



Supporting Information

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**Enantioselective Synthesis of α -Aryl Alkylamines by Rh(I)-
Catalyzed Additions of Arylboronic Acids to Aliphatic
Imines.**

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General Methods. The arylboronic acids were purchased from Aldrich and were recrystallized from water before use. 3-Acetylphenylboronic acid was purchased from Frontier Scientific, Inc. and was recrystallized from water before use. 3-Phenylpropionaldehyde was purchased from Acros and distilled prior to use. *N*-Diphenylphosphinoyl imines and *N*-tosyl imines were prepared from α -amido sulfone precursors immediately prior to use, according to literature procedures.¹ μ -Dichlorotetraethylene dirhodium(I) was purchased from Strem. Acetylacetonatobis(cyclooctene)rhodium was synthesized from rhodium trichloride (purchased from Strem) according to the literature procedures.² All commercially available chiral ligands were purchased from Strem and use without purification. Ligand **2** was synthesized according to the literature procedures.³ Toluene and 1,4-dioxane were passed through columns of activated alumina under nitrogen pressure and stored over MS 3Å in a nitrogen filled glove box. Triethylamine (NEt₃) was distilled over calcium hydride under an atmosphere of nitrogen immediately prior to use. K₃PO₄ was purchased from Aldrich and dried prior use. PS-DEAM resin (2.72 mmol/g) was purchased from Argonaut Technologies, Inc. Arylboronic acid additions reactions were performed in Kimble 5 mL microvials (Kimble product number 60700-5) equipped with PTFE stir-vanes (Kimble product number 749060-003) and capped with mini-inert seals (Kimble product number 749110-0022) and blue nylon caps (Kimble product number 410119-2015). The reaction solutions in vials were heated in an aluminum heating block (UC machine shop) and the temperature was maintained by an IKA stirrer/hotplate (RCT basic model) with a thermisor controller (ETS-D4).

Flash column chromatography was carried out either with Merck 60 230-240 mesh silica gel, or using a Biotage SP Flash Purification System (Biotage No. SP1-B1A) with Flash+ cartridges (Biotage No. FPK0-1107-16046). Enantiomeric excess determinations were performed on an Agilent 1100 series LC equipped with a Daicel AD or OD column and a multiwavelength detector. ^1H and $^{13}\text{C}\{^1\text{H}\}$ NMR chemical shifts are reported in ppm relative to either the residual solvent peak (^1H , ^{13}C) or TMS (^1H) as an internal standard. IR spectra were recorded as thin films on a Nicolet Avatar 360 FTIR spectrometer equipped with an attenuated total reflectance accessory or as KBr pellets on a Nicolet MAGNA-IR 850 spectrometer, and only partial data are listed. Melting points were determined on a Mel-Temp apparatus and are reported uncorrected. Mass spectrometry (HRMS) was carried out by the University of California at Berkeley Mass Spectrometry Facility.

Typical procedure for optimization screens.

Reactions were set up in a nitrogen filled glove box.

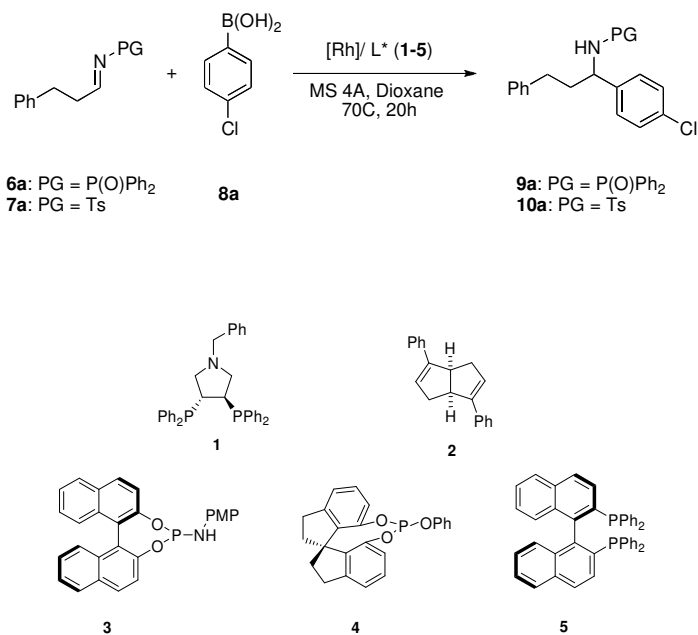
Procedure for entry 1. Imine **6a** (41.8 mg, 0.125 mmol, 1 equiv) was weighed into a vial with a stir-vane. The ligand **1** (3.6 mg, 0.0069 mmol, 0.055 equiv) was added as a solution in 0.25 mL of dioxane followed by a dioxane solution (0.25 mL) of acetylacetonatobis(cyclooctene)rhodium (2.6 mg, 0.0062 mmol, 0.05 equiv). The solution was stirred until it was homogeneous (1 min), and powdered, activated 4Å MS (200 mg) were added. Freshly distilled Et_3N (17 μL , 0.125 mmol, 1 equiv) was added. The vial was capped and heated to 70 °C in a heating block and a dioxane (0.5 mL) solution of **8a** (39.1 mg, 0.250 mmol, 2 equiv) was then added over 10 h. After 20 h, the reaction mixture was allowed to cool to rt. The mixture was extracted with EtOAc (2 x 5 mL) and

the combined organic layers were dried over Na₂SO₄. The crude material was purified by chromatography (10:1-0:1, Hexane/ EtOAc).

Procedure for entries 2-7 and 12-38, Table 1. To a vial containing a stir-vane, 4-chlorophenylboronic acid (39.1 mg, 0.250 mmol, 2 equiv) and powered 4Å molecular sieves (200 mg) was added a mixture of rhodium and the corresponding ligand (**1-5**) in 0.75 mL. The vial was then sealed using a mini-inert seal and a blue nylon cap, and the vial was removed from the glove box. The vial was then placed in a heating block with stirring and heated at 55-70 °C (aluminum block temperature) for 30-90 min. The reaction mixture was allowed to cool to rt and the vial was transferred to the glove box. To the vial was added the *N*-phosphinoyl or *N*-tosyl imine (0.125 mmol, 1 equiv) in 0.25 mL of dioxane. For entries 14-17 and 29-30, NEt₃ (0.25 mmol, 2 equiv) and imine were sequentially added. The vial was then heated at 55 °C (entries 13-16, 18, 20, 28 and 29) or 70 °C for 20 h. The reaction mixtures were allowed to cool to rt, filtered through a short pad of Celite and washed with CH₂Cl₂. The combined organic layers were washed with brine (1 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated. The conversion to product was determined by integration relative to 2,6-dimethoxytoluene, which was used as an internal standard. The crude residue was purified by silica gel chromatography or using Biotage flash purification, eluting with EtOAc/hexanes. For the reactions of *N*-phosphinoyl imine **6a**, enantiomeric excesses were determined by chiral HPLC analysis (Chiralpak OD-H, hexanes/EtOH 97/3, 0.9 mL min⁻¹): t_R = 18.8 min (major), t_R = 20.9 min (minor). For the reactions of *N*-tosyl imine **7a**, enantiomeric excesses were determined by chiral HPLC analysis (Chiralpak OD-H, hexanes/*i*-PrOH 80/20, 0.5 mL min⁻¹): t_R = 17.5 min (major), t_R = 32.6 min (minor).

Procedure for entries 7-11 and 39, Table 1. A solution of rhodium precatalyst and ligand **1** in dioxane (0.5 mL) was stirred for 90 min at 70 °C. Imine **6a** or **7a** (0.125 mmol) in dioxane (0.5 mL), chlorophenylboronic acid (39.1 mg, 0.250 mmol, 2 equiv), powdered 4Å molecular sieves (200 mg) and K₃PO₄ were successively added and the resulting mixture was stirred for 20 h at 70 °C. The reaction was quenched with water and extracted with CH₂Cl₂. The combined organic layers were washed with brine (1 mL). The combined organic layers were washed with brine (1 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated. The conversion to product was determined by integration relative to 2,6-dimethoxytoluene, which was used as an internal standard.

Table 1. Full results of catalyst optimization results.

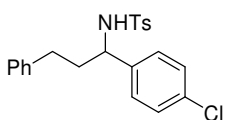


entry ^a	Imine	mol % [Rh]	mol% chiral ligand	Additive	Product	Yield (%) ^b	ee (%) ^c
1	6a	5 mol% [Rh(acac)(coe) ₂]	5.5 mol% 1	NEt ₃ (1 equiv)	9a	17	nd
2	6a	10 mol% [Rh(acac)(coe) ₂]	11 mol% 1	--	9a	74	59
3	6a	10 mol% [Rh(acac)(coe) ₂]	13 mol% 1	--	9a	70	76
4	6a	10 mol% [Rh(acac)(coe) ₂]	14 mol% 1	--	9a	71	86
5	6a	10 mol% [Rh(acac)(coe) ₂]	15 mol% 1	--	9a	51	86
6	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	--	9a	65	84
7 ^d	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	--	9a	70	90
8 ^e	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	--	9a	75	90
9 ^e	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	K ₃ PO ₄ (0.5 equiv)	9a	79	94
10 ^e	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	K ₃ PO ₄ (1 equiv)	9a	34	--
11 ^e	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 1	K ₃ PO ₄ (0.2 equiv)	9a	89	90
12 ^d	6a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 1	--	9a	53	81
13 ^f	6a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	5.5 mol% 1	--	9a	44	91
14 ^f	6a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	5.5 mol% 1	NEt ₃ (2 equiv)	9a	46	91
15 ^f	6a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	5.5 mol% 2	NEt ₃ (2 equiv)	9a	75	-80
16 ^f	6a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	7 mol% 2	NEt ₃ (2 equiv)	9a	70	-78
17 ^g	6a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	7 mol% 2	NEt ₃ (2 equiv)	9a	69	-60
18 ^f	6a	5 mol% [Rh(acac)(coe) ₂]	5.5 mol% 2	--	9a	37	-71
19 ^g	6a	5 mol% [Rh(acac)(coe) ₂]	5.5 mol% 2	--	9a	41	-56
20 ^f	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 2	--	9a	41	-74
21 ^g	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 2	--	9a	45	-60
22	6a	3 mol% [Rh(acac)(coe) ₂]	7.5 mol% 3	--	9a	46	6
23	6a	5 mol% [Rh(acac)(coe) ₂]	14 mol% 3	--	9a	40	14
24	6a	3 mol% [Rh(acac)(coe) ₂]	6 mol% 4	--	9a	36	7
25	6a	5 mol% [Rh(acac)(coe) ₂]	15 mol% 4	--	9a	35	10
26	6a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 5	--	9a	41	70
27	6a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 5	--	9a	36	74
28 ^f	7a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	5.5 mol% 2	NEt ₃ (2 equiv)	10a	79	-69
29 ^f	7a	5 mol% [RhCl(C ₂ H ₄) ₂] ₂	7 mol% 2	NEt ₃ (2 equiv)	10a	75	-67
30 ^g	7a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 2	--	10a	60	-63
31 ^g	7a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 2	--	10a	55	-62
32 ^g	7a	3 mol% [Rh(acac)(coe) ₂]	7.5 mol% 3	--	10a	85	19
33 ^g	7a	5 mol% [Rh(acac)(coe) ₂]	14 mol% 3	--	10a	80	23
34 ^g	7a	3 mol% [Rh(acac)(coe) ₂]	6 mol% 4	--	10a	40	12
35 ^g	7a	5 mol% [Rh(acac)(coe) ₂]	14 mol% 4	--	10a	37	15
36 ^g	7a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 5	--	10a	75	63
37 ^g	7a	5 mol% [Rh(acac)(coe) ₂]	7 mol% 5	--	10a	69	68
38 ^g	7a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 1	--	10a	78	93
39 ^e	7a	3 mol% [Rh(acac)(coe) ₂]	3.3 mol% 1	K ₃ PO ₄ (0.2 equiv)	10a	99	95

^aThe reaction was carried out with 1 equiv of imine and 2 equiv of boronic acid in the presence of the rhodium catalyst and the chiral ligand indicated in dioxane (0.12 M) at 70 °C. ^bYields were determined by ¹H-NMR, using 2,6-dimethoxytoluene as an internal standard in CDCl₃. ^cEnantiomeric excess of the pure product was determined by chiral HPLC. ^dThe imine was added to the reaction mixture after 90 min at 70 °C. ^eThe boronic acid and imine were added to the reaction mixture after 90 min at 70 °C. ^fThe reaction was carried out at 55 °C. ^gThe reactions were running at 70 °C.

Representative procedure for the enantioselective Rh(I)-catalyzed additions of arylboronic acids to *N*-tosyl imines.

A vial containing a stir-vane, acetylacetonatobis(cyclooctene)rhodium (1.5 mg, 0.00375 mmol, 0.03 equiv) and (*R*)-Deguphos (2.2 mg, 0.00425 mmol, 0.033 equiv) in 0.50 mL of dioxane is sealed using a mini-inert seal and a blue nylon cap, and removed from the glove box. The vial was then placed in a heating block with stirring and heated at 70 °C (aluminium block temperature) for 90 min. The reaction mixture was allowed to cool to rt and the vial was transferred to the glove box. To the vial was added the arylboronic acid **8** (0.250 mmol, 2 equiv) a mixture of *N*-tosyl imine (0.125 mmol, 1 equiv) in 0.50 mL of dioxane, K₃PO₄ (5.3 mg, 0.025, 0.2 equiv) and powered 4Å molecular sieves (200 mg), after transferring back to the heating block, the vial was then heated at 70 °C for 20 h. The reaction mixture was allowed to cool at rt. The reaction was quenched with water and extracted with CH₂Cl₂. The combined organic layers were washed with brine (1 mL). The organic layer was dried over Na₂SO₄, filtered, and concentrated. The crude material was taken up in THF (5 mL) and PS-DEAM resin (200 mg) and placed on a shaker for 10 h. The THF solution was filtered to remove the resin, and the resin was washed with ethyl acetate (4 x 10 mL). The filtrates were combined and concentrated. The conversion to product was determined by integration relative to 2,6-dimethoxytoluene, which was used as an internal standard. The crude residue was purified by flash column chromatography on a Biotage Flash+ cartridge with a gradient of EtOAc in hexanes to provide the tosylamides **10**.

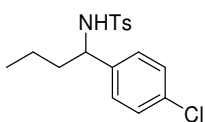


***N*-(1-(4-chlorophenyl)-1-phenylpropyl)-tosylamide (10a).** The

general procedure was followed and the product was isolated by flash

column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10a** as a white solid (47 mg, 94% yield). $R_f = 0.47$ (Hexane: EtOAc, 3:1). mp 138-140 °C. 95% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 18.5$ min (major), $t_R = 33.4$ min (minor).

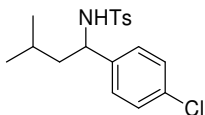
¹H NMR (400 MHz, CDCl₃): δ 7.55 (d, $J = 8.0$ Hz, 2H), 7.30-7.11 (m, 7H), 7.06 (dd, $J = 6.8, 1.6$ Hz, 2H), 6.99 (dd, $J = 8.4, 2.0$ Hz, 2H), 5.78-5.75 (m, 1H), 4.30 (q, $J = 7.6$ Hz, 1H), 2.58-2.51 (m, 2H), 2.42 (s, 3H), 2.13-2.10 (m, 1H), 2.09-1.97 (m, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.3, 140.6, 139.1, 137.4, 133.2, 129.4, 128.6, 128.5, 128.4, 128.1, 127.1, 126.1, 57.3, 38.8, 32.0, 21.5. HRMS (ES+) m/z : Calcd for C₂₂H₂₂ClNO₂S [M+Na]⁺ 422.0957; found 422.0965. IR (KBr, cm⁻¹) 3225, 1597, 1431, 1262, 810, 683.



***N*-(1-(4-chlorophenyl)butyl)-tosylamide (10b).** The general

procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10b** as a white solid (37.5 mg, 89% yield). $R_f = 0.44$ (Hexane: EtOAc, 3:1). mp 125-127 °C. 93% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 11.9$ min (major), $t_R = 14.6$ min (minor).

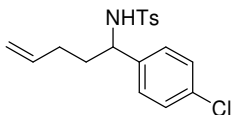
¹H NMR (400 MHz, CDCl₃): δ 7.54 (d, $J = 8.4$ Hz, 2H), 7.15 (d, $J = 8.4$ Hz, 2H), 7.11 (d, $J = 8.4$ Hz, 2H), 6.97 (d, $J = 8.4$ Hz, 2H), 5.31 (d, $J = 7.6$ Hz, 1H), 4.29 (q, $J = 7.2$ Hz, 1H), 2.41 (s, 3H), 1.77-1.60 (m, 2H), 1.33-1.13 (m, 2H), 0.87 (t, $J = 7.2$ Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.2, 139.5, 137.5, 132.9, 129.3, 128.4, 127.9, 126.9, 57.4, 39.5, 21.43, 18.9, 13.5. HRMS (FAB+) m/z : Calcd for C₁₇H₂₀ClNO₂S [MH]⁺ 338.0981; found 338.0977. IR (KBr, cm⁻¹) 3257, 1430, 1317, 1166, 811.



***N*-(1-(4-chlorophenyl)-3-methylbutyl)-tosylamide (10c).** The general

procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10c** as a white solid (42 mg, 96% yield). $R_f = 0.53$ (Hexane: EtOAc, 3:1). mp 93-95 °C. 91% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 10.3$ min (major), $t_R = 18.4$ min (minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.78 (d, $J = 8.8$ Hz, 2H), 7.07 (d, $J = 8.0$ Hz, 2H), 7.05 (dt, $J = 8.4, 2.4$ Hz, 2H), 6.94 (dt, $J = 8.4, 2.8$ Hz, 2H), 5.70 (d, $J = 8.0$ Hz, 1H), 4.30 (q, $J = 7.6$ Hz, 1H), 2.43 (s, 3H), 1.64-1.51 (m, 1H), 1.50-1.41 (m, 2H), 0.82 (d, $J = 6.4$ Hz, 3H), 0.79 (d, $J = 6.0$ Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.1, 139.7, 137.5, 132.9, 129.2, 128.3, 127.9, 127.1, 57.4, 46.6, 24.5, 22.3, 22.1, 21.4. HRMS (ES+) m/z : Calcd for C₁₈H₂₂ClNO₂S [M+Na]⁺ 374.0957; found 374.0960. IR (KBr, cm⁻¹) 3260, 1595, 1398, 1087, 374, 679.

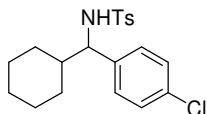


***N*-(1-(4-chlorophenyl)pent-4-enyl)-tosylamide (10d).** The general

procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 20% EtOAc in hexanes, affording **10d** as a white solid (38 mg, 87% yield). $R_f = 0.50$ (Hexane: EtOAc, 3:1). mp 99-101 °C. 98% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 9.4$ min (major), $t_R = 11.6$ min (minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.55 (d, $J = 8.4$ Hz, 2H), 7.13 (d, $J = 8.0$ Hz, 2H), 7.10 (dt, $J = 8.4, 2.8$ Hz, 2H), 6.97 (dt, $J = 8.8, 2.0$ Hz, 2H), 5.86 (d, $J = 8.0$ Hz, 1H), 5.75-5.65 (m, 1H), 4.99-4.92 (m, 2H), 4.29 (q, $J = 5.4$ Hz, 1H), 2.41 (s, 3H), 2.03-0.7 (m, 4H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.2, 139.3, 137.5, 136.9, 133.9, 129.3, 128.5,

128.1, 127.0, 115.7, 57.2, 36.4, 29.9, 21.4. LRMS (ES+) m/z : Calcd for $C_{18}H_{20}ClNO_2S$ [M+Na]⁺ 372; found 372. IR (KBr, cm^{-1}) 3265, 1442, 1317, 1287, 1166, 1089, 835.

