



Supporting Information

© Wiley-VCH 2006

69451 Weinheim, Germany

Total Synthesis of Paliurine F

Mathieu Toumi, François Couty and Gwilherm Evano *

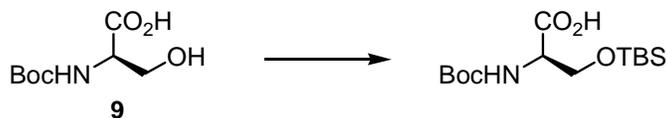
*Institut Lavoisier de Versailles, UMR CNRS 8180
Université de Versailles Saint-Quentin en Yvelines
45, avenue des Etats-Unis
78035 Versailles Cedex
France*

General information.....	S2
Experimental procedures.....	S3
Assignment of stereochemistry for Alcohol 11	S17
Paliurine F Spectral Data Compared to Reported Data	S18
¹ H and ¹³ C NMR spectra of intermediates.....	S22
¹ H and ¹³ C NMR spectra of synthetic paliurine F.....	S37

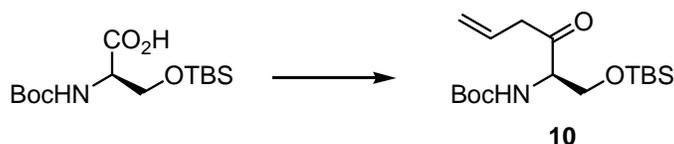
General Information. All reactions were carried out in oven or flame-dried glassware under an argon atmosphere employing standard techniques in handling air-sensitive materials. All solvents were reagent grade. Tetrahydrofuran (THF) was freshly distilled from sodium/benzophenone under argon immediately prior to use. Dichloromethane, toluene and acetonitrile were freshly distilled from calcium hydride. Triethylamine, diisopropylethylamine, *N*-methylmorpholine, *N,N'*-dimethylethylenediamine, diethylamine and 2,6-lutidine were distilled over calcium hydride. *N,N*-Dimethylformamide, HMPA, NMP and dimethylsulfoxide were distilled over calcium hydride. *n*-Butyllithium was purchased from Aldrich and standardized by titration with menthol/1,10-phenanthroline. Copper(I) iodide (99,999 % purity) was purchased from Aldrich and used as supplied. All other reagents were used as supplied. Unless otherwise noted, reactions were magnetically stirred and monitored by thin layer chromatography using Merck-Kieselgel 60F₂₅₄ plates. Flash chromatographies were performed with silica gel 60 (particle size 35-70 μm) supplied by SDS. Yields refer to chromatographically and spectroscopically pure compounds, unless otherwise noted. Proton NMR spectra were recorded using an internal deuterium lock at ambient temperature on a Bruker 300 MHz spectrometer. Internal references of δ_{H} 7.26 and δ_{H} 2.50 were respectively used for CDCl_3 and $\text{DMSO-}d_6$. Data are presented as follows: chemical shift (in ppm on the δ scale relative to $\delta_{\text{TMS}} = 0$), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, quint. = quintuplet, m = multiplet, br = broad), coupling constant (J/Hz) and integration. Resonances that are either partially or fully obscured are denoted obscured (obs.). Carbon-13 NMR spectra were recorded on a Varian 75 MHz spectrometer. Internal references of δ_{C} 77.16 and δ_{C} 39.52 were respectively used for CDCl_3 and $\text{DMSO-}d_6$. Infrared spectra were recorded on a Nicolet OPUS IR (impact 400D) spectrophotometer. Optical rotations were recorded on a Perkin Elmer 341 polarimeter at 589 nm and reported as follows: $[\alpha]_{\text{D}}^{20}$, concentration (c in g/100 mL) and solvent. Melting points were recorded on an Buchi B-545. UV spectra were recorded in methanol on a Shimadzu UV-160A. Mass spectra were obtained on a GCMS HP MS 5989B spectrometer.

Due to the presence of the *N*-Boc-pyrrolidine moiety, it was necessary to record ^1H and ^{13}C NMR spectra of most intermediates in $\text{DMSO-}d_6$ at temperatures ranging from 333 to 345K (see details below; the use of higher temperatures resulted in degradation of intermediates). Even at those temperatures, some ^{13}C peaks were poorly resolved and are denoted "broad" (br.).

Experimental Procedures



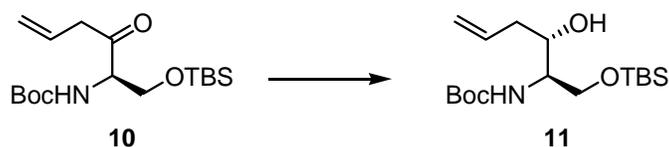
***N*-(*tert*-Butoxycarbonyl)-*O*-(*tert*-butyldimethylsilyl)-*D*-serine.**^{S1} To a solution of commercially available *N*-Boc-*D*-serine **9** (17.6 g, 86 mmol) in DMF (170 mL) were added at 0 °C imidazole (17.5 g, 258 mmol) and TBSCl (16.9 g, 112 mmol). The resulting mixture was slowly warmed to rt, stirred overnight, and poured into a mixture of 1M HCl (300 mL) and ether (600 mL) to hydrolyze the silyl ester. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were washed with brine, and extracted with 1M NaOH (120 mL). The aqueous layer was next acidified by slow addition of 1M HCl (200 mL) and extracted with ether. Combined organic layers were finally dried over MgSO₄, filtered and concentrated under vacuum to yield the desired product as a pale yellow and sticky oil (21.3 g, 67 mmol, 78 %). [α]_D²⁰ -15 (*c* 8.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 10.43 (br. s, 1H), 5.38 (d, *J* = 8.5 Hz, 1H), 4.35 (br. d, *J* = 8.5 Hz, 1H), 4.05 (A of ABX syst., *J* = 1.9, 10.1 Hz, 1H), 3.82 (B of ABX syst., *J* = 2.8, 10.1 Hz, 1H), 1.41 (s, 9H), 0.83 (s, 9H), 0.01, 0.00 (s, s, 3H, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 175.4, 155.7, 80.2, 63.6, 55.5, 28.3, 25.8, 18.3, -5.5, -5.6; IR (neat) ν_{\max} 3447, 2950, 2576, 1685, 1506 cm⁻¹; ESIMS (positive mode): 319.4, 262.3; HRMS (CI, NH₃) *m/z* calcd for C₁₄H₃₀NO₅Si [M+H]⁺ 320.1893, found 320.1889.



(*R*)-*N*-(*tert*-Butoxycarbonyl)-[1-(*tert*-butyl-dimethyl-silyloxymethyl)-2-oxo-pent-4-enyl]-amine **10.** To a solution of *N*-(*tert*-Butoxycarbonyl)-*O*-(*tert*-butyldimethylsilyl)-*D*-serine (5.0 g, 15.7 mmol) in THF (70 mL) was added dropwise at -10 °C a solution of *n*BuLi (1.6 M solution

^{S1} W. R. Ewing, M. M. Joullié, *Heterocycles* **1988**, 27, 2843-2850.

in hexanes, 9.8 mL, 15.7 mmol). The resulting thick gelatinous suspension was stirred at -10 °C for 30 minutes, cooled to -78 °C and treated with a solution of allylmagnesium bromide (0.8 M solution in ether, 45.0 mL, 36.0 mmol).^{S2} The obtained light grey slurry was stirred for 1 hour at 78 °C, warmed to rt over 1 hour, stirred at this temperature for 30 minutes and poured into a mixture of saturated NH₄Cl solution, ice and ether. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum to yield the desired product **10** as a colorless oil (5.3 g, 15.4 mmol, 98 %). This unstable product was immediately used without further purification in the next step. $[\alpha]_{\text{D}}^{20}$ -55 (*c* 2.0, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 5.82-5.98 (m, 1H), 5.45 (d, *J* = 6.9 Hz, 1H), 5.07-5.18 (m, 2H), 4.30 (app. quint., *J* = 3.6 Hz, 1H), 4.04 (A of ABX syst., *J* = 2.8, 10.4 Hz, 1H), 3.79 (B of ABX syst., *J* = 4.0, 10.4 Hz, 1H), 3.34 (A' of A'B'X' syst., *J* = 6.8, 17.6 Hz, 1H), 3.25 (B' of A'B'X' syst., *J* = 6.8, 17.6 Hz, 1H), 1.42 (s, 9H), 0.82 (s, 9H), 0.01 (s, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 205.9, 155.4, 130.0, 119.2, 79.9, 63.4, 61.0, 45.0, 28.4, 25.8, 18.2, -5.5; IR (neat) ν_{max} 3441, 2955, 1705, 1639, 1485, 1250, 1163, 840 cm⁻¹; ESIMS (positive mode): 366.3, 329.4, 310.3, 268.3; HRMS (CI, NH₃) *m/z* calcd for C₁₇H₃₄NO₄Si [M+H]⁺ 344.2257, found 344.2259.

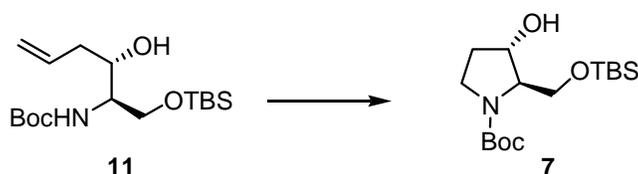


(1*R*,2*S*)-*N*-tert-Butoxycarbonyl-[1-(tert-butyl-dimethyl-silyloxymethyl)-2-hydroxy-pent-4-enyl]-amine **11.**^{S3} A solution of **10** (5.3 g, 15.4 mmol) in absolute ethanol (160 mL) was treated with sodium borohydride (1.15 g, 30.4 mmol) at -78 °C. The resulting mixture was stirred for 80 minutes, carefully quenched by addition of a saturated aqueous solution of NH₄Cl, warmed to rt, concentrated under vacuum and diluted with 1M NaOH solution and ether. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum to yield the

^{S2} P. Mazerolles, P. Boussaguet, V. Huc, *Organic Syntheses* **1999**, 76, 221-224.

^{S3} For assignment of relative stereochemistry, see p. S17.

desired product **11** as a colorless oil (5.2 g, 15.0 mmol, 98 %). Diastereoisomeric excess was determined by analysis of crude ^1H NMR spectra and was found to be higher than 95%. $[\alpha]_{\text{D}}^{20}$ -30 (*c* 2.0, CHCl_3); ^1H NMR (300 MHz, CDCl_3): δ 5.69-5.85 (m, 1H), 5.19 (d, $J = 8.3$ Hz, 1H), 5.02-5.11 (m, 2H), 3.89 (A of ABX syst., $J = 2.6, 10.5$ Hz, 1H), 3.72 (B of ABX syst., $J = 2.4, 10.5$ Hz, 1H), 3.64 (app. quint., $J = 6.2$ Hz, 1H), 3.44-3.53 (br. m, 1H), 3.10 (d, $J = 7.2$ Hz, 1H), 2.27 (app. t, $J = 6.7$ Hz, 2H), 1.36 (s, 9H), 0.82 (s, 9H), 0.00 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 155.7, 134.6, 117.7, 79.3, 72.6, 63.1, 53.9, 39.3, 28.4, 25.8, 18.1, -5.6; IR (neat) ν_{max} 3375, 2971, 2879, 1716, 1491, 1173, 840 cm^{-1} ; ESIMS (positive mode): 713.6, 384.4, 368.4, 329.4, 268.3, 246.3; HRMS (CI, NH_3) m/z calcd for $\text{C}_{17}\text{H}_{36}\text{NO}_4\text{Si}$ $[\text{M}+\text{H}]^+$ 346.2414, found 346.2407.

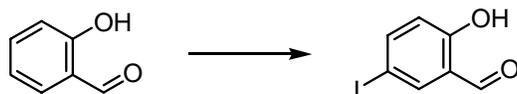


(1*R*,2*S*)-1-*tert*-Butoxycarbonyl-2-(*tert*-butyl-dimethyl-silyloxymethyl)-3-hydroxy-

pyrrolidine 7. To a solution of **10** (3.5 g, 10.1 mmol) in THF (150 mL) and water (10 mL) was added osmium tetroxide (4% wt solution in water, 3.2 mL, 0.5 mmol) and sodium periodate (8.65 g, 40.4 mmol). The resulting white slurry was vigorously stirred at rt for 90 minutes and quenched by addition of a 10% wt aqueous solution of sodium thiosulfate. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were successively washed with 10% wt aqueous sodium thiosulfate solution and brine, dried over MgSO_4 , filtered and concentrated under vacuum to yield the intermediate *N*-Boc-aminal as a complex mixture of rotamers and diastereoisomers.

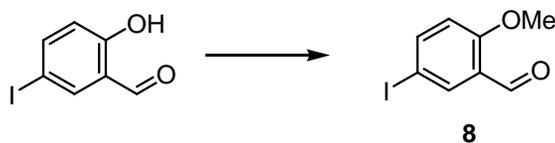
The resulting brownish oil was dissolved in dry dichloromethane, treated with triethylsilane (1.9 mL, 12.0 mmol) and $\text{BF}_3 \cdot \text{OEt}_2$ (1.3 mL, 12.0 mmol) dropwise at -78 $^\circ\text{C}$, and stirred for 3h30 while carefully keeping the temperature below -75 $^\circ\text{C}$. The reaction mixture was finally quenched by addition of a saturated aqueous solution of NaHCO_3 (20 mL), warmed to rt and diluted with water. The organic layer was separated and the aqueous layer was extracted with dichloromethane. The combined organic layers were washed brine, dried over MgSO_4 , filtered

and concentrated. The crude residue was purified by flash chromatography over silica gel (petroleum ether/AcOEt: 8/2) to yield the cyclized product **7** as a white solid (2.4 g, 7.2 mmol, 72 %). Mp: 110 °C; $[\alpha]_{\text{D}}^{20}$ -38 (*c* 2.3, CHCl₃); ¹H NMR (300 MHz, DMSO-*d*₆, 343 K): δ 4.66 (s, 1H), 4.18 (br. s, 1H), 3.67 (app. q, *J* = 6.8 Hz, 1H), 3.53 (br. s, 2H), 3.39 (td, *J* = 9.8, 7.3 Hz, 1H), 3.23 (td, *J* = 9.8, 2.6 Hz, 1H), 1.92-2.04 (m, 1H), 1.62-1.73 (m, 1H), 1.42 (s, 9H), 0.88 (s, 9H), 0.05, 0.03 (s, s, 3H, 3H); ¹³C NMR (75 MHz, DMSO-*d*₆, 343 K): δ 153.4, 77.7, 71.7 (br.), 66.6, 61.8 (br.), 44.4, 31.4 (br.), 27.8, 25.3, 17.4, -5.9, -6.0; IR (KBr) ν_{max} 3375, 2925, 2756, 2592, 1675, 1424, 1096, 835 cm⁻¹; ESIMS (positive mode): 354.3, 298.3, 254.2; HRMS (CI, NH₃) *m/z* calcd for C₁₆H₃₄NO₄Si [M+H]⁺ 332.2257, found 332.2252.

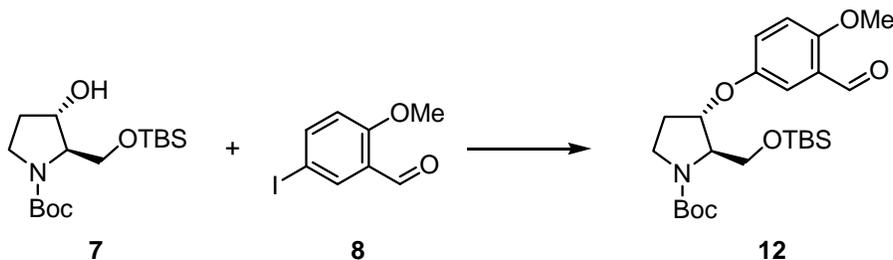


2-Hydroxy-5-iodo-benzaldehyde.^{S4} To a solution of salicylaldehyde (20.0 g, 164 mmol) in glacial acetic acid (200 mL) was added iodine monochloride (9.2 mL, 181 mmol). The resulting brown mixture was stirred at rt for 2 days and at 40 °C for a day, concentrated under vacuum, and the residue was diluted with water and dichloromethane. The organic layer was separated and the aqueous layer was extracted with dichloromethane. The combined organic layers were successively washed with 10% wt aqueous sodium thiosulfate solution and brine, dried over MgSO₄, filtered and concentrated under vacuum to yield a yellow-brown solid which was stored in the dark (38.0 g, 153 mmol, 93%). Mp: 83 °C; ¹H NMR (300 MHz, CDCl₃): δ 10.98 (s, 1H), 9.87 (d, *J* = 0.3 Hz, 1H), 7.88 (d, *J* = 2.2 Hz, 1H), 7.80 (dd, *J* = 2.2, 8.7 Hz, 1H), 6.84 (d, *J* = 8.7 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃): δ 195.5, 161.3, 145.4, 141.9, 122.6, 120.3, 80.5; IR (KBr) ν_{max} 3216, 1670, 1460, 1270, 1265, 1142 cm⁻¹; ESIMS (positive mode): 248.3, 219.3, 127.5; HRMS (CI, NH₃) *m/z* calcd for C₇H₅IO₂ [M]⁺ 247.9334, found 247.9329.

^{S4} procedure adapted from: Y. J. Cho, K. Y. Rho, S. R. Keum, S. H. Kim, C. M. Yoon, *Synth. Commun.* **1999**, *29*, 2061-2068.

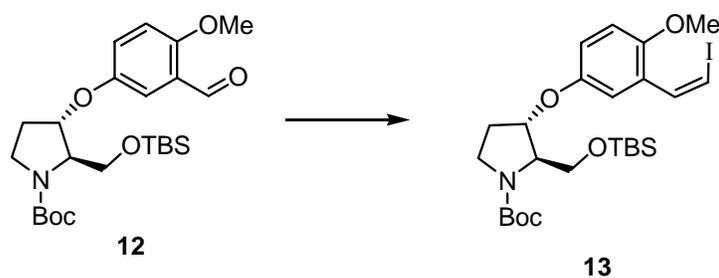


5-Iodo-2-methoxy-benzaldehyde 8. To a solution of 2-hydroxy-5-iodo-benzaldehyde (29.6 g, 119 mmol) in dry acetone (700 mL) was added potassium carbonate (24.9 g, 180 mmol) and dimethylsulfate (12.5 mL, 132 mmol). The resulting orange mixture was refluxed for 2 hours, cooled to rt, concentrated in vacuo and diluted with water and CH₂Cl₂. The aqueous layer was extracted with CH₂Cl₂, combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated. The crude residue was finally washed with pentane to give the desired ether as a brown solid which was stored in the dark (31.2 g, 180 mmol, quant.). Mp: 145 °C; ¹H NMR (300 MHz, CDCl₃): δ 10.25 (s, 1H), 7.98 (d, *J* = 2.3 Hz, 1H), 7.71 (dd, *J* = 2.3, 8.8 Hz, 1H), 6.70 (d, *J* = 8.8 Hz, 1H), 3.83 (s, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 188.3, 161.5, 144.1, 136.9, 126.5, 114.3, 83.0, 55.9; IR (KBr) ν_{max} 2960, 2865, 1689, 1474 cm⁻¹; ESIMS (positive mode): 262.5, 244.3, 202.7; HRMS (CI, NH₃) *m/z* calcd for C₈H₇IO₂ [M+H]⁺ 261.9491, found 261.9490.



