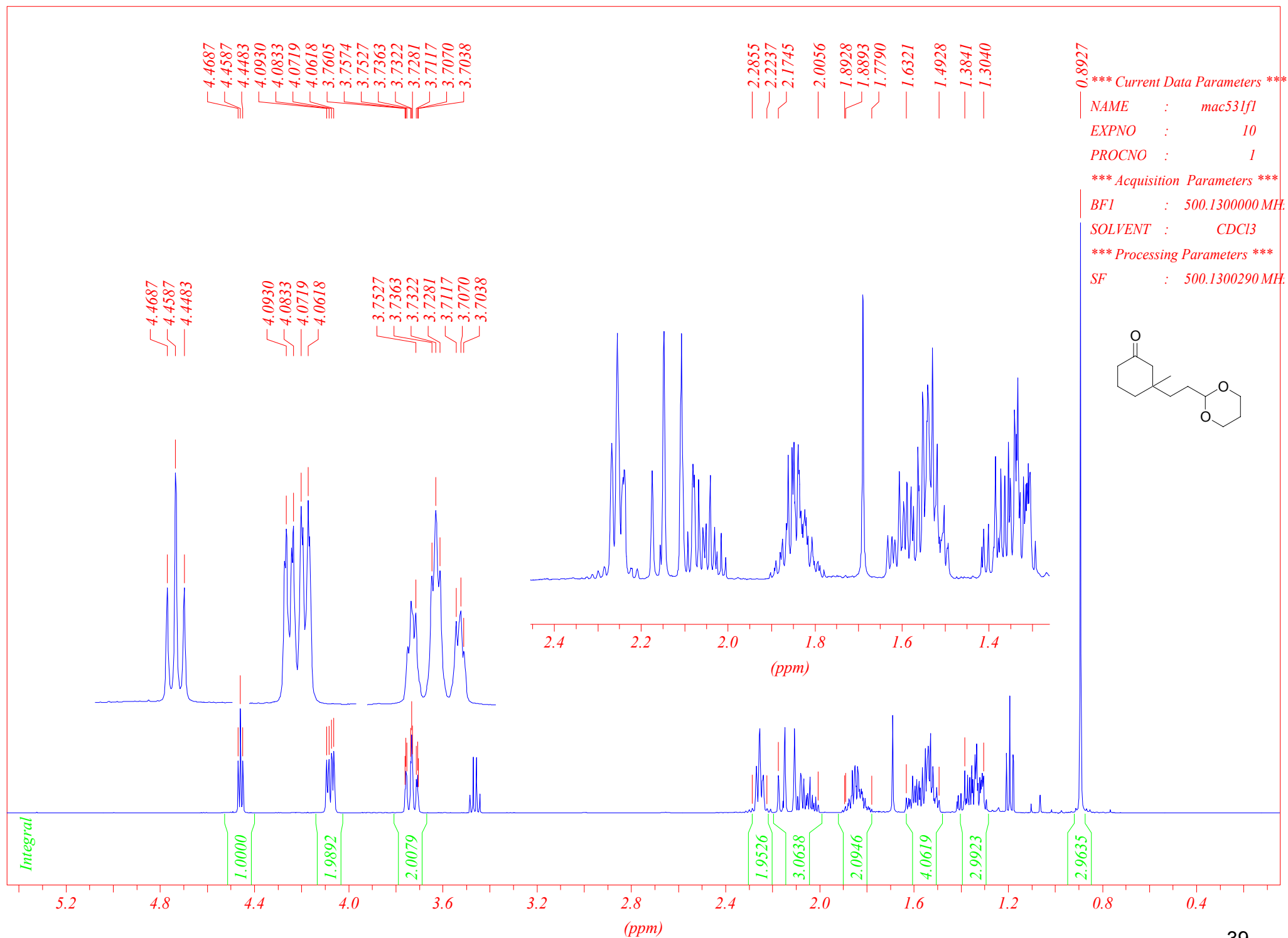
SOLVENT : CDCl3

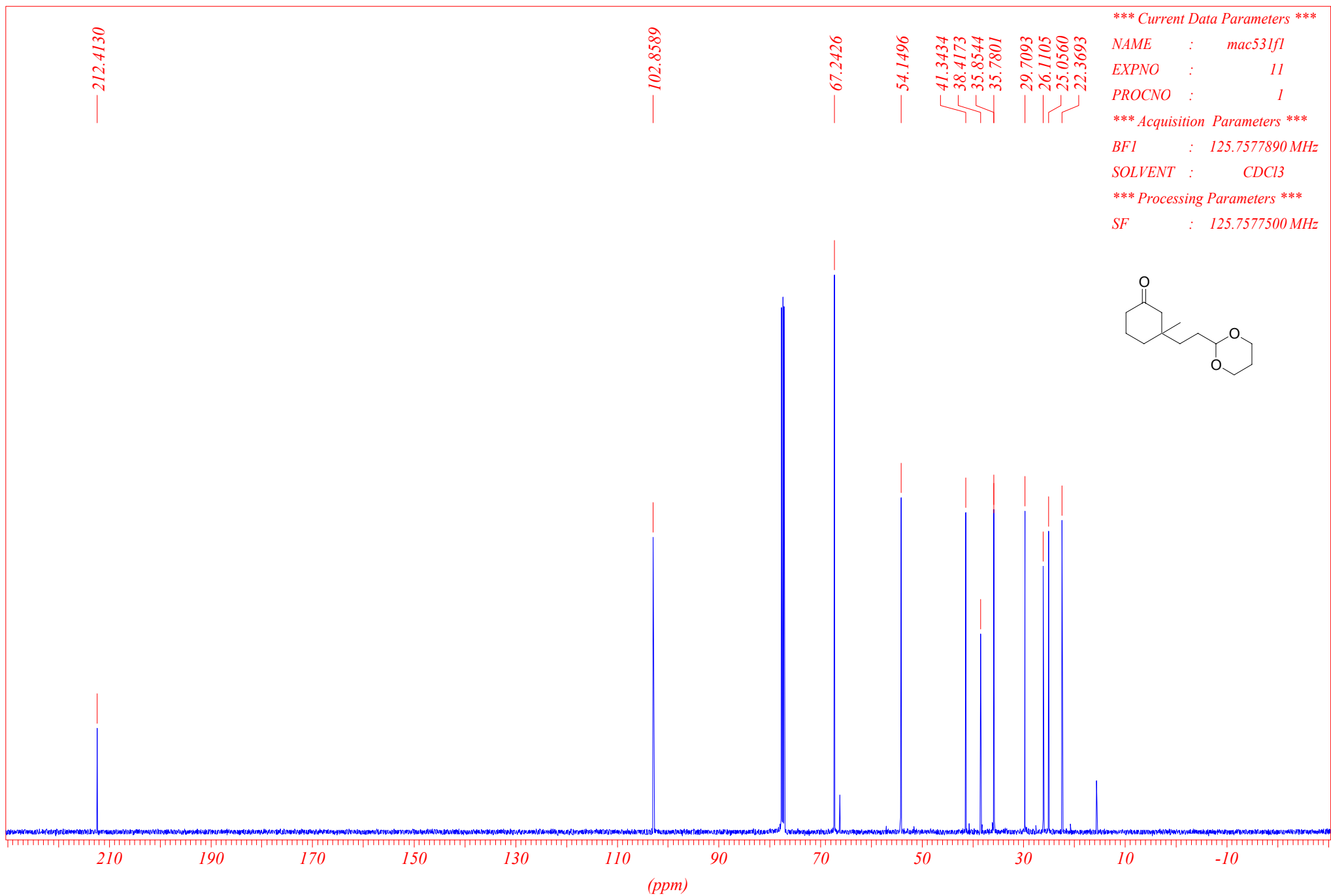