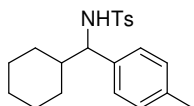


***N*-(1-(4-chlorophenyl)cyclohexylmethyl)-tosylamide (10e).** The

general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient

of 0% to 30% EtOAc in hexanes, affording **10e** as a white solid (38 mg, 80% yield). R_f = 0.52 (Hexane: EtOAc, 3:1). mp 155-157 °C. 96% ee (Chiralpak OD-H, hexanes/IPA 85/15, 0.5 mL min⁻¹, λ = 230 nm, t_R = 12.5 min (major), t_R = 19.7 min (minor).

¹H NMR (400 MHz, CDCl₃): δ 7.44 (d, J = 8.0 Hz, 2H), 7.08-7.05 (m, 4H), 6.85 (dd, J = 8.0, 2.0 Hz, 2H), 4.92 (d, J = 8.0 Hz, 1H), 4.02 (t, J = 8.0 Hz, 1H), 2.36 (s, 3H), 1.88-1.72 (m, 2H), 1.61-1.50 (m, 3H), 1.28-0.83 (m, 6H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.9, 143.0, 137.3, 129.1, 128.8, 126.9, 125.3, 124.8, 63.1, 43.4, 29.6, 29.4, 26.0, 25.7, 21.2. HRMS (ES+) m/z : Calcd for $C_{20}H_{24}ClNO_2S$ [M+Na]⁺ 400.1114; found 400.1123. IR (KBr, cm^{-1}) 3220, 1317, 1287, 685, 667.

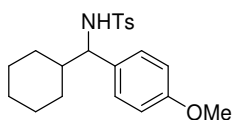


***N*-(1-(4-methylphenyl)(cyclohexyl)methyl)-tosylamide (10f).** The

general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10f** as a white solid (32 mg, 71% yield). R_f = 0.51 (Hexane: EtOAc, 3:1). mp 149-151 °C. 96% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, λ = 230 nm, t_R = 10.0 min (major), t_R = 15.3 min (minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.53 (dd, J = 8.4, 2.0 Hz, 2H), 7.06 (d, J = 8.0 Hz, 2H), 6.91 (d, J = 8.0 Hz, 2H), 6.84 (d, J = 8.0 Hz, 2H), 5.75 (d, J = 8.8 Hz, 1H), 4.02 (t, J = 8.4 Hz, 1H), 2.46 (s, 3H), 2.36 (s, 3H), 2.02 (m, 1H), 1.75 (m, 1H), 1.63-1.45 (m, 3H),

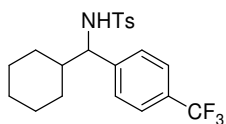
1.33 (m, 1H), 1.21-0.82 (m, 5H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 142.5, 137.7, 136.9, 136.4, 129.0, 128.6, 127.0, 126.9, 63.3, 43.6, 29.7, 29.5, 26.2, 25.9, 21.4, 21.0. HRMS (ES+) m/z : Calcd for $\text{C}_{21}\text{H}_{27}\text{NO}_2\text{S}$ $[\text{M}+\text{Na}]^+$ 380.1660; found 380.1667. IR (KBr, cm^{-1}) 3220, 1317, 1287, 685, 667.



***N*-(1-(4-methoxyphenyl) (cyclohexyl)methyl)-tosylamide (**10g**).**

The general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10g** as a white solid (34.5 mg, 74% yield). R_f = 0.39 (Hexane: EtOAc, 3:1). mp 156-158 °C. 90% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min^{-1} , λ = 230 nm, t_R = 11.4 min (major), t_R = 18.5 min (minor)).

^1H NMR (400 MHz, CDCl_3): δ 7.50 (d, J = 8.0 Hz, 2H), 7.09 (d, J = 7.6 Hz, 2H), 6.86 (dd, J = 8.8, 2.0 Hz, 2H), 6.66 (dd, J = 8.4, 2.0 Hz, 2H), 5.10 (d, J = 8.4 Hz, 1H), 4.00 (t, J = 8.0 Hz, 1H), 3.77 (s, 3H), 2.37 (s, 3H), 1.97 (m, 1H), 1.75 (m, 1H), 1.63-1.44 (m, 3H), 1.32 (d, J = 13.2 Hz, 1H), 1.20-0.79 (m, 5H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 158.7, 142.6, 137.8, 132.1, 129.1, 128.1, 127.1, 113.5, 63.0, 55.2, 43.8, 29.7, 29.6, 26.2, 25.9, 21.4. LRMS (ES+) m/z : Calcd for $\text{C}_{21}\text{H}_{27}\text{NO}_3\text{S}$ $[\text{M}+\text{Na}]^+$ 396; found 396. IR (KBr, cm^{-1}) 3261, 1496, 1317, 1270, 1089, 705.



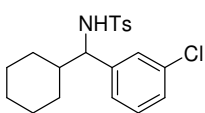
***N*-(1-(4-trifluoromethylphenyl) (cyclohexyl)methyl)-tosylamide**

(10h**).** The general procedure was followed and the product was

isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10h** as a white solid (46 mg, 89% yield). R_f = 0.50 (Hexane: EtOAc, 3:1). mp 152-154 °C. 91% ee (Chiralpak

OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, λ = 230 nm, t_R = 9.7 min (major), t_R = 19.7 min (minor)).

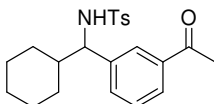
¹H NMR (400 MHz, CDCl₃): δ 7.48 (d, J = 8.0 Hz, 2H), 7.31 (d, J = 8.0 Hz, 2H), 7.07 (d, J = 8.0 Hz, 2H), 7.02 (d, J = 8.0 Hz, 2H), 6.10 (d, J = 8.8 Hz, 1H), 4.14 (t, J = 8.4 Hz, 1H), 2.33 (s, 3H), 2.01 (m, 1H), 1.74 (m, 1H), 1.61-1.56 (m, 3H), 1.27 (d, J = 13.6 Hz, 1H), 1.18-0.81 (m, 5H). For the ¹³C{¹H} NMR spectrum see page S28. HRMS (ES+) m/z : Calcd for C₂₁H₂₄NO₂F₃S [M+Na]⁺ 434.1378; found 434.1389. IR (KBr, cm⁻¹) 3278, 1443, 1321, 1164, 808.



***N*-(1-(3-chlorophenyl)(cyclohexyl)methyl)-tosylamide (10i).** The

general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10i** as a white solid (35.5 mg, 75% yield). R_f = 0.48 (Hexane: EtOAc, 3:1). mp 111-113 °C. 90% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, λ = 230 nm, t_R = 14.8 min (major), t_R = 18.1 min (minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.50 (d, J = 8.4 Hz, 2H), 7.11-7.08 (m, 4H), 6.92-6.90 (m, 1H), 6.80 (s, 1H), 5.39 (d, J = 8.4 Hz, 1H), 4.02 (t, J = 8.0 Hz, 1H), 2.37 (s, 3H), 1.97 (m, 1H), 1.76 (m, 1H), 1.68-1.55 (m, 3H), 1.30 (m, 1H), 1.17-0.87 (m, 5H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.1, 141.9, 137.3, 133.9, 129.4, 129.2, 127.3, 127.0, 126.9, 125.2, 63.0, 43.4, 29.7, 29.3, 26.0, 25.8, 21.4. HRMS (ES+) m/z : Calcd for C₂₀H₂₄ClNO₂S [M+Na]⁺ 400.1114; found 400.1120. IR (KBr, cm⁻¹) 3271, 1442, 1287, 722, 667.

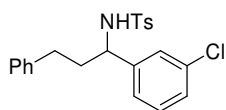


***N*-(1-(3-acetylphenyl)(cyclohexyl)methyl)-tosylamide (10j).** The

general procedure was followed and the product was isolated by flash

column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 40% EtOAc in hexanes, affording **10j** as a white solid (39 mg, 81% yield). $R_f = 0.27$ (Hexane: EtOAc, 3:1). mp 137-139 °C. 89% ee (Chiralpak OD-H, hexanes/IPA 80/20, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 15.4$ min (major), $t_R = 25.5$ min (minor)).

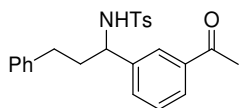
¹H NMR (400 MHz, CDCl₃): δ 7.75-7.71 (m, 1H), 7.49 (m, 3H), 7.31-7.25 (m, 2H), 7.02 (d, $J = 8.4$ Hz, 2H), 5.76 (d, $J = 8.4$ Hz, 1H), 4.14 (t, $J = 8.0$ Hz, 1H), 2.52 (s, 3H), 2.32 (s, 3H), 1.97 (m, 1H), 1.74 (m, 1H), 1.62-1.50 (m, 3H), 1.28 (m, 1H), 1.18-0.89 (m, 5H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 197.9, 142.9, 140.6, 137.6, 136.8, 132.0, 129.2, 128.4, 127.0, 126.97, 126.92, 63.2, 43.6, 29.7, 29.3, 26.6, 26.1, 25.8, 21.4. HRMS (ES+) m/z : Calcd for C₂₂H₂₇ClNO₃S [M+Na]⁺ 408.1609; found 408.1618. IR (KBr, cm⁻¹) 3270, 1690, 1440, 1316, 1168, 813, 685.



***N*-(1-(3-chlorophenyl)-1-phenylpropyl)-tosylamide (10k).** The

general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 0% to 30% EtOAc in hexanes, affording **10k** as a colorless oil (37 mg, 74% yield). $R_f = 0.41$ (Hexane: EtOAc, 3:1). 93% ee (Chiralpak OD-H, hexanes/IPA 85/25, 0.5 mL min⁻¹, $\lambda = 230$ nm, $t_R = 19.1$ min (major), $t_R = 34.3$ min (minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.51 (d, $J = 8.4$ Hz, 2H), 7.27-7.04 (m, 9H), 6.95 (t, $J = 4.0$ Hz, 1H), 6.86 (s, 1H), 5.43 (d, $J = 8.0$ Hz, 1H), 4.26 (q, $J = 7.6$ Hz, 1H), 2.59-2.43 (m, 2H), 2.37 (s, 3H), 2.10-1.67 (m, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 143.3, 142.5, 140.5, 137.2, 134.2, 129.8, 129.3, 128.4, 128.3, 127.5, 127.0, 126.9, 126.1, 124.7, 57.3, 38.8, 32.0, 21.4. HRMS (ES+) m/z : Calcd for C₂₂H₂₂ClNO₂S [M+Na]⁺ 422.0957; found 422.0968. IR (KBr, cm⁻¹) 3235, 1435, 1136, 1090, 805, 710.



***N*-(1-(3-acetylphenyl)-1-phenylpropyl)-tosylamide (10l).** The

general procedure was followed and the product was isolated by

flash column chromatography on a Biotage Flash+ cartridge with a

gradient of 0% to 40% EtOAc in hexanes, affording **10l** as a white solid (41mg, 80%

yield). R_f = 0.22 (Hexane: EtOAc, 3:1). mp 108-110 °C. 90% ee (Chiralpak OD-H,

hexanes/IPA 75/25, 0.5 mL min⁻¹, λ = 230 nm, t_R = 16.8 min (major), t_R = 30.8 min

(minor)).

¹H NMR (400 MHz, CDCl₃): δ 7.72 (dt, J = 8.8, 1.6 Hz, 1H), 7.58 (s, 1H), 7.51 (d, J = 8.0 Hz, 2H), 7.31-7.15 (m, 5H), 7.06-7.30 (m, 4H), 5.97 (d, J = 8.0 Hz, 1H), 4.37 (q, J = 7.6 Hz, 1H), 2.63-2.52 (m, 2H), 2.50 (s, 3H), 2.31 (s, 3H), 2.17-1.94 (m, 2H). ¹³C{¹H}

NMR (100 MHz, CDCl₃): δ 197.9, 143.1, 141.5, 140.6, 137.6, 137.1, 131.6, 129.4, 128.8,

128.5, 128.4, 127.4, 127.0, 126.5, 126.1, 57.7, 39.0, 32.1, 26.6, 21.4. HRMS (ES+) m/z :

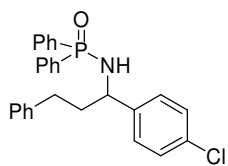
Calcd for C₂₄H₂₅NO₃S [M+Na]⁺ 430.1453; found 430.1460. IR (KBr, cm⁻¹) 3255, 1685,

1431, 1160, 809, 70870.

Representative procedure for the enantioselective Rh(I)-catalyzed additions of arylboronic acids to *N*-phosphinoyl imines.

A vial containing a stir-vane, acetylacetonatobis(cyclooctene)rhodium (2.6 mg, 0.0062 mmol, 0.05 equiv) and (*R,R*)-Deguphos (4.6 mg, 0.00875 mmol, 0.07 equiv) in 0.50 mL of dioxane is sealed using a mini-inert seal and a blue nylon cap, and removed from the glove box. The vial was then placed in a heating block with stirring and heated at 70 °C (aluminium block temperature) for 90 min. The reaction mixture was allowed to cool to rt and the vial was transferred to the glove box. To the vial was added the arylboronic acid **8** (0.250 mmol, 2 equiv) a mixture of *N*-phosphinoyl imine (0.125 mmol, 1 equiv) in 0.50

mL of dioxane, K_3PO_4 (5.3 mg, 0.025, 0.2 equiv) and powered 4Å molecular sieves (200 mg), after transferring back to the heating block, the vial was then heated at 70 °C for 20 h. The reaction mixture was allowed to cool at rt. The reaction was quenched with water and extracted with CH_2Cl_2 . The combined organic layers were washed with brine (1 mL). The organic layer was dried over Na_2SO_4 , filtered, and concentrated. The crude material was taken up in THF (5 mL) and PS-DEAM resin (200 mg) and placed on a shaker for 10 h. The THF solution was filtered to remove the resin, and the resin was washed with ethyl acetate (4 x 10 mL). The filtrates were combined and concentrated. The conversion to product was determined by integration relative to 2,6-dimethoxytoluene, which was used as an internal standard. The crude residue was purified by flash column chromatography on a Biotage Flash+ cartridge with a gradient of EtOAc in hexanes to provide the phosphoramides **9**.



***N*-(1-(4-chlorophenyl)-1-phenylpropyl)-diphenylphosphoramidate**

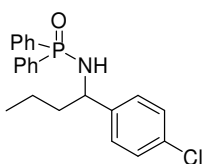
(9a). Procedure for the reaction performed on 5 mmol scale (entry 7, Table 1). A solution of $Rh(acac)(coe)_2$ (105.6 mg, 0.25 mmol, 0.05

equiv) and (*R*)-Deguphos (185.4 mg, 0.35 mmol, 0.07 equiv, mmol in dioxane (20 mL) was prepared in a glovebox and then was stirred for 90 min at 70 °C. Imine **6a** (1.67 g, 5 mmol, 1 equiv) in dioxane (20 mL), 4-chlorophenylboronic acid (1.56 g, 10 mmol, 2 equiv), powered 4Å molecular sieves (8 g) and K_3PO_4 (212 mg, 1 mmol, 0.2 equiv) were successively added and the resulting mixture was stirred for 20 h at 70 °C. The reaction mixture was diluted with water and extracted with CH_2Cl_2 . The crude material was taken up in THF (200 mL) and PS-DEAM resin (13.8 g, 1.58 mmol/g) and shaken for 16 h. The

THF solution was filtered and the resin was washed with ethyl acetate (5x). The product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 10% to 95% EtOAc in hexanes, affording **9a** as a white solid (2.05 g, 92% yield). $R_f = 0.62$ (Hexane: EtOAc, 1:10). mp 192-194 °C. 90% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 0.9 mL min⁻¹, $\lambda = 230$ nm, $t_R = 18.8$ min (major), $t_R = 20.9$ min (minor)).

The resulting resin was treated with a 90:5:5 THF/water/acetic acid mixture (150 mL) for 12 h. The liquid phase was drained and the resin rinsed with the above cleavage mixture (1x) and THF (3x). The combined filtrates were concentrated and dried to afford 670 mg of 4-chlorophenylboronic acid (43%).

¹H NMR (400 MHz, CDCl₃): δ 7.89-7.84 (m, 2H), 7.77-7.72 (m, 2H), 7.55-7.10 (m, 15H), 4.22 (q, $J = 7.2$ Hz, 1H), 3.31 (dd, $J = 9.6, 6.0$ Hz, 1H), 2.62-2.52 (m, 2H), 2.35-2.29 (m, 1H), 2.15-2.12 (m, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 142.14, 142.10, 141.1, 133.5, 132.9, 132.5, 132.4, 131.9, 131.8, 128.7, 128.6, 128.5, 128.49, 128.41, 128.3, 128.0, 125.9, 54.9, 40.8, 32.3. ³¹P (162 MHz, CDCl₃): δ 9.6. HRMS (FAB+) m/z : Calcd for C₂₇H₂₅ClNOP [MH]⁺ 446.1441; found 446.1438. IR (KBr, cm⁻¹) 2930, 1491, 1436, 1178, 1124, 1014, 827, 750, 697.



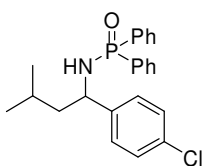
N-(1-(4-chlorophenyl)butyl)-diphenylphosphoramidate (9b). The

general procedure was followed and the product was isolated by flash

column chromatography on a Biotage Flash+ cartridge with a gradient

of 10% to 95% EtOAc in hexanes, affording **9b** as a white solid (43 mg, 89% yield). $R_f = 0.50$ (Hexane: EtOAc, 1:10). mp 155-157 °C. 86% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 0.8 mL min⁻¹, $\lambda = 230$ nm, $t_R = 14.4$ min (major), $t_R = 18.5$ min (minor)).

^1H NMR (400 MHz, CDCl_3): δ 7.88-7.85 (m, 2H), 7.77-7.72 (m, 2H), 7.53-7.14 (m, 6H), 7.10 (d, J = 8.0 Hz, 2H), 6.91 (d, J = 8.8 Hz, 2H), 4.16 (q, J = 7.6 Hz, 1H), 3.37 (dd, J = 9.2, 6.0 Hz, 1H), 1.98-1.71 (m, 2H), 1.31-1.16 (m, 2H), 0.85 (t, J = 7.2 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 142.6, 133.6, 132.7, 132.6, 132.5, 131.9, 131.8, 128.6, 128.5, 128.4, 128.3, 127.9, 55.0, 41.8, 19.4, 13.8. ^{31}P (162 MHz, CDCl_3): δ 9.9. HRMS (FAB+) m/z : Calcd for $\text{C}_{22}\text{H}_{23}\text{ClNOP}$ $[\text{MH}]^+$ 384.1284; found 384.1284. IR (KBr, cm^{-1}) 3214, 1491, 1435, 1181, 1125, 1014, 819, 745, 692.

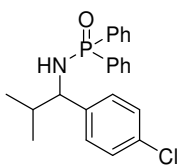


***N*-(1-(4-chlorophenyl)-3-methylbutyl)-diphenylphosphoramidate**

(9c). The general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a

gradient of 10% to 95% EtOAc in hexanes, affording **9c** as a colorless oil (40 mg, 80% yield). R_f = 0.51 (Hexane: EtOAc, 1:10). 81% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 0.9 mL min^{-1} , λ = 230 nm, t_R = 9.6 min (major), t_R = 15.4 min (minor).