(2*R*,3*S*)-1-(*tert*-Butoxycarbonyl)-2-(*tert*-butyldimethylsilyloxy)methyl-3-(3-formyl-4-methoxy-phenoxy)-pyrrolidine 12. A 15 mL pressure tube was charged with 5-iodo-2-methoxy-benzaldehyde **8** (1.3 g, 5.0 mmol), alcohol **7** (1.8 g, 5.4 mmol), cesium carbonate (3.25 g, 10.0 mmol), 1,10-phenanthroline (180 mg, 1.0 mmol) and copper(I) iodide (95 mg, 0.5 mmol). Toluene (3 mL) was added, the pressure tube was closed and the brownish suspension was heated to 125°C for 24 hours. Another portion of 5-iodo-2-methoxy-benzaldehyde **8** (650 mg, 2.5 mmol) was then added and the reaction mixture was heated for an additional 24 hours and cooled to rt. Crude reaction mixture was finally filtrated over a plug of silica gel (washed with

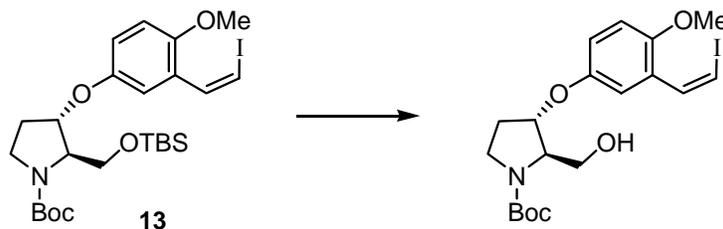
AcOEt), concentrated and purified by flash chromatography over silica gel (gradient from CH₂Cl₂ to CH₂Cl₂/EtOH: 95/5) to yield the aryl ether as a pale yellow sticky oil (1.9 g, 4.1 mmol, 75 %); $[\alpha]_D^{20}$ -5 (*c* 1.2, CHCl₃); ¹H NMR (300 MHz, DMSO-*d*₆, 345 K): δ 10.33 (s, 1H), 7.27 (d, *J* = 3.1 Hz, 1H), 7.22-7.25 (m, 1H), 7.13 (d, *J* = 8.8 Hz, 1H), 4.82 (d, *J* = 4.1 Hz, 1H), 3.78-3.93 (m, 2H), 3.88 (s, 3H), 3.58-3.63 (m, 1H), 3.34-3.51 (m, 2H), 2.23 (app. qt, *J* = 9.8, 4.7 Hz, 1H), 2.10 (app. dd, *J* = 13.4, 6.2 Hz, 1H), 1.42 (s, 9H), 0.88 (s, 9H), 0.06 (s, 6H); ¹³C NMR (75 MHz, DMSO-*d*₆, 345 K): δ 186.9, 155.2, 152.2, 149.4, 123.8, 123.0, 113.0, 112.5, 78.8 (br.), 77.2, 62.6, 60.7 (br.), 55.0, 43.3, 27.0 (br.), 26.7, 24.2, 16.4, -7.1; IR (neat) ν_{\max} 2883, 2755, 1694, 1490, 1398 cm⁻¹; ESIMS (positive mode): 970.0, 953.7, 599.5, 583.5, 488.4; HRMS (CI, NH₃) *m/z* calcd for C₂₄H₃₉NO₆Si [M]⁺ 465.2547, found 465.2552.



[2*R*,3*S*,3(3*Z*)]-1-(*tert*-Butoxycarbonyl)-2-(*tert*-butyldimethylsilyloxy)methyl-3-[3-(2-iodovinyl)-4-methoxy-phenoxy]-pyrrolidine **13.** To a suspension of methyltriphenylphosphonium iodide^{S5} (1.05 g, 1.98 mmol) in THF (11 mL) was added dropwise at rt a solution of NaHMDS (2.0 M solution in THF, 990 μ L, 1.98 mmol). The resulting red-orange solution was stirred at rt for 20 min and cooled to -78 °C before adding HMPA (1.3 mL) and a solution of **12** (700 mg, 1.5 mmol) in THF (7 mL). The reaction mixture was stirred at -78 °C for two hours and quenched at -78 °C by addition of a saturated aqueous solution of NaHCO₃. The mixture was warmed to rt, diluted with Et₂O and filtered through a plug of Celite[®] which was thoroughly washed with ether. The biphasic filtrate was separated and the organic layer was dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (AcOEt/petroleum ether: 1/9) to give the desired vinyl iodide **13** as a yellow oil (860 mg, 1.46 mmol, 97 %). Diastereoisomeric excess was determined by analysis of crude ¹H NMR

^{S5} M. C. Hillier, A. T. Price, A. I. Meyers, *J. Org. Chem.* **2001**, *66*, 6037-6045.

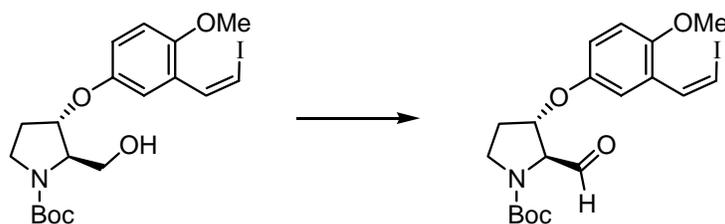
spectra and was found to be higher than 95%. $[\alpha]_{\text{D}}^{20}$ -4 (*c* 1.2, CHCl_3); ^1H NMR (300 MHz, $\text{DMSO-}d_6$, 333 K): δ 7.27 (d, $J = 8.5$ Hz, 1H), 7.14 (d, $J = 2.5$ Hz, 1H), 6.86-6.87 (m, 2H), 6.66 (d, $J = 8.5$ Hz, 1H), 4.68 (d, $J = 4.1$ Hz, 1H), 3.74 (A of ABX syst., $J = 3.2, 10.8$ Hz, 1H), 3.68-3.73 (m, 1H), 3.66 (s, 3H), 3.45-3.55 (m, 1H), 3.25-3.37 (m, 2H), 2.07-2.20 (m, 1H), 1.92-1.98 (m, 1H), 1.32 (s, 9H), 0.77 (s, 3H), 0.79 (s, 6H), 0.03 (s, 3H), -0.12 (s, 3H); ^{13}C NMR (75 MHz, $\text{DMSO-}d_6$, 333 K): δ 153.3, 151.1, 149.6, 134.2, 126.3, 117.0, 115.9, 112.4, 82.5, 80.3, 78.4, 63.6, 61.7 (br.), 55.8, 44.4, 28.5 (br.), 27.8, 25.4, 17.4, -3.6, -5.9; IR (neat) ν_{max} 2960; 1685; 1486; 1394; 1235; 1117; 830 cm^{-1} ; CIMS (NH_3 gas): 590; 534; 476; HRMS (CI, NH_3) m/z calcd for $\text{C}_{25}\text{H}_{40}\text{INO}_5\text{Si}$ $[\text{M}]^+$ 589.1720, found: 589.1724.



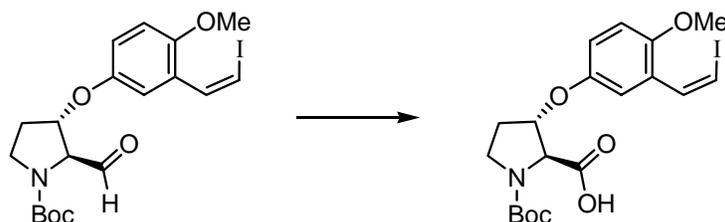
[2*R*,3*S*,3(3*Z*)]-1-(*tert*-Butoxycarbonyl)-2-hydroxymethyl-3-[3-(2-iodovinyl)-4-methoxy-

phenoxy]-pyrrolidine. A solution of **13** (860 mg, 1.46 mmol) in THF (18 mL) was treated with a solution of TBAF (1M solution in THF, 2.2 mL, 2.2 mmol) at -10 °C. The resulting light yellow mixture was warmed to rt over 50 minutes and quenched with water. The aqueous layer was extracted with Et_2O and the combined organic layers were washed with brine, dried over MgSO_4 , filtered and concentrated to give the crude unprotected alcohol (contaminated with *tert*-butyldimethyl-silanol) as an orange oil (694 mg, 1.46 mmol, quant.) which was used without further purification in the next step. An analytical sample was purified by flash chromatography over silica gel (EtOH/DCM : 3/97). $[\alpha]_{\text{D}}^{20}$ -7 (*c* 1.1, CHCl_3); ^1H NMR (300 MHz, $\text{DMSO-}d_6$, 333 K): δ 7.37 (d, $J = 8.5$ Hz, 1H), 7.25 (d, $J = 2.3$ Hz, 1H), 6.98-6.99 (m, 2H), 6.76 (d, $J = 8.5$ Hz, 1H), 4.78-4.90 (m, 2H), 3.82-3.86 (app. dd, $J = 3.7, 7.6$ Hz, 1H), 3.76 (s, 3H), 3.59-3.71 (m, 1H), 3.31-3.43 (m, 3H), 2.14-2.28 (m, 1H), 1.99-2.09 (m, 1H), 1.41 (s, 9H); ^{13}C NMR (75 MHz, $\text{DMSO-}d_6$, 333 K): δ 153.5, 150.9, 149.7, 134.2, 126.3, 116.7, 115.7, 112.4, 82.5, 79.4 (br.), 78.2, 64.1, 60.3 (br.), 55.8, 44.2, 28.5 (br.), 27.9; IR (neat) ν_{max} 3406, 2950, 1675, 1491, 1414,

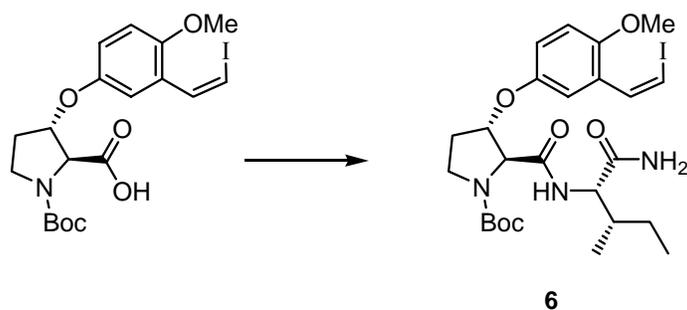
1214, 1035, 866, 764 cm^{-1} ; ESIMS (positive mode): 514.3, 498.2; HRMS (CI, NH_3) m/z calcd for $\text{C}_{19}\text{H}_{27}\text{INO}_5$ $[\text{M}+\text{H}]^+$ 476.0934, found: 476.0921.



[2R,3S,3(Z)]-1-(tert-Butoxycarbonyl)-2-formyl-3-[3-(2-iodovinyl)-4-methoxy-phenoxy]-pyrrolidine. DMSO (265 μL , 3.7 mmol) was added to a solution of oxalyl chloride (270 μL , 3.1 mmol) in dichloromethane (10 mL) at $-78\text{ }^\circ\text{C}$. The resulting solution was stirred at $-78\text{ }^\circ\text{C}$ for 30 minutes and a solution of [2R,3S,3(Z)]-1-(tert-butoxycarbonyl)-2-hydroxymethyl-3-[3-(2-iodovinyl)-4-methoxy-phenoxy]-pyrrolidine (694 mg, 1.46 mmol) in dichloromethane (15 mL) was added dropwise *via* cannula. The reaction mixture was stirred for 40 minutes at $-78\text{ }^\circ\text{C}$ before adding triethylamine dropwise (880 μL , 6.3 mmol), and the mixture was warmed to $-10\text{ }^\circ\text{C}$ over 2 hours. The reaction was next quenched at $-10\text{ }^\circ\text{C}$ with water and diluted with CH_2Cl_2 . The aqueous layer was extracted with CH_2Cl_2 and the combined organic layers were washed with brine, dried over MgSO_4 , filtered and concentrated to give the desired aldehyde as an orange oil (691 mg, 1.46 mmol, quant.) which was used without purification in the next step. An analytical sample was purified by flash chromatography over silica gel (AcOEt/petroleum ether: 35/65) to give a colorless oil. $[\alpha]_{\text{D}}^{20}$ -27 (c 1.3, CHCl_3); ^1H NMR (300 MHz, $\text{DMSO}-d_6$, 335 K): δ 9.60 (s, 1H), 7.38 (d, $J = 8.5$ Hz, 1H), 7.30 (d, $J = 2.3$ Hz, 1H), 7.00-7.01 (m, 2H), 6.78 (d, $J = 8.5$ Hz, 1H), 5.00 (br. s, 1H), 4.35 (app. s, 1H), 3.77 (s, 3H), 3.50-3.57 (m, 2H), 2.03-2.12 (m, 2H), 1.40 (s, 9H); ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$, 333 K): δ 198.7, 151.4, 150.8 and 149.9 (rotamers), 149.7, 134.1, 126.4, 117.1, 116.3, 112.3, 82.9, 79.2, 78.1 (br.), 70.1, 55.8, 44.3, 29.6 (br.), 27.7; IR (neat) ν_{max} 2971, 1726, 1675, 1482, 1388, 1276, 1045, 748 cm^{-1} ; ESIMS (positive mode): 512.1, 496.1, 440.0, 312.8, 311.2; HRMS (CI, NH_3) m/z calcd for $\text{C}_{19}\text{H}_{25}\text{INO}_5$ $[\text{M}+\text{H}]^+$ 474.0777, found: 474.0773.



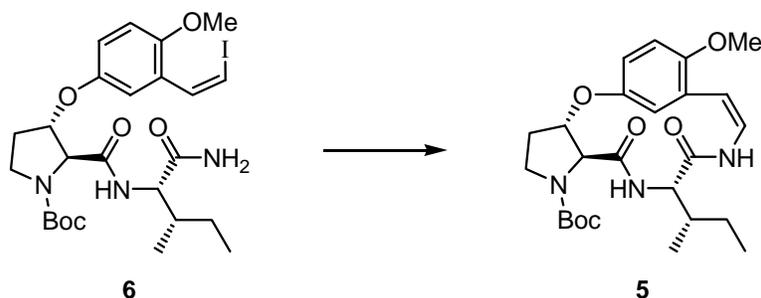
[2R,3S,3(Z)]-1-(tert-Butoxycarbonyl)-3-[3-(2-iodovinyl)-4-methoxyphenoxy]-proline. To a solution of [2R,3S,3(Z)]-1-(tert-butoxycarbonyl)-2-formyl-3-[3-(2-iodovinyl)-4-methoxyphenoxy]-pyrrolidine (691 mg, 1.46 mmol) in a mixture of THF (5 mL) and *tert*-butanol (14 mL) was added 2-methylprop-2-ene (90 %, 1.6 mL, 13.2 mmol), followed by a solution of sodium chlorite (80 %, 400 mg, 3.5 mmol) and sodium dihydrogen phosphate dihydrate (485 mg, 3.1 mmol) in water (10 mL). The yellow reaction mixture was then stirred for 1 hour, carefully quenched with a 1M HCl solution and diluted with ether. The aqueous layer was extracted with ether, combined organic layers were dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (EtOH/DCM: 6/94) to give the free carboxylic acid as a yellow oily solid (486 mg, 0.99 mmol, 68 % over three steps). $[\alpha]_D^{20}$ -24 (c 1, CHCl₃); ¹H NMR (300 MHz, DMSO-*d*₆, 345 K): δ 7.38 (d, *J* = 8.5 Hz, 1H), 7.29 (d, *J* = 2.3 Hz, 1H), 7.01-7.02 (m, 2H), 6.77 (d, *J* = 8.5 Hz, 1H), 4.90 (br. s, 1H), 4.28 (app. s, 1H), 3.77 (s, 3H), 3.44-3.60 (m, 2H), 2.09-2.19 (m, 2H), 1.39 (br. s, 9H); ¹³C NMR (75 MHz, DMSO-*d*₆, 345 K): δ 170.7, 153.0, 151.3, 149.2, 134.2, 126.5, 116.7, 116.2, 112.5, 82.7, 80.1 (br.), 78.8, 64.5 (br.), 55.8, 44.0, 28.9 (br.), 27.7; IR (neat) ν_{\max} 3441, 2919, 1731, 1680, 1419, 1214 cm⁻¹; HRMS (CI, NH₃) *m/z* calcd for C₁₉H₂₅INO₆ [M+H]⁺ 490.0727, found: 490.0733.



[2*R*,3*S*,3(3*Z*)]-1-(*tert*-Butoxycarbonyl)-3-[3-(2-iodovinyl)-4-methoxy-phenoxy]-prolyl]-isoleucinamide **6.**

To a solution of [2*R*,3*S*,3(3*Z*)]-1-(*tert*-butoxycarbonyl)-3-[3-(2-iodovinyl)-4-methoxy-phenoxy]-proline (1.09 g, 2.23 mmol) and isoleucinamide acetate^{S6} (424 mg, 2.23 mmol) in DMF (20 mL) was added 1-hydroxybenzotriazole (HOBt, 316 mg, 2.34 mmol). 1-(3-Dimethylaminopropyl)-3-ethyl-carbodiimide hydrochloride (EDC, 470 mg, 2.45 mmol) and *N*-methylmorpholine (620 μ L, 5.60 mmol) were next added at 0 °C and the solution was stirred for 16 hours while progressively warmed to rt. The yellow reaction mixture was quenched with water and diluted with ether. The aqueous layer was extracted with ether and the combined organic layers were successively washed with a 1M HCl aqueous solution, saturated aqueous solution of NaHCO₃ and brine, dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (AcOEt/petroleum ether: 9/1) to give the desired peptide **6** (992 mg, 1.65 mmol, 75 %) as a pale yellow solid. Mp: 87 °C; $[\alpha]_{\text{D}}^{20}$ -31 (*c* 1, CHCl₃); ¹H NMR (300 MHz, DMSO-*d*₆, 343 K): δ 7.65 (d, *J* = 8.5 Hz, 1H), 7.38 (d, *J* = 8.5 Hz, 1H), 7.28 (d, *J* = 2.3 Hz, 1H), 7.00-7.06 (m, 2H), 6.80-7.20 (br. s, 2H), 6.77 (d, *J* = 8.5 Hz, 1H), 4.83 (br. s, 1H), 4.36, 4.43 (rotamers, s, s, 1H), 4.19 (dd, *J* = 6.7, 8.4 Hz 1H), 3.80, 3.77 (rotamers, s, s, 3H), 3.46-3.56 (m, 2H), 2.05-2.10 (m, 2H), 1.72-1.84 (m, 1H), 1.47-1.50 (m, 1H), 1.41 (s, 9H), 1.05-1.20 (m, 1H), 0.87 (t, *J* = 6.4 Hz, 3H), 0.84 (d, *J* = 7.3 Hz, 3H); ¹³C NMR (75 MHz, DMSO-*d*₆, 343 K): δ 172.1, 168.6, 153.5, 151.2, 149.3, 134.1, 126.3, 116.8, 116.0, 112.5, 83.7, 79.5 (br.), 79.0, 65.7 (br.), 56.6, 55.9, 44.3, 36.4, 28.8 (br.), 27.7, 23.9, 15.1, 10.6; IR (KBr) ν_{max} 3314, 2960, 2720, 1680, 1486, 1394, 1214, 1168, 1025 cm⁻¹; ESIMS (positive mode): 624.2, 640.2; HRMS (CI, NH₃) *m/z* calcd for C₂₅H₃₇IN₃O₆ [M+H]⁺ 602.1727, found: 602.1741.

^{S6} T. Moriguchi, T. Yanagi, M. Kunimori, T. Wada, M. Sekine, *J. Org. Chem.* **2000**, *65*, 8229-8238.