*** Processing Parameters ***

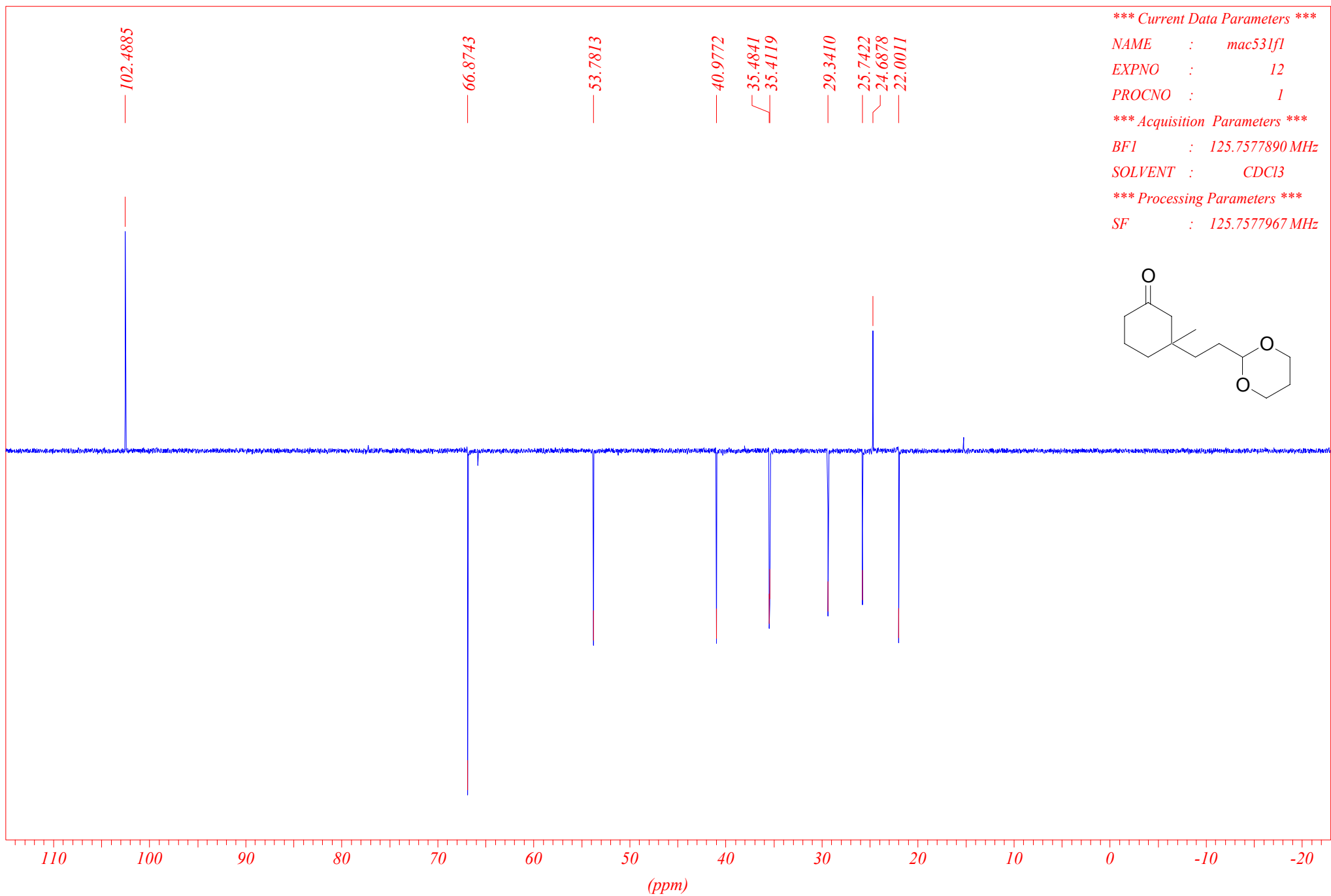
