



Supporting Information

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New Tricks for the Carboxylic Acid. Efficient Synthesis and Resolution of Pyrrolizidines

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General

All reactions were carried out under an argon atmosphere in absolute solvents with syringe and Schlenk techniques in oven-dried glassware. THF was distilled under argon from lead/sodium in the presence of benzophenone. Bulk solvents for chromatography and extraction were distilled prior to use. Reagents were obtained from commercial sources and used without further purification unless otherwise stated. TLC was performed on E. Merck precoated plates (SiO₂ 60 F₂₅₄, layer thickness 0.2 mm), and chromatography was performed with E. Merck SiO₂ 60 (0.040–0.063 mm) in the flash mode with a nitrogen pressure of 0.2 bar.

Melting points were determined with a Büchi 510 apparatus and are uncorrected. ¹H and ¹³C NMR spectra were recorded on a Varian VXR 300, a Varian Mercury 300, a Varian Inova 400 and a Varian Unity 500 instrument. ¹H chemical shifts are reported in delta (δ) units in parts per million (ppm) relative to the singlet at 7.26 ppm for *d*-chloroform (residual CHCl₃) and the singlet (0.00 ppm) for TMS. ¹³C Chemical shifts are reported in ppm relative to the central line of the triplet at 77.0 ppm for *d*-chloroform. Splitting patterns are designated as s, singlet; d, doublet; t, triplet; q, quartet; qt, quintet; m, multiplet; and l, large and combinations thereof. Coupling constants are recorded in Hertz (Hz). IR spectra were recorded neat on a Perkin-Elmer PE 1759 FT instrument; absorptions are given in wavenumbers (cm⁻¹). Low resolutions mass spectra were recorded on a Varian MAT 212 S instrument using either electron impact ionisation (EI, 70 eV), or chemical ionisation (CI, CH₄, isobutane or NH₃). High resolution mass spectra were recorded either on a Varian MAT 95 mass spectrometer or on a Micromass LCT Spectrometer (ESI, TOF). Optical rotations were measured with a Perkin-Elmer model 241 polarimeter in a 1-dm cell and the sodium D line (589 nm) at the temperature, solvent, and concentration indicated using solvents of Merck UVASOL-quality. Elemental analyses were performed by the Microanalytical Laboratory of the Institute of Organic Chemistry.

Data sets were collected with a Nonius KappaCCD diffractometer. Programs used: data collection COLLECT (Nonius B.V., 1998), data reduction Denzo-SMN (Z. Otwinowski, W. Minor, *Methods in Enzymology*, **1997**, 276, 307-326), absorption correction Denzo (Z. Otwinowski, D. Borek, W. Majewski, W. Minor, *Acta Cryst.* **2003**, A59, 228-234), structure solution SHELXS-97 (G.M. Sheldrick, *Acta Cryst.* **1990**, A46, 467-473), structure refinement SHELXL-97 (G.M. Sheldrick, Universität Göttingen, 1997), graphics SCHAKAL (E. Keller, Universität Freiburg, 1997).

CCDC 623489 and 632725 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge at www.ccdc.cam.ac.uk/conts/retrieving.html [or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: (internat.) +44(1223)336-033, E-mail: deposit@ccdc.cam.ac.uk].

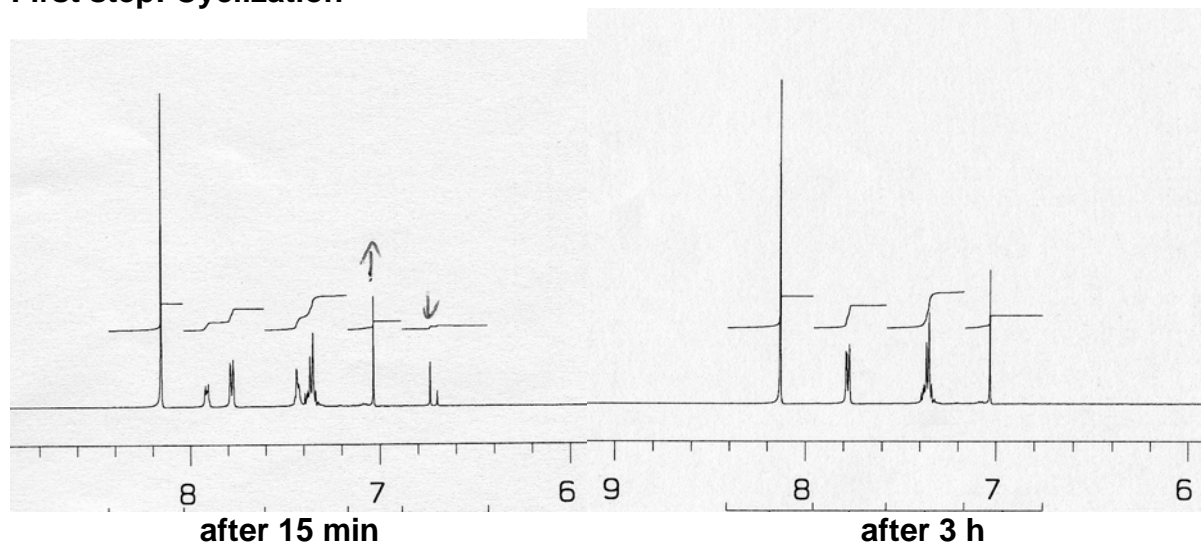
General procedure for the cyclization of maleimidoalkanoic acids (in NMR solvents)

To a suspension of pre-**7d** (131 mg, 0.51 mmol) in CDCl_3 (1,3 mL) is added Et_3N (180 mg, 1.78 mmol) upon which the solution became clear. After completed formation of the amide (NMR analysis), TBSOTf (402 mg, 1.52 mmol) is slowly added with a syringe. At the end of the addition the solution has turned red brown and it further darkens as the reaction proceeds. After 3 h, the reaction is diluted with CDCl_3 (10 mL) and extracted with 1N HCl (in order to remove the Et_3N). The aqueous layer is back extracted with 10 mL CDCl_3 . The combined organic layers are dried (Na_2SO_4), filtered and evaporated in vacuo. The solid residue is suspended in d^4 -MeOH (10 mL) and a catalytic amount of K_2CO_3 is added. As the reaction proceeds, a clear solution is obtained. After completion (NMR), solvent is evaporated in vacuo. The residue is portioned between 1N HCl and CDCl_3 (10 mL each). The layers are separated and the organic layer is dried (Na_2SO_4). From this solution, a crude NMR was obtained (see next page). The crude products can be further purified by flash chromatography (EtOAc/pentane 1:1). In large scale, these reactions were carried out in undeuterated solvents.

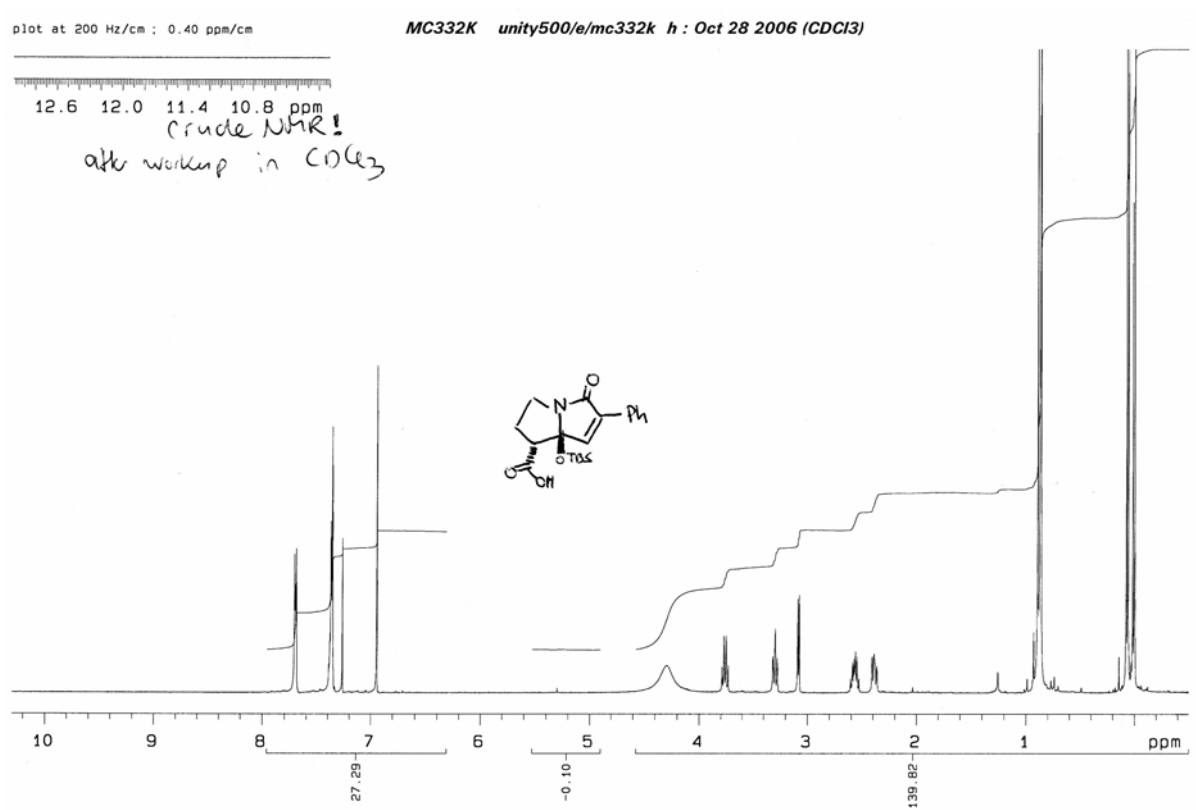
General Procedure for the Oxa-Michael lactonization

A solution of acid (\pm)-**7b** (20 mmol) and the cinchona alkaloid (10 mmol) in the appropriate solvent (15 mL) are stirred for the time indicated in table 1. The reaction progress can be monitored by removing a small sample, evaporating the solvent and taking an NMR in CDCl_3 . Upon desired conversion, the reaction mixture is extracted with 0.1M HCl. The organic layer is dried, filtered and evaporated in vacuo to give a solid residue. This material is triturated in hot pentane. This procedure is eventually repeated once. The residue consists of the racemic starting material (\pm)-**7b**. The pentane solution is evaporated to give the lactone **9** and enantiopure acid **7b**, which are easily separated by a short column (ethyl acetate/pentane 1:1→ethyl acetate).

First step: Cyclization



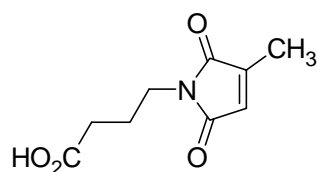
Second step: hydrolysis



Synthesis of the cyclization precursors

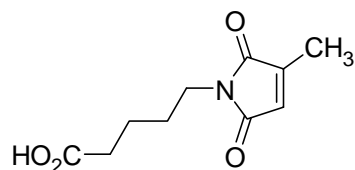
The maleimide carboxylic acid precursors (pre-7x) for the TBSOTf-mediated cyclization were synthesized according to the procedure of Rich et al.^[1] and – unless described in the literature – completely characterized.

4-(3-methyl-2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)butanoic acid (pre-7b)



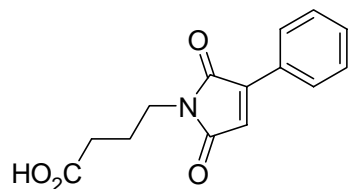
Colorless solid; mp 61-64 °C; ¹H NMR (500 MHz, CDCl₃) δ 6.32 (q, *J* = 1.8 Hz, 1H), 3.55 (t, *J* = 6.7 Hz, 2H), 2.35 (t, *J* = 7.3 Hz, 2H), 2.06 (d, *J* = 2.1 Hz, 3H), 1.90 (qn, *J* = 7.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 178.6, 171.8, 170.9, 145.7, 127.2, 36.9, 31.1, 23.6, 10.9; IR 1709, 1446, 1288, 1215, 926, 871, 750, 713 cm⁻¹; MS (EI) 197 (M⁺, 30), 179 (53), 151 (25), 138 (59), 137 (33), 124 (100); Anal calcd for C₉H₁₁NO₄: C, 54.82; H, 5.62; N, 7.10; Found: C, 54.74; H, 5.61; N, 7.01.