^1H NMR (400 MHz, CDCl_3): δ 7.86-7.82 (m, 2H), 7.73-7.71 (m, 2H), 7.49-7.27 (m, 6H), 7.22 (d, J = 8.4 Hz, 2H), 7.06 (d, J = 8.4 Hz, 2H), 4.16 (t, J = 8.4 Hz, 1H), 3.20-3.16 (m, 1H), 1.79-1.74 (m, 1H), 1.64-1.51 (m, 1H), 1.50-1.47 (m, 1H), 0.83 (d, J = 6.4 Hz, 3H), 0.80 (d, J = 6.8 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 142.7, 133.5, 132.6, 132.5, 132.4, 131.9, 128.65, 128.60, 128.5, 128.4, 128.2, 127.9, 53.4, 49.2, 24.9, 22.7, 22.1. HRMS (ES+) m/z : Calcd for $\text{C}_{23}\text{H}_{25}\text{ClNOP}$ $[\text{M}+\text{Na}]^+$ 420.1260; found 420.1269. IR (KBr, cm^{-1}) 3020, 1455, 1180, 1123, 1014, 750, 697.

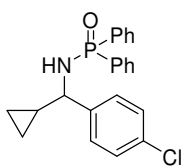


***N*-(1-(4-chlorophenyl)-2-methylpropyl)-diphenylphosphoramidate**

(9d). The general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a

gradient of 10% to 95% EtOAc in hexanes, affording **9d** as a white solid (36 mg, 75% yield). $R_f = 0.53$ (Hexane: EtOAc, 1:10). mp 170-172 °C. 84% ee (Chiralpak AD-H, hexanes/IPA 90/10, 1.0 mL min⁻¹, $\lambda = 222$ nm, $t_R = 17.6$ min (minor) $t_R = 20.7$ min (major).

¹H NMR (400 MHz, CDCl₃): δ 7.85-7.77 (m, 2H), 7.68-7.61 (m, 2H), 7.47-7.25 (m, 6H), 7.20 (d, $J = 11.2$ Hz, 2H), 6.97 (d, $J = 11.2$ Hz, 2H), 3.88-3.85 (m, 1H), 3.35-3.25 (m, 1H), 1.97-1.95 (m, 1H), 0.99 (d, $J = 8.8$ Hz, 3H), 0.79 (d, $J = 9.2$ Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 141.6, 134.1, 133.6, 132.8, 132.6, 132.5, 132.2, 131.94, 131.86, 131.8, 131.7, 128.7, 128.6, 128.4, 128.2, 128.1, 60.9, 35.6, 19.32, 19.29. HRMS (FAB+) m/z : Calcd for C₂₂H₂₃ClNOP [MH]⁺ 384.1284; found 384.1283. IR (KBr, cm⁻¹) 2955, 1436, 1185, 1123, 1014, 693.



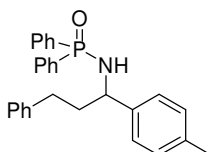
***N*-(1-(4-chlorophenyl)-2-methylpropyl)-diphenylphosphoramidate**

(9e). The general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a

gradient of 10% to 95% EtOAc in hexanes, affording **9e** as a white solid (38 mg, 79% yield). $R_f = 0.55$ (Hexane: EtOAc, 1:10). mp 149-151 °C. 95% ee (Chiralpak AD-H, hexanes/IPA 90/10, 1.0 mL min⁻¹, $\lambda = 210$ nm, $t_R = 19.5$ min (minor), $t_R = 21.6$ min (major).

¹H NMR (400 MHz, CDCl₃): δ 7.95-7.89 (m, 2H), 7.78-7.74 (m, 2H), 7.55-7.23 (m, 10H), 3.68 (c, $J = 9.6$ Hz, 1H), 3.42 (t, $J = 5.6$ Hz, 1H), 1.25-1.20 (m, 1H), 0.60-0.47 (m, 2H), 0.41 (sext, $J = 4.8$ Hz, 1H), 0.25 (sext, $J = 4.8$ Hz, 1H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 141.9, 132.8, 132.4, 132.3, 132.2, 132.1, 132.0, 131.9, 131.8, 128.6, 128.5, 128.4, 128.3, 61.5, 19.4, 5.3, 3.9. HRMS (FAB+) m/z : Calcd for C₂₂H₂₁ClNOP [MH]⁺

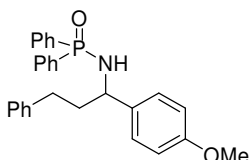
382.1127; found 382.1118. IR (KBr, cm^{-1}) 3203, 1491, 1436, 1183, 1124, 1014, 748, 692.



***N*-(1-(4-tolyl)-1-phenylpropyl)-diphenylphosphoramidate (9f).** The

general procedure was followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 10% to 95% EtOAc in hexanes, affording **9f** as a white solid (38 mg, 71% yield). R_f = 0.65 (Hexane: EtOAc, 1:10). mp 165-167 °C. 91% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 1.0 mL min^{-1} , λ = 210 nm, t_R = 9.7 min (major), t_R = 19.7 min (minor).

^1H NMR (400 MHz, CDCl_3): δ 7.88-7.82 (m, 2H), 7.80-7.77 (m, 2H), 7.52-7.09 (m, 15H), 4.19 (m, 1H), 3.42 (td, J = 9.6, 6.8 Hz, 1H), 2.59-2.47 (m, 2H), 2.39 (s, 3H), 2.40-2.32 (m, 1H), 2.18-2.06 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 141.9, 140.5, 140.4, 136.9, 133.9, 132.6, 132.5, 131.9, 131.8, 131.7, 131.4, 129.3, 128.5, 128.43, 128.38, 128.30, 126.4, 125.8, 55.3, 41.0, 32.4, 21.1. LRMS (FAB+) m/z : Calcd for $\text{C}_{28}\text{H}_{28}\text{ClNOP} [\text{MH}]^+$ 426.1987; found 426.1981. IR (KBr, cm^{-1}) 3143, 1437, 1193, 1109, 1068, 723, 695.

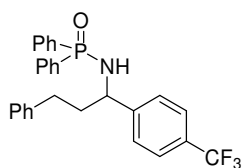


***N*-(1-(4-methoxyphenyl)-1-phenylpropyl)-**

diphenylphosphoramidate (9g). The general procedure was

followed and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 10% to 95% EtOAc in hexanes, affording **9g** as a white solid (37.5 mg, 68% yield). R_f = 0.58 (Hexane: EtOAc, 1:10). mp 180-182 °C. 86% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 0.9 mL min^{-1} , λ = 210 nm, t_R = 20.0 min (major), t_R = 22.5 min (minor).

^1H NMR (400 MHz, CDCl_3): δ 7.90-7.86 (dd, $J = 12.0, 7.2$ Hz, 2H), 7.81-7.76 (dd, $J = 12.0, 7.2$ Hz, 2H), 7.54-7.16 (m, 11H), 7.13 (d, $J = 8.8$ Hz, 2H), 6.88 (d, $J = 8.4$ Hz, 2H), 4.21-4.15. (m, 1H), 3.85 (s, 3H), 3.29-3.25 (m, 1H), 2.62-2.46 (m, 2H), 2.39-2.32 (m, 1H), 2.20-2.02 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3): δ 158.7, 141.5, 135.62, 135.56, 132.7, 132.6, 132.5, 131.9, 131.8, 131.7, 131.4, 128.6, 128.44, 128.37, 128.32, 128.26, 127.7, 125.8, 55.3, 55.0, 40.9, 32.4. LRMS (ES+) m/z : Calcd for $\text{C}_{28}\text{H}_{28}\text{NO}_2\text{P}$ $[\text{M}+\text{Na}]^+$ 464; found 464. IR (KBr, cm^{-1}) 3209, 1512, 1437, 1247, 183, 1030, 830, 750, 697.



***N*-(1-(4-trifluoromethylphenyl)-1-phenylpropyl)-**

diphenylphosphoramidate (9h). The general procedure was followed

and the product was isolated by flash column chromatography on a Biotage Flash+ cartridge with a gradient of 10% to 95% EtOAc in hexanes, affording **9h** as a white solid (51 mg, 85% yield). $R_f = 0.67$ (Hexane: EtOAc, 1:10). mp 190-192 °C.

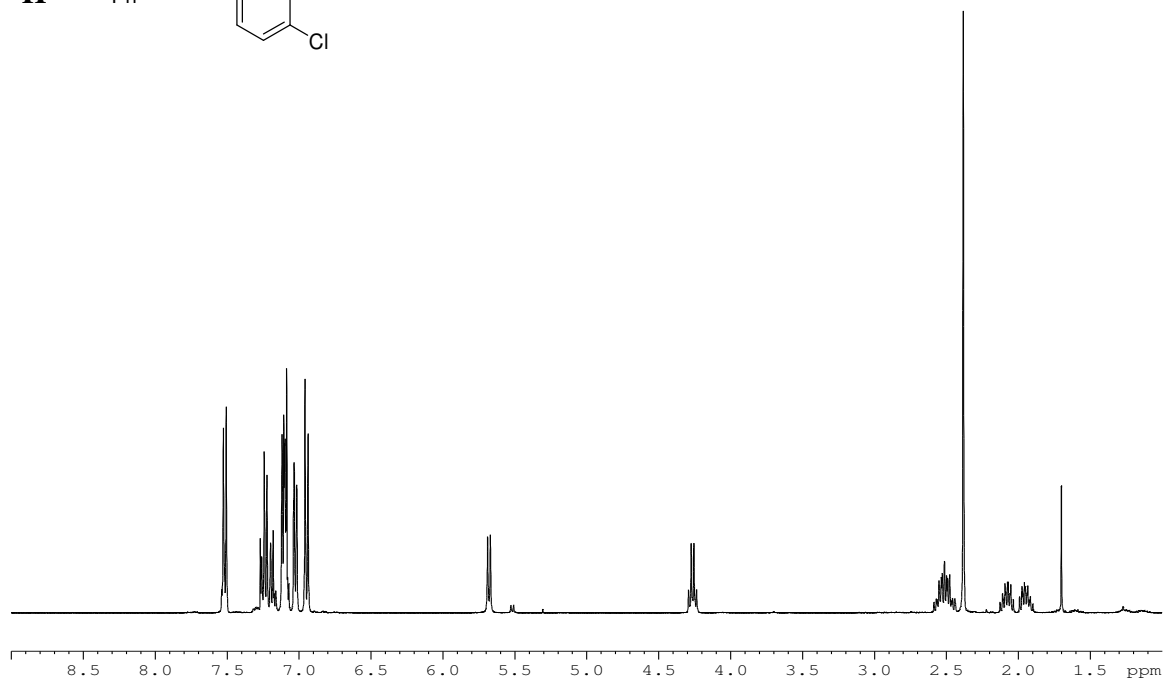
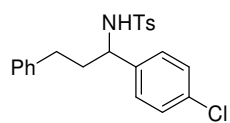
82% ee (Chiralpak OD-H, hexanes/EtOH 97/3, 0.9 mL min^{-1} , $\lambda = 230$ nm, $t_R = 16.9$ min (major), $t_R = 19.6$ min (minor))

^1H NMR (400 MHz, CDCl_3): δ 7.90-7.84 (m, 2H), 7.74-7.72 (m, 2H), 7.56-7.10 (m, 13H), 7.11 (d, $J = 7.2$ Hz, 2H), 4.45-4.25. (m, 1H), 3.55 (dd, $J = 10.0, 6.4$ Hz, 1H), 2.66-2.62 (m, 1H), 2.57-2.54 (m, 1H), 2.34-2.30 (m, 1H), 2.18-2.12 (m, 1H). For the $^{13}\text{C}\{^1\text{H}\}$ NMR spectrum see page 40. HRMS (FAB+) m/z : Calcd for $\text{C}_{28}\text{H}_{25}\text{NOF}_3\text{P}$ $[\text{MH}]^+$ 480.1704; found 480.1704. IR (KBr, cm^{-1}) 3187, 1512, 1438, 1327, 1180, 1122, 1018, 752, 698.

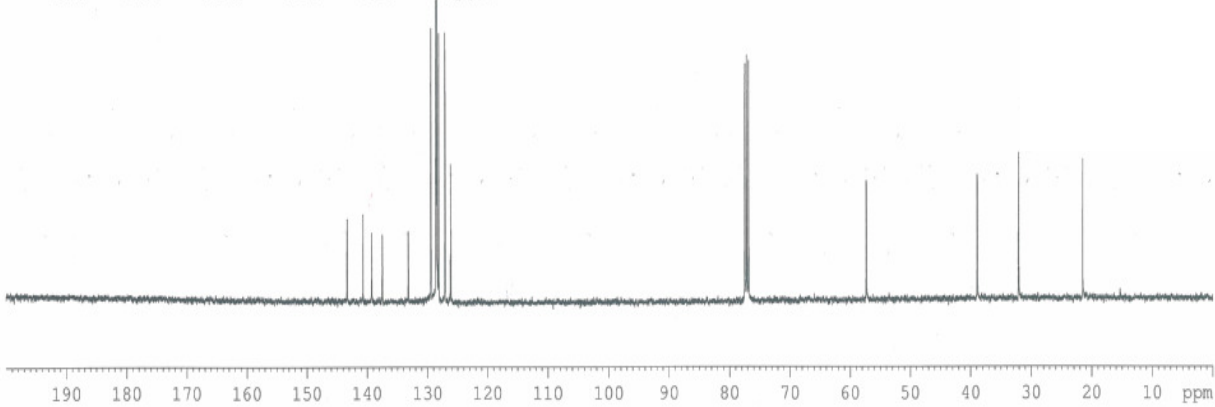
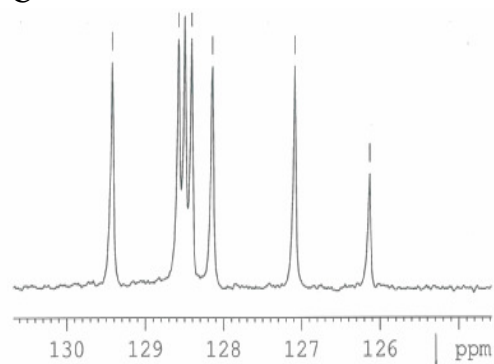
References

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- (3) (a) Henry, P. M.; Davies, M.; Ferguson, G.; Philips, S.; Restivo, R. *Chem. Commun.* **1974**, 112. (b) Lemke, K.; Ballschuh, S.; Kunath, A.; Theil, F. *Tetrahedron: Asymmetry* **1997**, *8*, 2051. (c) Wang, Z. Q.; Feng, C. G.; Xu, M. H.; Lin, G. Q. *J. Am. Chem. Soc.* **2007**, *129*, 5336.

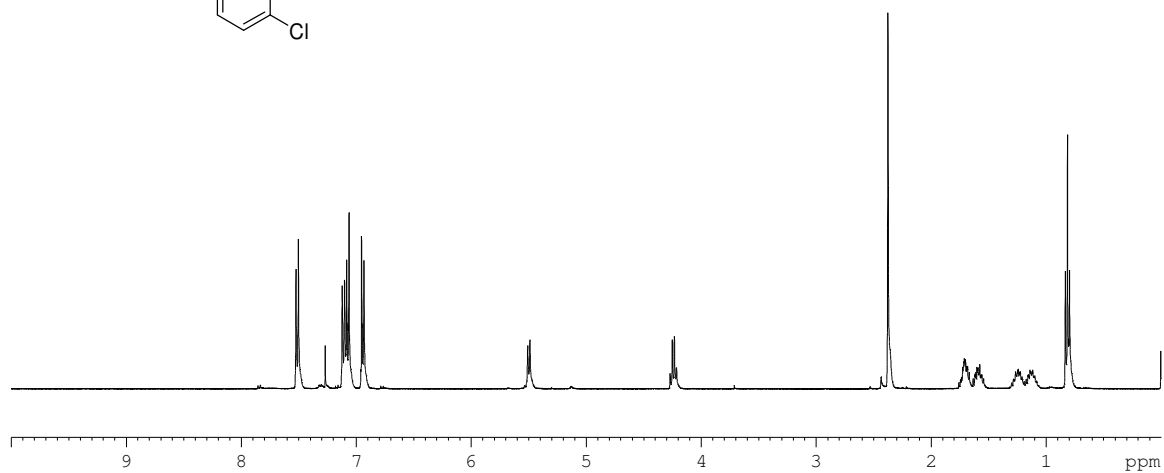
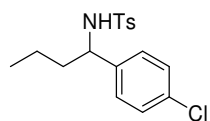
10a
¹H



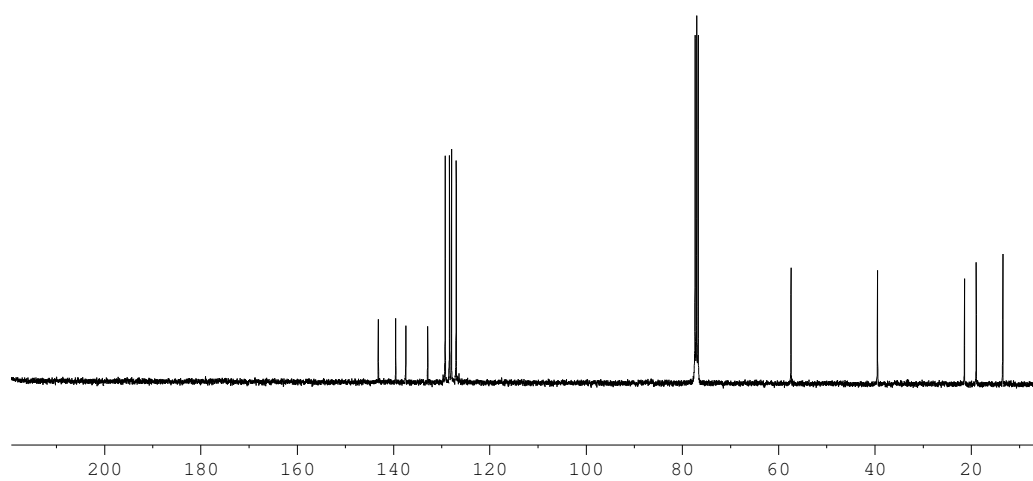
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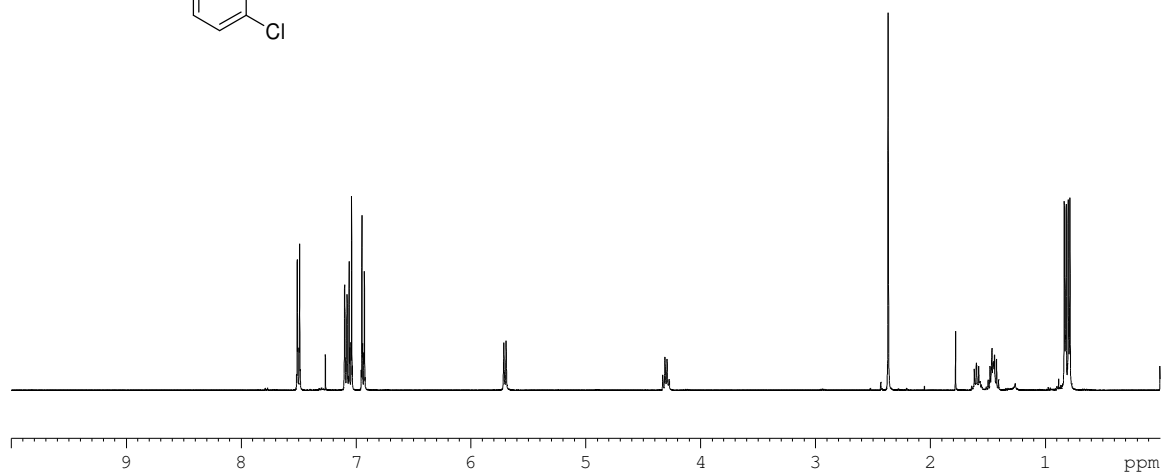
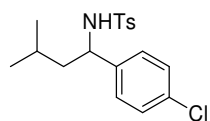
10b
¹H