SF : 400.1300210 MHz

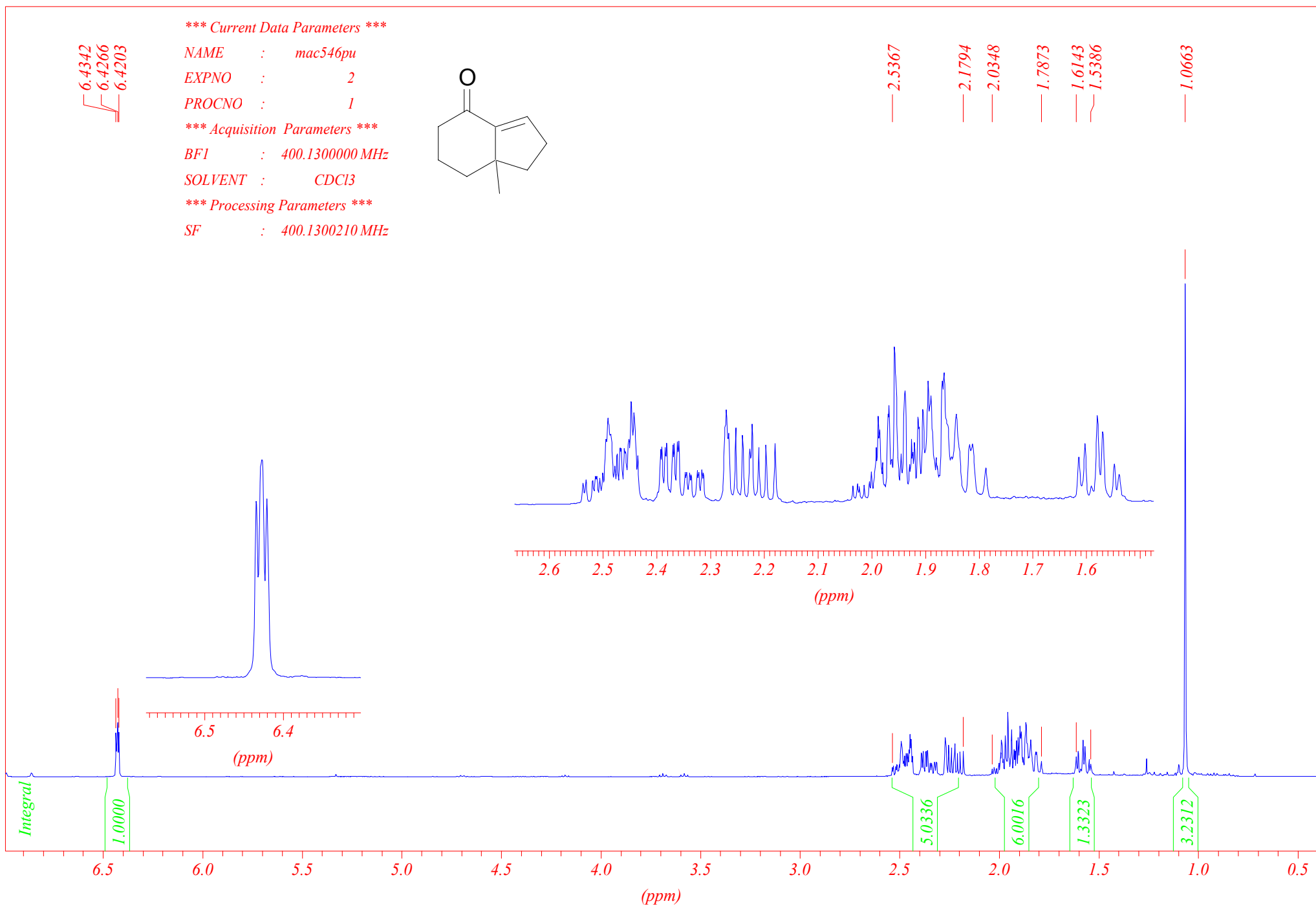