5-(3-methyl-2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)pentanoic acid (pre-7c)



Colorless solid; mp 51-54 °C; ¹H NMR (500 MHz, CDCl₃) δ 6.30 (q, *J* = 1.8 Hz, 1H), 3.49 (t, *J* = 6.4 Hz, 2H), 2.35 (t, *J* = 7.0 Hz, 2H), 2.05 (d, *J* = 1.8 Hz, 3H), 1.66-1.54 (m, 4H); ¹³C NMR (125 MHz, CDCl₃) δ 179.3, 171.8, 170.9, 145.5, 127.2, 37.3, 33.3, 27.8, 21.6, 10.9; IR (KBr) 3099, 2946, 1707, 1444, 1407, 1256, 1209, 1043, 908, 871 cm⁻¹; MS (EI) 211 ([M]⁺, 34), 193 (20), 165 (34), 137 (21), 124 (100), 96 (18); Anal calcd for C₁₀H₁₃NO₄: C, 56.86; H, 6.20; N, 6.63; Found: C, 56.85; H, 6.01; N, 6.59.

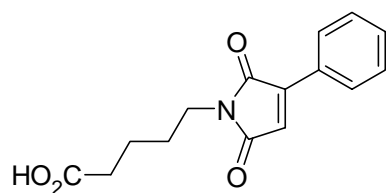
4-(2,5-dioxo-3-phenyl-2,5-dihydro-1H-pyrrol-1-yl)butanoic acid (pre-7d)



Colorless solid; mp 129-132 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.93-7.89 (m, 2H), 7.47-7.38 (m, 3H), 6.74 (s, 1H), 3.64 (t, *J* = 6.7 Hz, 2H), 2.38 (t, *J* = 7.3 Hz, 2H), 1.95 (qn, *J* = 7.0 Hz, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 178.6, 170.6, 170.4, 143.7, 131.1, 128.9 (2C), 128.6 (3C), 123.6, 37.0, 31.2, 23.5; IR 1702, 1610, 1445, 1407, 1288, 791 cm⁻¹; MS (EI) 259 (M⁺, 24), 85 (98), 83 (89), 69 (100), 55 (93); Anal calcd for C₁₄H₁₃NO₄: C, 64.86; H, 5.05; N, 5.40; Found: C, 65.11; H, 4.96; N, 5.40.

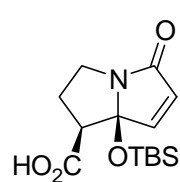
¹ D. H. Rich, P. D. Gesellchen, A. Tong, A. Cheung, C. K. Buckner, *J. Med. Chem.* **1975**, *18*, 1004.

5-(2,5-dioxo-3-phenyl-2,5-dihydro-1H-pyrrol-1-yl)pentanoic acid (pre-7f)



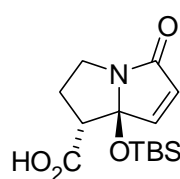
Colorless solid; mp 99-101 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.95-7.88 (m, 2H), 7.48-7.43 (m, 3H), 6.73 (s, 1H), 3.64-3.56 (m, 2H), 2.44-2.36 (m, 2H), 1.75-1.60 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.9, 170.5, 170.3, 143.6, 131.0, 128.8 (2C), 128.6, 128.5 (2C), 123.7, 37.5, 33.4, 28.0, 21.8; IR (KBr) 1702, 1443, 1408, 1250, 794 cm^{-1} ; MS (EI) 273 (M^+ , 14), 181 (71), 153 (45), 109 (42), 105 (100), 55 (26); Anal calcd for $\text{C}_{15}\text{H}_{15}\text{NO}_4$: C, 65.92; H, 5.53; N, 5.13 Found: C, 65.98; H, 5.47; N, 5.01.

(1S,7aS)-7a-(tert-butyltrimethylsilyloxy)-5-oxo-2,3,5,7a-tetrahydro-1H-pyrrolizine-1-carboxylic acid (7a)



Colorless solid; mp 58-61 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.24 (d, $J = 5.8$ Hz, 1H), 6.02 (d, $J = 5.8$ Hz, 1H), 3.55-3.48 (m, 1H), 3.39-3.33 (m, 1H), 2.90-2.80 (m, 1H), 2.60 (dd, $J = 11.9, 7.3$ Hz, 1H), 2.48-2.41 (m, 1H), 0.83 (s, 9H), 0.08 (s, 3H), 0.00 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 173.8, 173.3, 150.4, 126.8, 98.5, 52.0, 41.0, 39.4, 25.3 (3C), 17.9, -3.6, -4.2; IR (KBr) 2937, 2861, 1733, 1670, 1381, 1344, 1143, 836 cm^{-1} ; MS (EI) 297 (M^+ , 1), 282 ($[\text{M}-\text{CH}_3]^+$, 1) 240 ($[\text{M}-t\text{Bu}]^+$, 100), 196 (39), 166 ($[\text{M}-\text{TBSO}]^+$, 4), 75 (51); Anal calcd for $\text{C}_{14}\text{H}_{23}\text{NO}_4\text{Si}$: C, 56.54; H, 7.79; N, 4.71; Found: C, 56.91; H, 7.96; N, 4.72

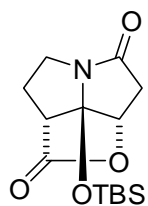
(1R,7aS)-7a-(tert-butyltrimethylsilyloxy)-5-oxo-2,3,5,7a-tetrahydro-1H-pyrrolizine-1-carboxylic acid (7b)



This compound has been described previously.^[2]

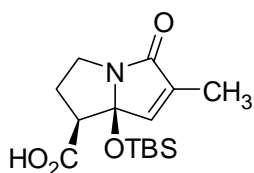
² T. H. Lambert, S. J. Danishefsky, *J. Am. Chem. Soc.* **2006**, *128*, 426.

(2aR,2a¹R,7aS)-2a¹-(tert-butyldimethylsilyloxy)tetrahydrofuro[4,3,2-gh]pyrrolizine-2,6(2aH,2a¹H)-dione (9)



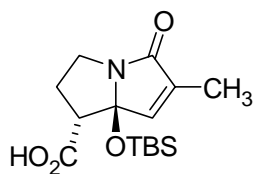
Colorless solid; mp 100-103 °C; R_f 0.41 (pentane/EtOAc 4:1); $[\alpha]_D^{23}$ -26.1 (c 1.13 CHCl_3); ^1H NMR (400 MHz, CDCl_3) δ 4.79 (d, $J = 4.4$ Hz, 1H), 3.81 (dddd, $J = 2.5, 6.6, 9.0, 15.7$ Hz, 1H), 3.23 (m, 1H), 3.16 (dd, $J = 2.2, 8.2$ Hz, 1H), 3.01 (d, $J = 4.4$ Hz, 1H), 2.55 (d, $J = 17.8$ Hz, 1H), 2.47 (m, 2H), 0.89 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 174.9, 173.6, 102.7, 80.5, 49.7, 42.8, 38.6, 29.8, 25.4 (3C), 17.8, -3.4, -3.7; IR (CHCl_3) 2953, 2858, 1785, 1719, 1363, 1328, 1135, 1032, 839, 777 cm^{-1} ; MS (EI): m/z (%) 438 ($[\text{M}^+ - 57]$, 100), 75 (19); HRMS calcd for $\text{C}_{10}\text{H}_{14}\text{NO}_4\text{Si}$ ($\text{M}^+ - 57$), 240.06921; found, 240.06921.

(1S,7aS)-7a-(tert-butyldimethylsilyloxy)-6-methyl-5-oxo-2,3,5,7a-tetrahydro-1H-pyrrolizine-1-carboxylic acid (10a)



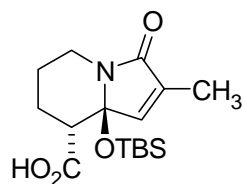
Colorless solid; mp 88-91 °C; ^1H NMR (500 MHz, CDCl_3) δ 6.85 (q, $J = 1.5$ Hz, 1H), 3.52-3.45 (m, 1H), 3.39-3.33 (m, 1H), 2.88-2.77 (m, 1H), 2.61 (dd, $J = 12.2, 7.3$ Hz, 1H), 2.44-2.37 (m, 1H), 1.86 (d, $J = 1.8$ Hz, 3H), 0.82 (s, 9H), 0.04 (s, 3H), -0.03 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 174.6, 174.3, 143.1, 135.4, 96.7, 52.3, 41.1, 29.2, 25.4, 17.9, 10.7, -3.6, -4.2; IR (KBr) 1705, 1131, 1074, 841, 779 cm^{-1} ; MS (EI) 311 (M^+ , 2), 254 ($[\text{M}-t\text{Bu}]^+$, 100), 210 (51), 180 ($[\text{M}-\text{TBSO}]^+$, 6), 136 (15), 75 (50); Anal calcd for $\text{C}_{15}\text{H}_{25}\text{NO}_4\text{Si}$: C, 57.85; H, 8.09; N, 4.50; Found: C, 57.85; H, 7.89; N, 4.42.

(1R,7aS)-7a-(tert-butyldimethylsilyloxy)-6-methyl-5-oxo-2,3,5,7a-tetrahydro-1H-pyrrolizine-1-carboxylic acid (10b)



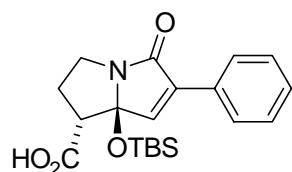
Colorless solid; mp 141-143 °C; ^1H NMR (400 MHz, CDCl_3) δ 6.58 (q, $J = 1.6$ Hz, 1H), 3.76-3.66 (m, 1H), 3.30-3.22 (m, 1H), 3.06 (d, $J = 6.6$ Hz, 1H), 2.64-2.53 (m, 1H), 2.50-2.42 (m, 1H), 1.80 (d, $J = 1.6$ Hz, 3H), 0.85 (s, 9H), 0.03 (s, 3H), -0.02 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.6, 174.3, 140.2, 136.7, 98.7, 52.5, 41.9, 31.0, 25.5 (3C), 17.9, 10.7, -3.4, -4.0; IR (KBr) 2953, 2860, 1739, 1676, 1473, 1440, 1411, 1382, 1254, 1093, 839 cm^{-1} ; MS (EI) 311 (M^+ , 2), 296 ($[\text{M}-\text{CH}_3]^+$, 1), 254 ($[\text{M}-t\text{Bu}]^+$, 100), 236 (50), 210 (42), 75 (51); Anal calcd for $\text{C}_{15}\text{H}_{25}\text{NO}_4\text{Si}$: C, 57.85; H, 8.09; N, 4.50; Found: C, 57.88; H, 7.99; N, 4.46.

(8*R*,8*aS*)-8*a*-(*tert*-butyldimethylsiloxy)-2-methyl-3-oxo-3,5,6,7,8,8*a*-hexahydroindolizine-8-carboxylic acid (10c)



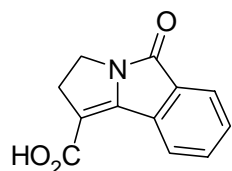
Colorless solid; mp 125-127 °C; ¹H NMR (400 MHz, CDCl₃) δ 6.51 (q, *J* = 1.7 Hz, 1H), 4.07 (dd, *J* = 11.7, 5.5 Hz, 1H), 2.99-2.87 (m, 2H), 2.22-2.11 (m, 1H), 1.96-1.88 (m, 2H) 1.86 (d, *J* = 1.7 Hz, 3H), 1.54-1.46 (m, 1H), 0.88 (s, 9H), 0.01 (s, 3H), -0.09 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 175.4, 168.0, 139.7, 136.9, 86.5, 48.6, 35.7, 25.7 (3C), 22.0, 19.6, 18.2, 10.8, -2.6, -3.8; IR (KBr) 3280, 2935, 2859, 1734, 1685, 1651, 1437, 1060, 837, 670 cm⁻¹; MS (EI) 325 (M⁺, 5), 310 ([M-CH₃]⁺, 2), 268 [M-*t*Bu]⁺, 97), 250 (91), 224 (70), 176 (46), 75 (100); Anal calcd for C₁₆H₂₇NO₄Si: C, 59.04; H, 8.36; N, 4.30; Found: C, 58.80; H, 8.19; N, 4.14.

(1*R*,7*aS*)-7*a*-(*tert*-butyldimethylsilyloxy)-5-oxo-6-phenyl-2,3,5,7*a*-tetrahydro-1*H*-pyrrolizine-1-carboxylic acid (10d)



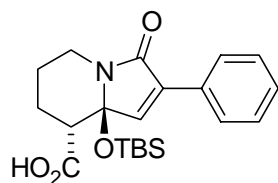
Colorless solid; mp 170-172 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.71-7.67 (m, 2H), 7.38-7.33 (m, 3H), 6.93 (s, 1H), 3.76 (q, *J* = 9.1 Hz, 1H), 3.29 (t, *J* = 9.4 Hz, 1H), 3.08 (d, *J* = 6.1 Hz, 1H), 2.61-2.52 (m, 1H), 2.41-2.34 (m, 1H), 0.86 (s, 9H), 0.05 (s, 3H), -0.01 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 175.5, 172.4, 139.1, 137.7, 130.4 (d, *J* = 2.1 Hz), 129.2, 128.5 (2C), 127.6 (2C), 97.9, 52.6, 42.1, 30.7, 25.5 (3C), 17.8, -3.3, -4.1; IR (KBr) 2939, 2860, 1735, 1667, 1623, 1385, 1255, 1197, 1091, 911, 840, 786, 686 cm⁻¹; MS (EI) 373 (M⁺, 1), 358 ([M-CH₃]⁺, 2), 316 ([M-*t*Bu]⁺, 100), 75 (89); EA Anal calcd for C₂₀H₂₇NO₄Si: C, 64.31; H, 7.29; N, 3.75; Found: C, 64.33; H, 7.47; N, 3.68.