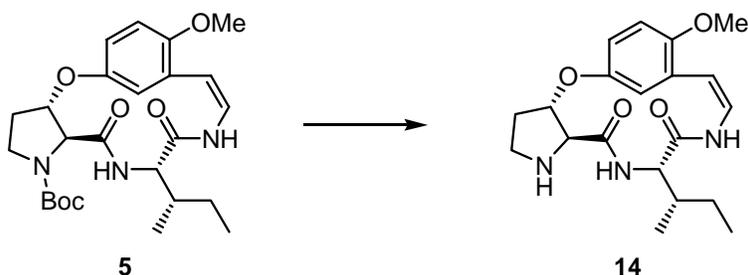


cyclopeptide core 5. A 100 mL flask was charged with iodo-amide **6** (350 mg, 0.59 mmol), copper(I) iodide (23 mg, 0.12 mmol) and cesium carbonate (285 mg, 0.88 mmol). The flask was evacuated under high vacuum, backfilled with argon and closed with a rubber septa. Dry and degassed THF (78 mL) and *N,N'*-dimethylethylene-1,2-diamine (26 μ L, 0.24 mmol) were next added, the rubber septa was replaced by a glass stopper and the light blue suspension was heated to 60°C for 20 hours. The reaction mixture was cooled to rt and filtrated over a plug of silica gel (washed with AcOEt) and concentrated. The crude residue was purified by flash chromatography over silica gel (gradient from Et₂O/EtOH: 99/1 to Et₂O/EtOH: 95/5) to give the recovered starting peptide **6** (70 mg, 0.12 mmol, 20 %) and the desired cyclized product **5** (193 mg, 0.41 mmol, 70 %, 89 % based on recovered starting material) as a white solid. Mp: 188 °C; $[\alpha]_D^{20}$ -436 (*c* 0.54, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 8.43 (d, *J* = 11.2 Hz, 1H), 7.22 (d, *J* = 5.0 Hz, 1H), 6.87 (dd, *J* = 6.9, 11.2 Hz, 1H), 6.83 (d, *J* = 9.2 Hz, 1H), 6.74 (dd, *J* = 2.9, 8.9 Hz, 1H), 6.65 (d, *J* = 2.9 Hz, 1H), 5.85 (d, *J* = 9.1 Hz, 1H), 5.43 (dt, *J* = 2.7, 8.1 Hz, 1H), 4.25 (app. t, *J* = 4.6 Hz, 1H), 4.15 (d, *J* = 2.6 Hz, 1H), 3.70-3.78 (m, 1H), 3.73 (s, 3H), 3.30-3.39 (m, 1H), 2.39-2.50 (m, 1H), 2.14-2.24 (m, 1H), 2.00-2.10 (m, 1H), 1.38 (s, 9H), 1.05-1.32 (m, 2H), 0.93 (d, *J* = 7.0 Hz, 3H), 0.81 (t, *J* = 9.8 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 171.4, 167.6, 155.3, 151.6, 151.4, 124.5, 121.7, 117.8, 113.9, 111.5, 106.7, 81.2, 78.0, 64.9, 60.4, 56.2, 46.0, 35.3, 32.2, 28.4, 24.6, 16.2, 11.9; IR (KBr) ν_{\max} 3319, 2919, 1706, 1650, 1506, 1404, 1214, 1163, 1122, 1030, 764 cm⁻¹; CIMS (NH₃ gas): 491.0, 473.0, 418.0 ; HRMS (CI, NH₃) *m/z* calcd for C₂₅H₃₆N₃O₆ [M+H]⁺ 474.2604, found: 474.2607.

Alternate procedure for macrocyclization using CuTc. A 15 mL pressure tube was charged with iodo-amide **6** (50 mg, 0.083 mmol), copper(I) thiophenecarboxylate^{S7} (CuTc, 5.0 mg, 0.025

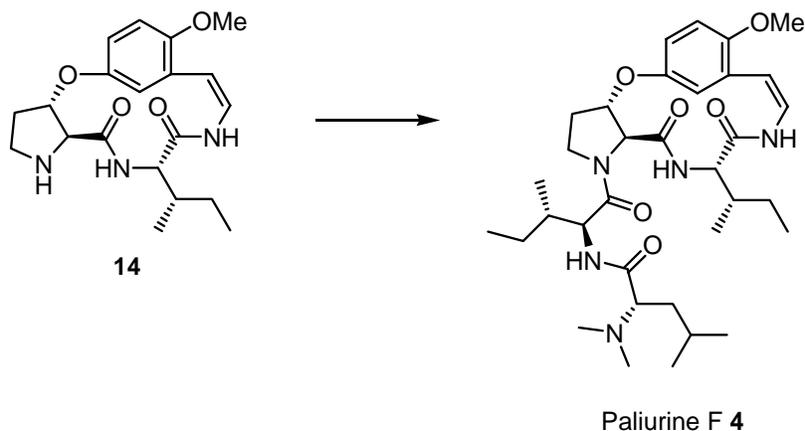
^{S7} prepared according to G. D. Allred, L. S. Liebeskind, *J. Am. Chem. Soc.* **1996**, *118*, 2748-2749.

mmol) and cesium carbonate (40.7 mg, 0.125 mmol). The pressure was evacuated under high vacuum, backfilled with argon and closed with a rubber septa. Dry and degassed NMP (11.5 mL) was next added and the tube was sealed and the yellow suspension was heated to 90°C for 20 hours. The reaction mixture was cooled to rt, and diluted with ether and water. The aqueous layer was extracted with ether and the combined organic layers were washed brine, dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (gradient from Et₂O/EtOH: 99/1 to Et₂O/EtOH: 95/5) to give the desired cyclized product **5** (23.5 mg, 0.050 mmol, 60 %) as a white solid.



deprotected cyclopeptide core 14. To a solution of **5** (100 mg, 0.21 mmol) in dichloromethane (4.7 mL) were added at -10 °C 2,6-lutidine (25 μ L, 21.1 mmol) and a solution of trimethylsilyl trifluoromethanesulfonate (1.4 M solution in dichloromethane, 600 μ L, 84.0 mmol). The resulting light pink solution was stirred for 1 hour while progressively warmed to 0 °C. The mixture was next hydrolyzed at 0 °C by addition of a saturated aqueous solution of NaHCO₃ and diluted with dichloromethane. The aqueous layer was extracted with dichloromethane and the combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (AcOEt/EtOH/Et₃N: 85/14/1) to give the desired *N*-deprotected macrocycle **14** (63 mg, 0.17 mmol, 80 %) as a white solid. Mp: 218 °C; $[\alpha]_D^{20}$ -452 (*c* 0.49, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 8.50 (br. s, 1H), 6.91 (dd, *J* = 9.2, 11.3 Hz, 1H), 6.92 (d, *J* = 9.1 Hz, 1H), 6.82 (dd, *J* = 2.9, 9.0 Hz, 1H), 6.65 (br. s, 1H), 5.95 (d, *J* = 8.9 Hz, 1H), 5.09-5.11 (m, 1H), 4.32 (br. s, 1H), 3.79 (s, 3H), 3.43 (br. s, 1H), 3.13-3.20 (m, 1H), 2.90-2.95 (m, 1H), 2.16-2.26 (m, 3H), 1.95-2.03 (br. m, 1H), 1.37-1.47 (m, 1H), 1.02-1.14 (m, 1H), 0.94 (d, *J* = 6.9 Hz, 3H), 0.87 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 173.0, 167.0 (br.), 151.2, 151.1, 124.1, 121.5, 117.5, 113.8, 111.1, 107.2, 80.9,

69.6, 60.1 (br.), 56.1, 47.5 (br.), 35.6 (br.), 32.0, 25.3, 16.0, 11.7; ESIMS (positive mode): 412.3, 396.4, 374.4, 359.4, 346.4 ; HRMS (CI, NH₃) m/z calcd for C₂₀H₂₈N₃O₄ [M+H]⁺ 374.2080, found: 374.2069.



Paliurine F 4. To a cooled solution of *N*-Fmoc-L-isoleucine (90 mg, 0.25 mmol) and 1-hydroxyazabenzotriazole (HOAt, 42 mg, 0.31 mmol) in DMF (2 mL) were added *O*-(7-azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HATU, 90 mg, 0.24 mmol) and diisopropylamine (125 μ L, 0.72 mmol) at 0 °C. The resulting yellow solution was stirred at 0 °C for 20 minutes and added dropwise *via* cannula to a cooled solution of **14** (63 mg, 0.17 mmol) in DMF (2 mL) at 0 °C. The flask containing the activated acid was rinsed with an additional portion of DMF (2 mL) which was cannulated into the solution of the amine. The resulting yellow solution was warmed to rt and stirred overnight. The mixture was quenched at 0 °C with a saturated aqueous solution of NH₄Cl and diluted with ether. The aqueous layer was extracted with ether and the combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated to give an orange oil which was used in the next step without further purification.

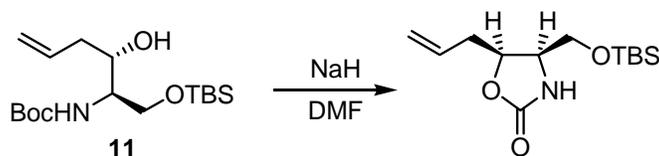
This oil was dissolved in CH₃CN (2.5 mL) and diethylamine was added (630 μ L, 6.1 mmol) at rt. After 40 minutes, the crude yellow mixture was concentrated under vacuum and residue was immediately engaged in the next step without further purification.

To a cooled solution of *N,N*-dimethyl-L-leucine (50 mg, 0.25 mmol) and 1-hydroxyazabenzotriazole (HOAt, 44 mg, 0.32 mmol) in DMF (2 mL) were added

O-(7-azabenzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HATU, 90 mg, 0.24 mmol) and diisopropylethylamine (125 μ L, 0.72 mmol) at 0 °C. The resulting yellow solution was stirred at 0 °C for 20 minutes and added dropwise *via* cannula to a solution of the previously deprotected amine in DMF (2 mL) at 0 °C. The flask containing the activated acid was rinsed with an additional portion of DMF (2 mL) which was cannulated into the solution of the amine. The yellow solution was warmed to rt and stirred overnight. The mixture was hydrolyzed at 0 °C with a saturated aqueous solution of NH₄Cl and diluted with ether. The aqueous layer was extracted with ether and the combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated. The crude residue was purified by flash chromatography over silica gel (AcOEt/EtOH: 99.5/0.5) to give the desired synthetic paliurine F **4** (61 mg, 97 μ mol, 57 % over three steps) as a white solid. *R*_f: 0.26 (Merck-Kiesegel 60F₂₅₄ TLC plates, AcOEt/EtOH: 99/1); Mp: 192 °C {Lit.: unreported}; [α]_D²⁰ -468 (*c* 1.1, CH₃CN) {Lit.: natural paliurine F: [α]_D²⁰: -323 (*c* 1.0, CH₃CN)}^{S8}; ¹H NMR (300 MHz, CDCl₃):^{S9} δ 8.46 (d, *J* = 11.4 Hz, 1H), 7.41 (d, *J* = 9.0 Hz, 1H), 7.20 (d, *J* = 5.0 Hz, 1H), 6.93 (dd, *J* = 9.2, 11.3 Hz, 1H), 6.88 (d, *J* = 9.2 Hz, 1H), 6.80 (dd, *J* = 2.9, 9.0 Hz, 1H), 6.69 (d, *J* = 2.9 Hz, 1H), 5.92 (d, *J* = 9.0 Hz, 1H), 5.53 (dt, *J* = 3.2, 7.3 Hz, 1H), 4.56 (app. t, *J* = 8.5 Hz, 1H), 4.49 (d, *J* = 3.2 Hz, 1H), 4.38 (dt, *J* = 2.2, 8.5 Hz, 1H), 4.27 (app. t, *J* = 4.4 Hz, 1H), 3.78 (s, 3H), 3.55 (app. dt, *J* = 6.6, 10.8 Hz, 1H), 2.89 (dd, *J* = 5.3, 8.6 Hz, 1H), 2.59 (ddt, *J* = 13.1, 2.3, 7.1 Hz, 1H), 2.24-2.38 (m, 1H), 2.25 (s, 6H), 2.02-2.15 (m, 1H), 1.65-1.83 (m, 1H), 1.47-1.60 (m, 1H), 1.40-1.49 (m, 1H), 1.36-1.46 (m, 1H), 1.31-1.39 (m, 1H), 1.03-1.18 (m, 2H), 1.06-1.10 (m, 1H), 0.96 (d, *J* = 7.0 Hz, 3H), 0.93 (d, *J* = 6.4 Hz, 3H), 0.91 (d, *J* = 6.3 Hz, 3H), 0.87 (t, *J* = 7.5 Hz, 3H), 0.85 (t, *J* = 7.5 Hz, 3H), 0.82 (d, *J* = 6.6 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃):^{S9} δ 174.0, 172.0, 170.4, 167.2, 151.6, 151.2, 124.4, 121.6, 117.8, 113.9, 111.3, 106.8, 76.9, 67.4, 64.5, 60.4, 56.1, 53.8, 46.9, 42.4, 37.5, 37.1, 35.2, 32.7, 26.0, 24.8, 24.6, 23.4, 22.2, 16.3, 15.4, 11.8, 10.8; UV λ_{\max} (log ϵ): 220 (4.62), 296 (4.09), 320 (4.39); IR (KBr) ν_{\max} 3406, 3324, 2955, 2883, 1690, 1640, 1506, 1429, 1261, 1225, 1178, 1035, 820, 764 cm⁻¹ CIMS (NH₃ gas): 628, 487, 416, 402, 374, 324, 227; HRMS (FAB) *m/z* calcd for C₃₄H₅₄N₅O₆ [M+H]⁺ 628.4074, found: 628.4061.

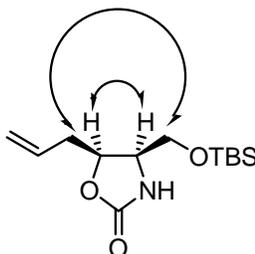
^{S8} H. Y. Lin, C. H. Chen, B. J. You, K. C. S. C. Liu, S. S. Lee, *J. Nat. Prod.* **2000**, *63*, 1338-1343.

^{S9} See Paliurine F Spectral Data Compared to Reported Data page S18.

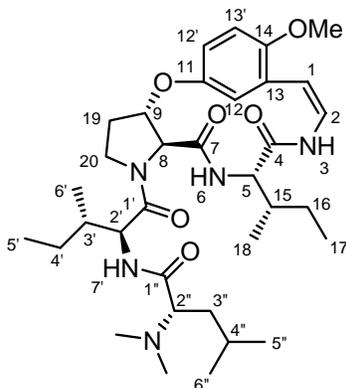
Assignment of stereochemistry for Alcohol **11**.

(4*R*,5*S*)-5-Allyl-4-(tert-butyl-dimethyl-silyloxymethyl)-oxazolidin-2-one. A solution of **11** (200 mg, 0.58 mmol) in DMF (6 mL) was treated with sodium hydride (60% wt in mineral oil, 25.5 mg, 0.64 mmol) at 0°C. The resulting mixture was slowly warmed to rt over 1 hour and quenched by addition of a saturated aqueous solution of NH₄Cl. The organic layer was separated and the aqueous layer was extracted with ether. The combined organic layers were washed with brine, dried over MgSO₄, filtered and concentrated under vacuum. The crude residue was purified by flash chromatography over silica gel (AcOEt/petroleum ether: 35/65) to give the corresponding oxazolidinone (69 mg, 0.25 mmol, 43 %) as a colorless oil. $[\alpha]_D^{20}$ 19 (*c* 0.4, CHCl₃); ¹H NMR (300 MHz, CDCl₃): δ 6.05 (s, 1H), 5.69-5.90 (m, 1H), 5.07-5.19 (m, 2H), 4.66 (td, *J* = 7.9, 5.6 Hz, 1H), 3.76-3.86 (m, 1H), 3.60-3.70 (m, 2H), 2.53-2.63 (m, 1H), 2.41-2.50 (m, 1H), 0.87 (s, 9H), 0.05 (s, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 159.6, 133.1, 118.3, 78.4, 61.7, 56.6, 33.5, 25.8, 18.2, -5.4; IR (neat) ν_{\max} 3375, 2963, 1716 1164 cm⁻¹; HRMS (CI, NH₃) *m/z* calcd for C₁₃H₂₆NO₃Si [M+H]⁺ 272.1682, found 272.1671.

Selected nuclear Overhauser effects:



Paliurine F Spectral Data Compared to Reported Data

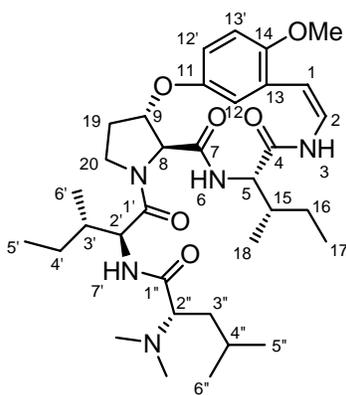
¹H NMR (Synthetic at 300 MHz; Natural at 400 MHz)

Paliurine F

Position	Synthetic		Natural		$\Delta\delta$
	δ	multiplicity	δ	multiplicity	
1 (1H)	5.91	d, $J = 9.0$ Hz	5.91	d, $J = 9.0$ Hz	0.00
2 (1H)	6.92	dd, $J = 11.3, 9.2$ Hz	6.92	dd, $J = 11.3, 9.0$ Hz	0.00
3 (1H)	8.45	d, $J = 11.4$ Hz	8.44	d, $J = 11.3$ Hz	0.01
5 (1H)	4.26	app. t, $J = 4.4$ Hz	4.26	dd, $J = 4.4, 4.8$ Hz	0.00
6 (1H)	7.19	d, $J = 5.0$ Hz	7.19	d, $J = 4.8$ Hz	0.00
8 (1H)	4.48	d, $J = 3.2$ Hz	4.48	d, $J = 3.2$ Hz	0.00
9 (1H)	5.52	dt, $J = 3.2, 7.3$ Hz	5.52	dt, $J = 3.2, 7.2$ Hz	0.00
12 (1H)	6.68	d, $J = 2.9$ Hz	6.68	d, $J = 2.9$ Hz	0.00
12' (1H)	6.79	dd, $J = 9.0, 2.9$ Hz	6.79	dd, $J = 9.0, 2.9$ Hz	0.00
13' (1H)	6.87	d, $J = 9.2$ Hz	6.87	d, $J = 9.0$ Hz	0.00
15 (1H)	2.01-2.14	m	2.08	m	0.00
16 (1H)	1.02-1.17	m	1.09	m	0.01
16 (1H)	1.30-1.38	m	1.34	m	0.00
17 (3H)	0.86	t, $J = 7.5$ Hz	0.86	t, $J = 7.4$ Hz	0.00
18 (3H)	0.95	d, $J = 7.0$ Hz	0.95	d, $J = 7.0$ Hz	0.00
19 α (1H)	2.23-2.37	m	2.32	m	-0.02
19 β (1H)	2.58	ddt, $J = 13.1, 2.3, 7.1$ Hz	2.58	ddt, $J = 12.9, 2.3, 6.8$ Hz	0.00

20 α (1H)	4.37	dt, $J = 2.2, 8.5$ Hz	4.37	dt, $J = 2.1, 8.4$ Hz	0.00
20 β (1H)	3.54	app. dt, $J = 6.6, 10.8$ Hz	3.54	m	0.00
OMe (3H)	3.77	s	3.78	s	-0.01
2' (1H)	4.55	app. t, $J = 8.5$ Hz	4.55	dd, $J = 8.9, 8.5$ Hz	0.00
3' (1H)	1.65-1.83	m	1.77	m	-0.03
4' (1H)	1.05-1.09	m	1.06	m	-0.01
4' (1H)	1.39-1.48	m	1.44	m	0.00
5' (3H)	0.84	t, $J = 7.5$ Hz	0.84	t, $J = 7.5$ Hz	0.00
6' (3H)	0.81	d, $J = 6.6$ Hz	0.81	d, $J = 6.8$ Hz	0.00
7' (1H)	7.40	d, $J = 9.0$ Hz	7.41	d, $J = 8.9$ Hz	-0.01
2'' (1H)	2.88	dd, $J = 8.6, 5.3$ Hz	2.89	dd, $J = 8.4, 5.0$ Hz	-0.01
3'' (1H)	1.35-1.45	m	1.40	m	0.00
3'' (1H)	1.47-1.59	m	1.53	m	0.00
4'' (1H)	1.02-1.17	m	1.10	m	0.00
5'' (3H)	0.90	d, $J = 6.3$ Hz	0.90	d, $J = 6.4$ Hz	0.00
6'' (3H)	0.92	d, $J = 6.4$ Hz	0.92	d, $J = 6.5$ Hz	0.00
NMe ₂ (6H)	2.24	s	2.24	s	0.00