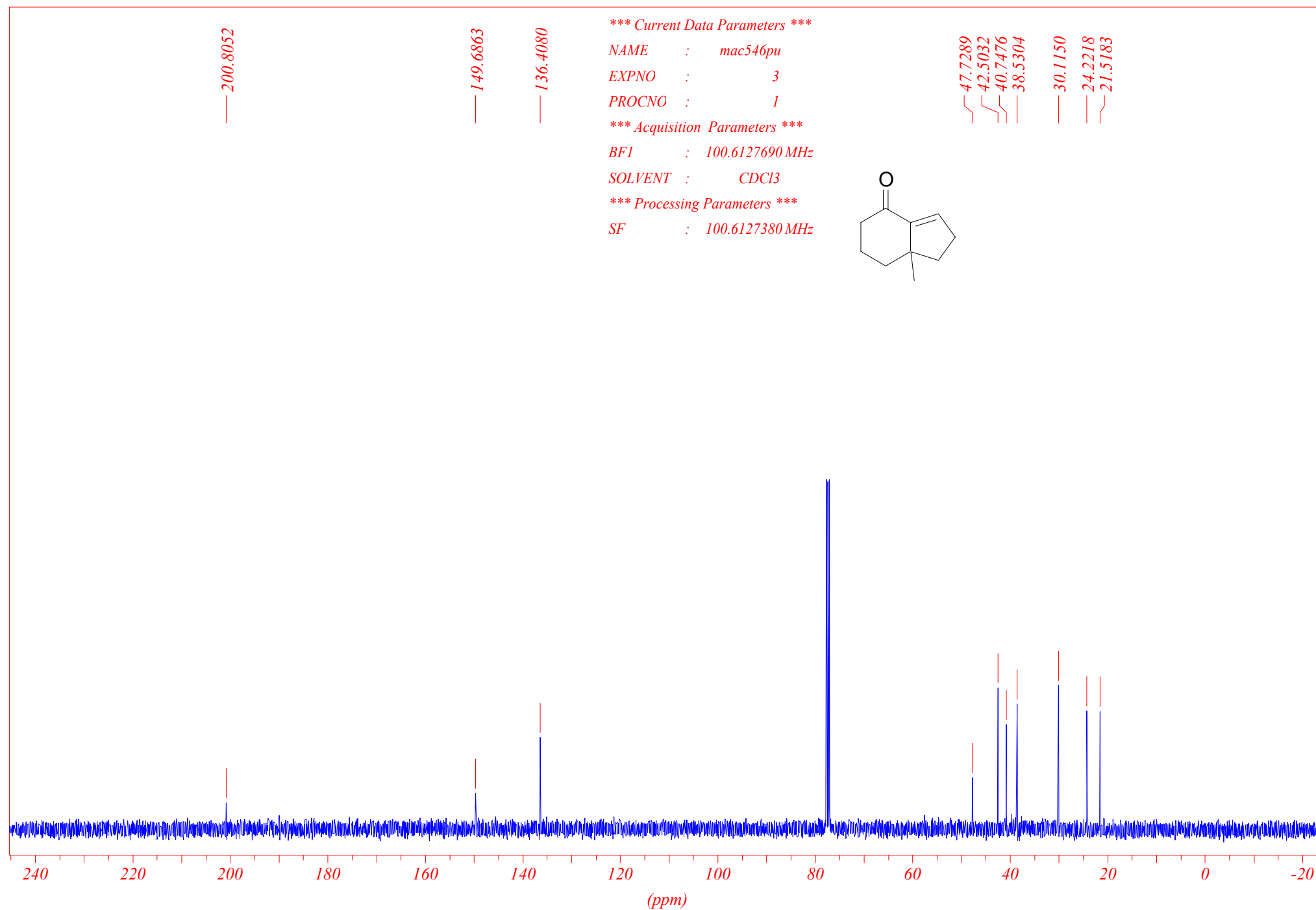












*** Current Data Parameters ***

NAME : mac546pu

EXPNO : 4

PROCNO : 1

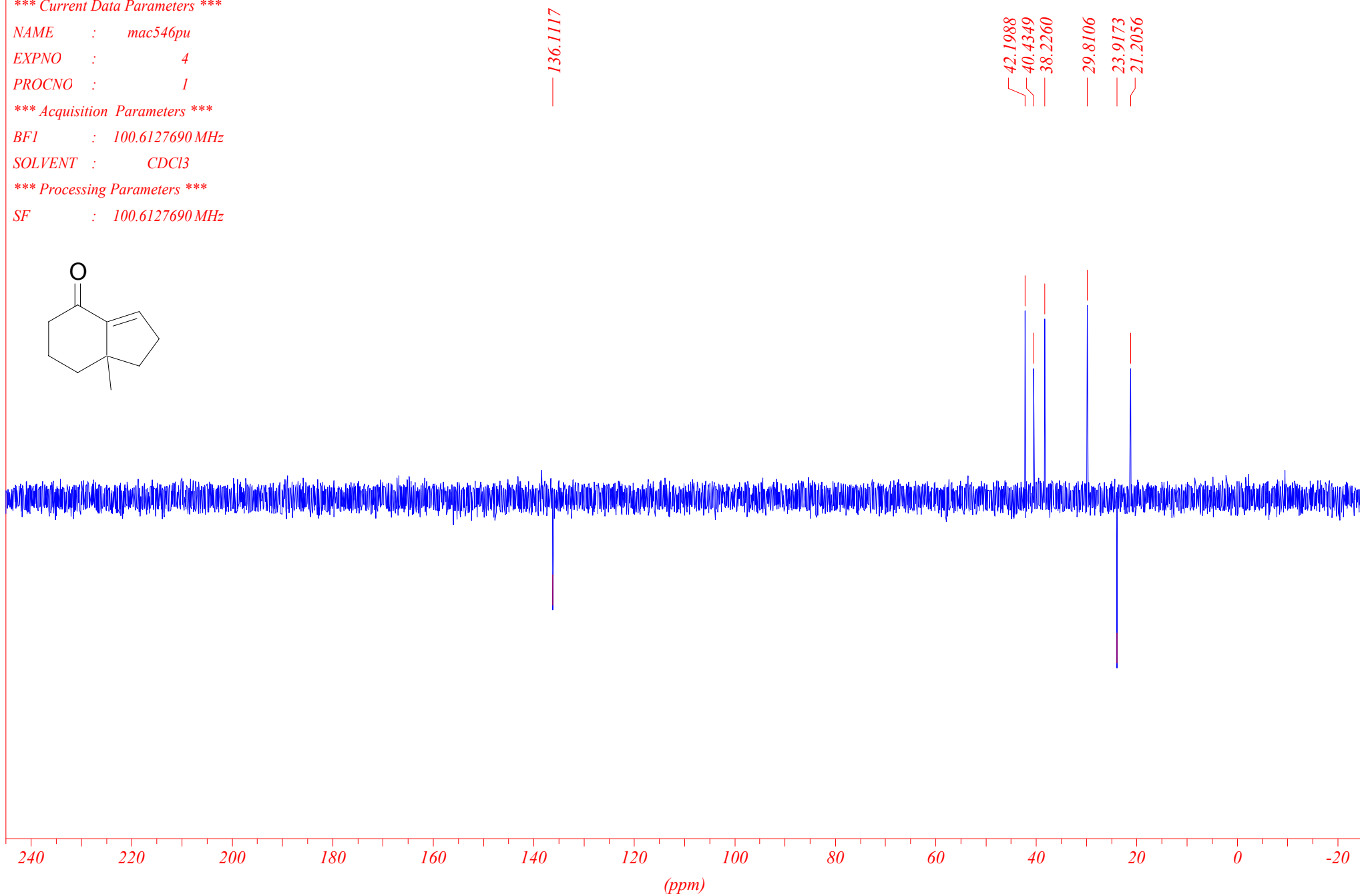
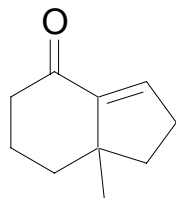
*** Acquisition Parameters ***