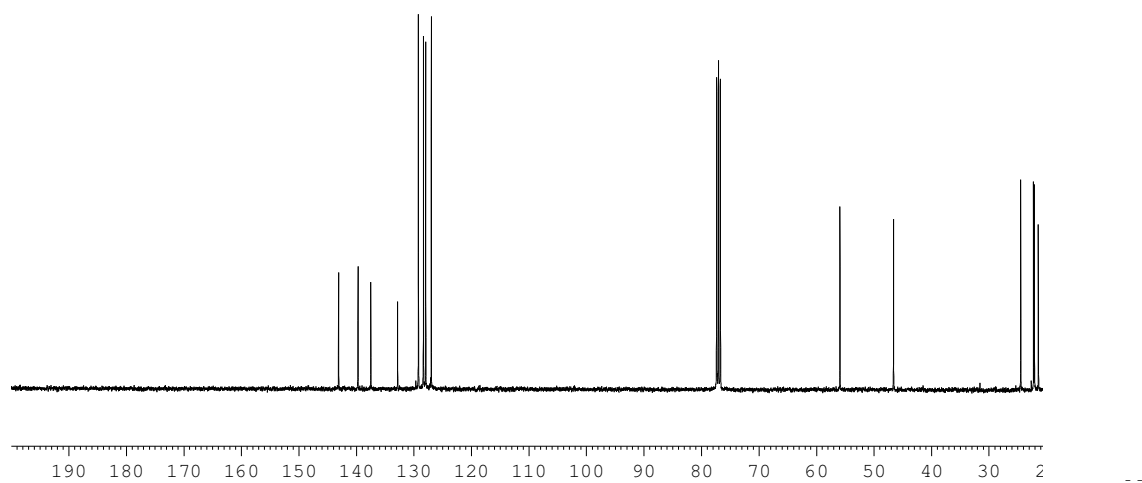
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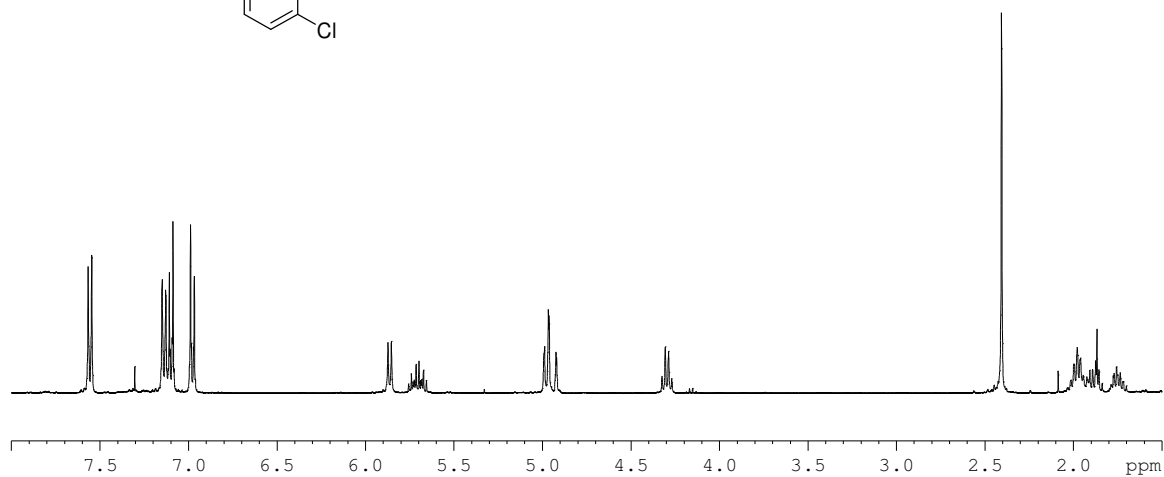
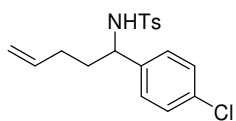
10c
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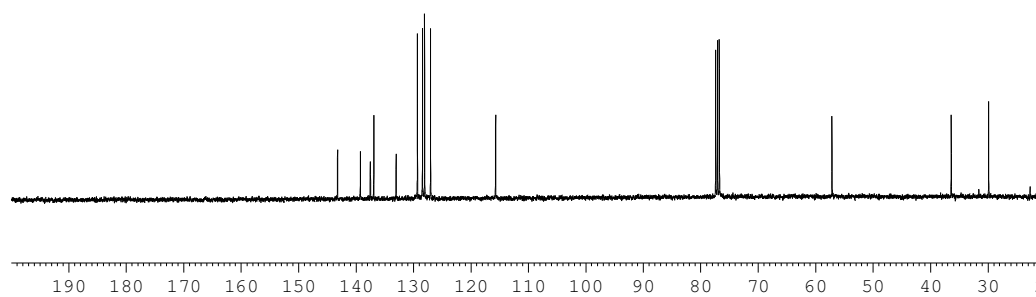
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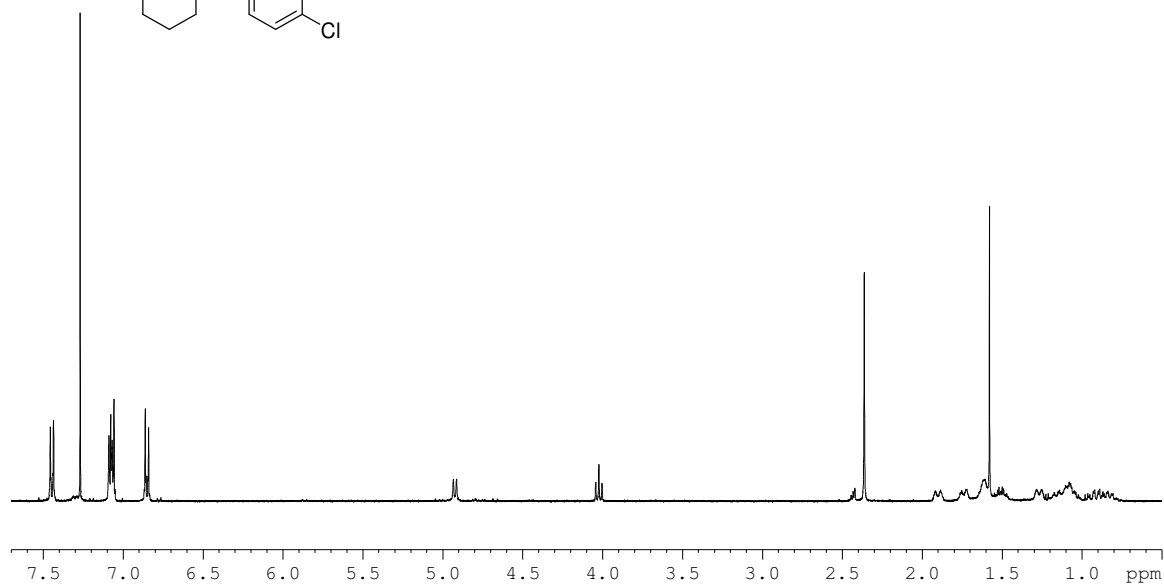
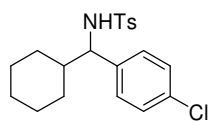
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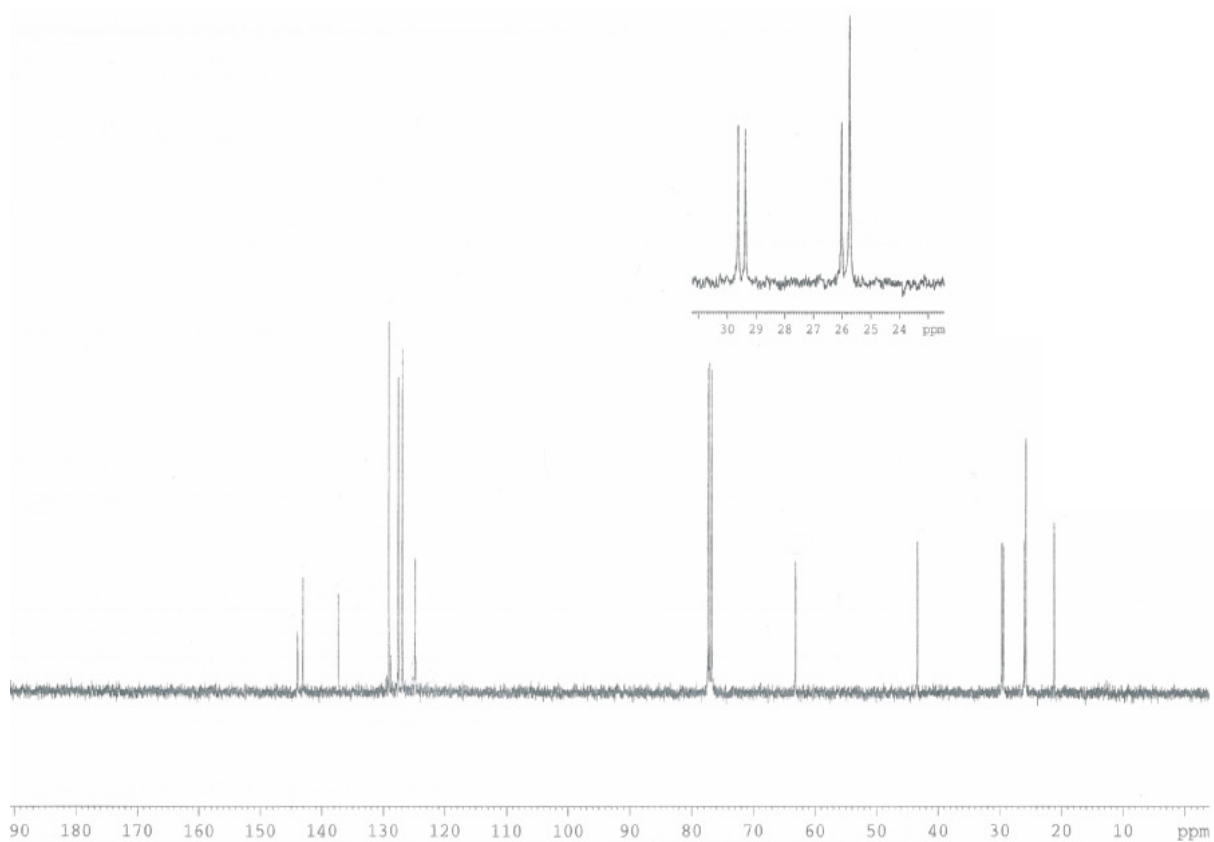
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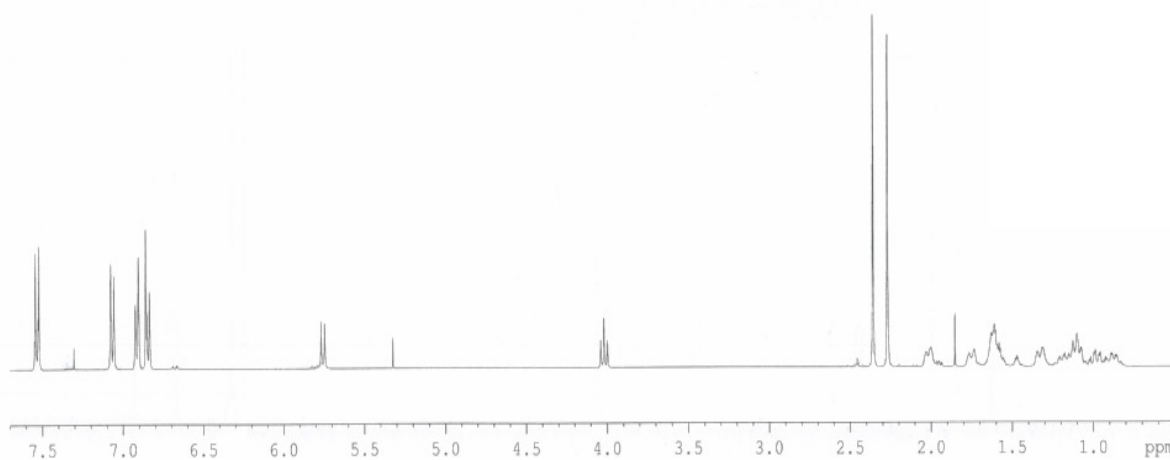
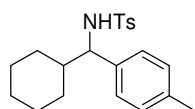
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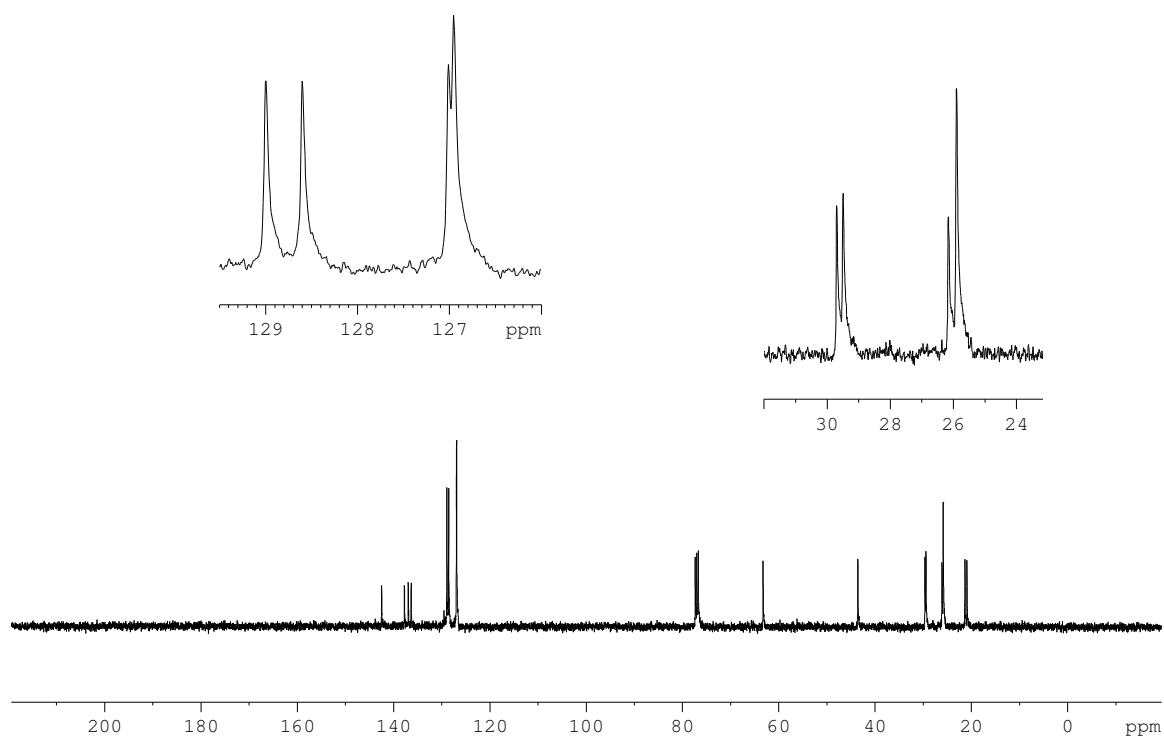
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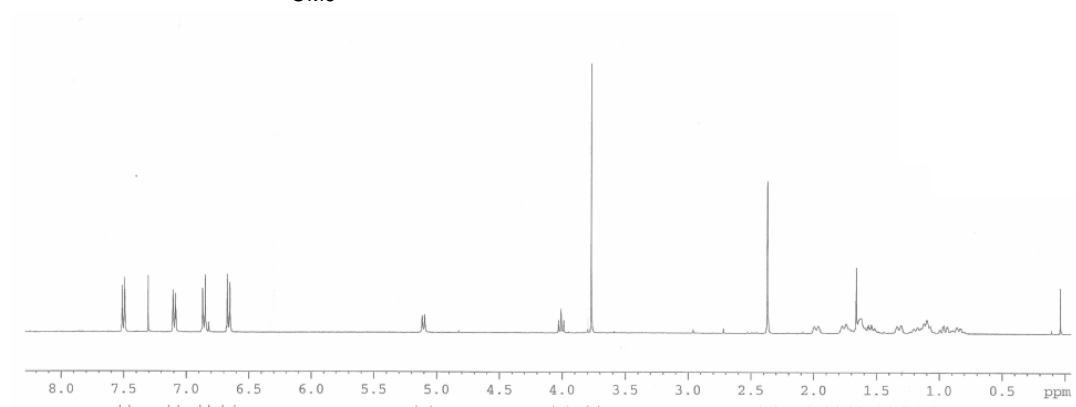
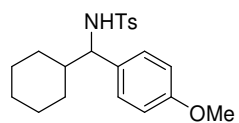
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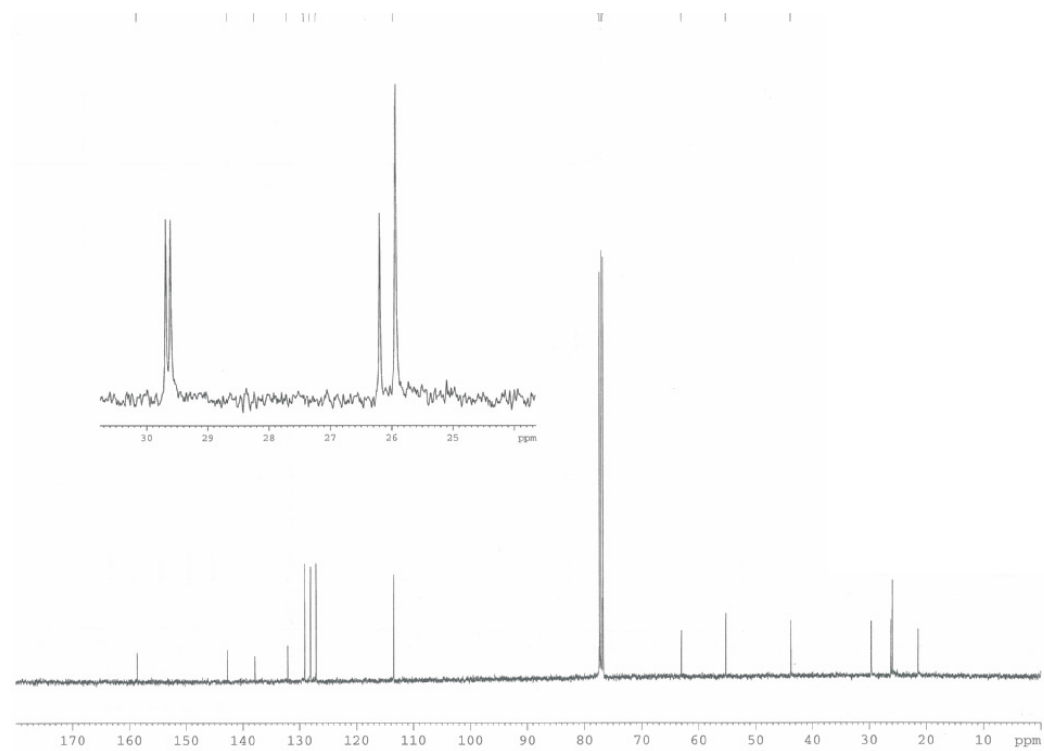
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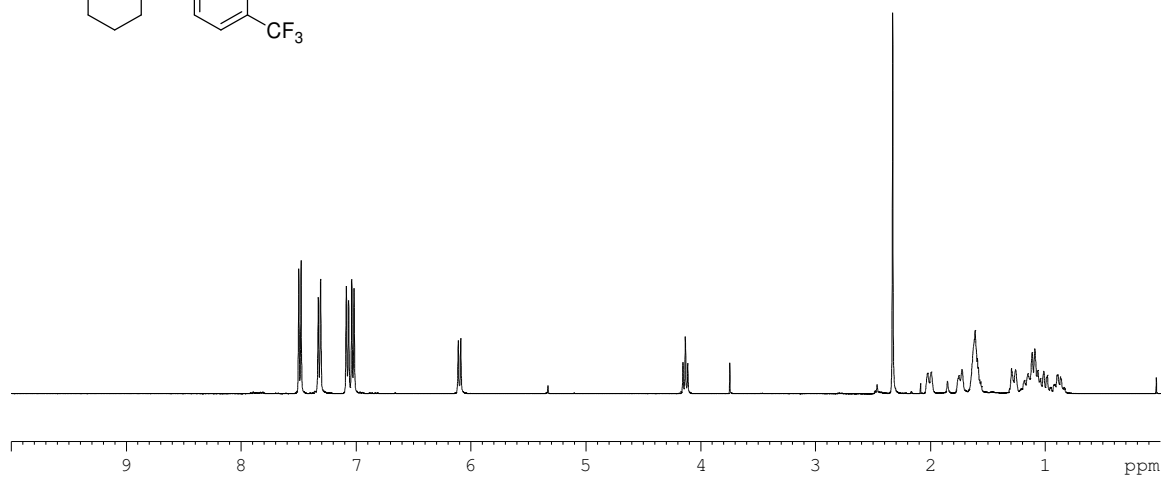
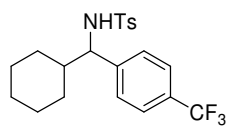
10g
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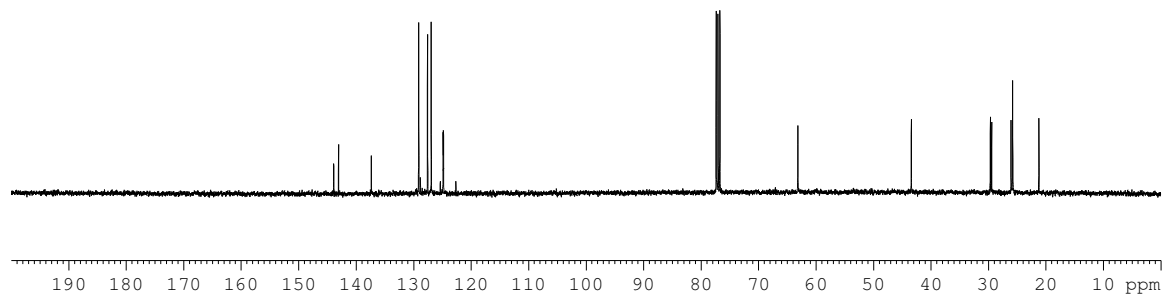
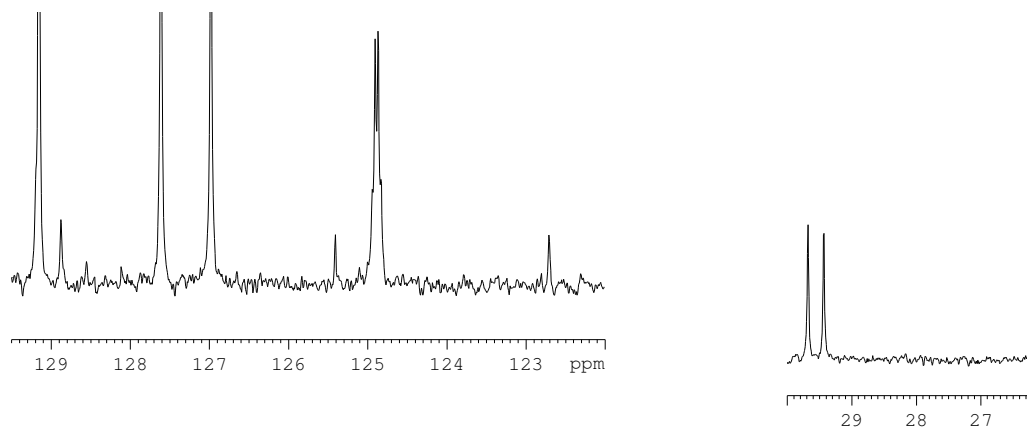
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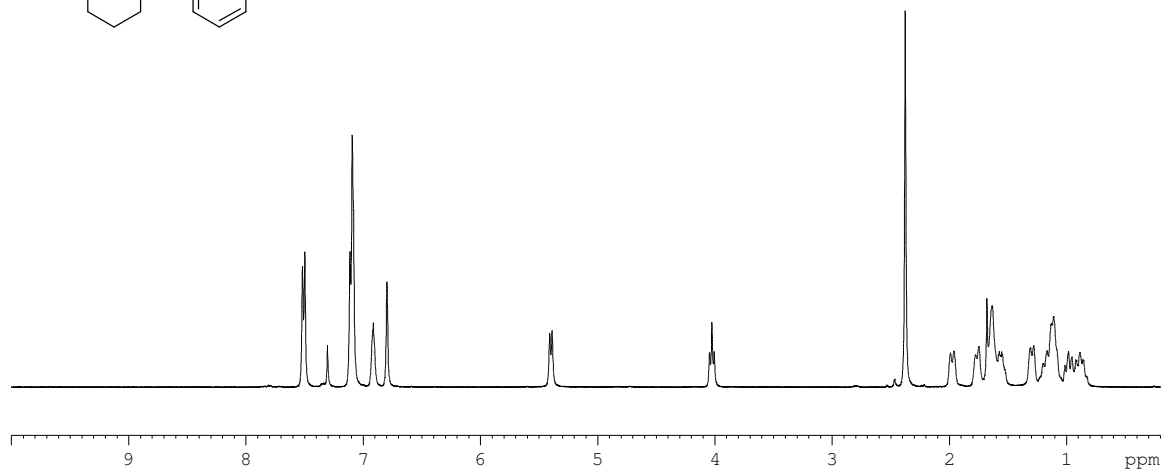
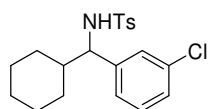
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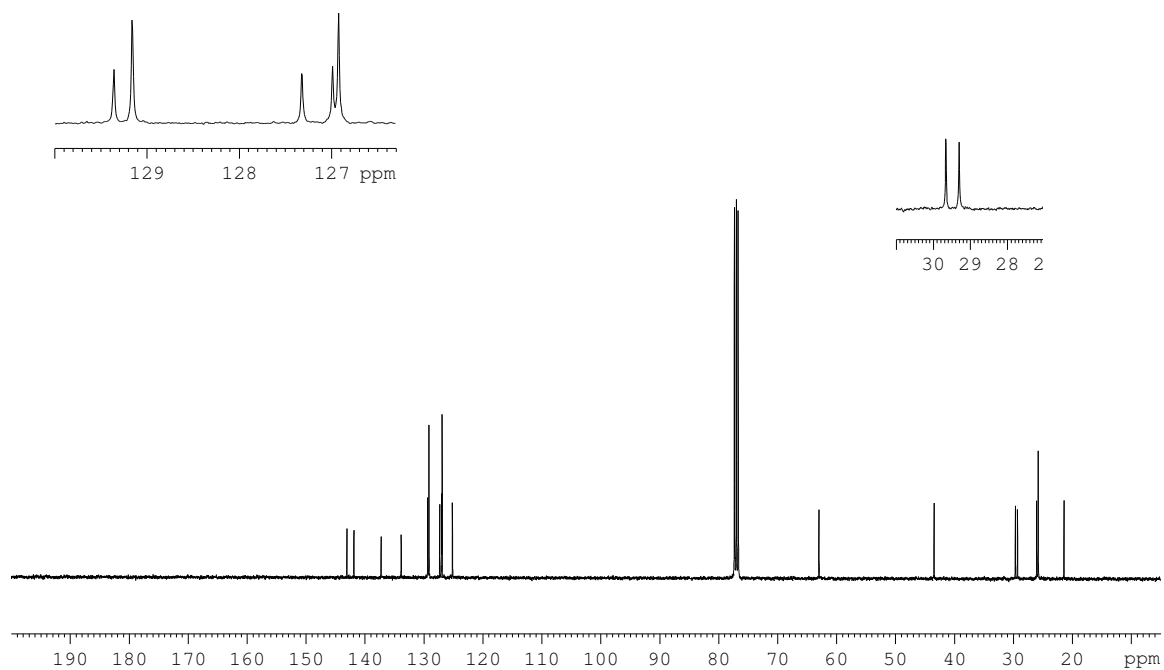
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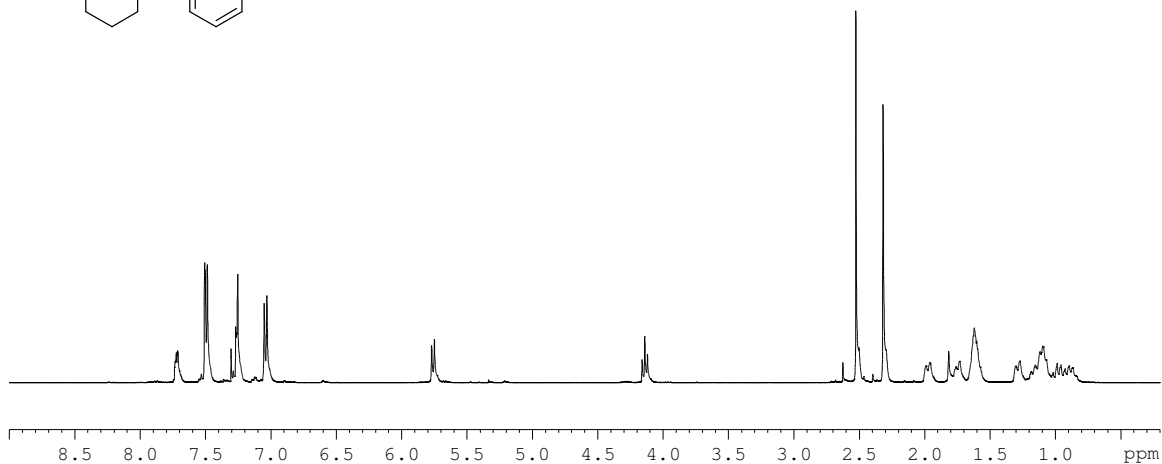
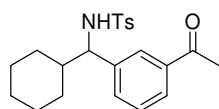
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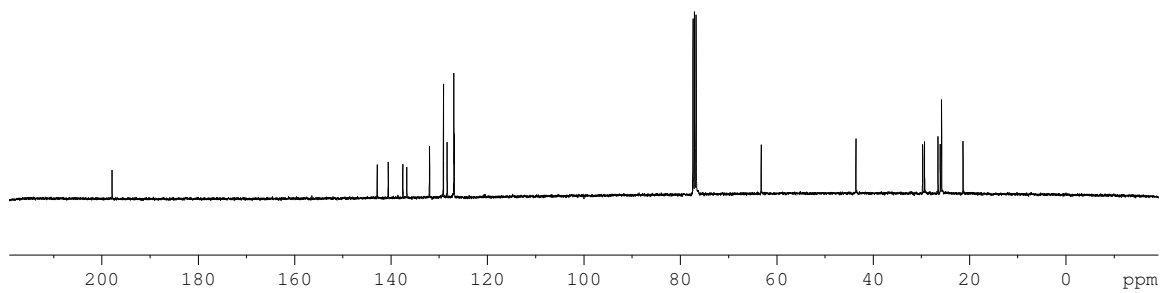
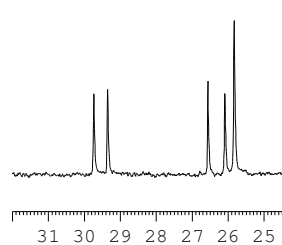
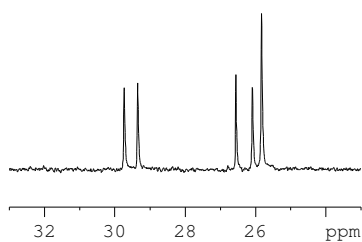
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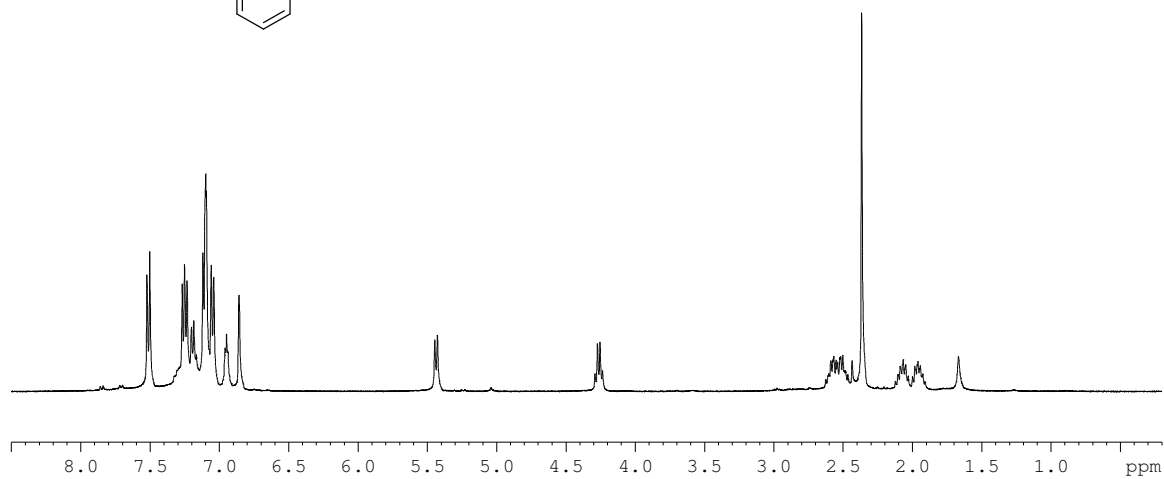
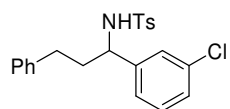
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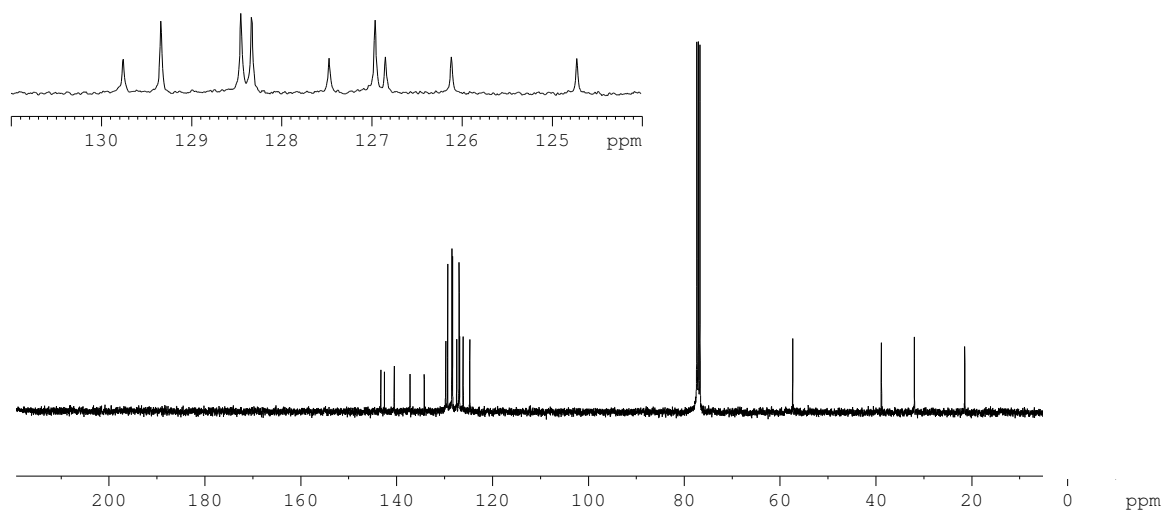
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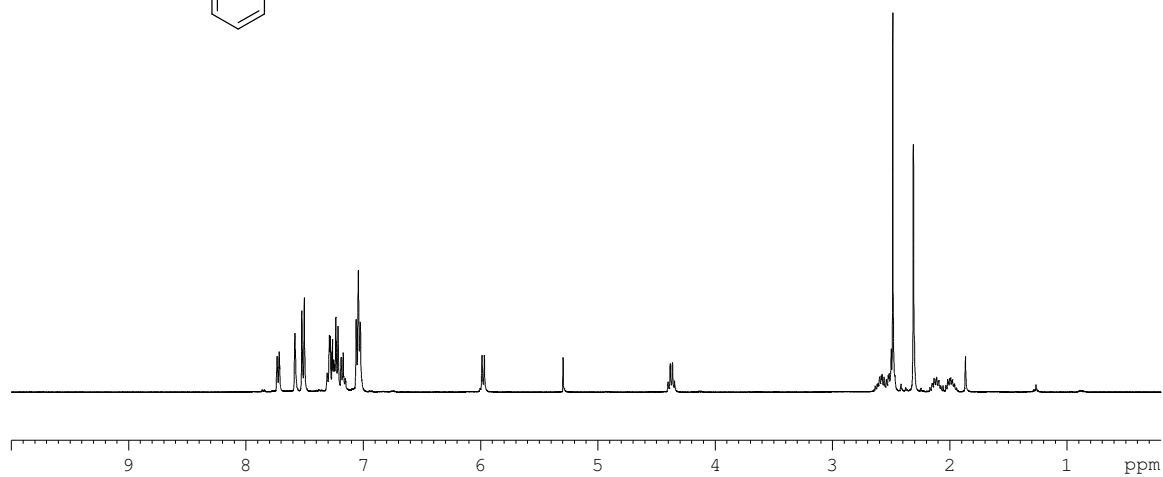
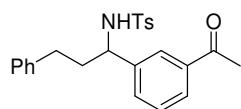
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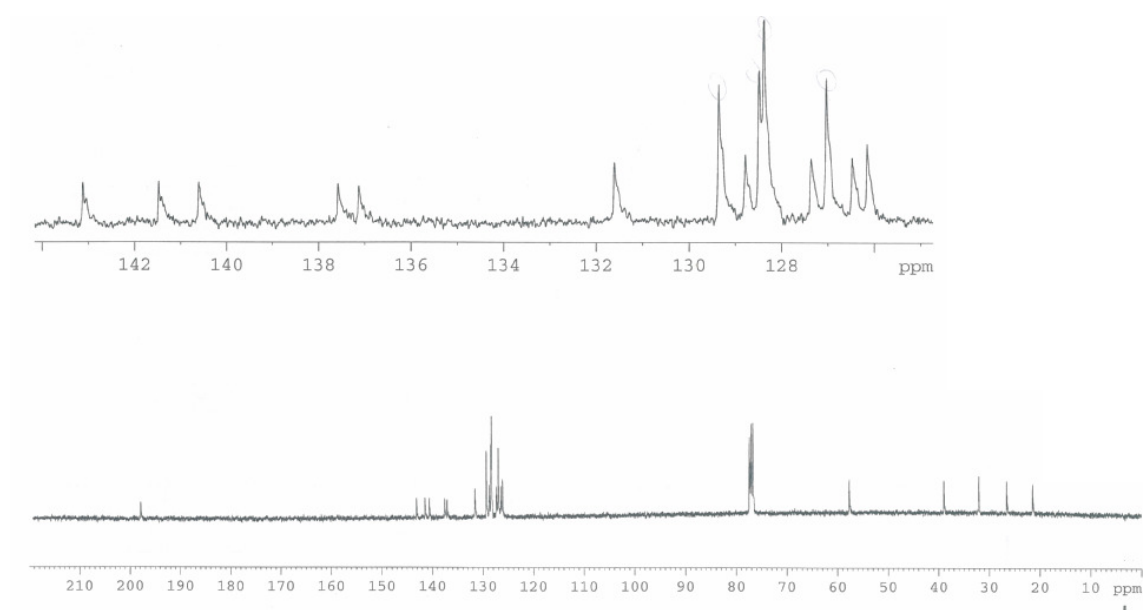
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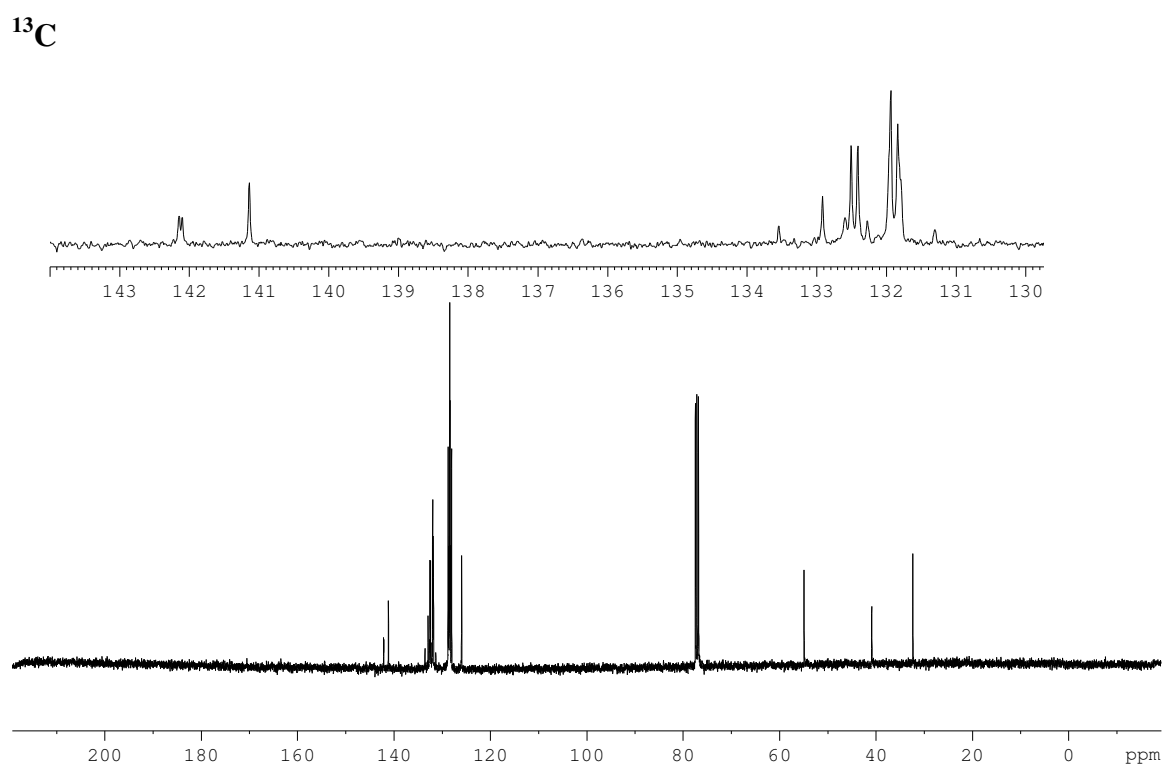
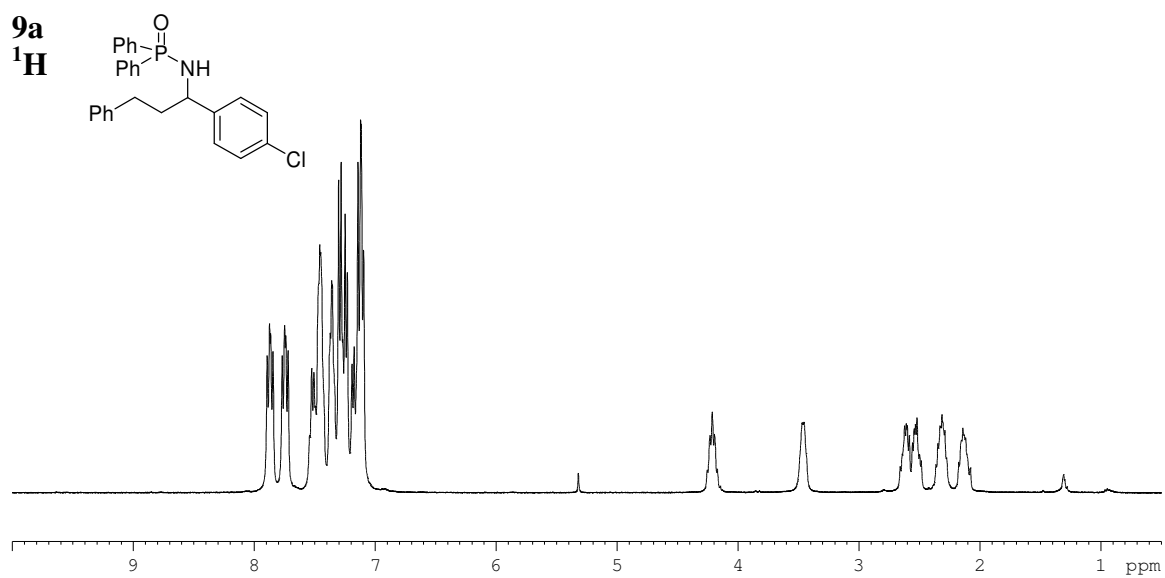