5-oxo-3,5-dihydro-2*H*-pyrrolo[2,1-*a*]isoindole-1-carboxylic acid (10e)



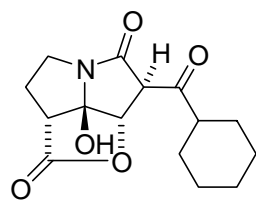
Colorless solid; mp 229-231 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 13.00 (sb, 1H), 8.56 (d, *J* = 8.55 Hz, 1H), 7.74-7.88 (m, 3H), 3.95 (t, *J* = 8.24 Hz, 2H), 3.33 (t, *J* = 8.24 Hz, 2H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 165.8, 162.7, 147.5, 136.0, 132.4, 131.7, 129.8, 126.6, 123.2, 111.9, 40.1, 34.5; IR (KBr) 1723, 1621, 1450, 1312, 1268, 1222, 750, 687 cm⁻¹; MS (EI) 215 (M⁺, 100), 170 ([M-CO₂H]⁺, 75), 142 (16), 115 (21); Anal calcd for C₁₂H₉NO₃: C, 66.97; H, 4.22; N, 6.51; Found: C, 66.77; H, 4.59; N, 6.48.

(8*R*,8*aS*)-8*a*-(*tert*-butyldimethylsilyloxy)-3-oxo-2-phenyl-3,5,6,7,8,8*a*-hexahydroindoline-8-carboxylic acid (10*f*)



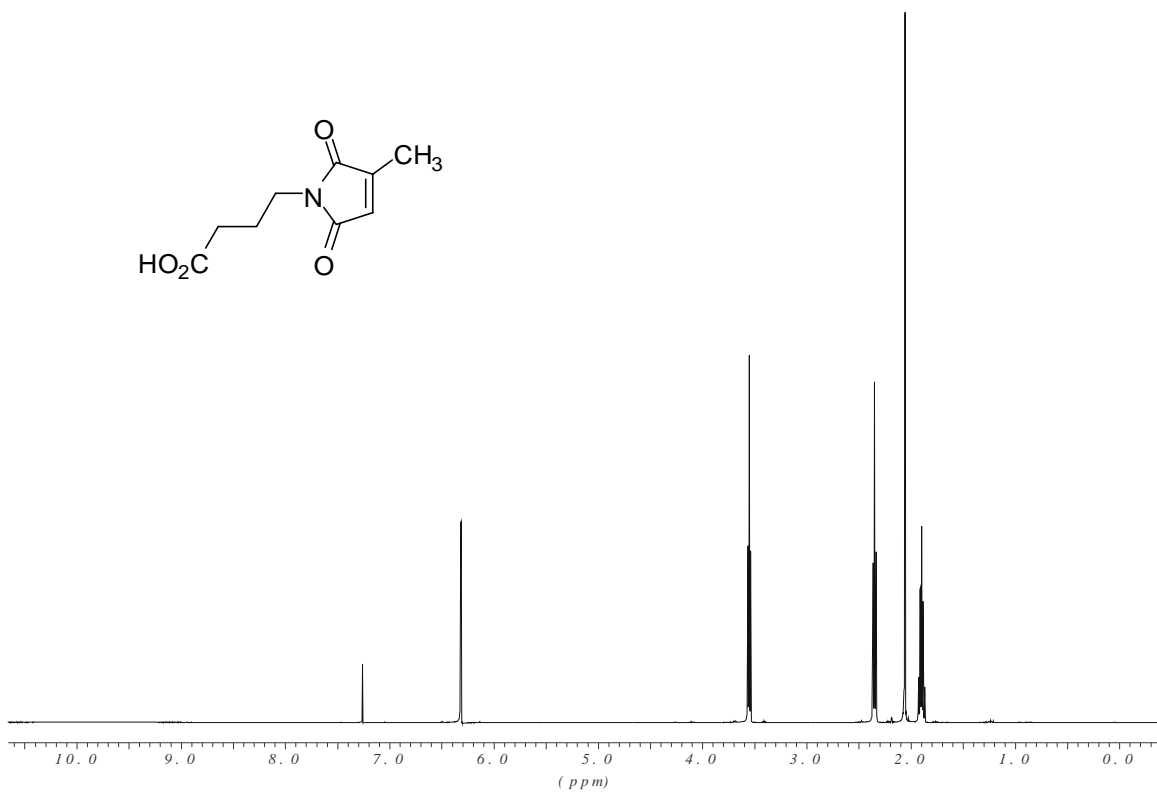
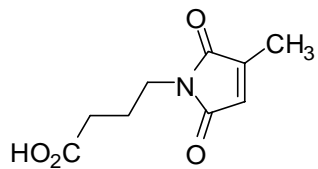
Colorless solid; mp 140-142 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.78-7.74 (m, 2H), 7.40-7.34 (m, 3H), 6.85 (s, 1H), 4.14 (dd, $J = 13.0, 5.4$ Hz, 1H), 3.02-2.88 (m, 2H), 2.22-2.12 (m, 1H), 1.92-1.75 (m, 2H), 1.57-1.44 (m, 1H), 0.89 (s, 9H), 0.02 (s, 3H), -0.07 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 175.7, 166.1, 139.0, 137.6, 130.7, 128.9, 128.4 (2C), 127.5 (2C), 85.8, 48.5, 35.7, 25.7 (3C), 22.0, 19.6, 18.2, -2.4 , -3.8 ; IR (KBr) 2932, 2863, 1739, 1672, 1428, 1164, 1068, 832 cm^{-1} ; MS (EI) 387 (M^+ , 1), 330 ($[\text{M}-t\text{Bu}]^+$, 19), 312 (12), 286 (16), 255 (19), 211 (48), 182 (18), 75 (100); Anal calcd for $\text{C}_{21}\text{H}_{29}\text{NO}_4\text{Si}$: C, 65.08; H, 7.54; N, 3.61; Found: C, 65.15; H, 7.38; N, 3.51.

(2*aR*,2*a*¹*R*,7*R*,7*aS*)-7-(Cyclohexanecarbonyl)-2*a*¹-hydroxytetrahydrofuro[4,3,2-*gh*]pyrrolizine-2,6(2*aH*,2*a*¹*H*)-dione (11)



Colorless solid; R_f 0.41 (pentane/EtOAc 1:1); $[\alpha]_D^{24} = -22.2$ (c 0.90 CHCl_3); ^1H NMR (500 MHz, CDCl_3) δ 5.01 (s), 4.73 (s), 4.18 (s), 3.87 (dddd, $J = 5.8, 9.2, 11.9, 15.0$ Hz), 3.78 (dddd, $J = 5.2, 8.9, 11.6, 15.0$ Hz), 3.44 (dddd, $J = 5.5, 9.8, 12.2, 15.3$ Hz), 3.29 (dddd, $J = 5.2, 9.8, 12.0, 15.0$ Hz), 3.22 (td, $J = 1.8, 7.3$ Hz), 2.94 (tdd, $J = 3.9, 11.3, 15.0$ Hz), 2.68 (m), 2.51 (m), 2.00 (m), 1.81 (m), 1.69 (m), 1.27 (m); ^{13}C NMR (125 MHz, CDCl_3) δ 209.6, 200.3, 174.8, 172.8, 167.1, 166.9, 100.9, 98.8, 83.9, 80.5, 62.7, 51.3, 48.1 (2C), 47.7, 42.6, 41.7, 30.3, 30.0, 29.7, 29.4, 29.1, 28.5, 28.3, 26.9, 25.6 (3C), 25.2, 24.9; IR (CHCl_3) 3241, 2929, 2858, 1802, 1722, 1690, 1326, 1309, 1217, 1164, 1042, 758 cm^{-1} ; MS (EI): m/z (%) 293 (M^+ , 5), 275 (8), 111 (58), 83 (100), 55 (36); HRMS calcd for $\text{C}_{15}\text{H}_{19}\text{NO}_5$ (M^+), 293.12632; found, 293.12632.

pre-7b



—178.5556
—171.8061
—170.8528

—145.6545

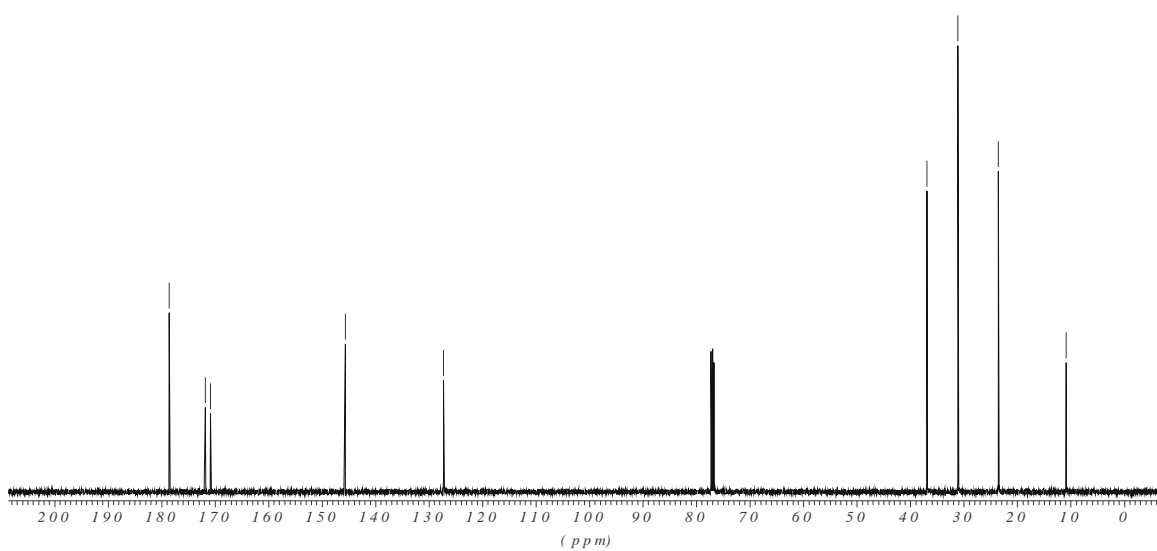
—127.2287

—36.9299

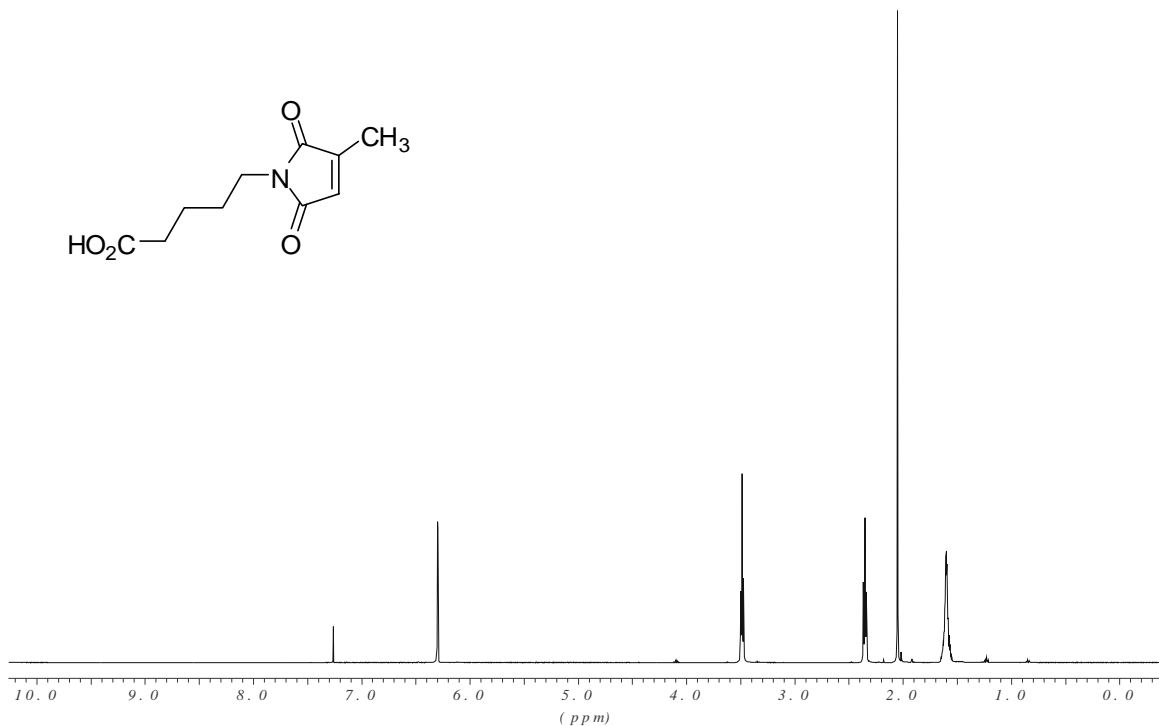
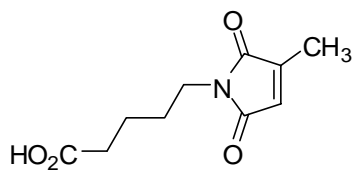
—31.1261