BF1 : 100.6127690 MHz

SOLVENT : CDCl₃

*** Processing Parameters ***

SF : 100.6127690 MHz



March: PROTON CDCl3 u guest 22

*** Current Data Parameters ***

NAME : sm171pur

EXPNO : 10

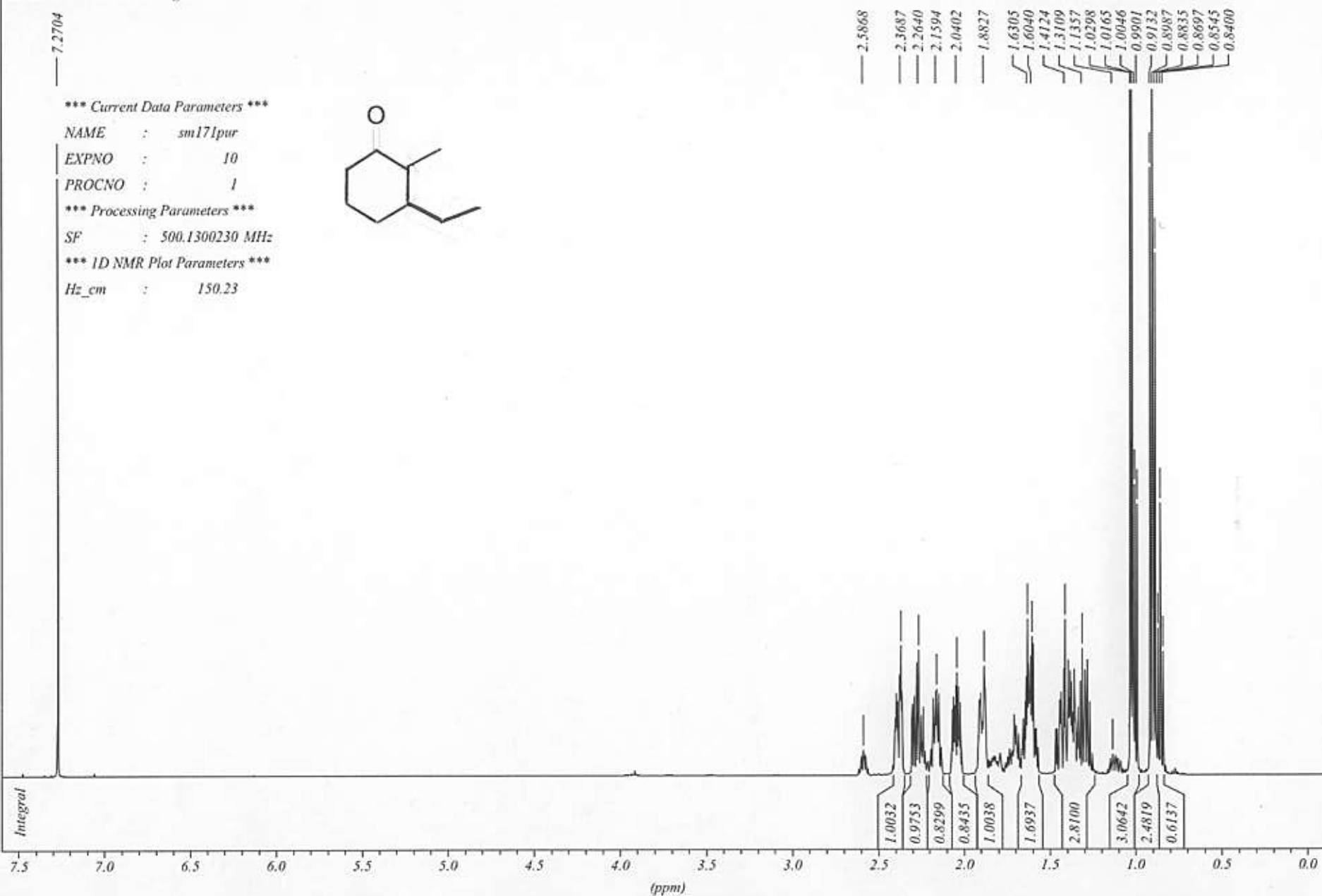
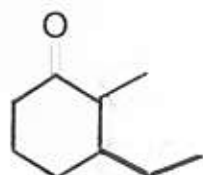
PROCNO : 1