10l
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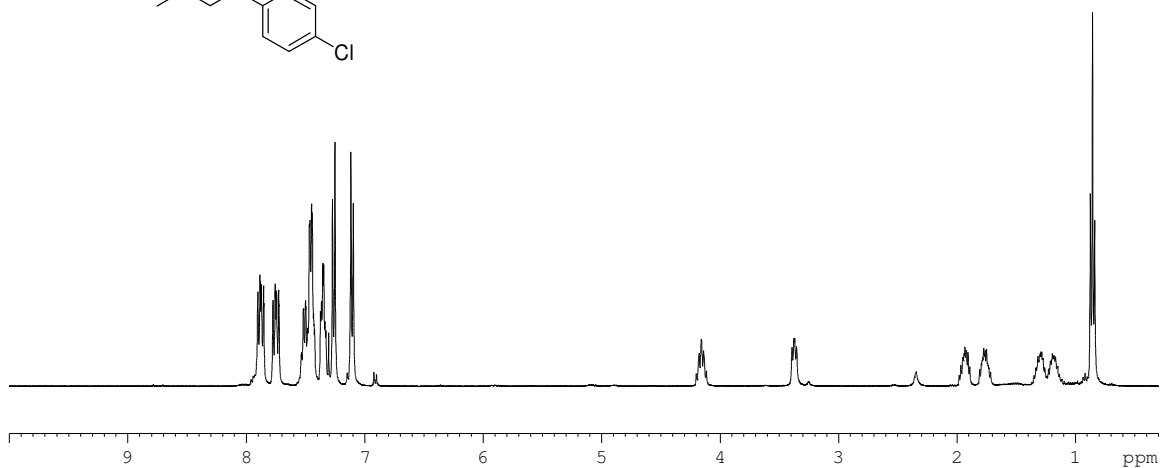
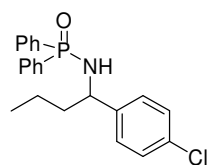