—23.5529

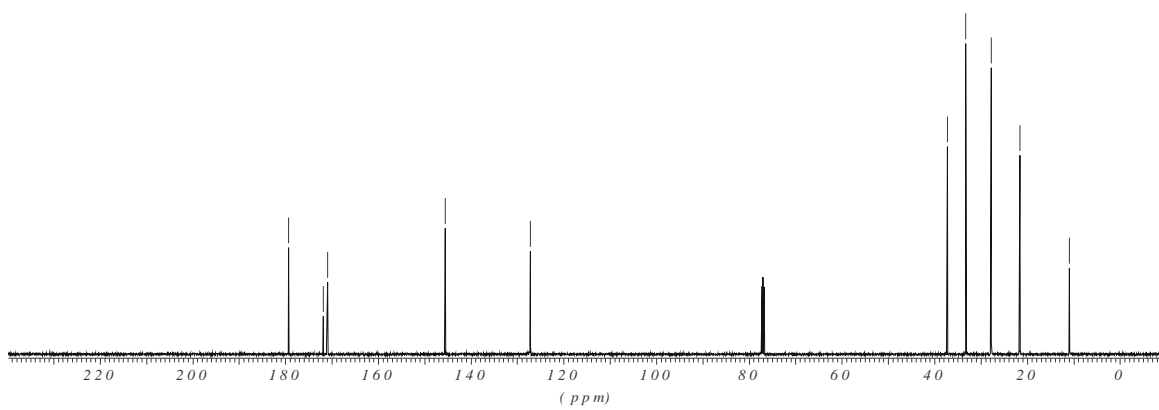
—10.9080



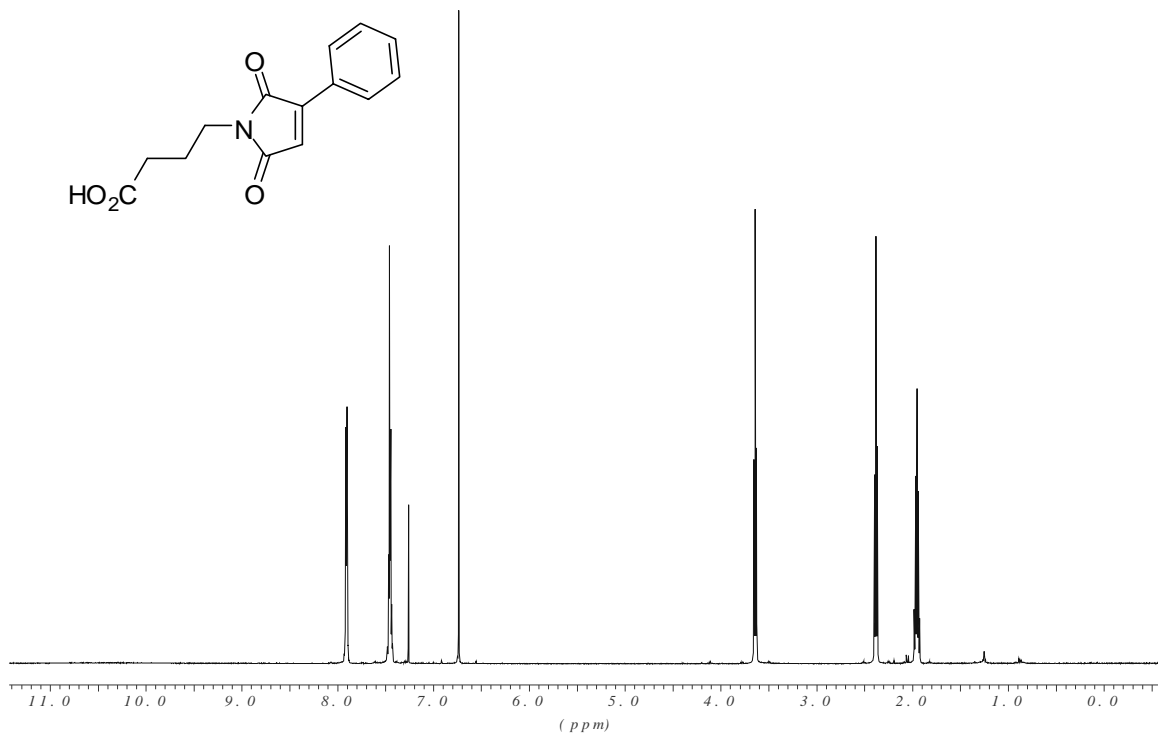
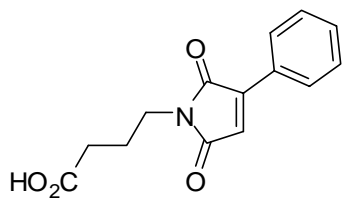
pre-7c



— 179.2802
— 171.8366
— 170.9138
— 145.5173
— 127.1982
— 37.2655
— 33.2844
— 27.8314
— 21.6462
— 10.8927



pre-7d

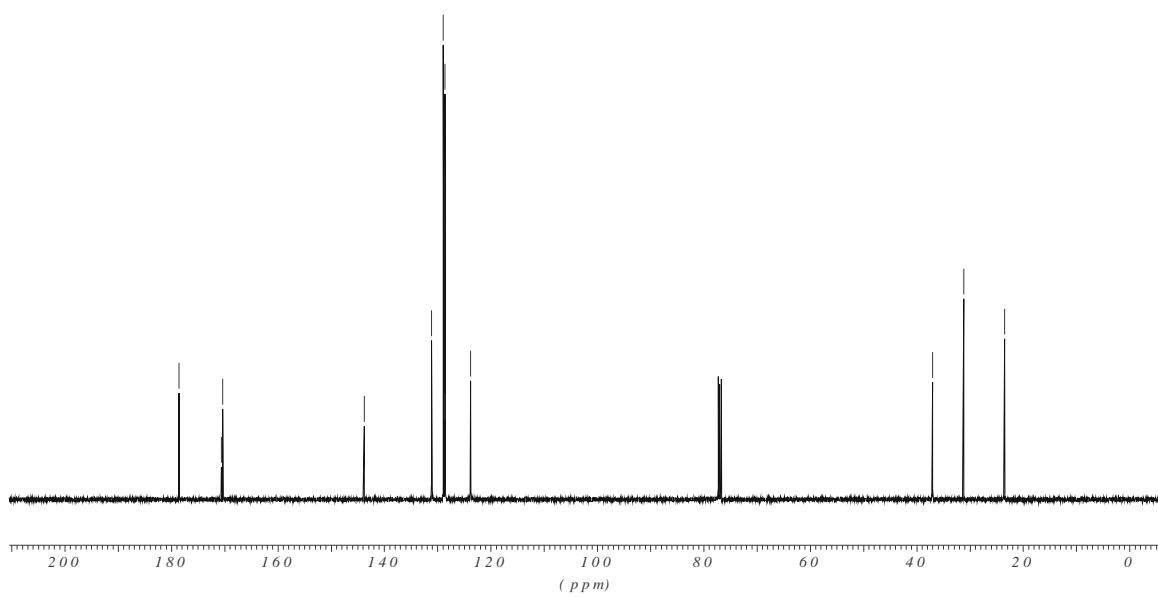


— 178.5938
— 170.5935
— 170.3570

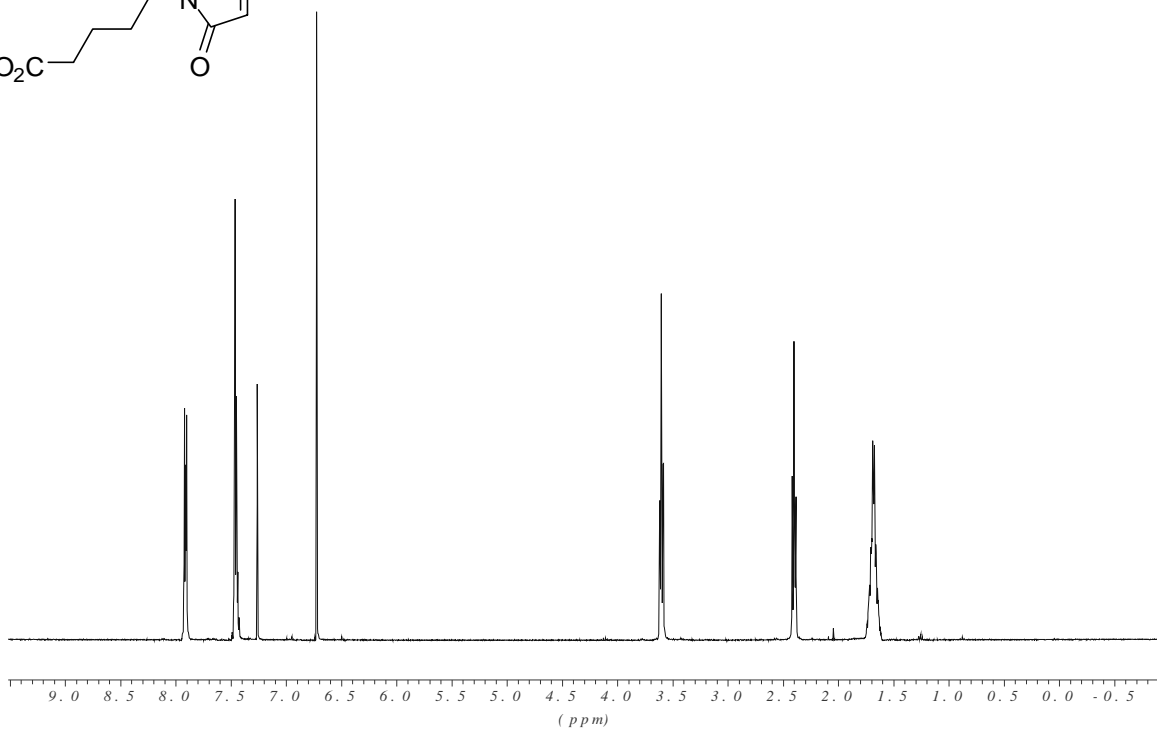
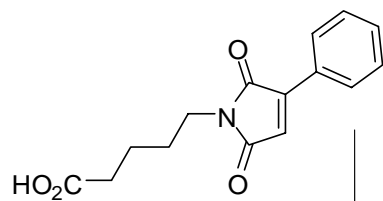
— 143.7479