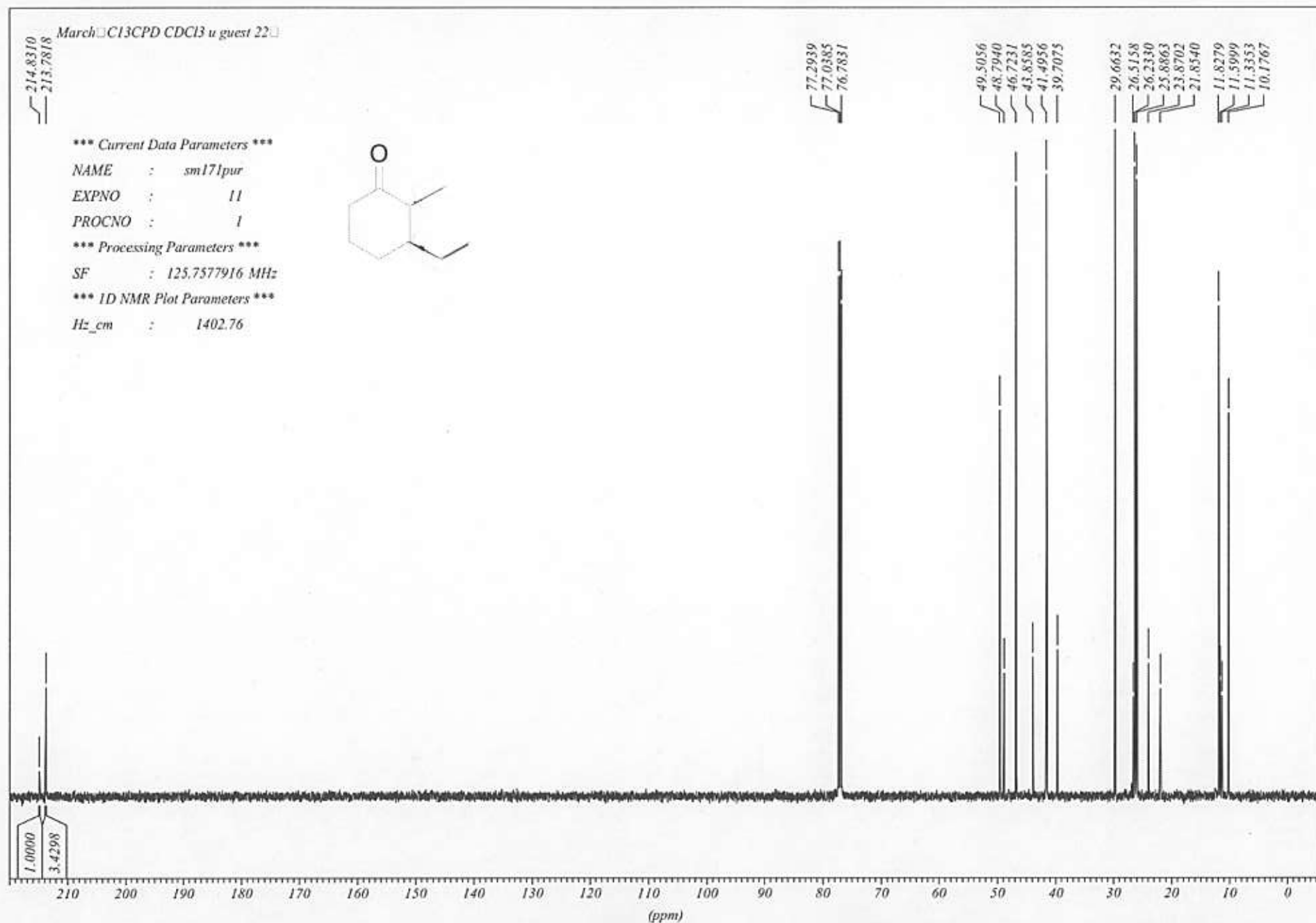
*** Processing Parameters ***

SF : 500.1300230 MHz

*** 1D NMR Plot Parameters ***

Hz_cm : 150.23





March C13DEPT135 CDCl3 u guest 22 SM171 pur