¹³C NMR (Synthetic at 75 MHz; Natural at 100 MHz)



Paliurine F

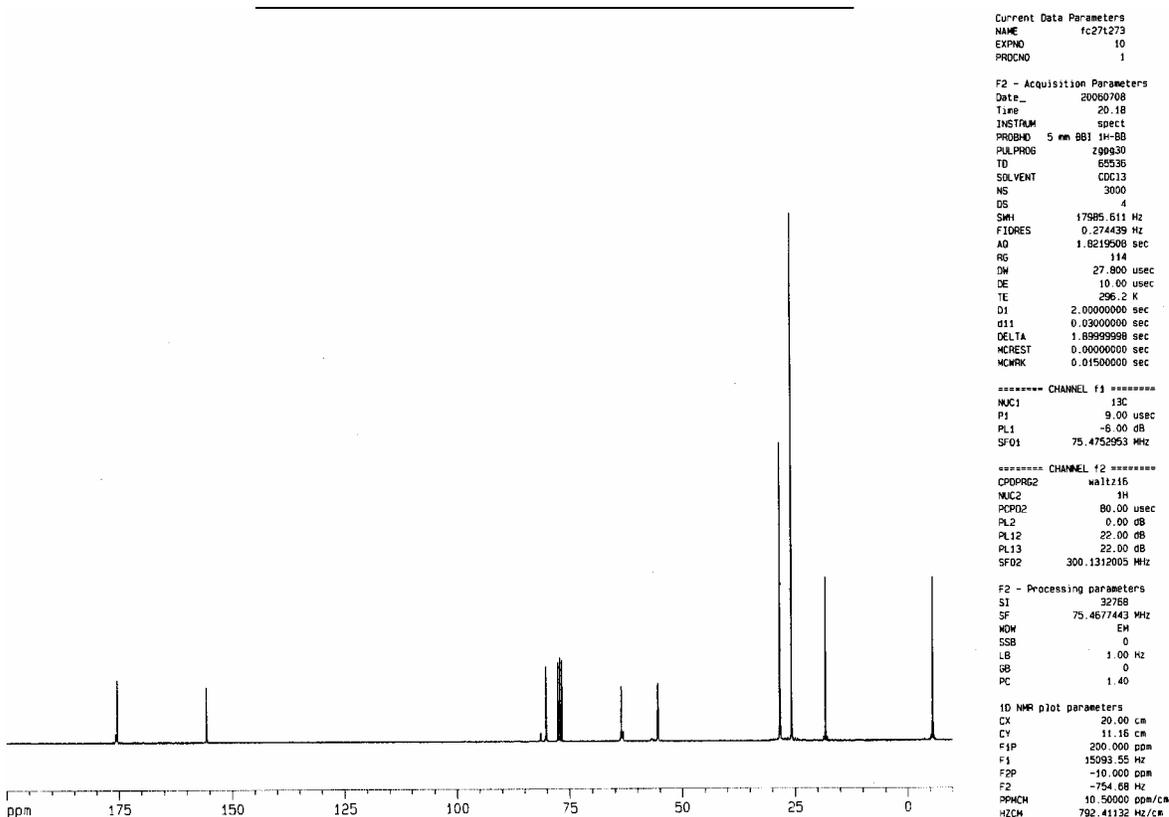
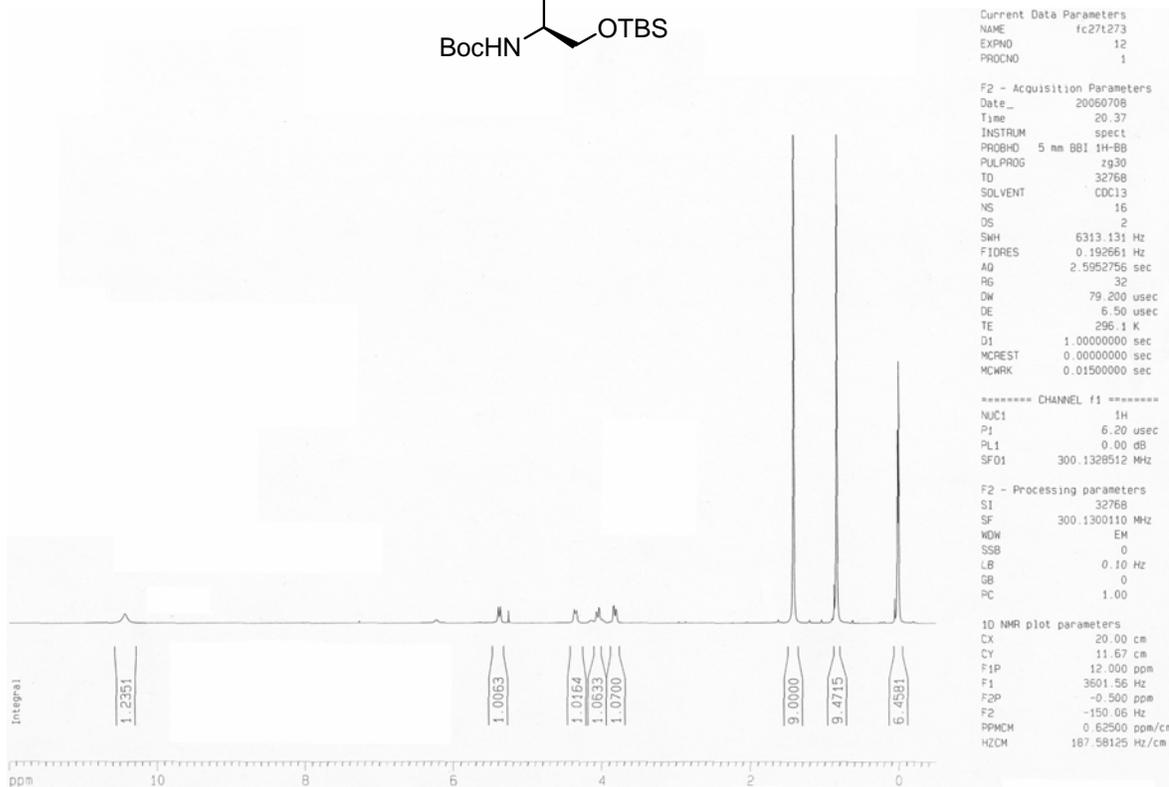
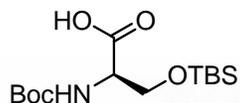
	Synthetic	Natural	
Position	δ	δ	$\Delta\delta$
1	106.6	106.8	-0.2
2	121.6	121.6	0.0
4	167.0	167.2	-0.2
5	60.3	60.4	-0.1
7	170.5	170.4	0.1
8	64.7	64.5	0.2
9	76.9	76.9	0.0
11	151.4	151.2	0.2
12	111.5	111.3	0.2
12'	117.8	117.8	0.0
13	124.8	124.4	0.4
13'	114.3	113.9	0.4
14	151.8	151.6	0.2
15	35.4	35.2	0.2
16	24.8	24.8	0.0
17	11.7	11.8	-0.1
18	16.2	16.3	-0.1
19	32.7	32.7	0.0
20	46.8	46.9	-0.1

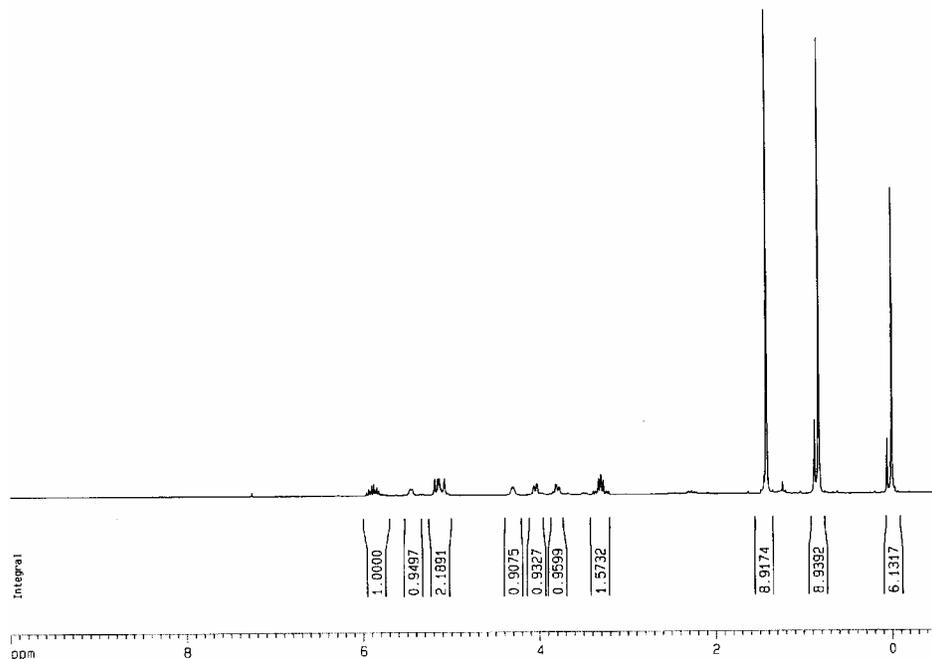
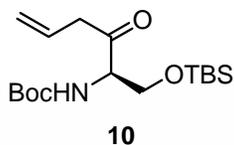
OMe	56.2	56.1	-0.1
1'	172.0	172.0	0.0
2'	53.9	53.8	0.1
3'	37.6	37.5	0.1
4'	24.7	24.6	0.1
5'	10.8	10.8	0.0
6'	15.4	15.4	0.0
1''	173.7	174.0	-0.3
2''	67.7	67.4	0.3
3''	37.2	37.1	0.1
4''	26.0	26.0	0.0
5''	23.3 ^a	23.4	-0.1
6''	22.2 ^a	22.2	0.0
NMe ₂	42.3	42.4	-0.1

^a both assignments can be interchanged

Supporting Information

**^1H and ^{13}C NMR spectra of intermediates
and synthetic paliurine F**





```

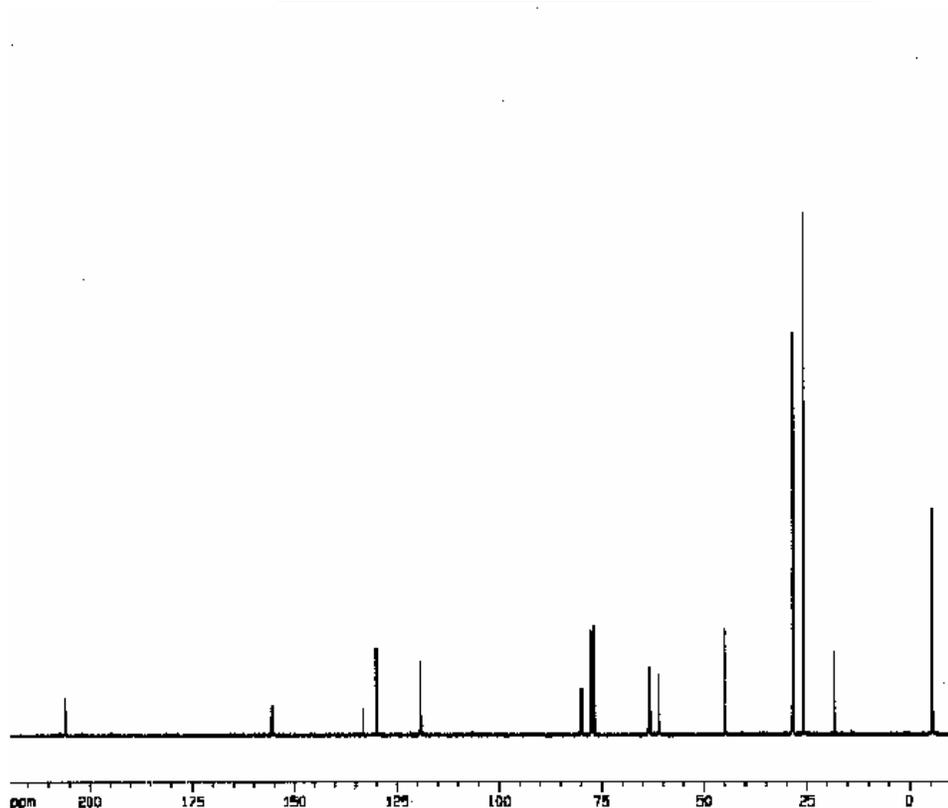
Current Data Parameters
NAME      1c27k275
EXPNO    14
PROCNO    1

F2 - Acquisition Parameters
Date_    20060709
Time     0.47
INSTRUM  spect
PROBHD   5 mm BBI 1H-88
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        16
DS        2
SWH       6172.833 Hz
FIDRES    0.094190 Hz
AQ        5.3084660 sec
RG        64
DW        81.000 usec
DE        10.00 usec
TE        296.1 K
D1        1.0000000 sec
MCREST    0.0000000 sec
MCNRK     0.0150000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        6.20 usec
PL1       0.00 dB
SFO1     300.1318534 MHz

F2 - Processing parameters
SI        32768
SF        300.1300149 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

ID NMR plot parameters
CX        20.00 cm
CY        11.71 cm
F1P       10.000 ppm
F1        3001.30 Hz
F2P       -0.500 ppm
F2        -150.06 Hz
PPMCM     0.52500 ppm/cm
HZCM      157.56825 Hz/cm
    
```



```

Current Data Parameters
NAME      1c27k275
EXPNO    14
PROCNO    1

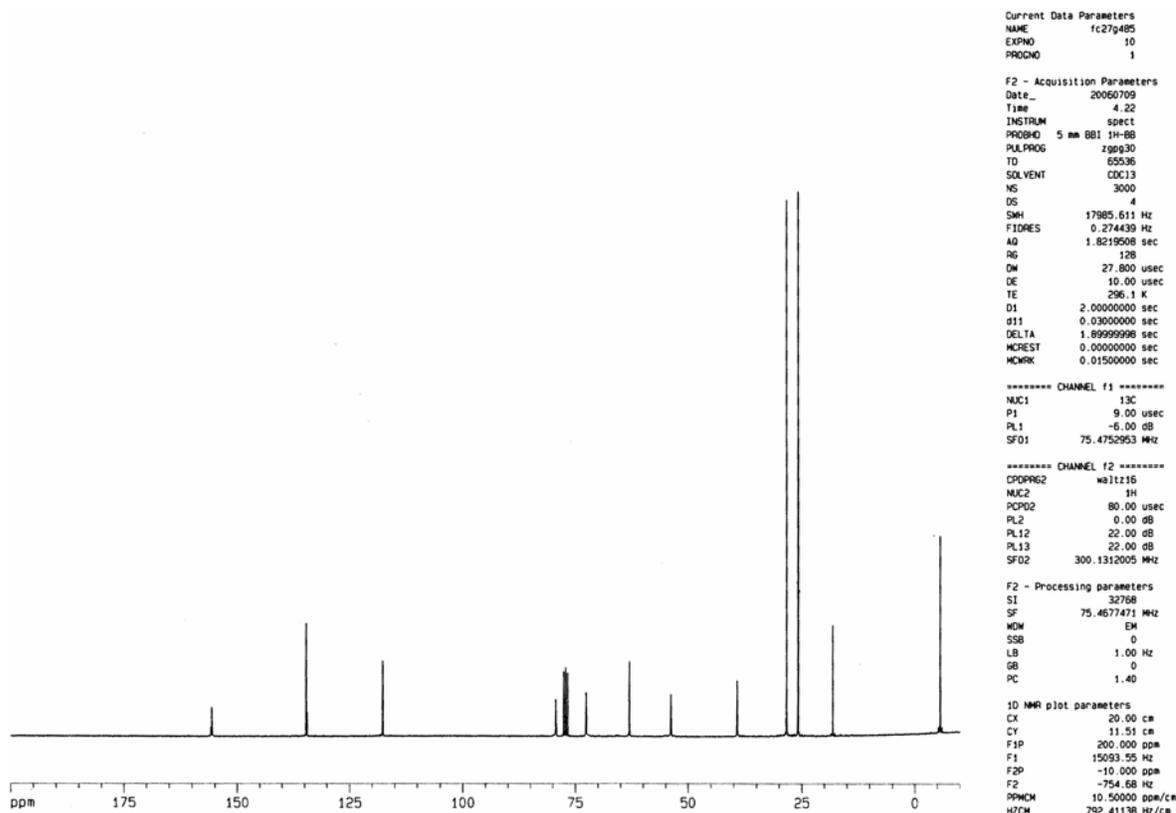
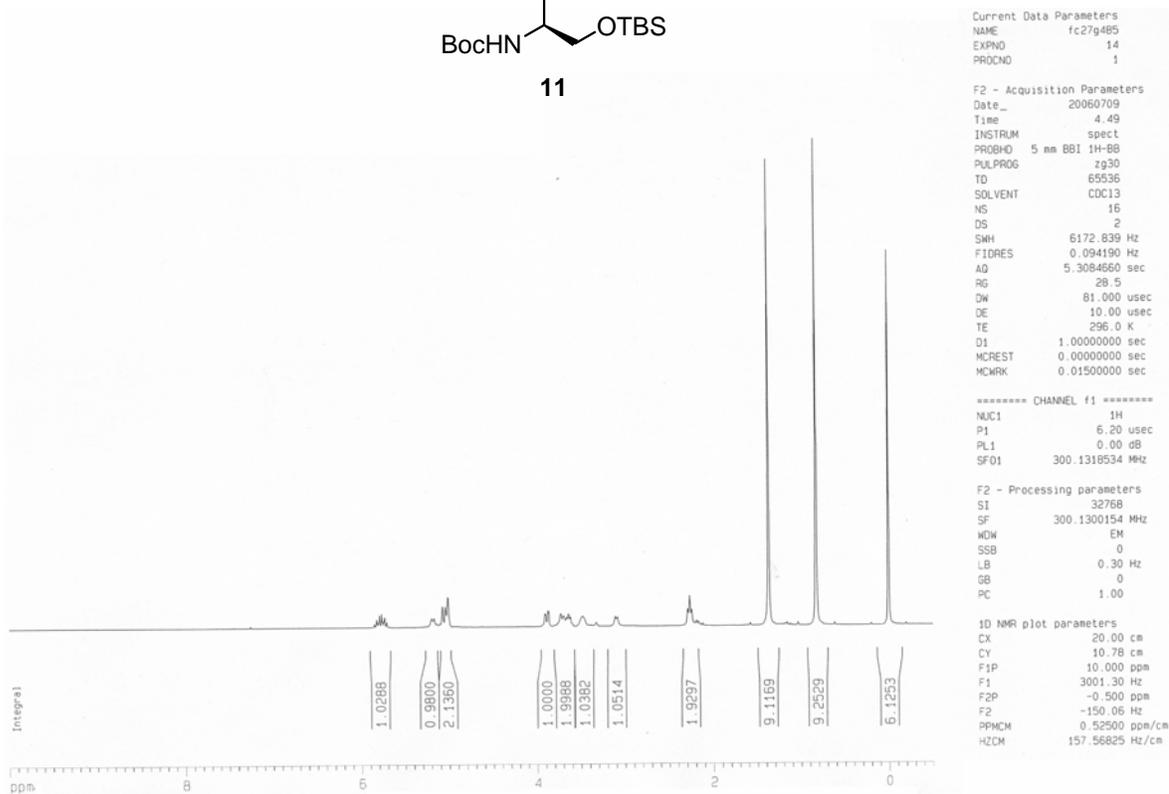
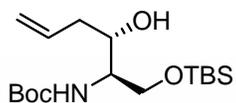
F2 - Acquisition Parameters
Date_    20060709
Time     0.81
INSTRUM  spect
PROBHD   5 mm BBI 1H-88
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        3000
DS        4
SWH       17385.841 Hz
FIDRES    0.274433 Hz
AQ        1.0219500 sec
RG        126
DW        27.800 usec
DE        10.00 usec
TE        296.1 K
D1        3.4000000 sec
M11       0.4000000 sec
DELTA     0.4000000 sec
MCREST    0.4000000 sec
MCNRK     0.4000000 sec

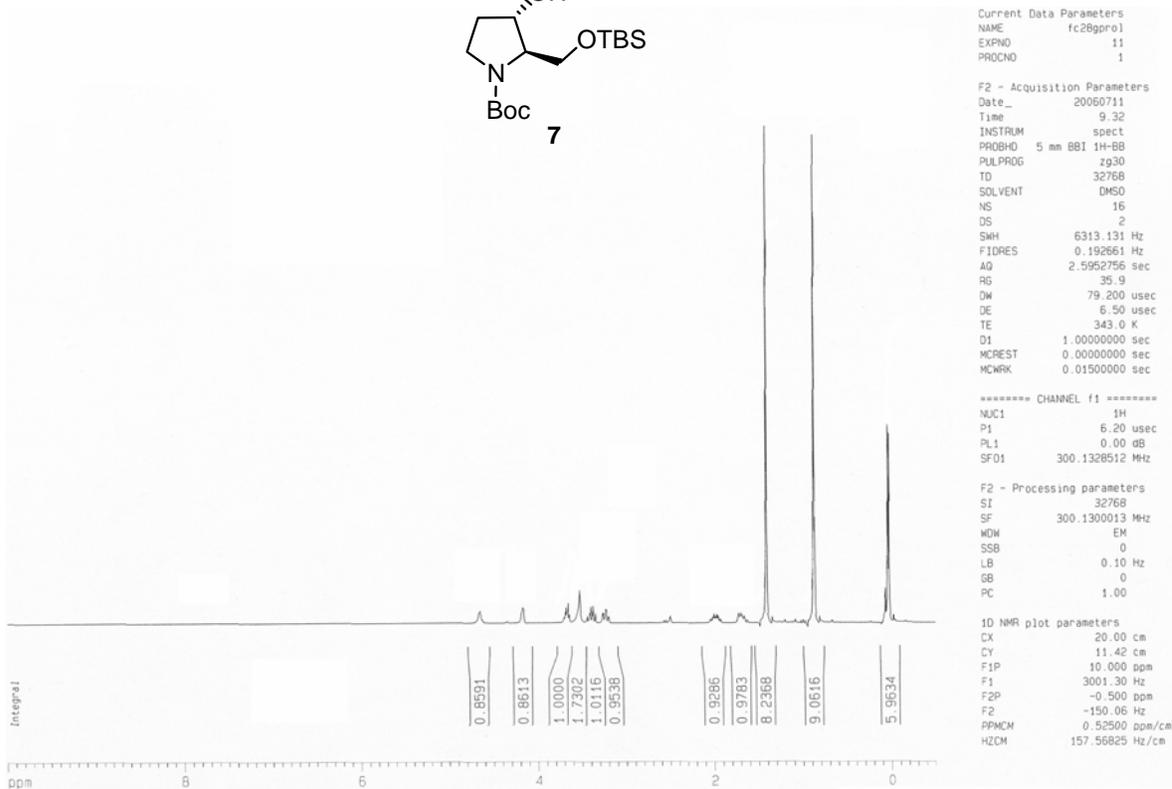
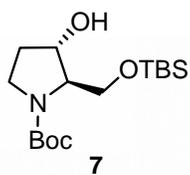
===== CHANNEL f1 =====
NUC1      13C
P1        9.00 usec
PL1       -0.90 dB
SFO1     125.4756453 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2      1H
P2P2     89.99 usec
P2F2     0.00 dB
PL12     21.00 dB
PL13     21.00 dB
SFO2     300.1318534 MHz

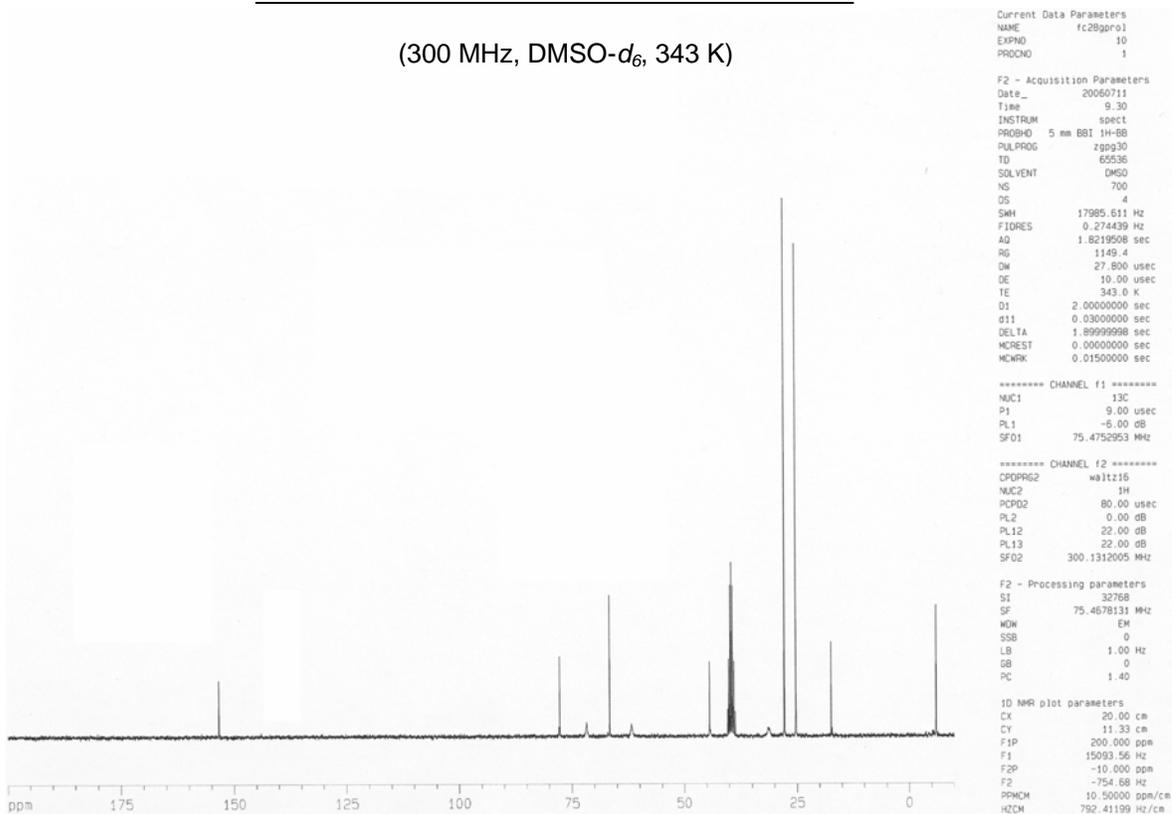
F2 - Processing parameters
SI        32768
SF        250.4677457 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.00

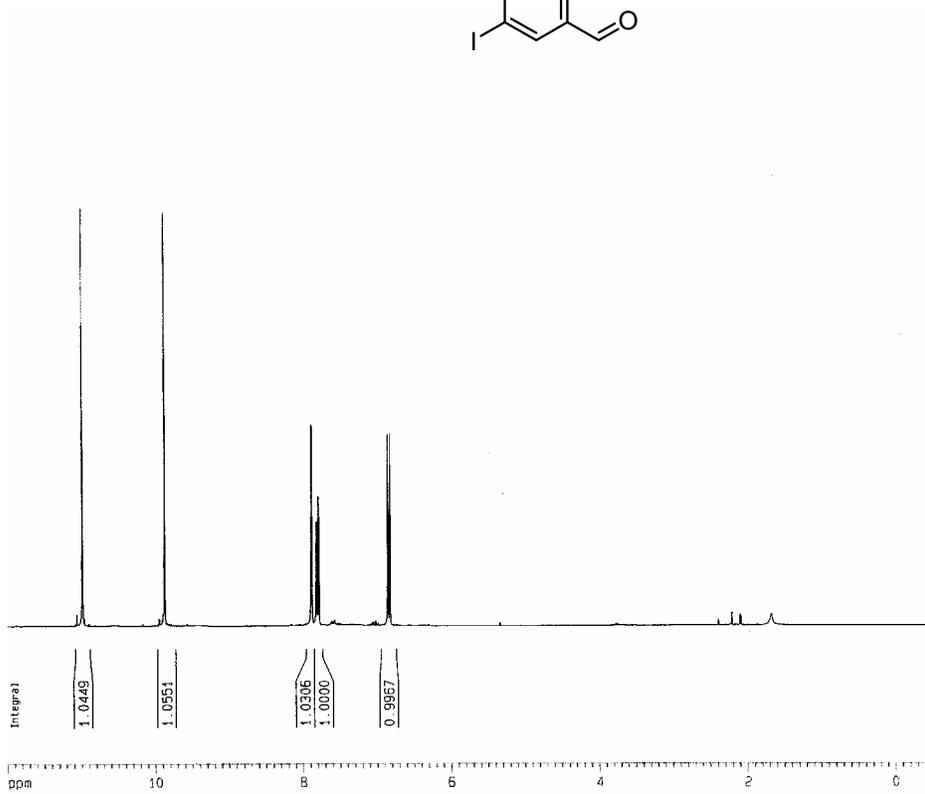
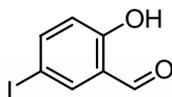
ID NMR plot parameters
CX        20.00 cm
CY        11.71 cm
F1P       100.000 ppm
F1        250.1300149 MHz
F2P       -16.000 ppm
F2        -794.060 Hz
PPMCM     11.26000 ppm/kHz
HZCM      960.87196 Hz/cm
    
```