— 131.1183
— 128.8989
— 128.6707
— 128.5786
— 123.7891

— 37.0367
— 31.1795
— 23.5300



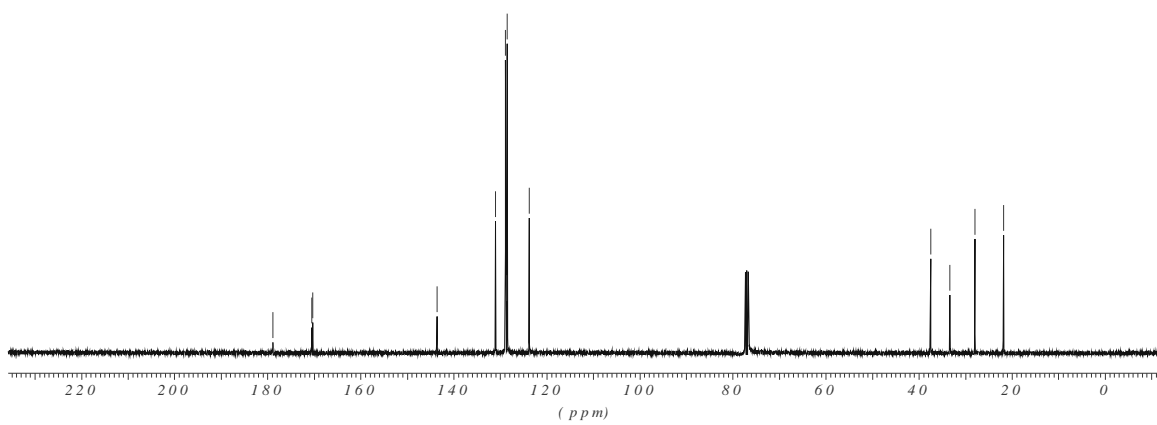
pre-7f



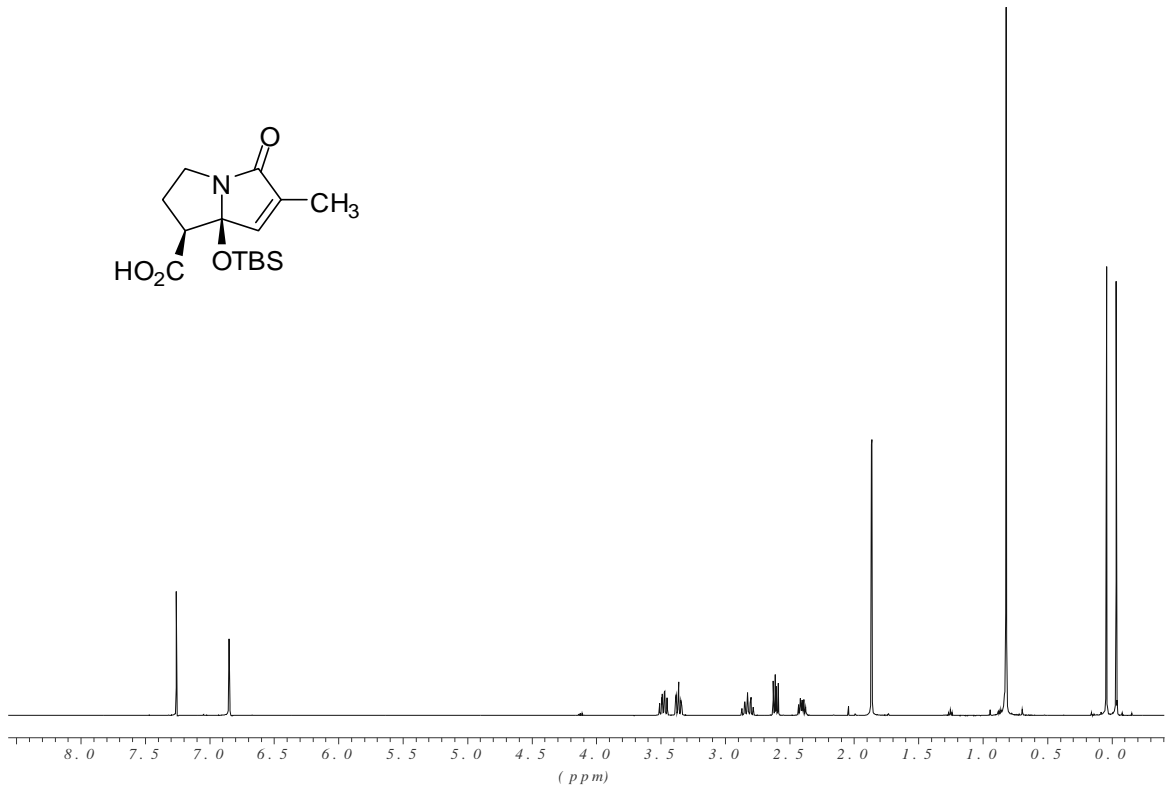
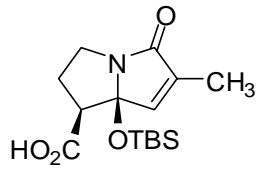
— 178.8853
— 170.5181
— 170.2830

— 143.6188
— 131.0111
— 128.8360
— 128.6272
— 123.5076
— 123.7439

— 37.4778
— 33.3663
— 27.9652
— 21.8434



7a



174.6279
174.2771

143.1225
135.4349

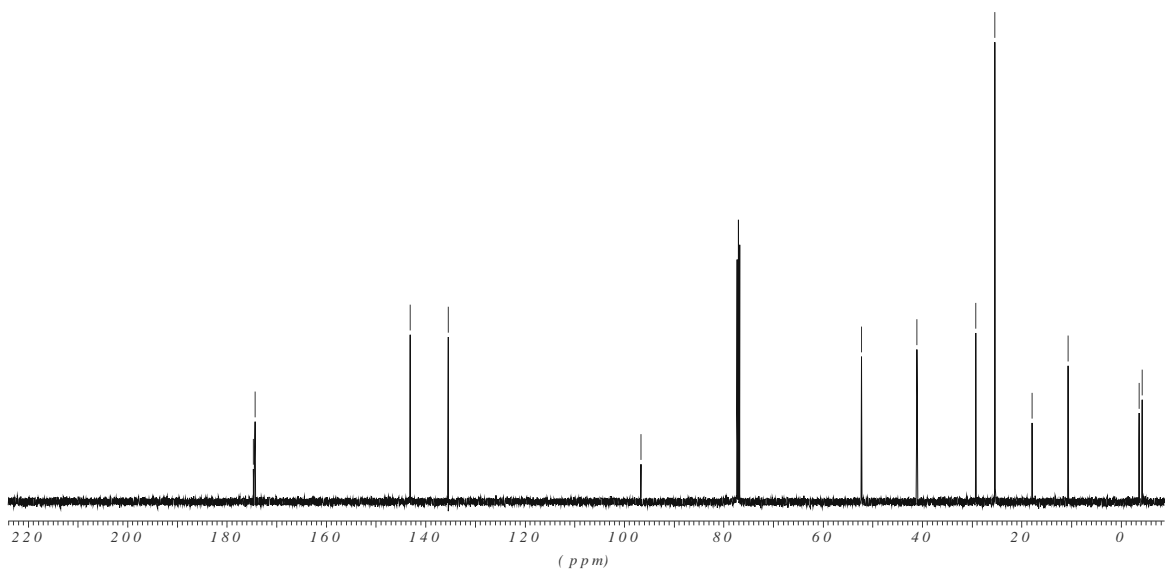
96.6537

52.2517
41.1093

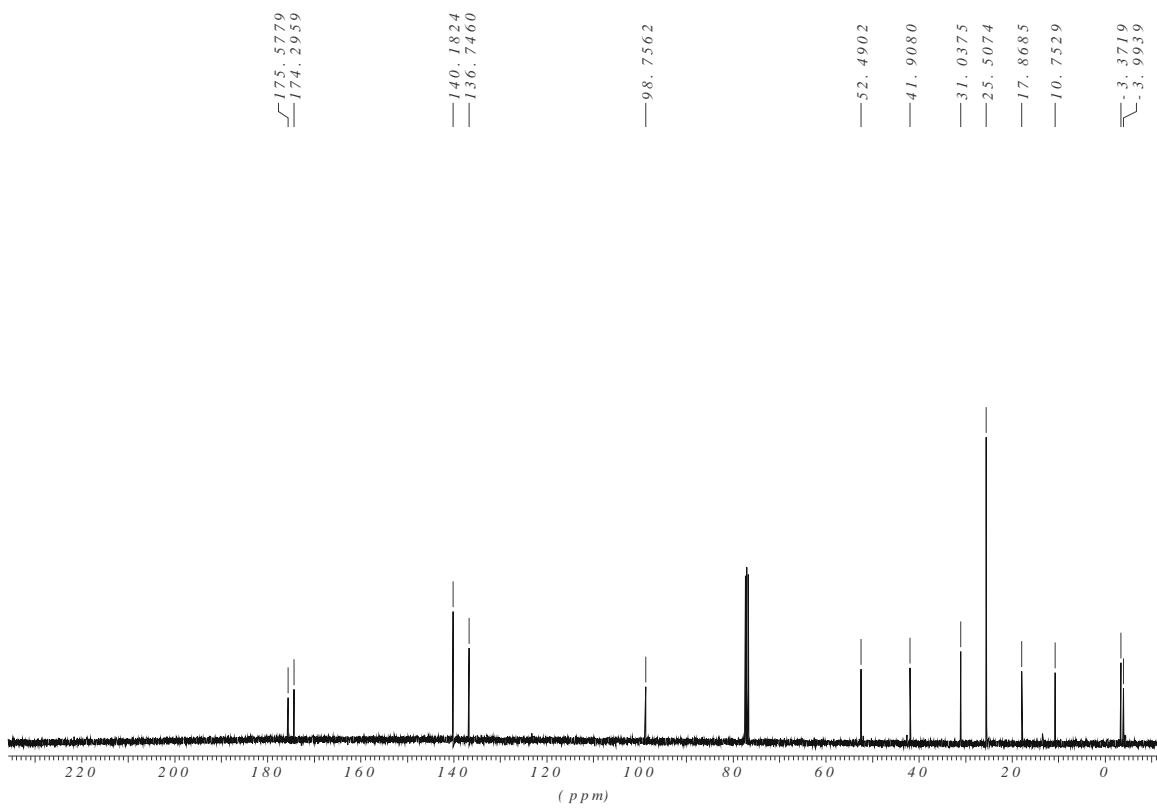
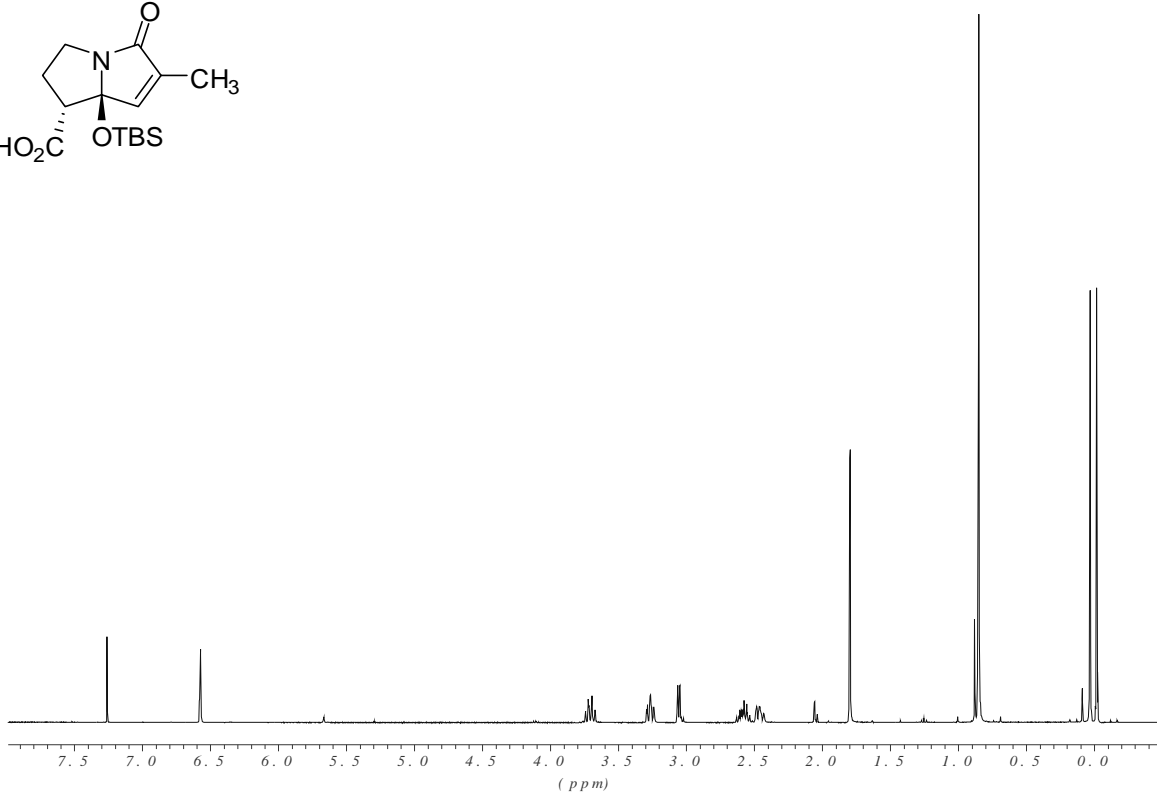
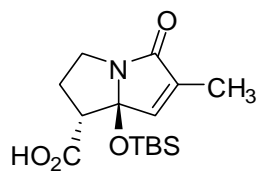
29.2194
25.3604