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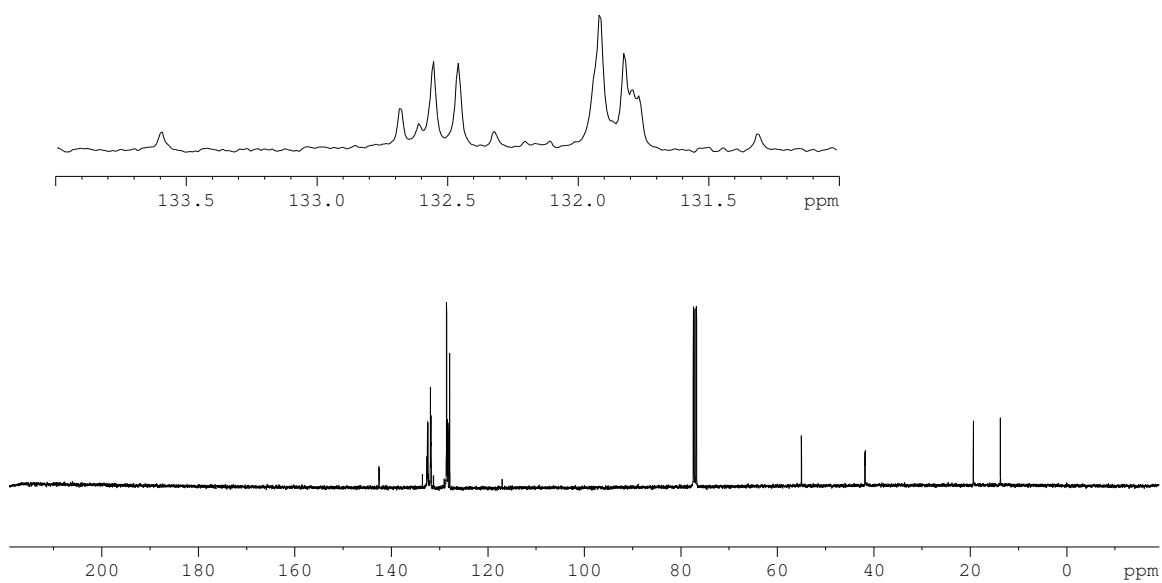


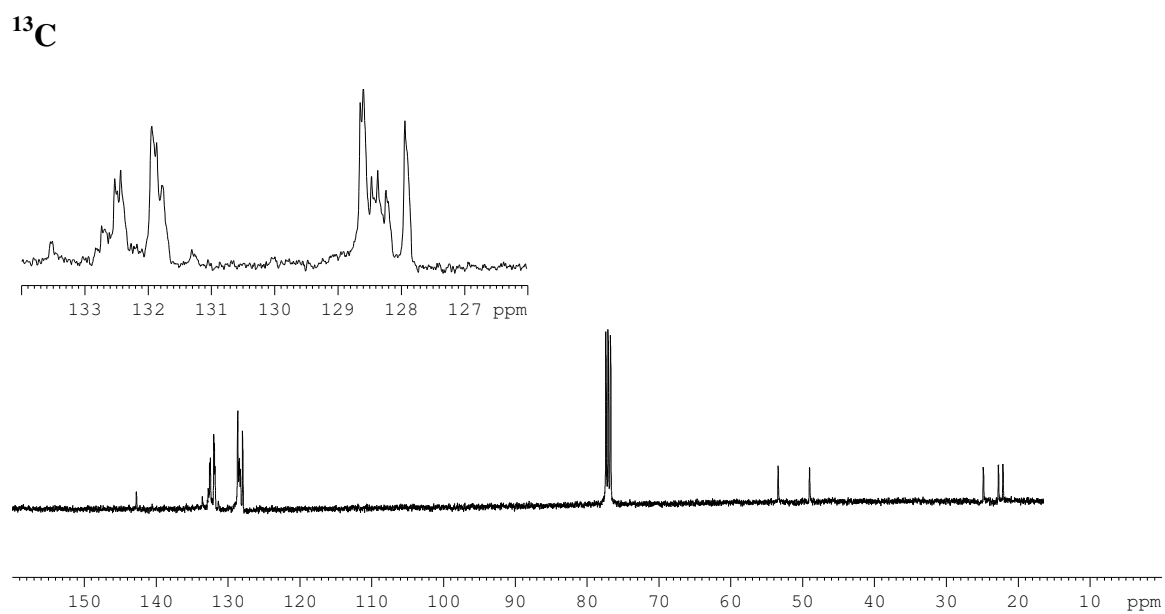
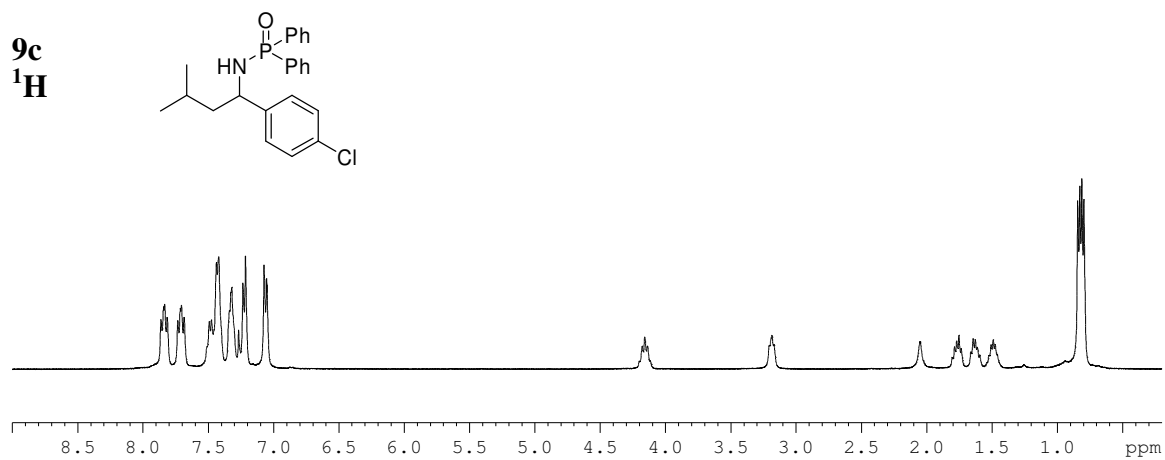


9b
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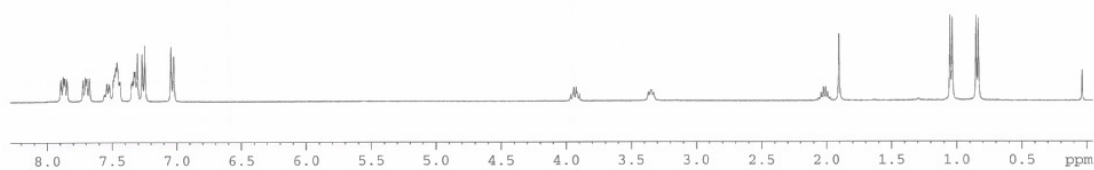
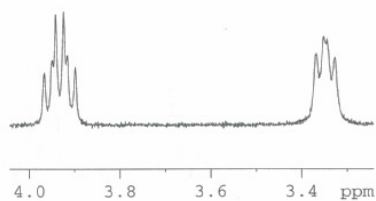
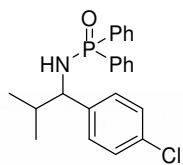


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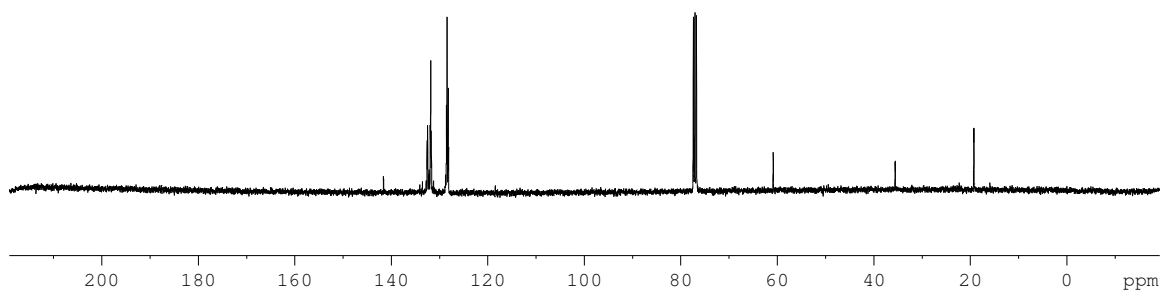
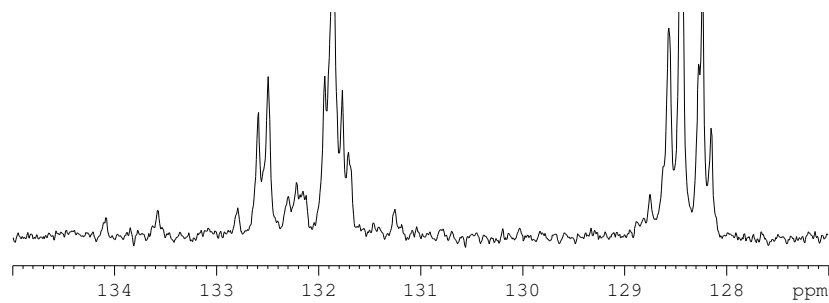