(300 MHz, DMSO-*d*₆, 343 K)





```

Current Data Parameters
NAME          fc43par1
EXPNO        11
PROCNO       1

F2 - Acquisition Parameters
Date_        20051030
Time         19:57
INSTRUM      spect
PROBHD       5 mm BBI 1H-BB
PULPROG      zg30
TD           32768
SOLVENT      CDCl3
NS           16
DS           2
SWH          6313.131 Hz
FIDRES      0.192661 Hz
AQ          2.5952756 sec
RG          456.1
DM          79.200 usec
DE          6.50 usec
TE          296.2 K
D1          1.0000000 sec
MCREST      0.0000000 sec
MCNRK       0.0150000 sec
    
```

```

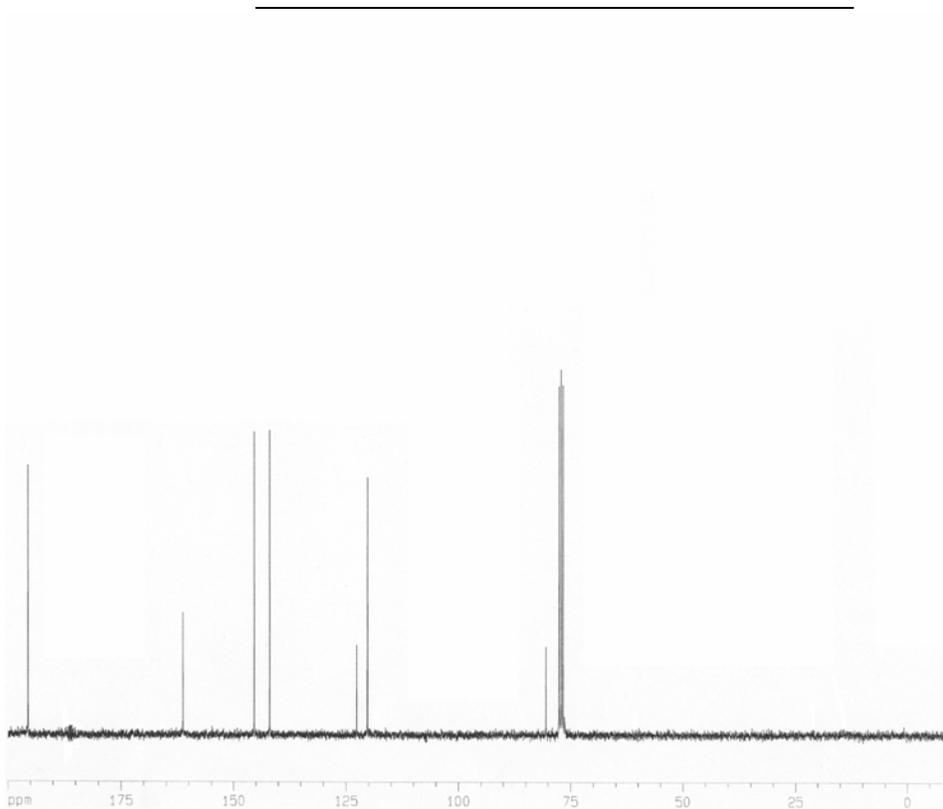
===== CHANNEL f1 =====
NUC1         1H
P1           6.20 usec
PL1          0.00 dB
SFO1        300.1320512 MHz
    
```

```

F2 - Processing parameters
SI           32768
SF          300.1300000 MHz
WDW          EM
SSB          0
LB           0.10 Hz
GB           0
PC           1.00
    
```

```

1D NMR plot parameters
CX           20.00 cm
CY           9.70 cm
F1P         12.000 ppm
F1          3601.56 Hz
F2P         -0.500 ppm
F2          -150.06 Hz
PPMCM       0.62500 ppm/cm
HZCM        187.58125 Hz/cm
    
```



```

Current Data Parameters
NAME          fc44par1
EXPNO        20
PROCNO       1
    
```

```

F2 - Acquisition Parameters
Date_        20051102
Time         8:11
INSTRUM      spect
PROBHD       5 mm BBI 1H-BB
PULPROG      zgpg30
TD           65536
SOLVENT      CDCl3
NS           1024
DS           4
SWH          17985.611 Hz
FIDRES      0.274439 Hz
AQ          1.8219508 sec
RG          143.7
DM          27.800 usec
DE          10.00 usec
TE          294.9 K
D1          2.0000000 sec
d11         0.0300000 sec
DELTA       1.89999998 sec
MCREST      0.0000000 sec
MCNRK       0.0150000 sec
    
```

```

===== CHANNEL f1 =====
NUC1         13C
P1           8.20 usec
PL1          -6.00 dB
SFO1        75.4752953 MHz
    
```

```

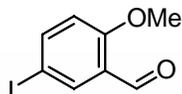
===== CHANNEL f2 =====
CPDPRG2      waltz16
NUC2         1H
PCPD2       80.00 usec
PL2          0.00 dB
PL12         22.00 dB
PL13         22.00 dB
SFO2        300.1312005 MHz
    
```

```

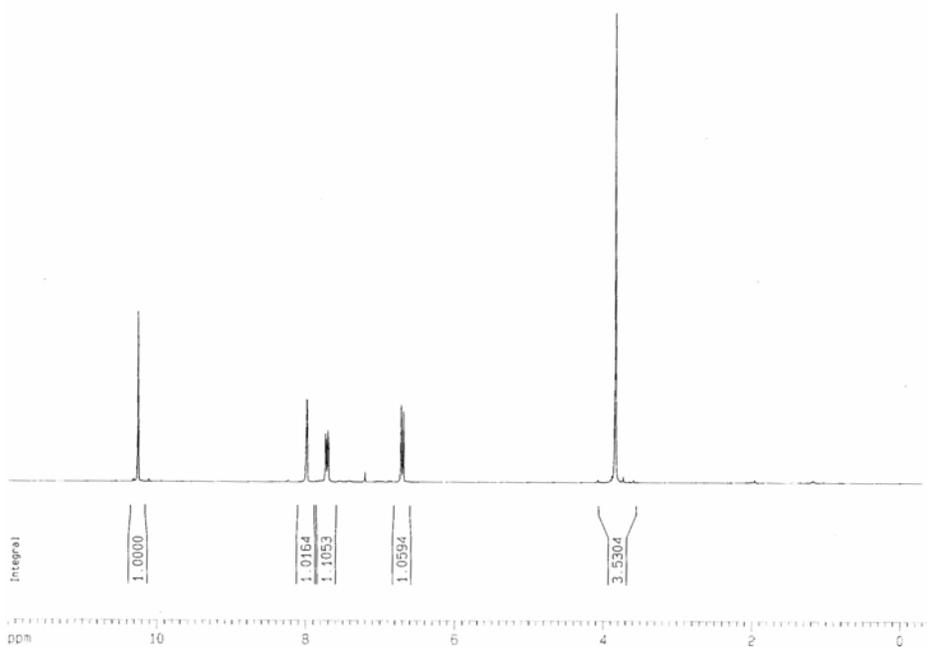
F2 - Processing parameters
SI           32768
SF          75.4677443 MHz
WDW          EM
SSB          0
LB           1.00 Hz
GB           0
PC           1.40
    
```

```

1D NMR plot parameters
CX           20.00 cm
CY           7.81 cm
F1P         200.000 ppm
F1          15093.55 Hz
F2P         -10.000 ppm
F2          -754.68 Hz
PPMCM       10.50000 ppm/cm
HZCM        792.41132 Hz/cm
    
```