17.8787
10.6944

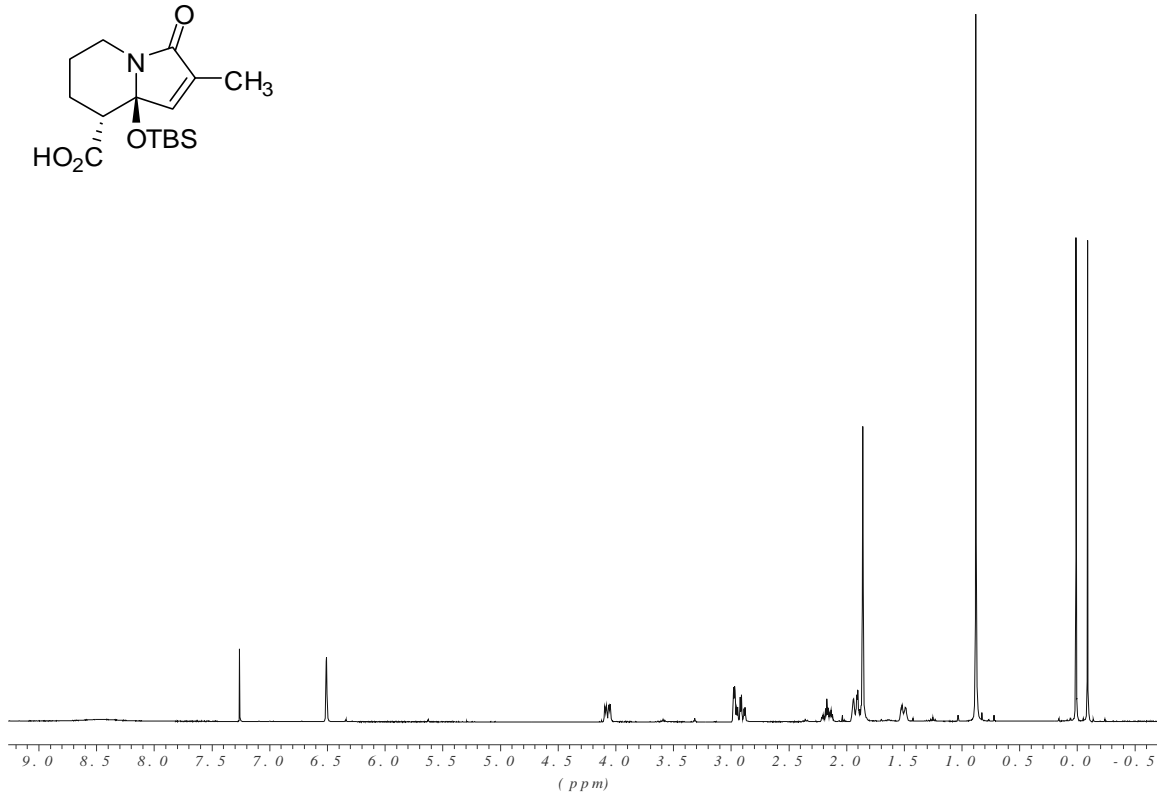
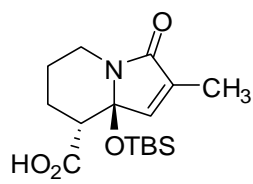
3.6130
4.2384



7b



7c



— 175.4034

— 167.9769

— 139.7500

— 136.8901

— 86.4975

— 48.5607

— 35.7103

— 25.7425

— 22.0027

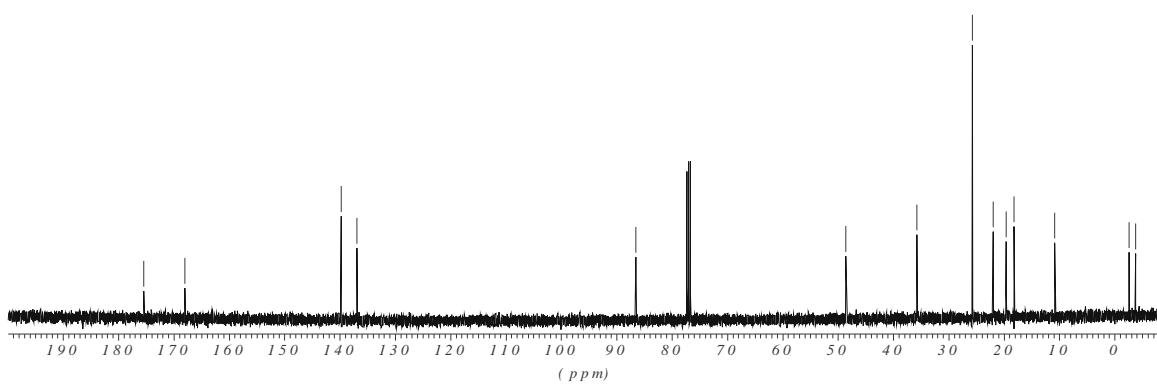
— 19.6435

— 18.1795

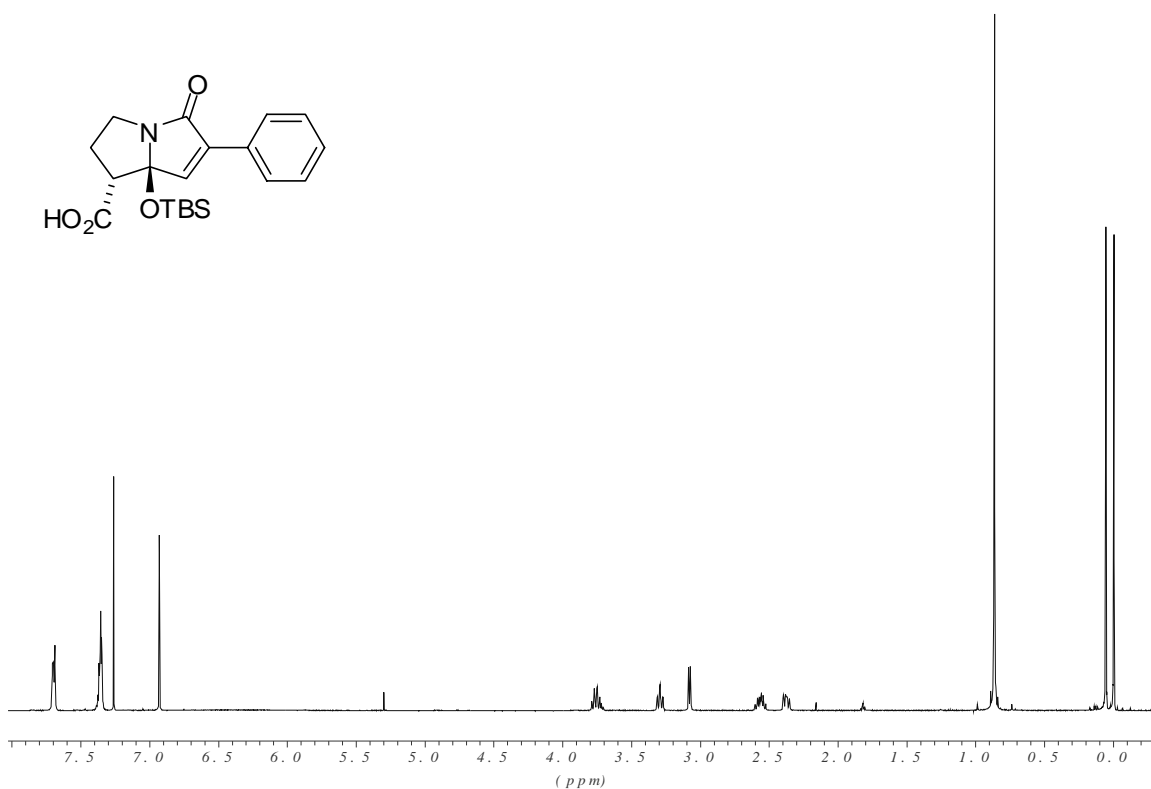
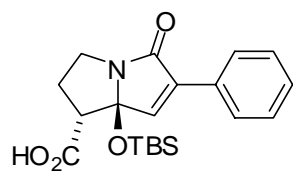
— 10.8364

— 2.5829

— 3.7739



7d



— 175.4669

— 172.3628

— 139.1414

— 137.6847

— 130.3861

— 130.3708

— 129.2192

— 128.5100

— 127.5566

— 97.8892

— 52.6407

— 42.1084

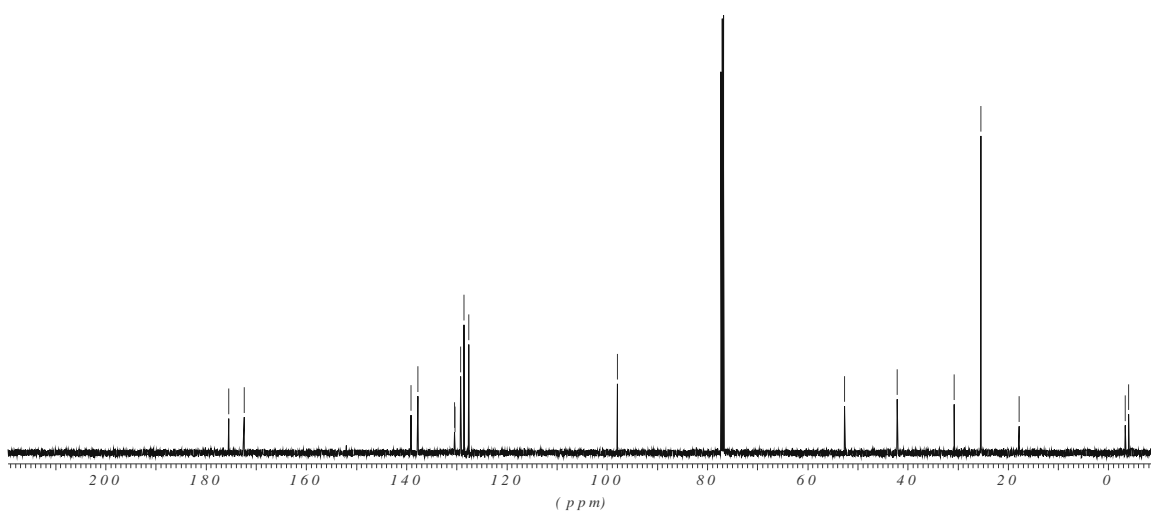
— 30.7447

— 25.4519

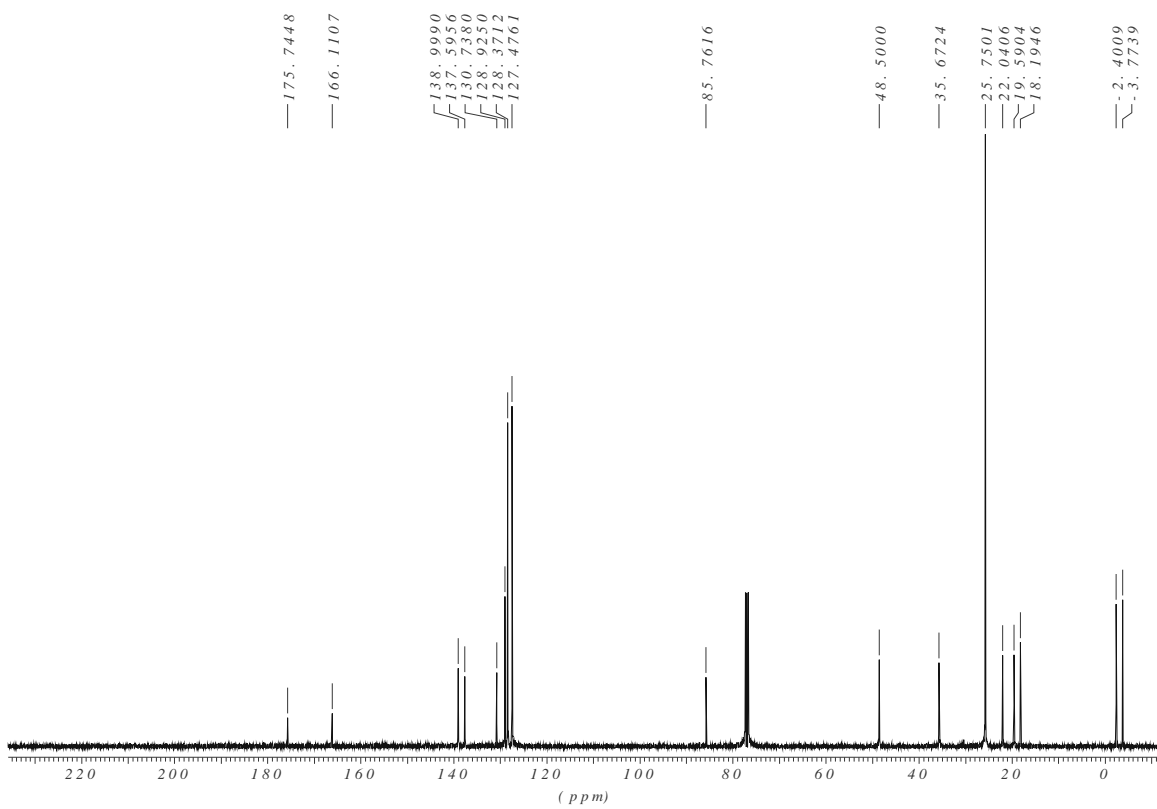
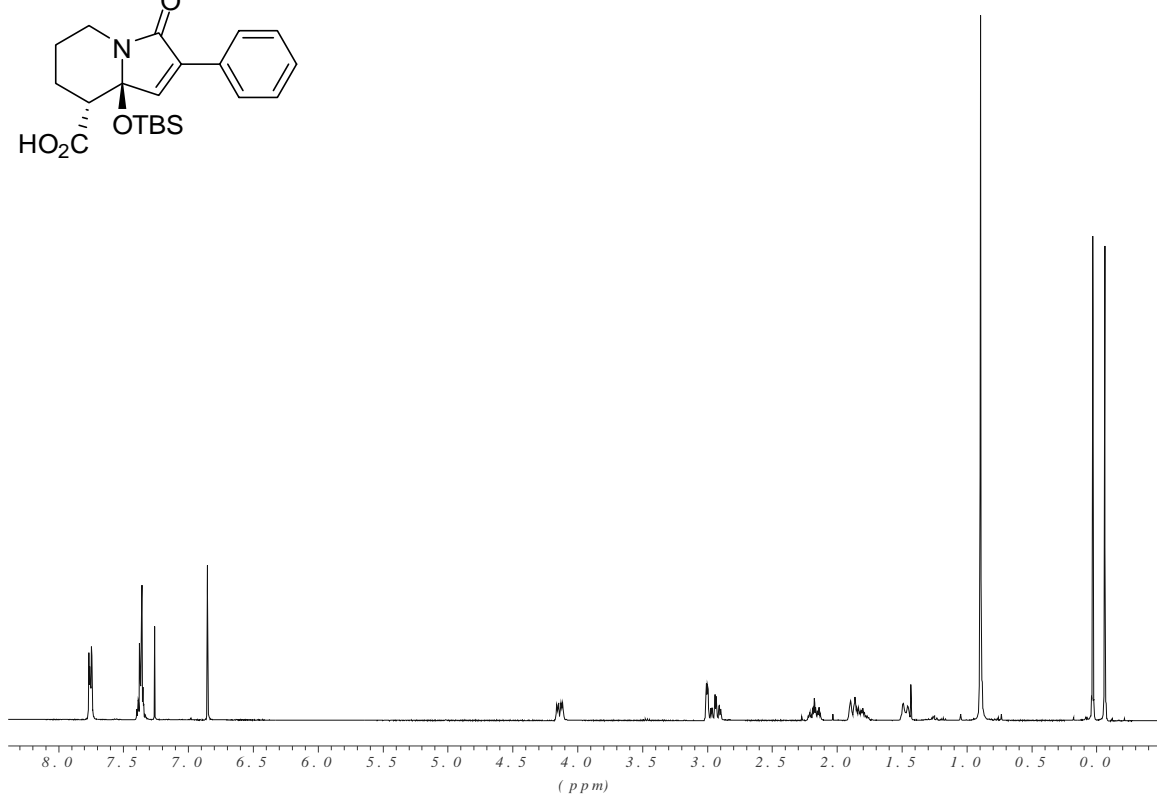
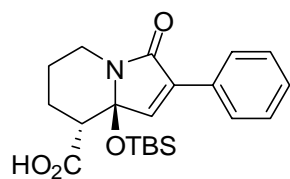
— 17.8101