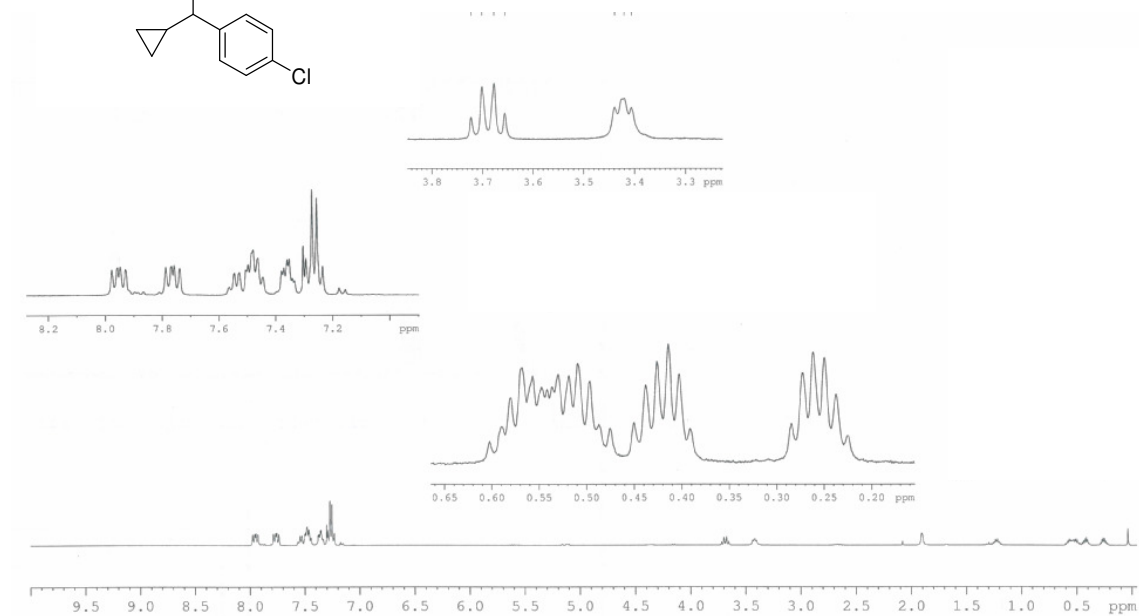
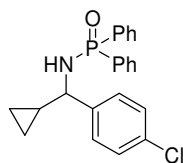
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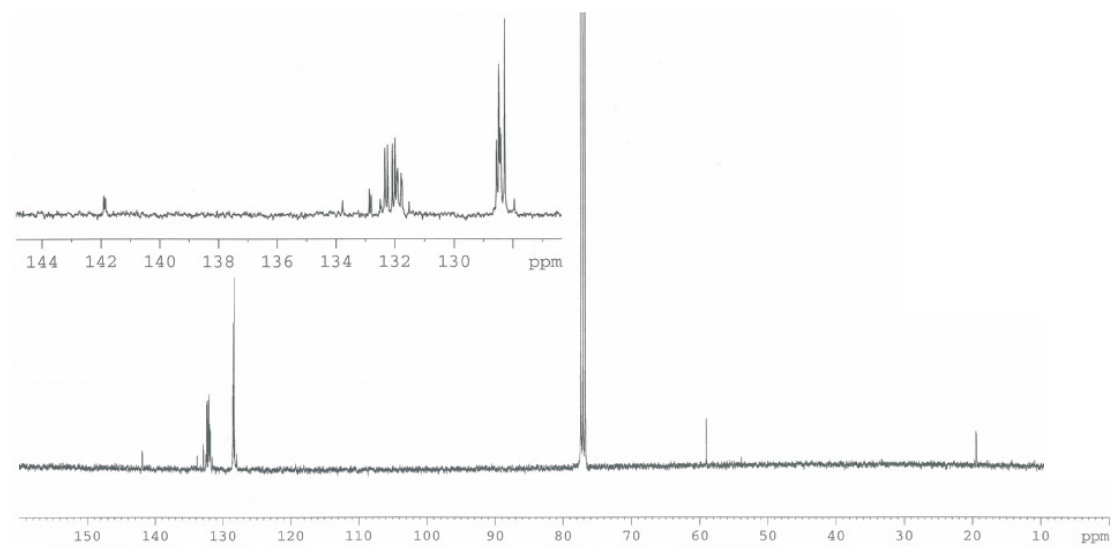
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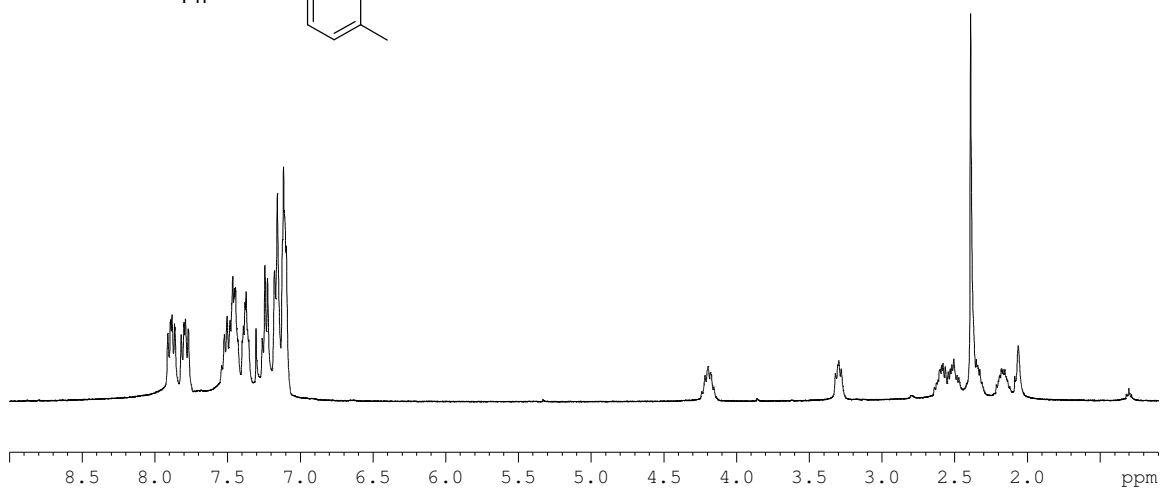
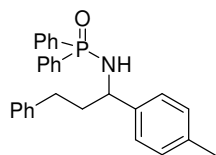
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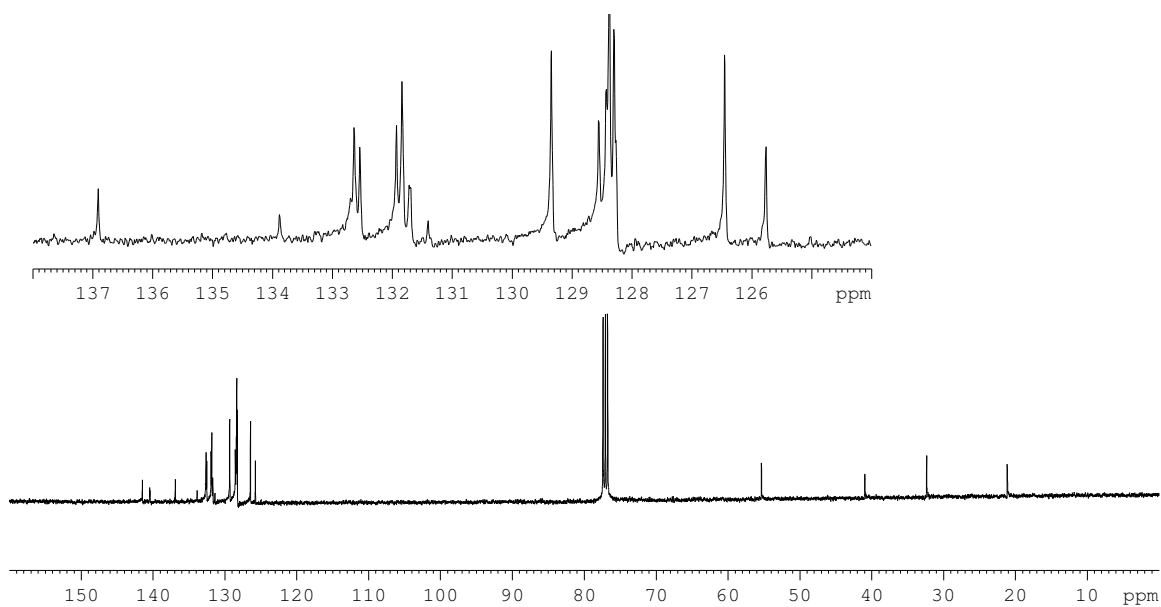
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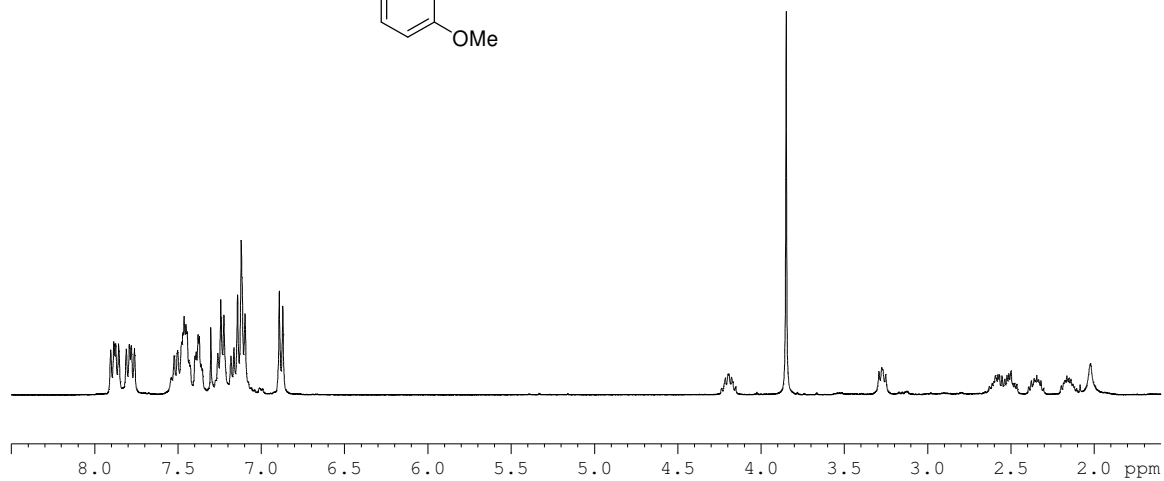
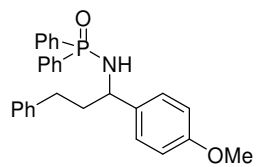
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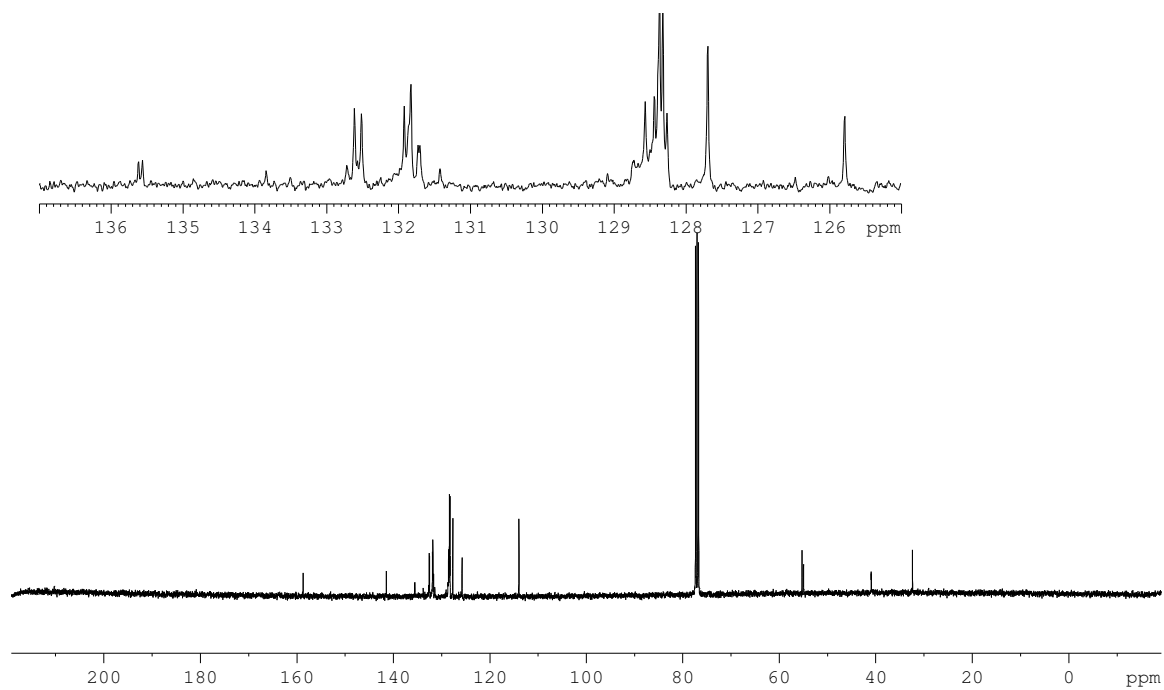
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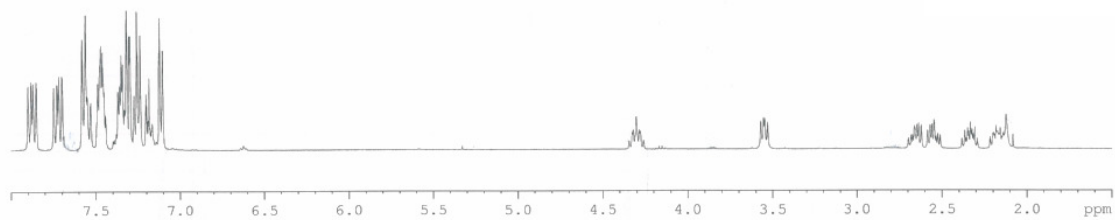
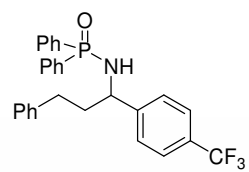
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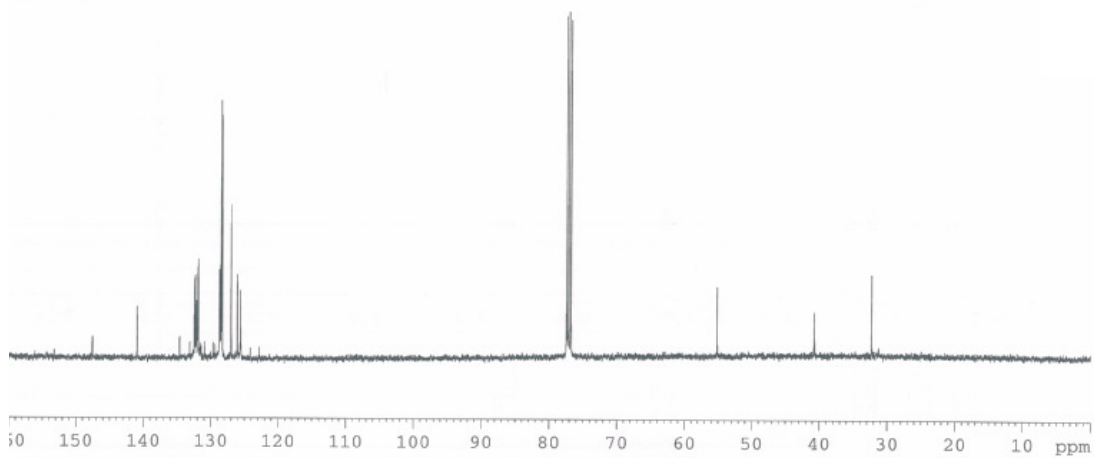
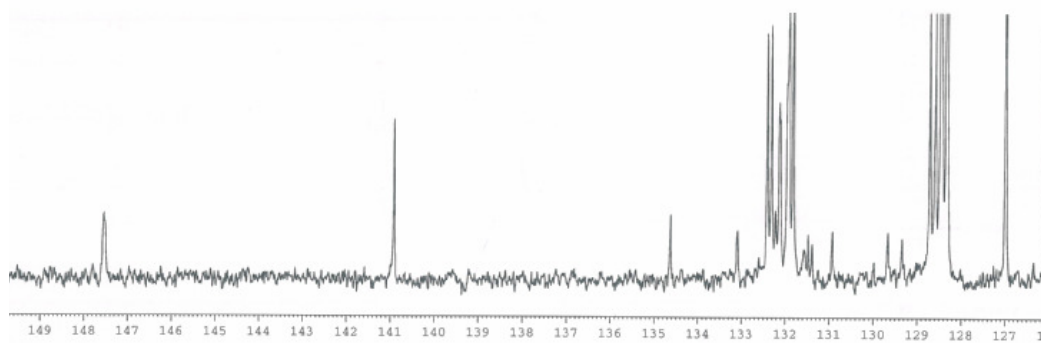
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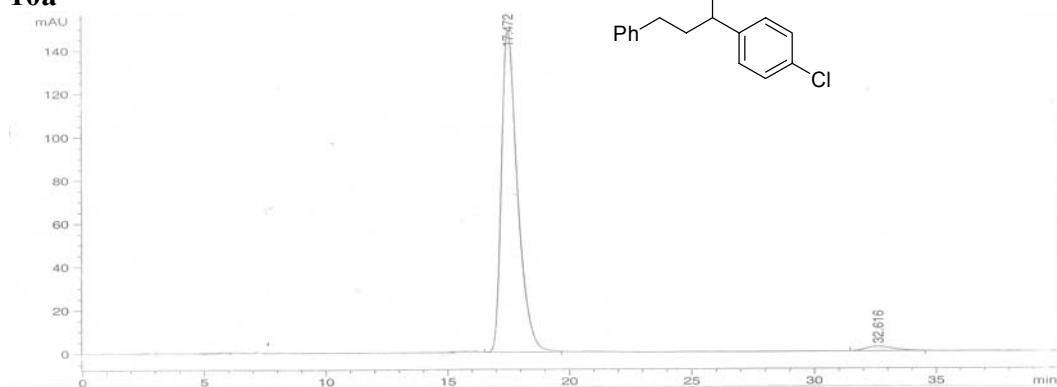
9h
¹H



¹³C



10a



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Area Percent Report
=====

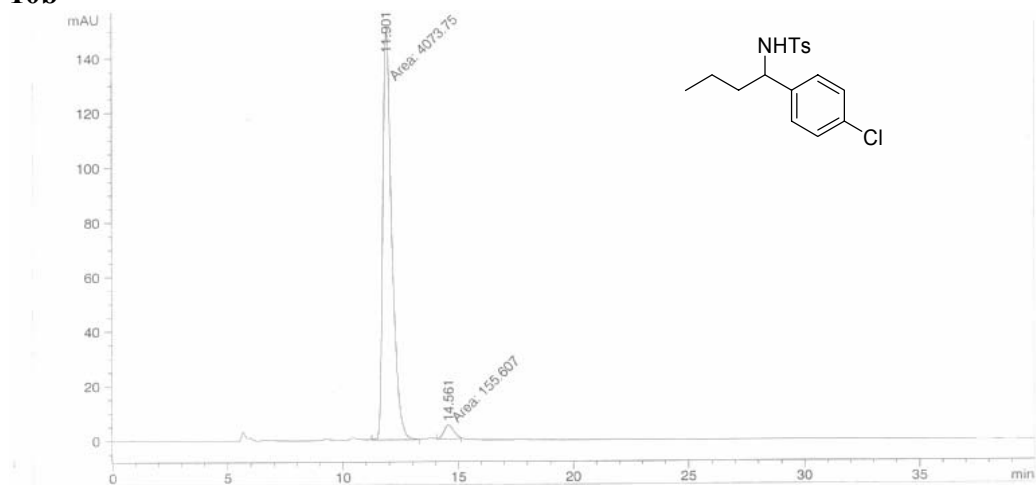
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Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.472	VB	0.6858	6822.37158	149.10960	97.4633
2	32.616	BB	1.0907	177.56927	2.09965	2.5367

Totals : 6999.94086 151.20926

10b



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Area Percent Report
=====

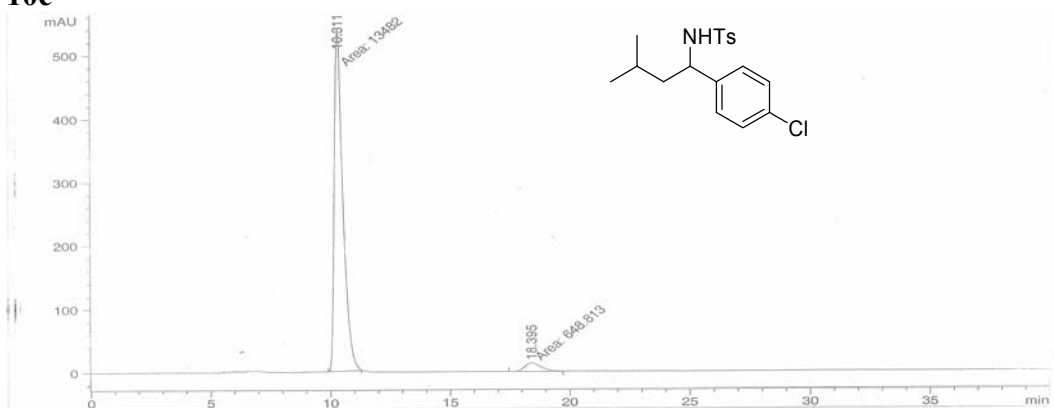
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Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.901	MM	0.4535	4073.75195	149.70436	96.3208
2	14.561	MM	0.5077	155.60712	5.10840	3.6792

Totals : 4229.35907 154.81276

10c



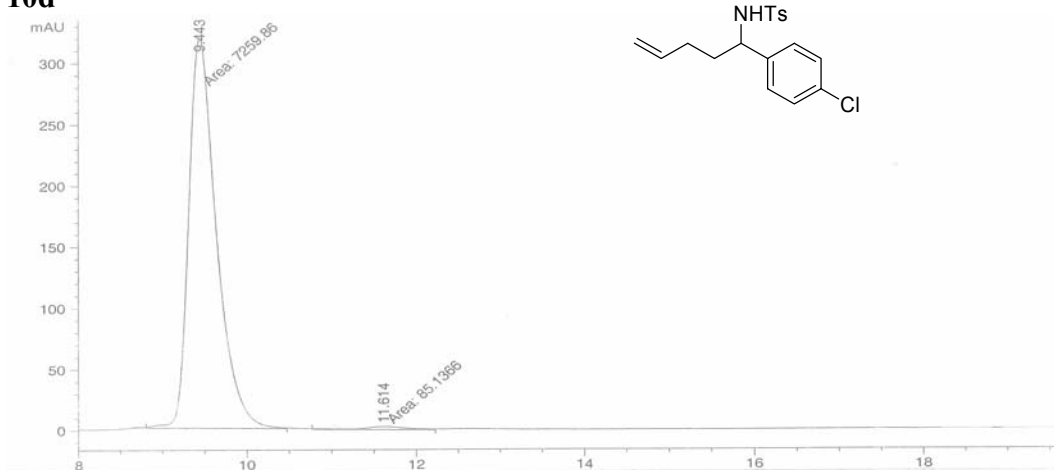
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Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.311	MM	0.4190	1.34820e4	536.21442	95.4085
2	18.395	MM	0.7953	648.81342	13.59645	4.5915
Totals :				1.41308e4	549.81087	