8



```

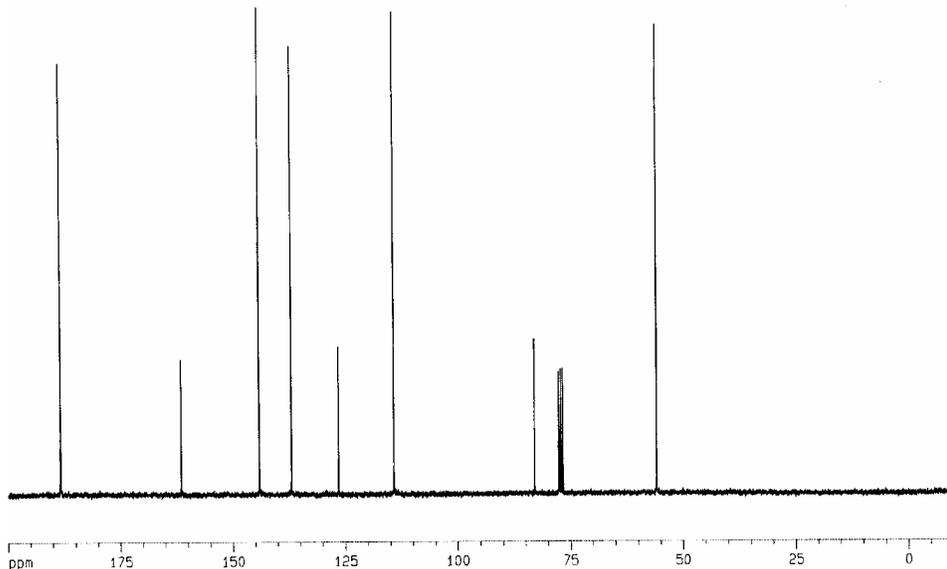
Current Data Parameters
NAME      fc50g338
EXPNO    12
PROCNO   1

F2 - Acquisition Parameters
Date_    20051223
Time     2.39
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zg30
TD        32768
SOLVENT  CDCl3
NS        16
DS        2
SWH       6313.131 Hz
FIDRES   0.192561 Hz
AQ        2.5952756 sec
RG        228.1
DW        79.200 usec
DE        6.50 usec
TE        297.4 K
D1        1.0000000 sec
MCREST   0.0000000 sec
MCMRK    0.0150000 sec

***** CHANNEL f1 *****
NUC1      1H
P1        6.20 usec
PL1       0.00 dB
SF01     300.1320512 MHz

F2 - Processing parameters
SI        32768
SF        300.1300332 MHz
WDW       EM
SSB       0
LB        0.10 Hz
GB        0
PC        1.00

1D NMR plot parameters
CX        20.00 cm
CY        10.10 cm
F1P       12.000 ppm
F1        3601.56 Hz
F2P       -0.500 ppm
F2        -150.07 Hz
PPMCH    0.62500 ppm/cm
HZCM     187.58128 Hz/cm
    
```



```

Current Data Parameters
NAME      fc24v009
EXPNO    10
PROCNO   1

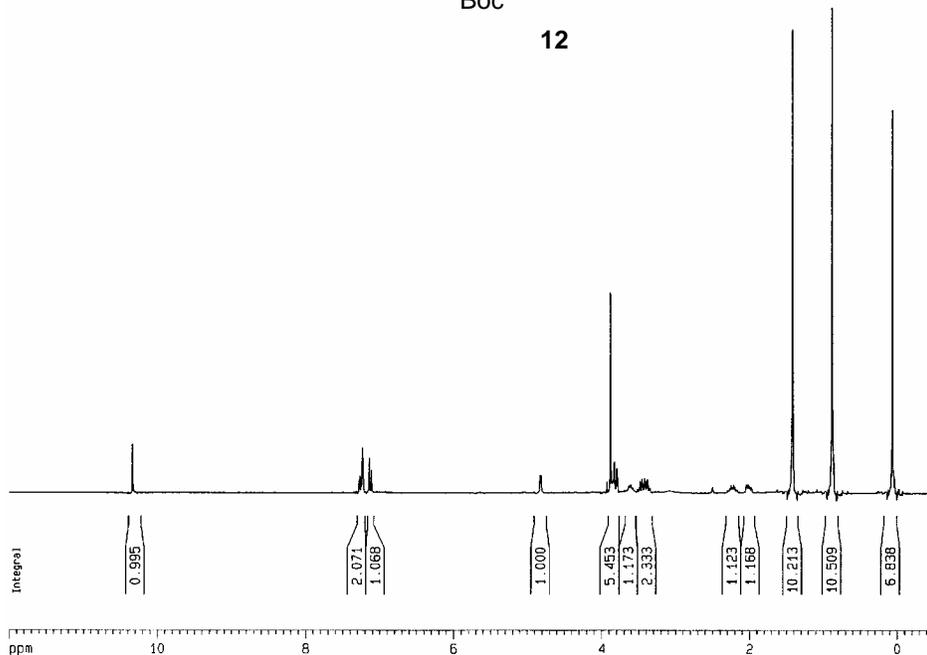
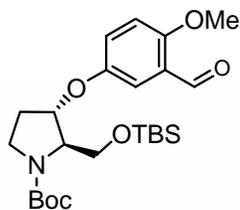
F2 - Acquisition Parameters
Date_    20060615
Time     10.15
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zgpg30
TD        65536
SOLVENT  CDCl3
NS        1024
DS        4
SWH       17985.611 Hz
FIDRES   0.274439 Hz
AQ        1.8219508 sec
RG        128
DW        27.000 usec
DE        10.00 usec
TE        297.1 K
D1        2.0000000 sec
d11      0.0300000 sec
DELTA    1.6999998 sec
MCREST   0.0000000 sec
MCMRK    0.0150000 sec

***** CHANNEL f1 *****
NUC1      13C
P1        9.00 usec
PL1       -6.00 dB
SF01     75.4752953 MHz

***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
PCPD2    80.00 usec
PL2       0.00 dB
PL12     22.00 dB
PL13     22.00 dB
SF02     300.1312005 MHz

F2 - Processing parameters
SI        32768
SF        75.4677487 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

1D NMR plot parameters
CX        20.00 cm
CY        10.26 cm
F1P       200.000 ppm
F1        15093.55 Hz
F2P       -10.000 ppm
F2        -754.68 Hz
PPMCH    10.50000 ppm/cm
HZCM     792.41138 Hz/cm
    
```



```

Current Data Parameters
NAME      fc28vb11P
EXPNO    11
PROCNO   1

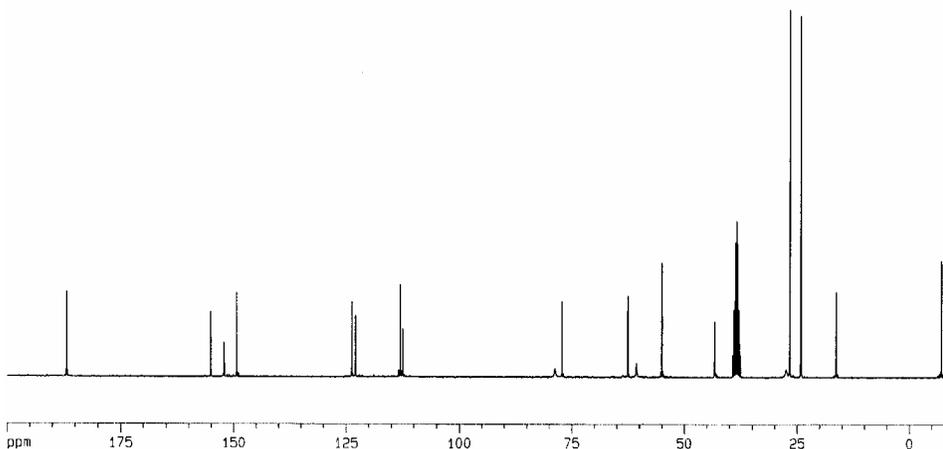
F2 - Acquisition Parameters
Date_    20060713
Time     17.29
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zg30
TD        32768
SOLVENT  DMSO
NS        16
DS        2
SWH       6313.131 Hz
FIDRES    0.192661 Hz
AQ        2.5952756 sec
RG         28.5
DM         79.200 usec
DE         6.50 usec
TE         345.0 K
D1         1.0000000 sec
MCREST    0.0000000 sec
MCMARK    0.0150000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        6.20 usec
PL1       0.00 dB
SFO1     300.1328512 MHz

F2 - Processing parameters
SI        32768
SF        300.1300011 MHz
WDW       EM
SSB       0
LB        0.10 Hz
GB        0
PC        1.00

1D NMR plot parameters
CX        20.00 cm
CY        10.58 cm
F1P       12.000 ppm
F1        3601.56 Hz
F2P       -0.500 ppm
F2        -150.06 Hz
PPMCM     0.62500 ppm/cm
HZCM      187.58125 Hz/cm
    
```

(300 MHz, DMSO-*d*₆, 345 K)



```

Current Data Parameters
NAME      fc28vb11P
EXPNO    10
PROCNO   1

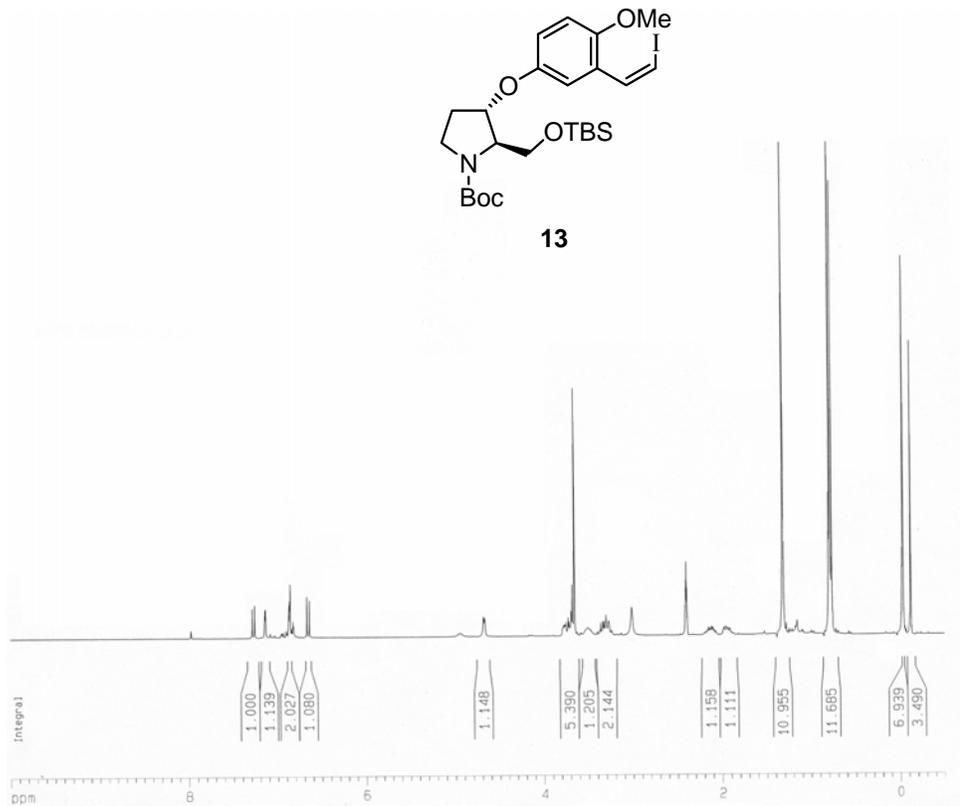
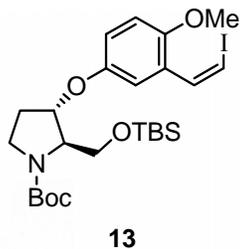
F2 - Acquisition Parameters
Date_    20060713
Time     12.35
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        4000
DS        4
SWH       17985.611 Hz
FIDRES    0.274459 Hz
AQ        1.6219508 sec
RG         812.7
DM         27.800 usec
DE         10.00 usec
TE         345.0 K
D1         4.0000000 sec
d11       0.0300000 sec
DELTA     3.9000010 sec
MCREST    0.0000000 sec
MCMARK    0.0150000 sec

===== CHANNEL f1 =====
NUC1      13C
P1        9.00 usec
PL1       -6.00 dB
SFO1     75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2      1H
PCPD2    80.00 usec
PL2       0.00 dB
PL12     22.00 dB
PL13     22.00 dB
SFO2     300.1312005 MHz

F2 - Processing parameters
SI        32768
SF        75.4678695 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

1D NMR plot parameters
CX        20.00 cm
CY         7.96 cm
F1P       200.000 ppm
F1        15093.58 Hz
F2P       -10.000 ppm
F2        -754.68 Hz
PPMCM     10.50000 ppm/cm
HZCM      792.41278 Hz/cm
    
```



```

Current Data Parameters
NAME fc20t207p dms0 333K
EXPNO 12
PROCNO 1

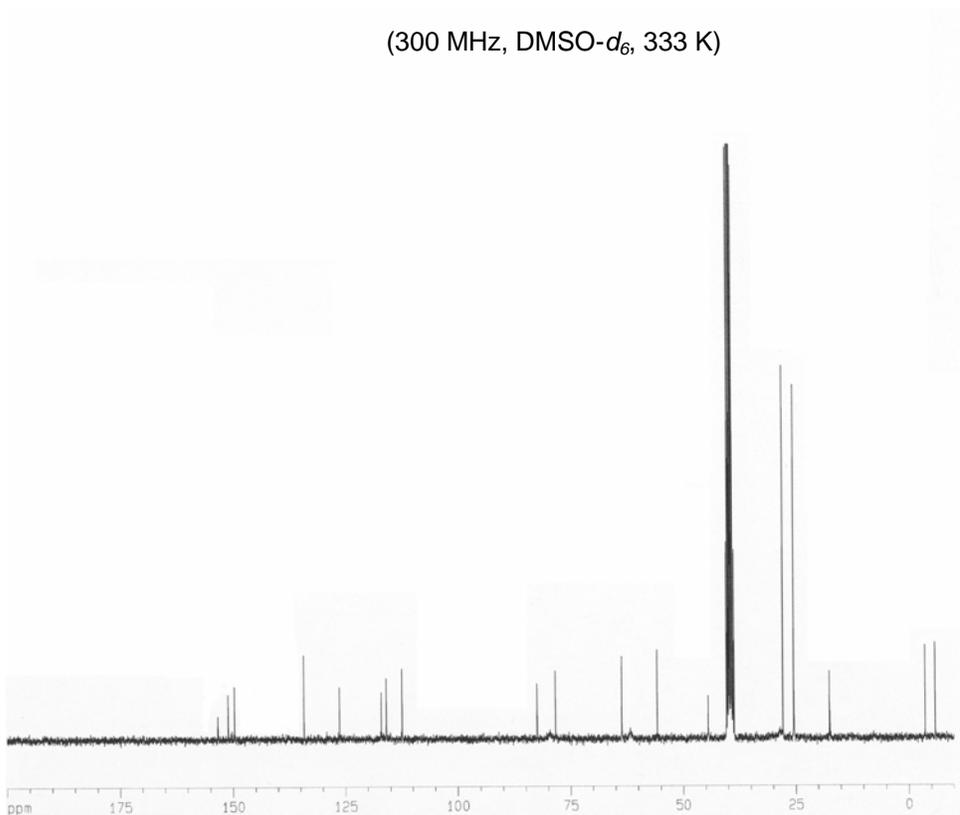
F2 - Acquisition Parameters
Date_ 20060520
Time 18.06
INSTRUM spect
PROBHD 5 mm BBI 1H-BB
PULPROG zg30
TD 32768
SOLVENT DMSO
NS 16
DS 2
SWH 6313.131 Hz
FIDRES 0.192661 Hz
AQ 2.5952756 sec
RG 181
DM 79.200 usec
DE 6.50 usec
TE 333.0 K
D1 1.00000000 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

***** CHANNEL f1 *****
NUC1 1H
P1 6.20 usec
PL1 0.00 dB
SFO1 300.1328512 MHz

F2 - Processing parameters
SI 32768
SF 300.1300302 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 20.00 cm
CY 14.37 cm
F1P 10.000 ppm
F1 3001.30 Hz
F2P -0.500 ppm
F2 -150.07 Hz
PPMCM 0.52500 ppm/cm
HZCM 157.56827 Hz/cm
    
```

(300 MHz, DMSO-*d*₆, 333 K)



```

Current Data Parameters
NAME fc20t207p dms0 333K
EXPNO 10
PROCNO 1

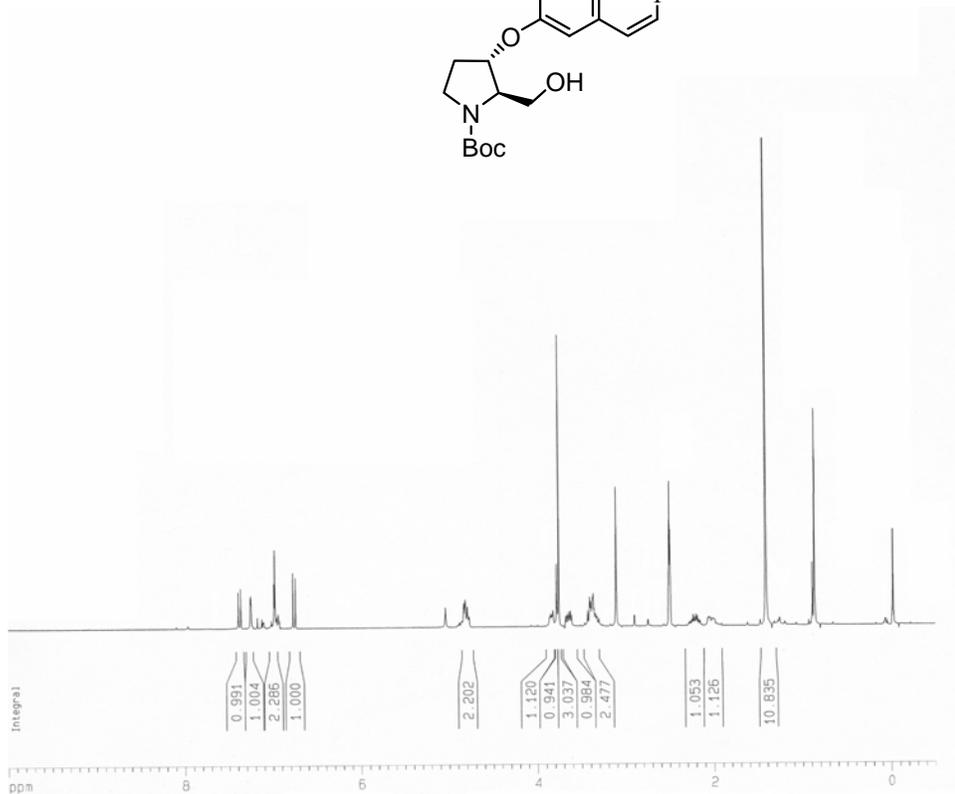
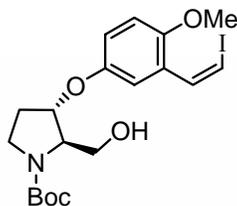
F2 - Acquisition Parameters
Date_ 20060520
Time 17.16
INSTRUM spect
PROBHD 5 mm BBI 1H-BB
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 5000
DS 4
SWH 17985.611 Hz
FIDRES 0.274439 Hz
AQ 1.8219508 sec
RG 143.7
DM 27.800 usec
DE 10.00 usec
TE 333.0 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999996 sec
MCREST 0.00000000 sec
MCWRK 0.01500000 sec

***** CHANNEL f1 *****
NUC1 13C
P1 9.00 usec
PL1 -6.00 dB
SFO1 75.4752953 MHz

***** CHANNEL f2 *****
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0.00 dB
PL12 22.00 dB
PL13 22.00 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4678113 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
CY 28.85 cm
F1P 200.000 ppm
F1 15093.56 Hz
F2P -10.000 ppm
F2 -754.68 Hz
PPMCM 10.50000 ppm/cm
HZCM 792.41199 Hz/cm
    
```



```

Current Data Parameters
NAME      fc21t209P
EXPNO    11
PROCNO   1

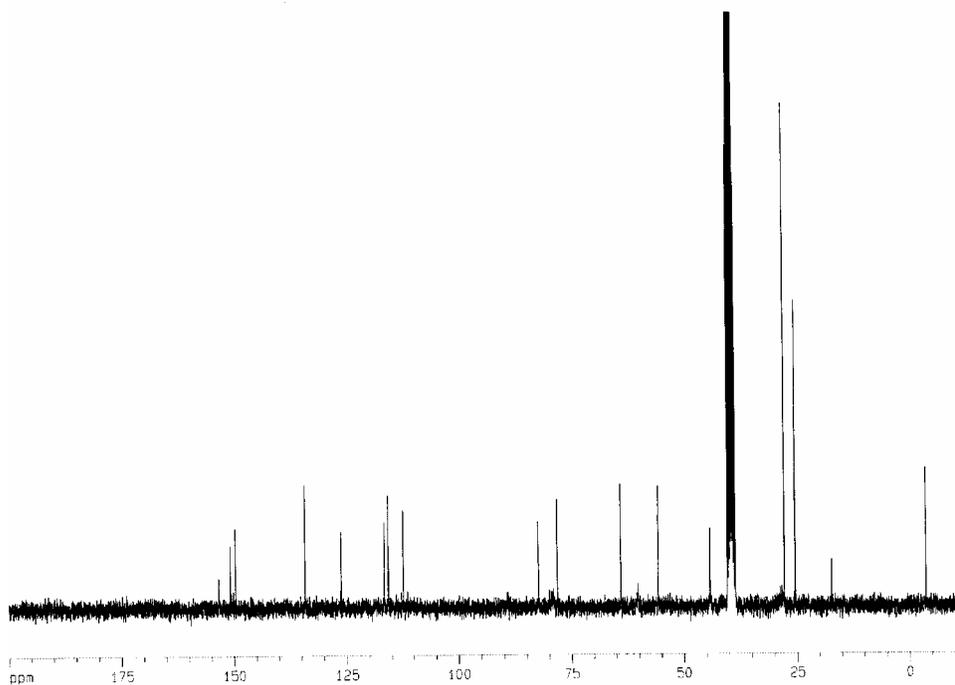
F2 - Acquisition Parameters
Date_    20060525
Time     16.30
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zg30
TD        32768
SOLVENT  DMSO
NS        15
DS        2
SWH       6313.131 Hz
FIDRES   0.192661 Hz
AQ        2.5952756 sec
RG         456.1
DW         79.200 usec
DE         6.50 usec
TE         333.0 K
D1         1.00000000 sec
MCREST   0.00000000 sec
MCWRK    0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1         6.20 usec
PL1        0.00 dB
SFO1      300.1328512 MHz

F2 - Processing parameters
SI         32768
SF         300.1300011 MHz
WDW        EM
SSB         0
LB          0.10 Hz
GB          0
PC          1.00

1D NMR plot parameters
CX         20.00 cm
CY         15.45 cm
F1P        10.000 ppm
F1          3001.30 Hz
F2P        -0.500 ppm
F2         -150.06 Hz
PPMCM      0.52500 ppm/cm
HZCM       157.56825 Hz/cm
    
```