— 3.3308

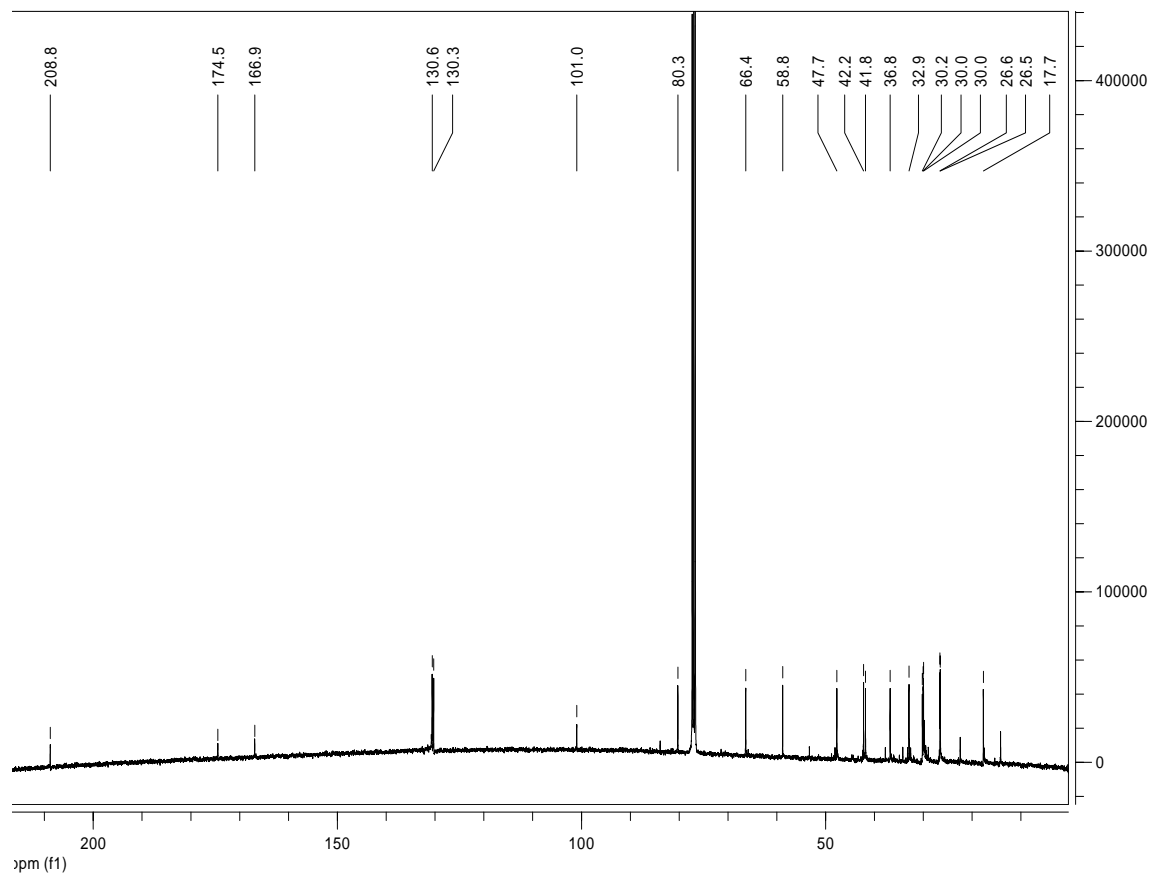
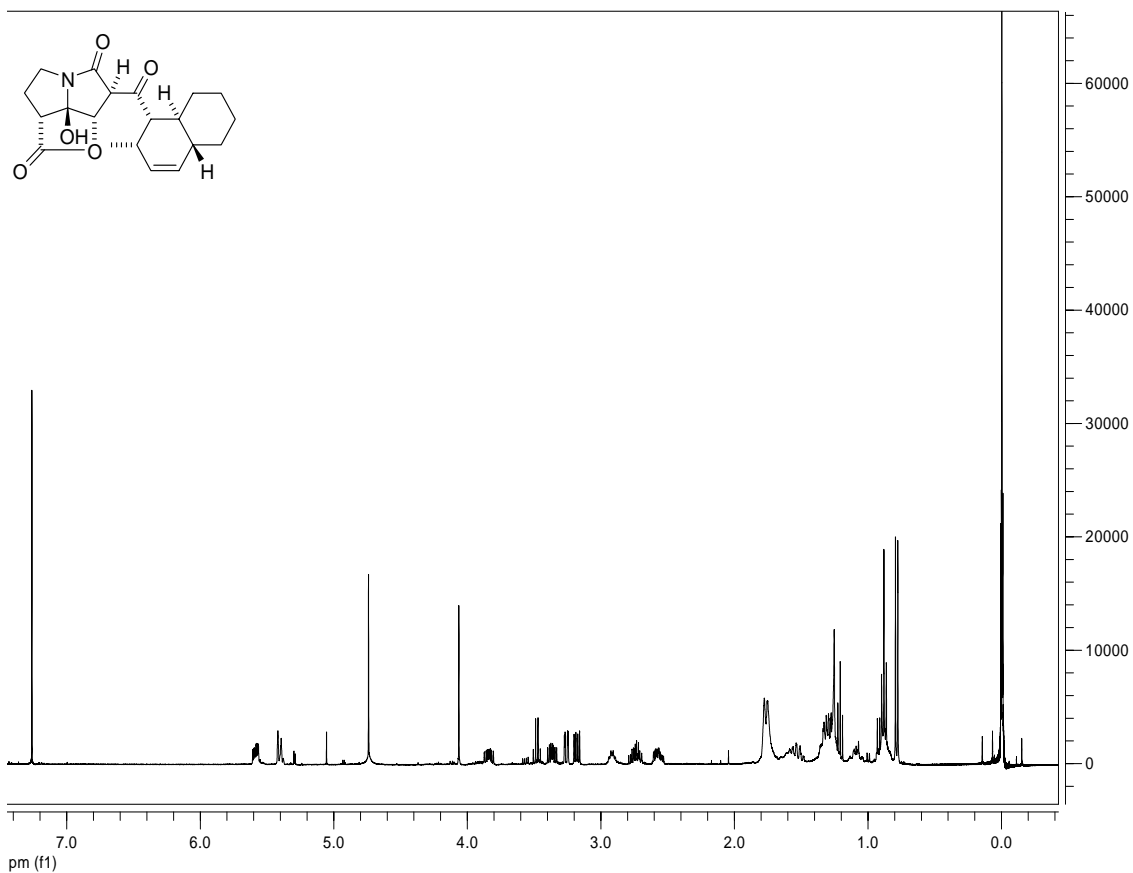
— 4.0783



7f



UCS1025A



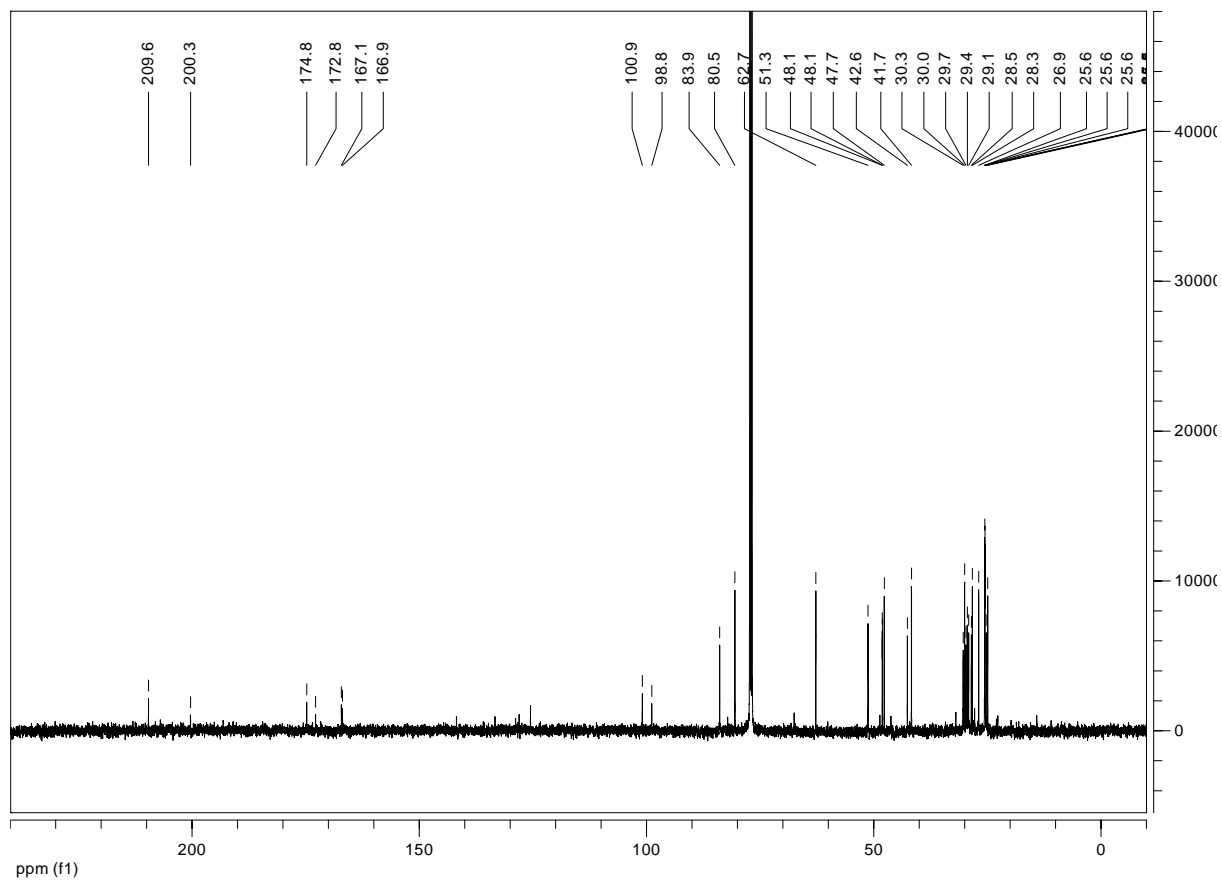
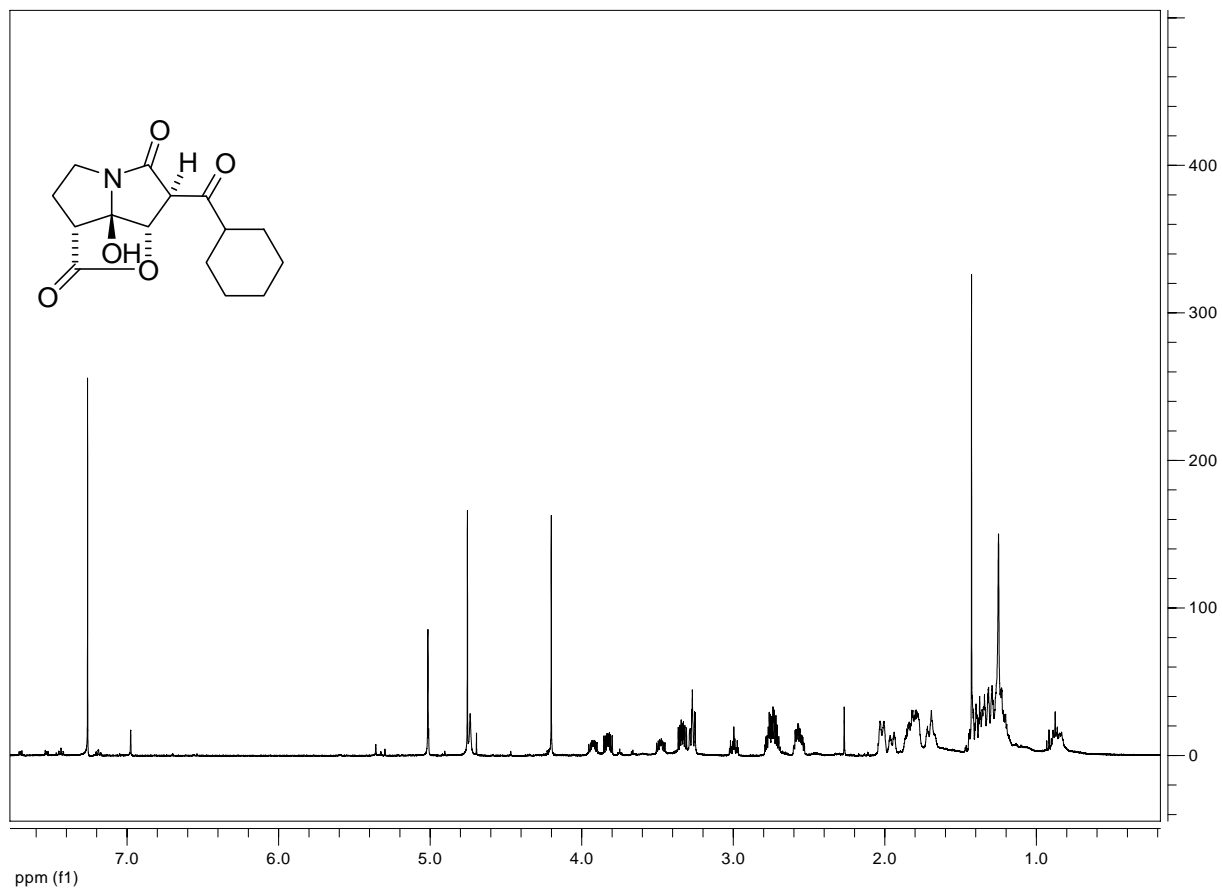
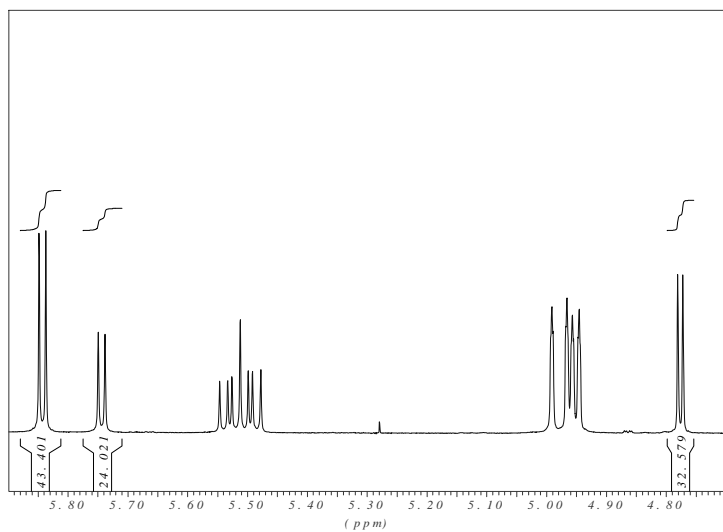
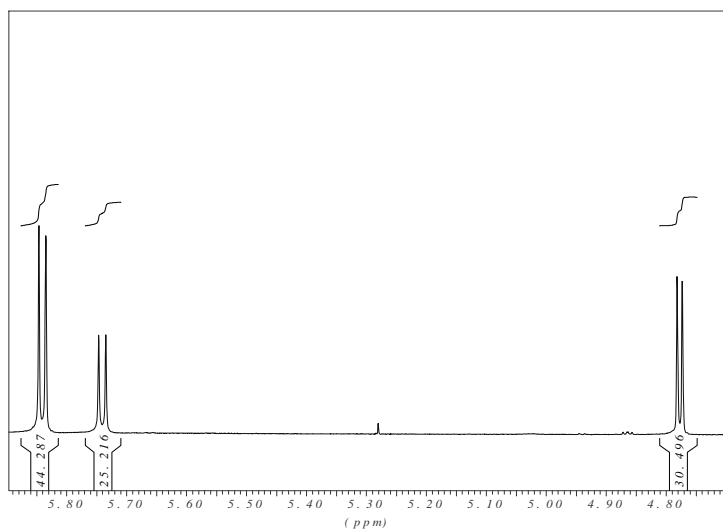


Table 1: Catalyst screening.

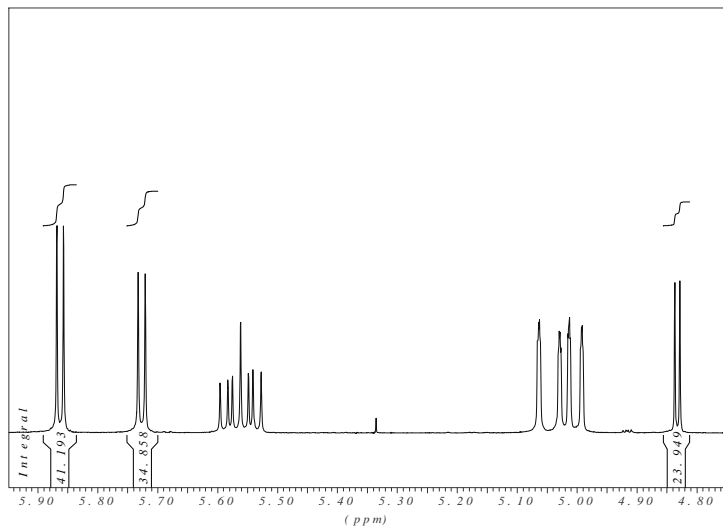
Entry 1 (Quinine, CDCl₃, 26 h, 25 °C)



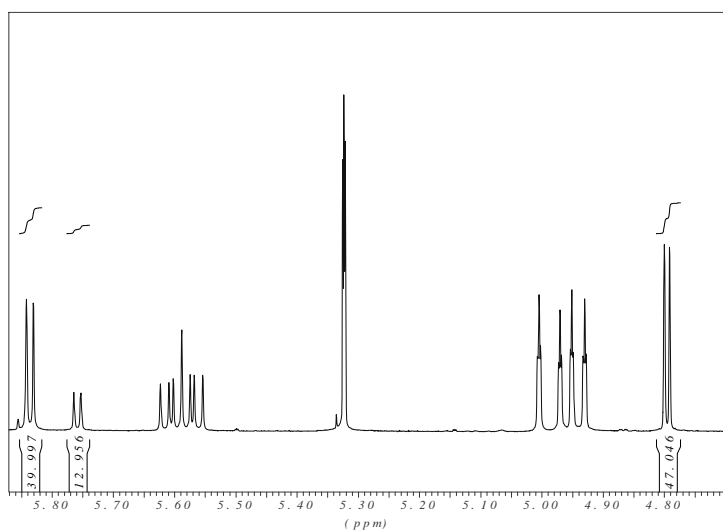
Entry 2 (Hydroquinine, CDCl₃, 26 h, 25 °C)



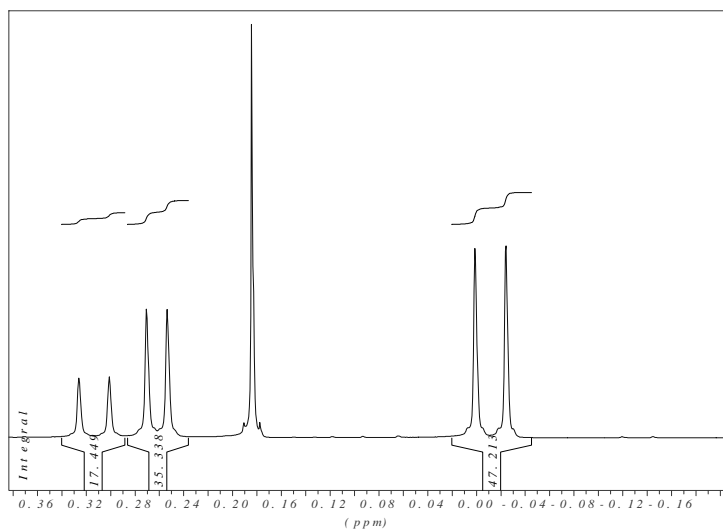
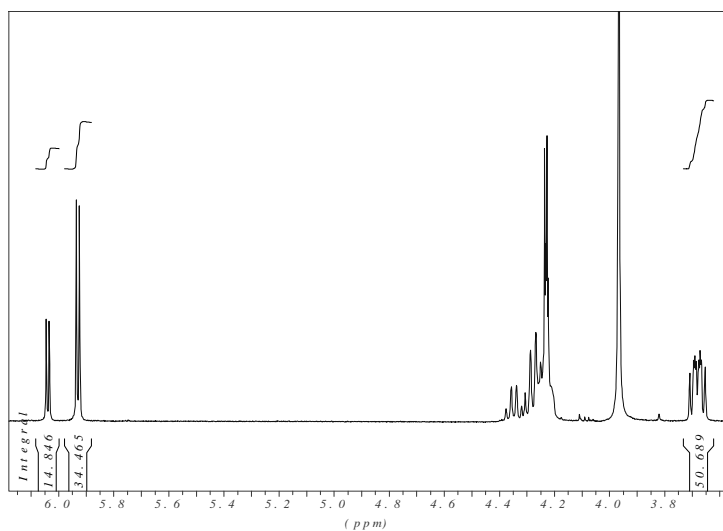
Entry 3 (Cinchonidin, CDCl₃, 26 h, 25 °C)



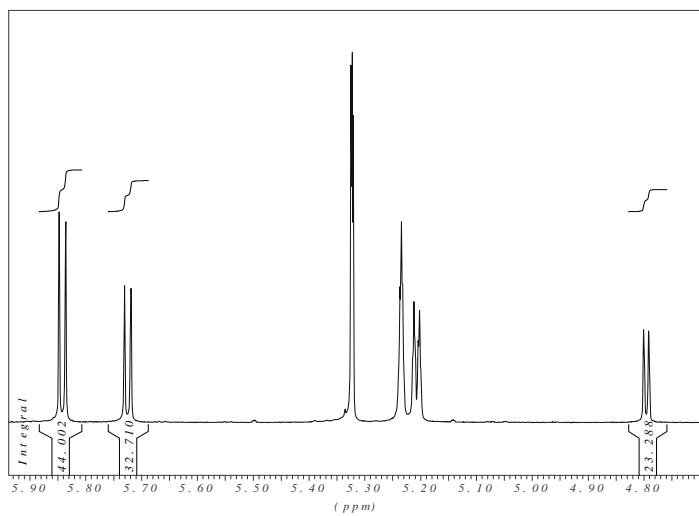
Entry 4 (Quinine, CD₂Cl₂, 26 h, 25 °C)



Entry 5 (Hydroquinine, C₆D₆, 26 h, 25 °C)



Entry 6 (Quinidine, CH₂Cl₂, 26 h, 25 °C)



Entry 7 (Cinchonidine, CH₂Cl₂, 26 h, 25 °C)