(300 MHz, DMSO-d₆, 333 K)



```

Current Data Parameters
NAME      fc21t209P
EXPNO    10
PROCNO   1

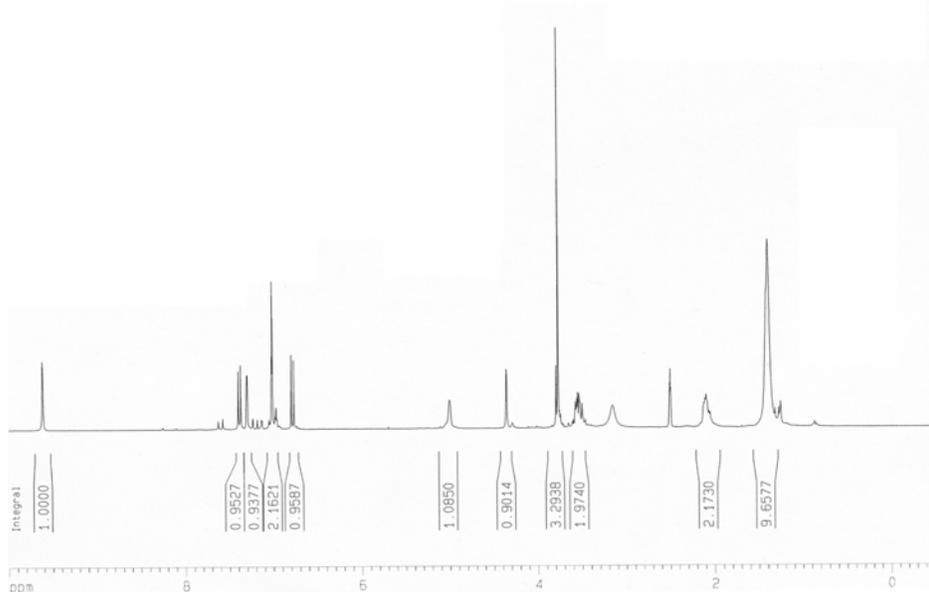
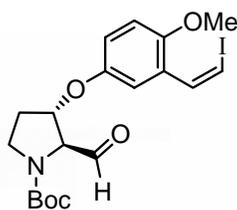
F2 - Acquisition Parameters
Date_    20060525
Time     16.28
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        5000
DS         4
SWH       17955.611 Hz
FIDRES   0.274436 Hz
AQ        1.8219508 sec
RG         128
DW         27.800 usec
DE         10.00 usec
TE         333.0 K
D1         2.00000000 sec
d11       0.03000000 sec
DELTA     1.89999998 sec
MCREST   0.00000000 sec
MCWRK    0.01500000 sec

===== CHANNEL f1 =====
NUC1      13C
P1         9.00 usec
PL1        -6.00 dB
SFO1      75.4752953 MHz

===== CHANNEL f2 =====
CPDPRG2  waltz16
NUC2      1H
PCPD2     86.00 usec
PL2        0.00 dB
PL12       22.00 dB
PL13       22.00 dB
SFO2      300.1312005 MHz

F2 - Processing parameters
SI         32768
SF         75.4678124 MHz
WDW        EM
SSB         0
LB          1.00 Hz
GB          0
PC          1.40

1D NMR plot parameters
CX         20.00 cm
CY         67.15 cm
F1P        200.000 ppm
F1          15093.55 Hz
F2P        -10.000 ppm
F2         -754.68 Hz
PPMCM      10.50000 ppm/cm
HZCM       792.41199 Hz/cm
    
```



```

Current Data Parameters
NAME      fc271211P
EXPNO    13
PROCNO   1

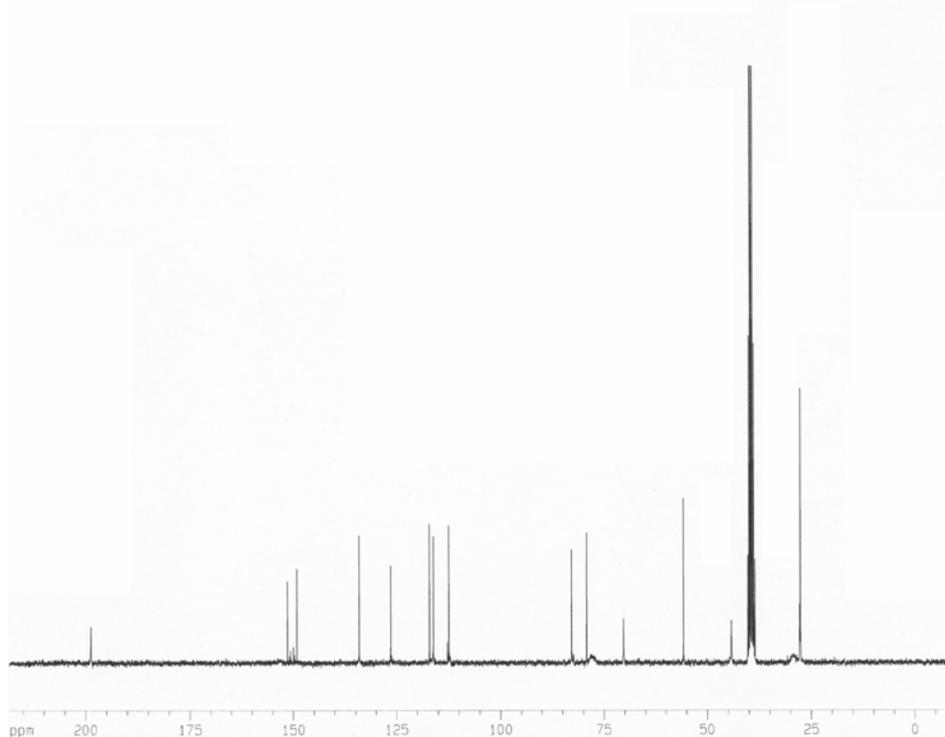
F2 - Acquisition Parameters
Date_    20060706
Time     15.28
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zg30
TD        65536
SOLVENT  DMSO
NS        16
DS        2
SWH       6172.839 Hz
FIDRES    0.094190 Hz
AQ        5.3084660 sec
RG         114
DW         81.000 usec
DE         10.00 usec
TE         335.0 K
D1         1.0000000 sec
MCREST    0.0000000 sec
MCWRK     0.0150000 sec

***** CHANNEL f1 *****
NUC1      1H
P1         6.20 usec
PL1        0.00 dB
SF01      300.1318534 MHz

F2 - Processing parameters
SI         32768
SF         300.1300000 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

1D NMR plot parameters
CX         20.00 cm
CY         8.65 cm
F1P        10.000 ppm
F1         3001.30 Hz
F2P        -0.500 ppm
F2         -150.06 Hz
PPMCM      0.52500 ppm/cm
HZCM       157.56825 Hz/cm
    
```

(300 MHz, DMSO-d₆, 335 K)



```

Current Data Parameters
NAME      fc271211P
EXPNO    10
PROCNO   1

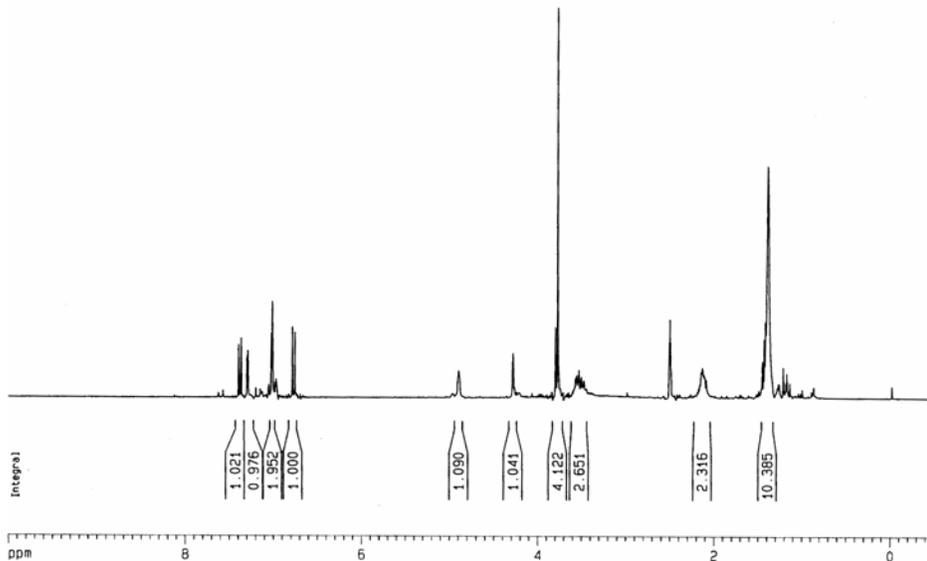
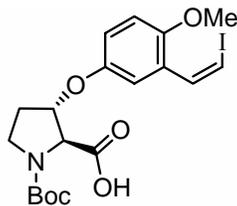
F2 - Acquisition Parameters
Date_    20060706
Time     11.21
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        5000
DS        4
SWH       17985.611 Hz
FIDRES    0.274439 Hz
AQ        1.8219508 sec
RG         128
DW         27.800 usec
DE         10.00 usec
TE         335.0 K
D1         2.0000000 sec
d11       0.0300000 sec
DELTA     1.8999999 sec
MCREST    0.0000000 sec
MCWRK     0.0150000 sec

***** CHANNEL f1 *****
NUC1      13C
P1         9.00 usec
PL1        -6.00 dB
SF01      75.4752953 MHz

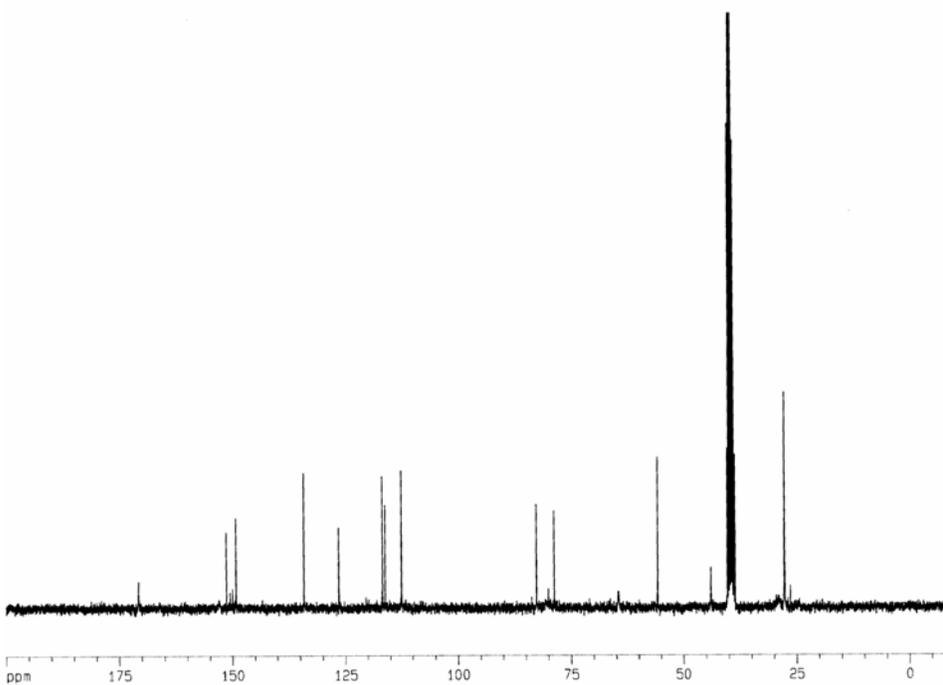
***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
PCPD2     80.00 usec
PL2        0.00 dB
PL12       22.00 dB
PL13       22.00 dB
SF02      300.1312005 MHz

F2 - Processing parameters
SI         32768
SF         75.4678102 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

1D NMR plot parameters
CX         20.00 cm
CY         15.81 cm
F1P        220.000 ppm
F1         16602.92 Hz
F2P        -10.000 ppm
F2         -754.68 Hz
PPMCM      11.50000 ppm/cm
HZCM       867.87982 Hz/cm
    
```



(300 MHz, DMSO-d₆, 345 K)



```

Current Data Parameters
NAME      fc28t213P
EXPNO    12
PROCNO   1

F2 - Acquisition Parameters
Date_    20060712
Time     19.53
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zg30
TD        32768
SOLVENT  DMSO
NS        16
DS        2
SWH       6313.131 Hz
FIDRES    0.192661 Hz
AQ        2.5952756 sec
RG         228.1
DM        79.200 usec
DE         6.50 usec
TE        345.0 K
D1        1.00000000 sec
MCREST    0.00000000 sec
MCMRK     0.01500000 sec

***** CHANNEL f1 *****
NUC1      1H
P1        6.20 usec
PL1       0.00 dB
SF01      300.1328512 MHz

F2 - Processing parameters
SI        32768
SF        300.1300009 MHz
WDW       EM
SSB       0
LB        0.10 Hz
GB        0
PC        1.00

1D NMR plot parameters
CX        20.00 cm
CY        8.42 cm
F1P       10.000 ppm
F1        3001.30 Hz
F2P       -0.500 ppm
F2        -150.05 Hz
PPHMC     0.52500 ppm/cm
HZCM      157.56825 Hz/cm
    
```

```

Current Data Parameters
NAME      fc28t213P
EXPNO    10
PROCNO   1

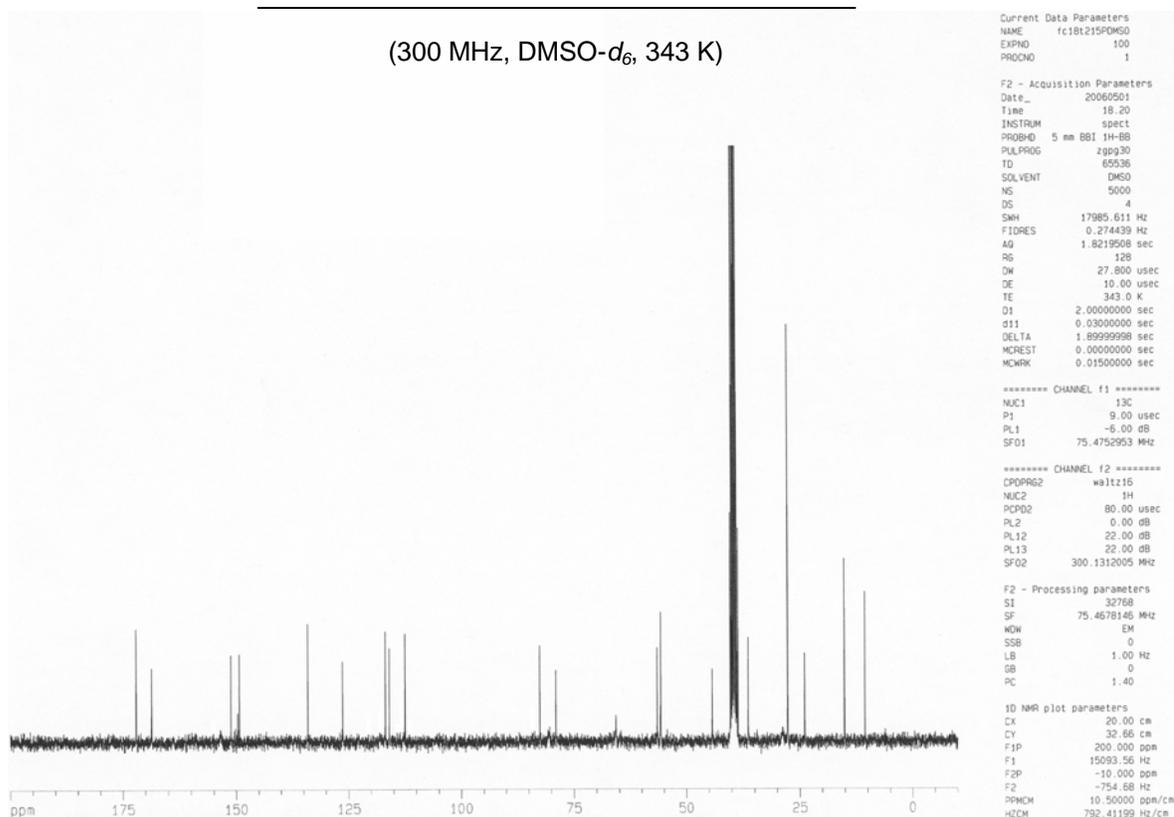
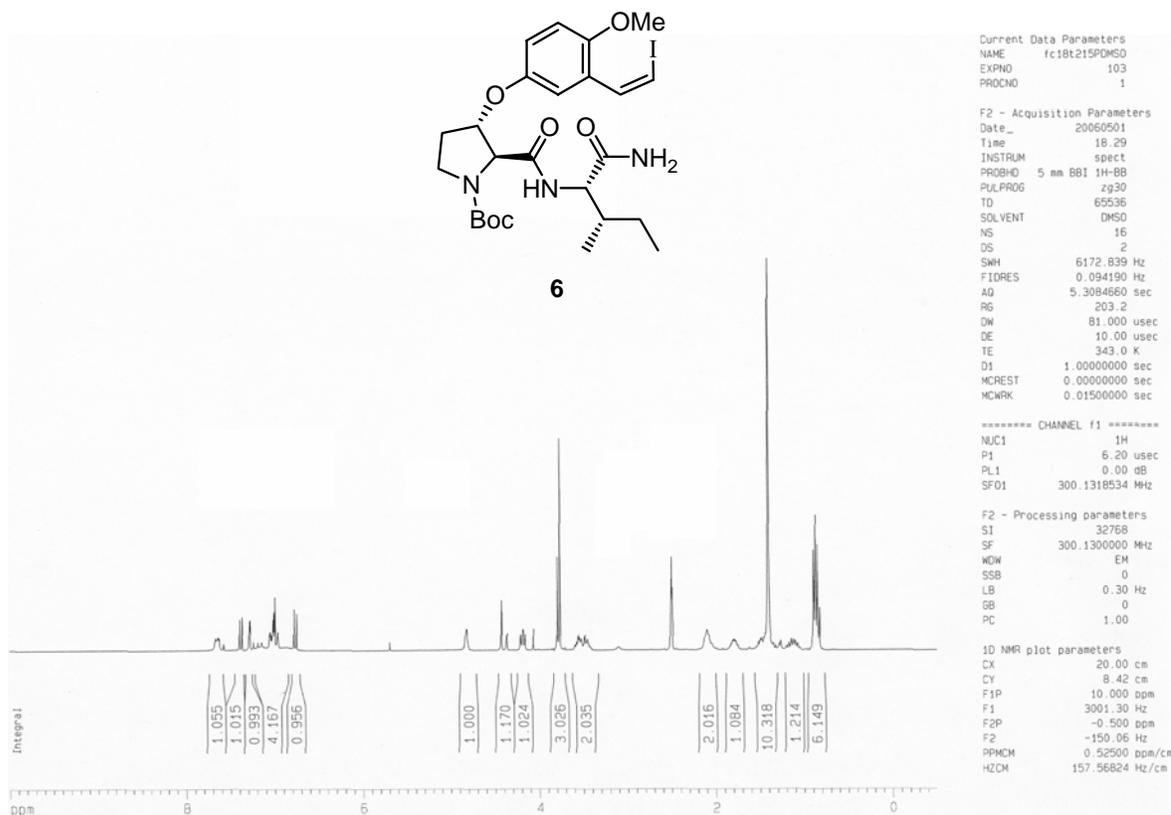
F2 - Acquisition Parameters
Date_    20060712
Time     16.18
INSTRUM  spect
PROBHD   5 mm BBI 1H-BB
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        5000
DS        4
SWH       17985.611 Hz
FIDRES    0.274439 Hz
AQ        1.8219508 sec
RG         128
DM        27.800 usec
DE         10.00 usec
TE        345.0 K
D1        2.00000000 sec
d11       0.03000000 sec
DELTA     1.89999998 sec
MCREST    0.00000000 sec
MCMRK     0.01500000 sec

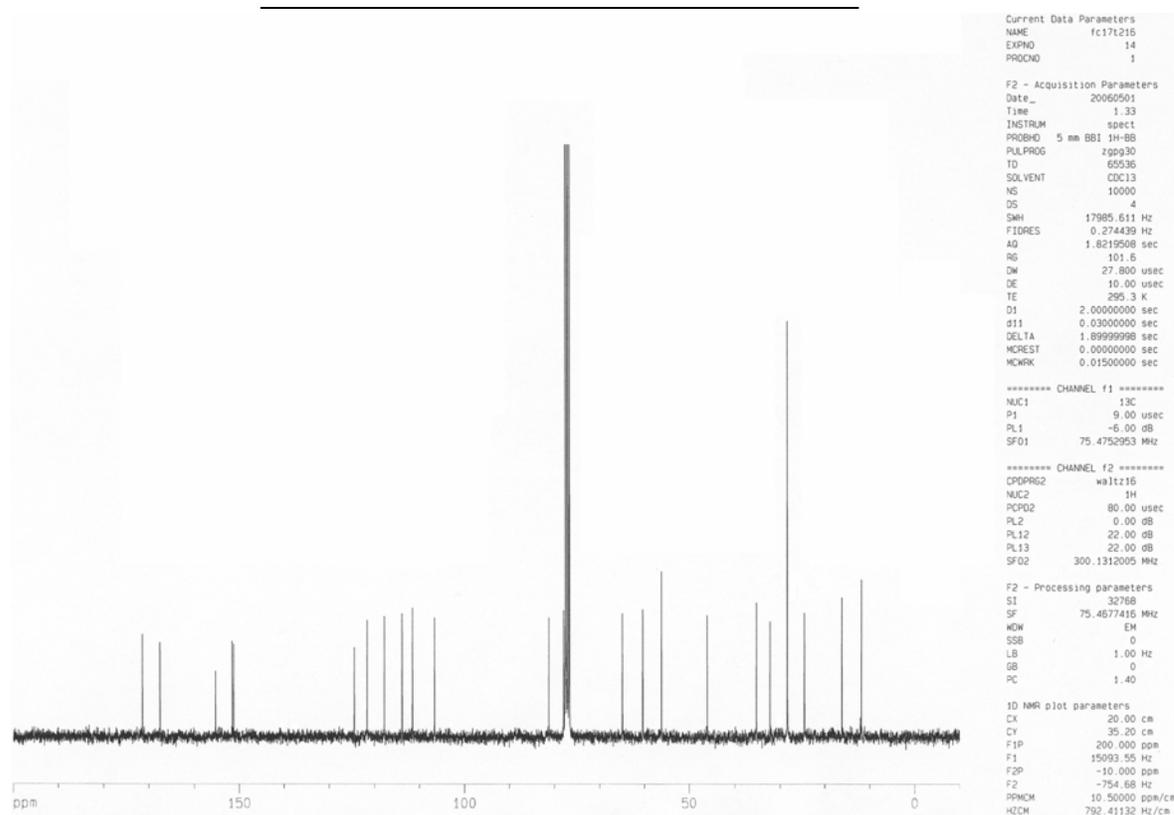
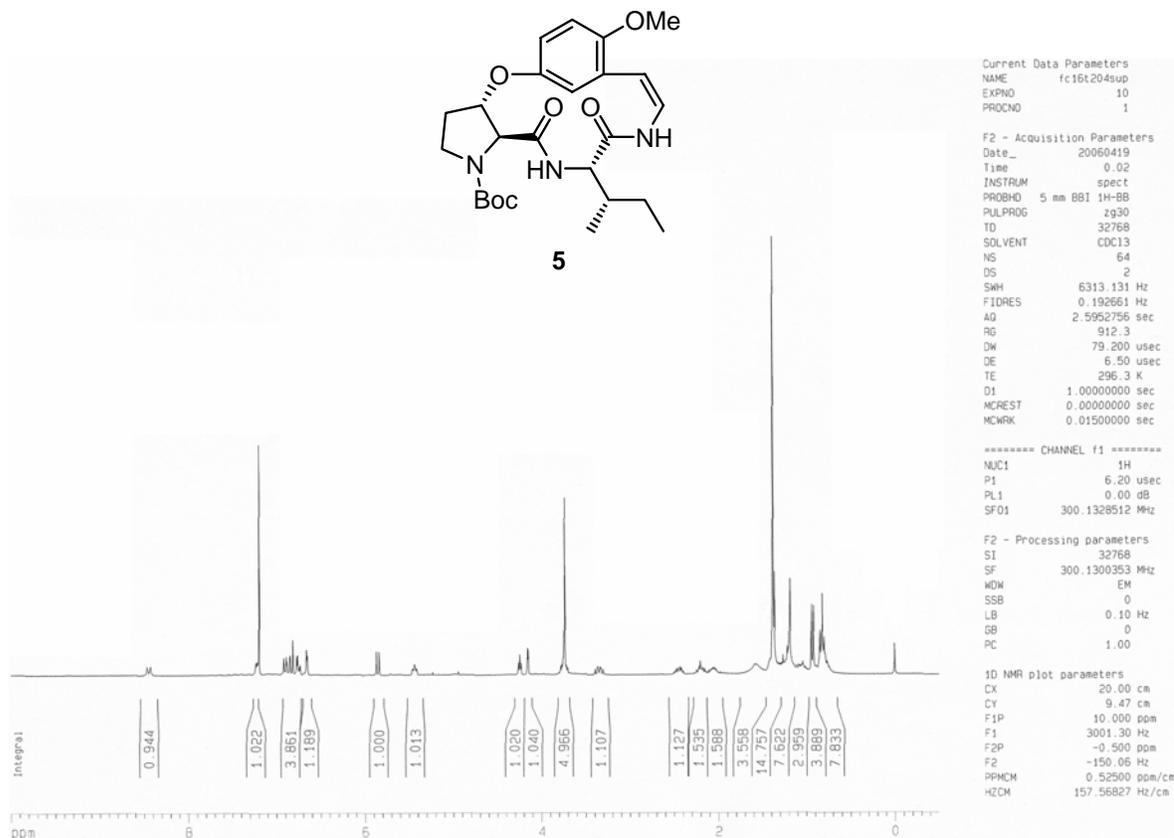
***** CHANNEL f1 *****
NUC1      13C
P1        9.00 usec
PL1       -6.00 dB
SF01      75.4752953 MHz

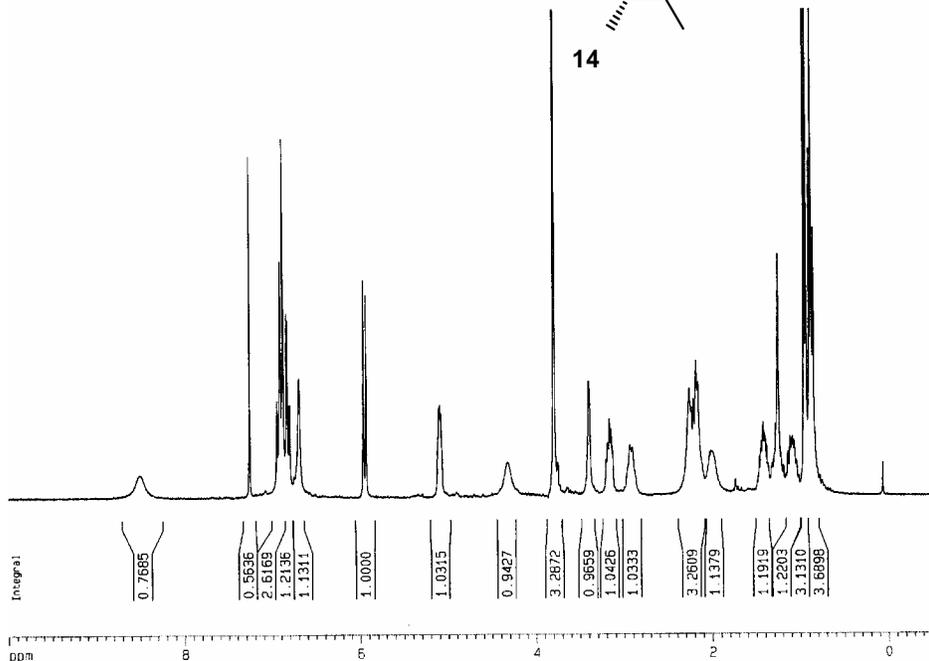
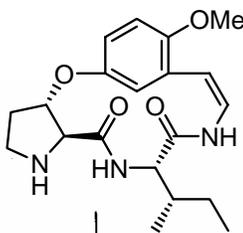
***** CHANNEL f2 *****
CPDPRG2  waltz16
NUC2      1H
PCPD2     80.00 usec
PL2       0.00 dB
PL12      22.00 dB
PL13      22.00 dB
SF02      300.1312005 MHz

F2 - Processing parameters
SI        32768
SF        75.4678141 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        1.40

1D NMR plot parameters
CX        20.00 cm
CY        23.56 cm
F1P       200.000 ppm
F1        15093.56 Hz
F2P       -10.000 ppm
F2        -754.68 Hz
PPHMC     10.50000 ppm/cm
HZCM      792.41199 Hz/cm
    
```







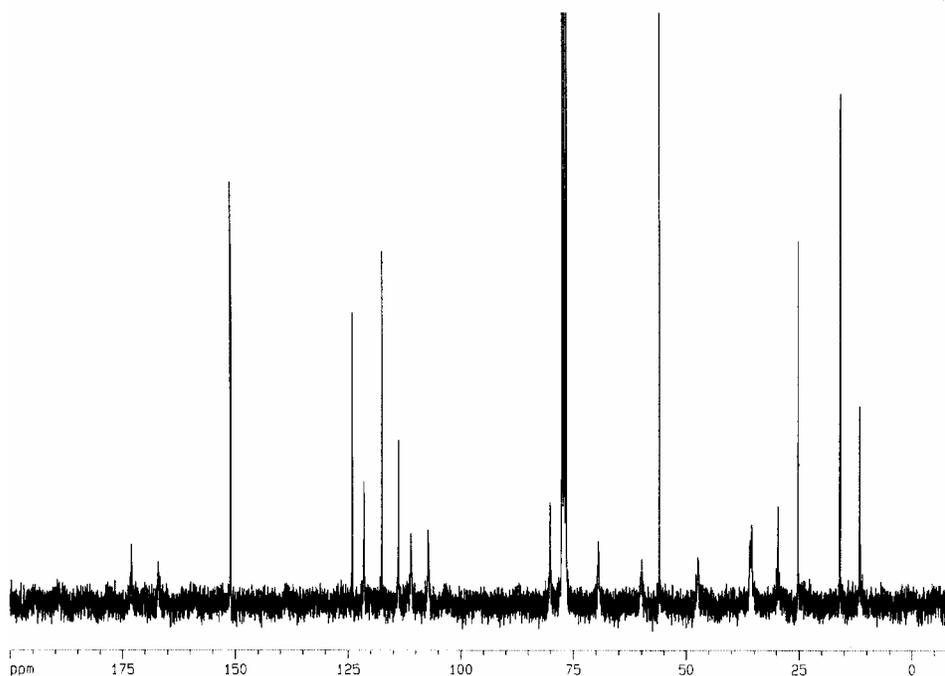
Current Data Parameters
 NAME fc24t249P
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060618
 Time 20.50
 INSTRUM spect
 PROBHD 5 mm BBI 1H-BB
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6313.134 Hz
 FIDRES 0.192664 Hz
 AQ 2.5952786 sec
 RG 456.1
 DW 79.200 usec
 DE 6.50 usec
 TE 296.2 K
 D1 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 1H
 P1 6.20 usec
 PL1 0.00 dB
 SF01 300.1320512 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300149 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 30.85 cm
 F1P 10.000 ppm
 F1 3001.30 Hz
 F2P -0.500 ppm
 F2 -150.07 Hz
 PPMCM 0.52500 ppm/cm
 HZCM 157.56825 Hz/cm



Current Data Parameters
 NAME fc24t249P
 EXPNO 10
 PROCNO 1

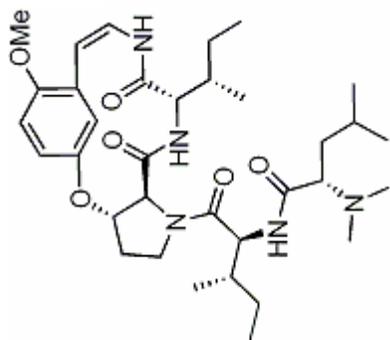
F2 - Acquisition Parameters
 Date_ 20060618
 Time 19.26
 INSTRUM spect
 PROBHD 5 mm BBI 1H-BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 20000
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 101.6
 DW 27.800 usec
 DE 16.00 usec
 TE 296.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 DELTA 1.89999998 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

***** CHANNEL f1 *****
 NUC1 13C
 P1 9.00 usec
 PL1 -6.00 dB
 SF01 75.4752953 MHz

***** CHANNEL f2 *****
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 22.00 dB
 PL13 22.00 dB
 SF02 300.1312005 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677490 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 CY 118.20 cm
 F1P 200.000 ppm
 F1 15093.55 Hz
 F2P -10.000 ppm
 F2 -754.68 Hz
 PPMCM 10.50000 ppm/cm
 HZCM 792.41138 Hz/cm



Current Data Parameters
 NAME fc26t253P
 EXPNO 11
 PROCNO 1

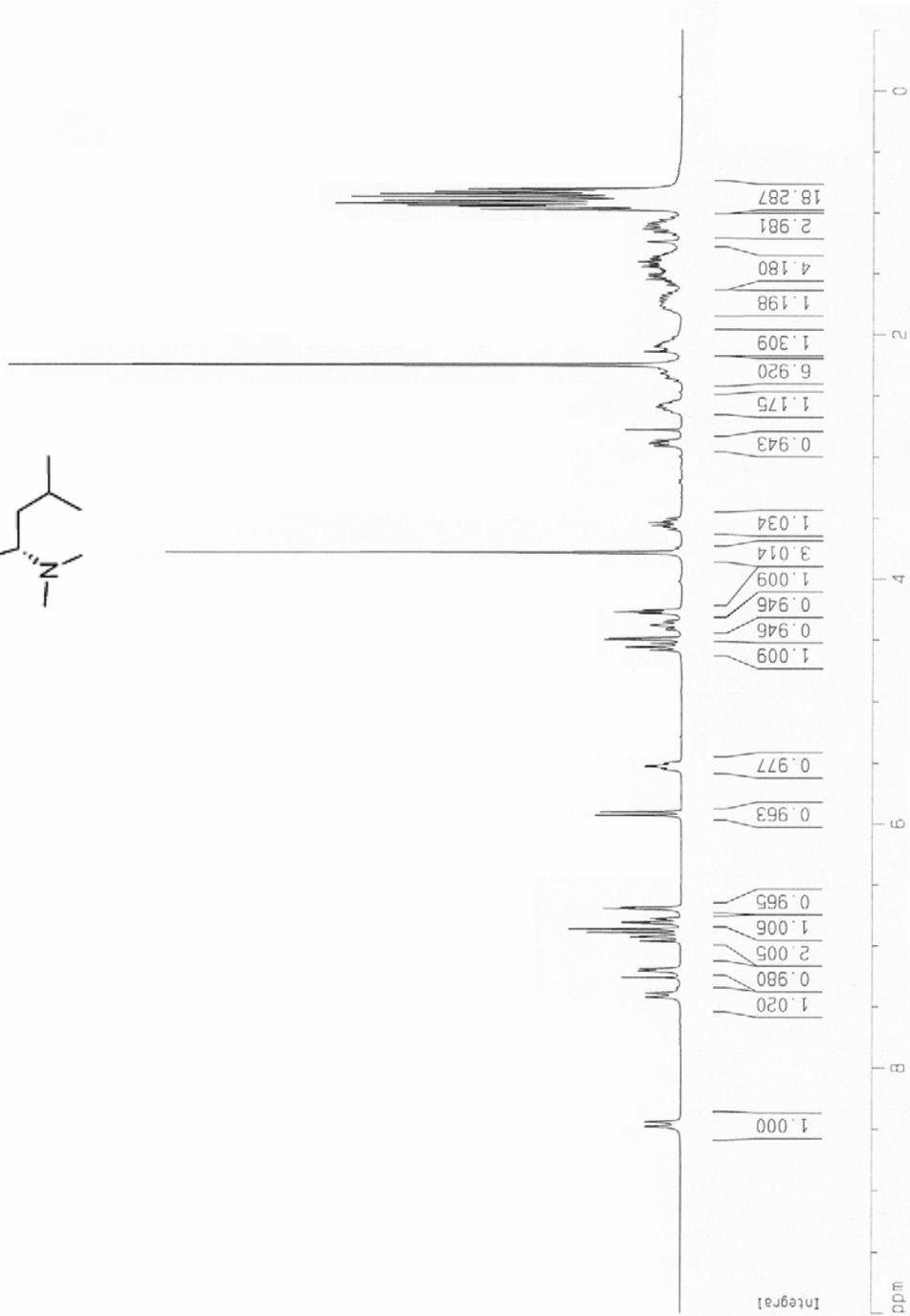
F2 - Acquisition Parameters

Date_ 20060702
 Time 11.15
 INSTRUM spect
 PROBHD 5 mm BBI 1H-BB
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6313.131 Hz
 FIDRES 0.192661 Hz
 AQ 2.5952756 sec
 RG 114
 DW 79.200 usec
 DE 6.50 usec
 TE 296.4 K
 D1 1.0000000 sec
 MCREST 0.0000000 sec
 MCMRK 0.0150000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 6.20 usec
 PL1 0.00 dB
 SF01 300.1328512 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300149 MHz
 MDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 CY 12.22 cm
 F1P 10.000 ppm
 F1 3001.30 Hz
 F2P -0.500 ppm
 F2 -150.06 Hz
 PPMCM 0.52500 ppm/cm
 HZCM 157.56625 Hz/cm



Current Data Parameters
 NAME fc26t253P
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20060702
 Time 11.13
 INSTRUM spect
 PROBHD 5 mm BBI 1H-8B
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 20000
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.274439 Hz
 AQ 1.8219508 sec
 RG 101.6
 DM 27.800 usec
 DE 10.00 usec
 TE 296.4 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 DELTA 1.8999998 sec
 HCREST 0.0000000 sec
 MCPRK 0.01500000 sec

***** CHANNEL f1 *****

NUC1 13C
 P1 9.00 usec
 PL1 -6.00 dB
 SF01 75.4752953 MHz

***** CHANNEL f2 *****

CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 22.00 dB
 PL13 22.00 dB
 SF02 300.1312005 MHz

F2 - Processing parameters

SI 32768
 SF 75.4677432 MHz
 NDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

10 NMR plot parameters

CX 20.00 cm
 CY 8.53 cm
 F1P 200.000 ppm
 F1 15093.55 Hz
 F2P -10.000 ppm
 F2 -754.68 Hz
 PPHCH 10.50000 ppm/cm
 HZCH 792.41132 Hz/cm