10d



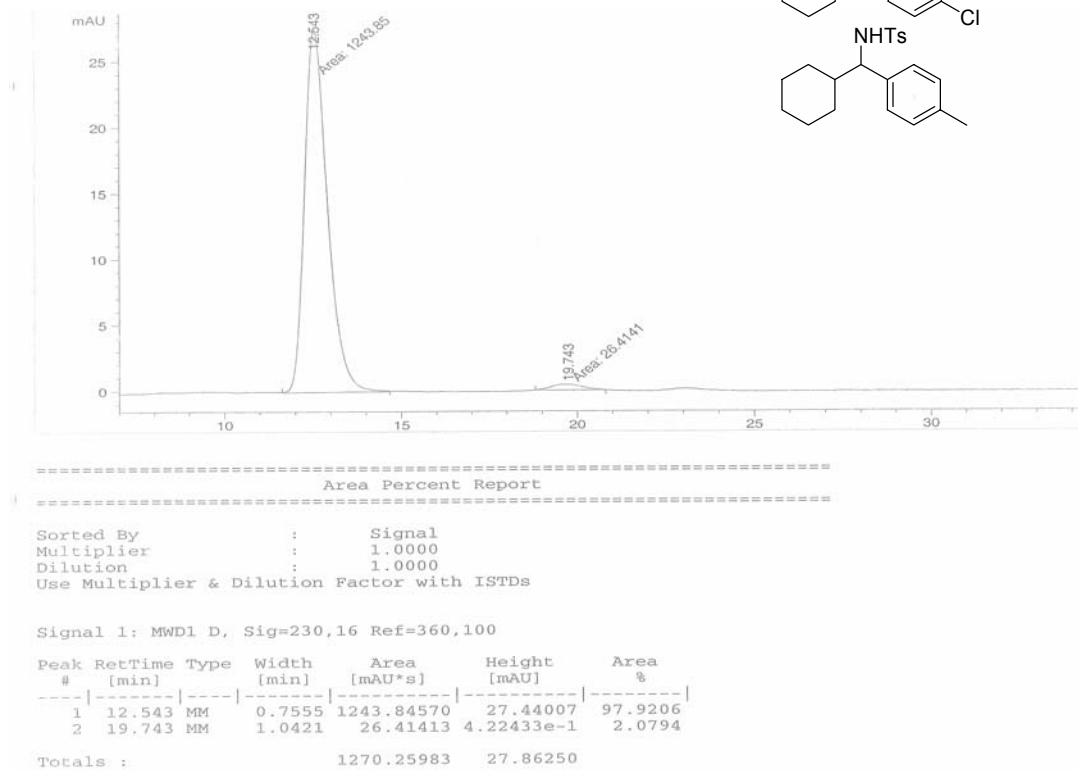
=====
Area Percent Report
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orted By : Signal
ultiplier : 1.0000
ilution : 1.0000
se Multiplier & Dilution Factor with ISTDs

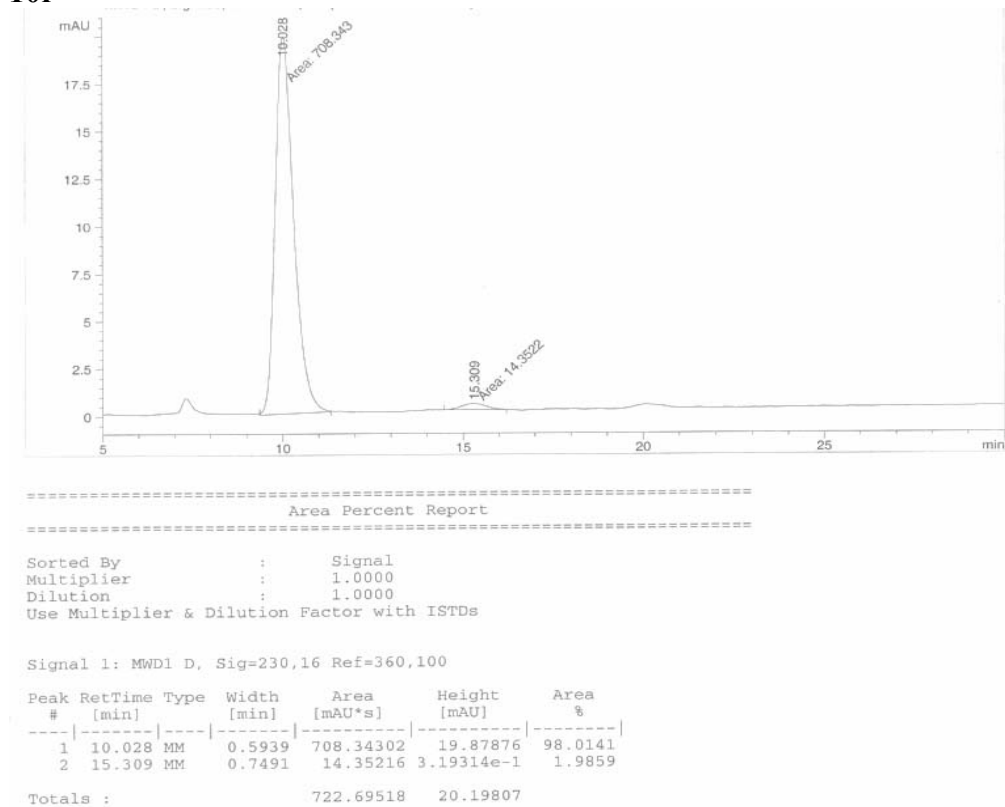
ignal 1: MWD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.443	MM	0.3802	7259.85596	318.25342	98.8409
2	11.614	MM	0.5685	85.13660	2.49602	1.1591
Totals :				7344.99256	320.74944	

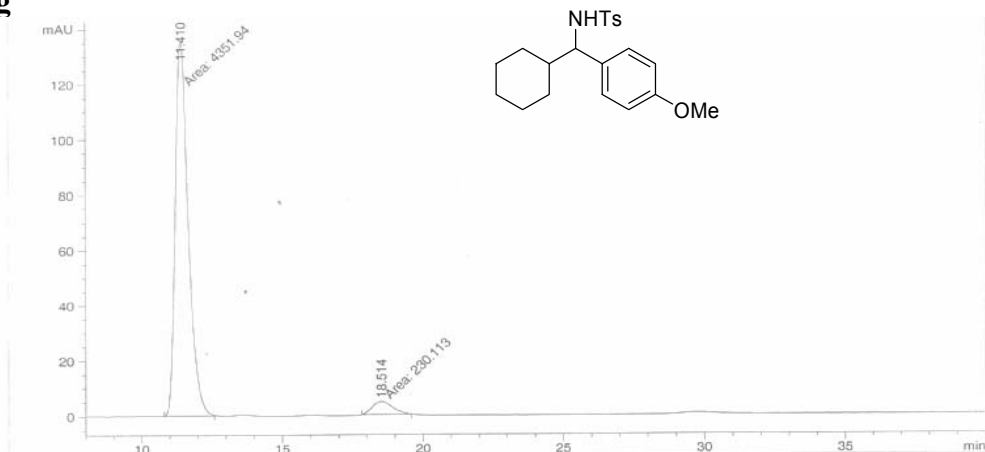
10e



10f



10g



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Area Percent Report
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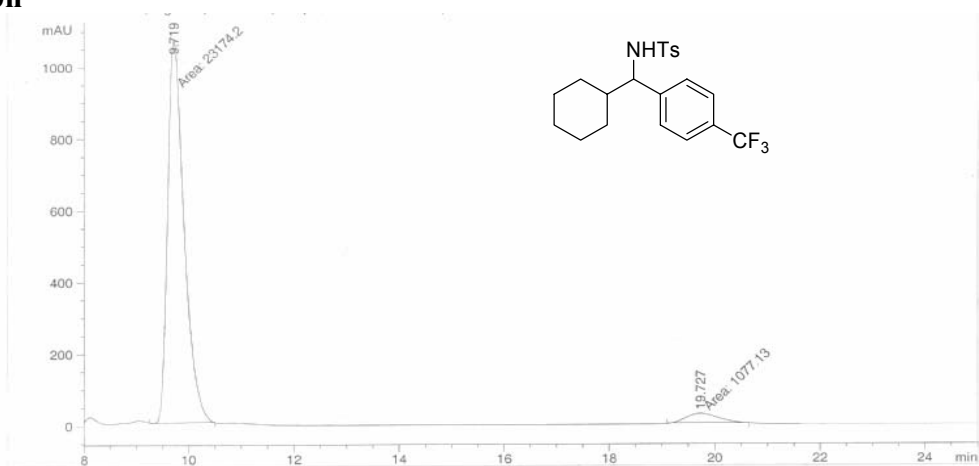
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.410	MM	0.5345	4351.93994	135.70813	94.9779
2	18.514	MM	0.8305	230.11317	4.61798	5.0221

Totals : 4582.05312 140.32611

10h



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Area Percent Report
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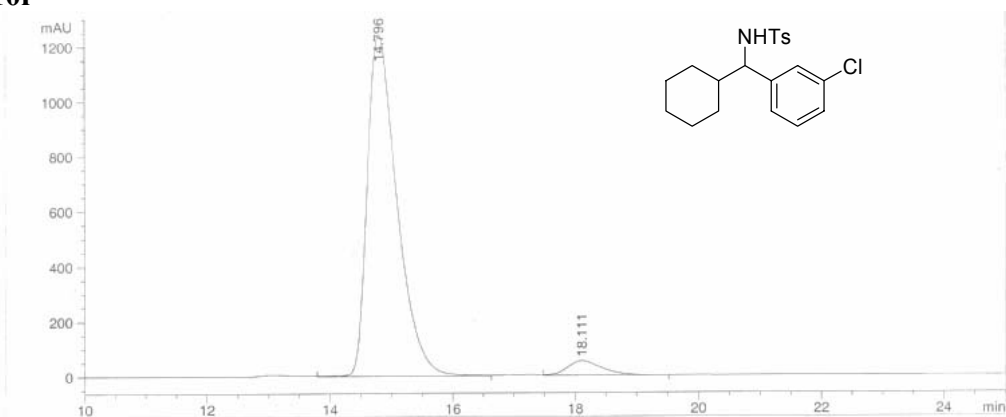
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.719	MM	0.3629	2.31742e4	1064.24548	95.5585
2	19.727	MM	0.6791	1077.13159	26.43612	4.4415

Totals : 2.42514e4 1090.68160

10i



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Area Percent Report
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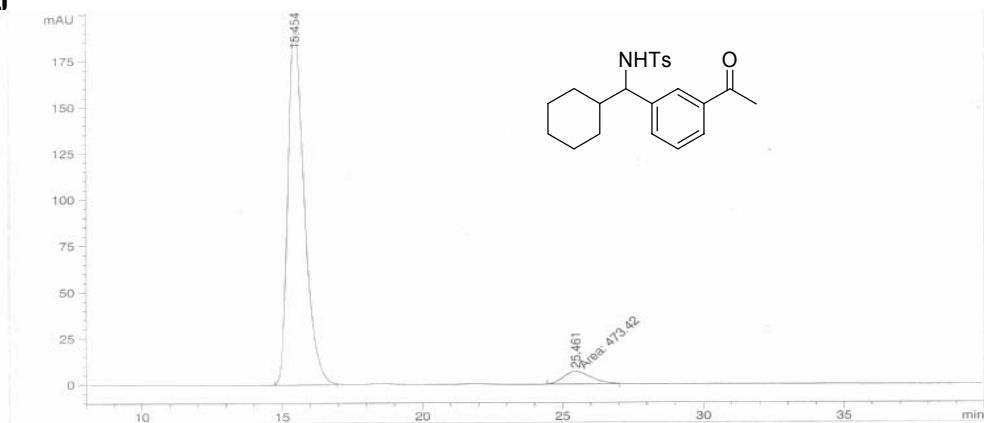
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.796	VV	0.5078	4.16516e4	1240.37769	95.1222
2	18.111	VB	0.6038	2135.85352	53.26540	4.8778

Totals : 4.37874e4 1293.64309

10j



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Area Percent Report
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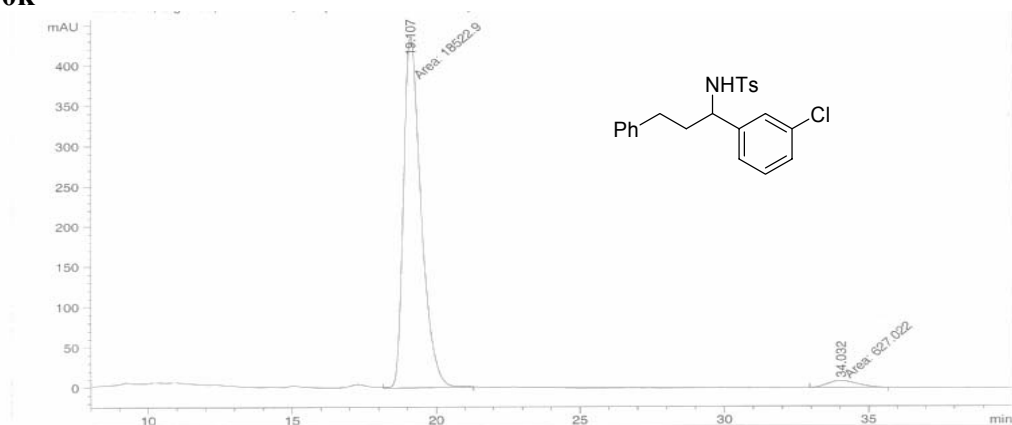
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.454	BB	0.6214	7857.95117	192.03940	94.3176
2	25.461	MM	1.1332	473.41977	6.96293	5.6824

Totals : 8331.37094 199.00233

10k



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Area Percent Report
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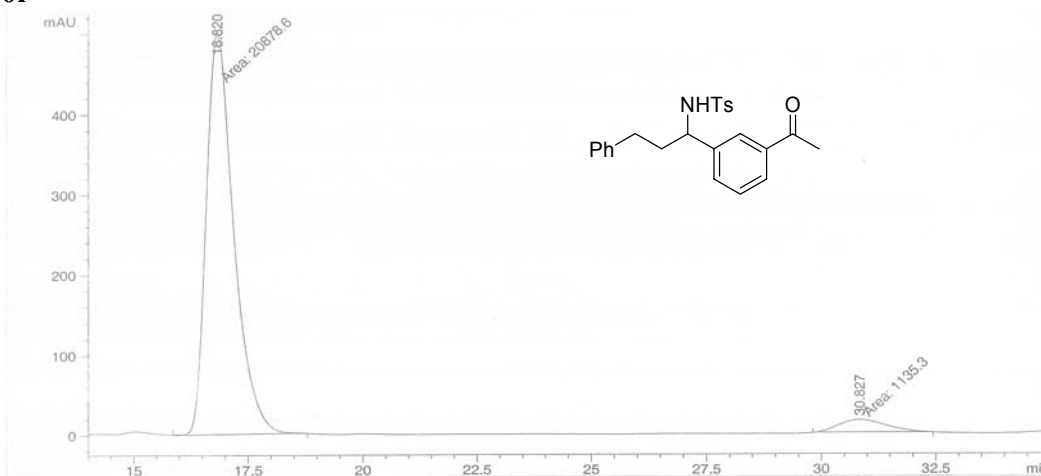
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.107	MM	0.7078	1.85229e4	436.17819	96.7257
2	34.032	MM	1.2384	627.02234	8.43858	3.2743

Totals : 1.91499e4 444.61677

10l



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Area Percent Report
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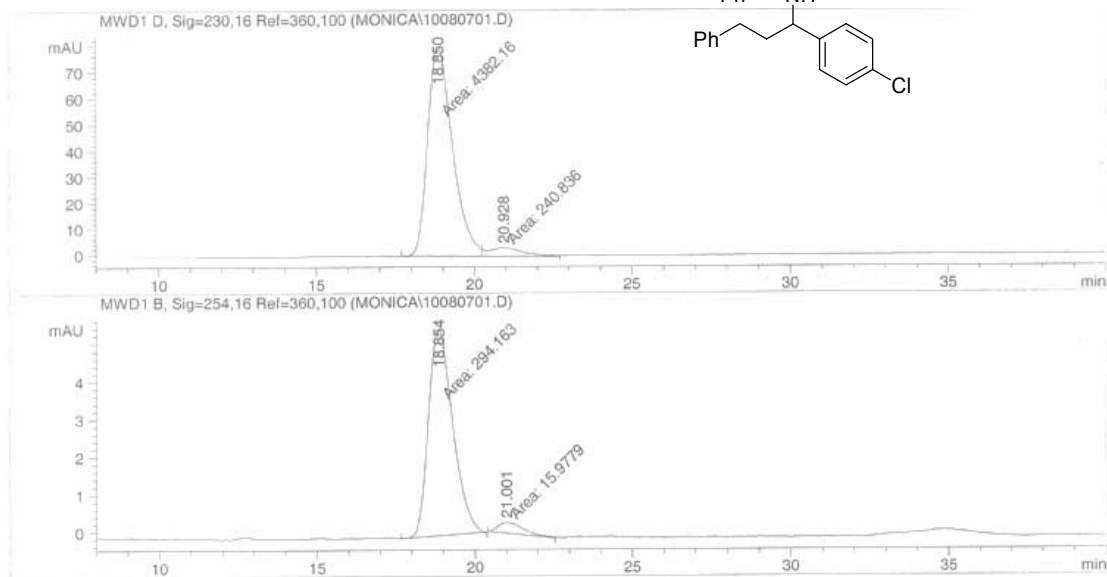
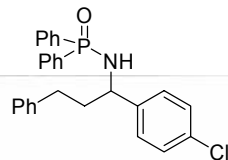
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.820	MM	0.6968	2.08786e4	499.36090	94.8428
2	30.827	MM	1.1962	1135.29797	15.81770	5.1572

Totals : 2.20139e4 515.17860

9a



Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.850	MF	0.9137	4382.16016	79.93278	94.7905
2	20.928	FM	1.2464	240.83580	3.22050	5.2095

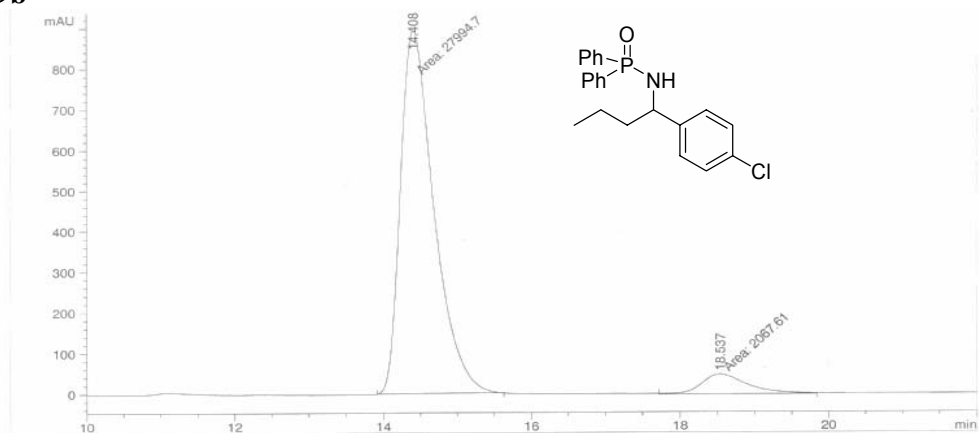
Totals : 4622.99596 83.15328

Signal 2: MWD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.854	MM	0.8968	294.16290	5.46691	94.8482
2	21.001	MM	0.9286	15.97789	2.86766e-1	5.1518

Totals : 310.14080 5.75367

9b



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Area Percent Report
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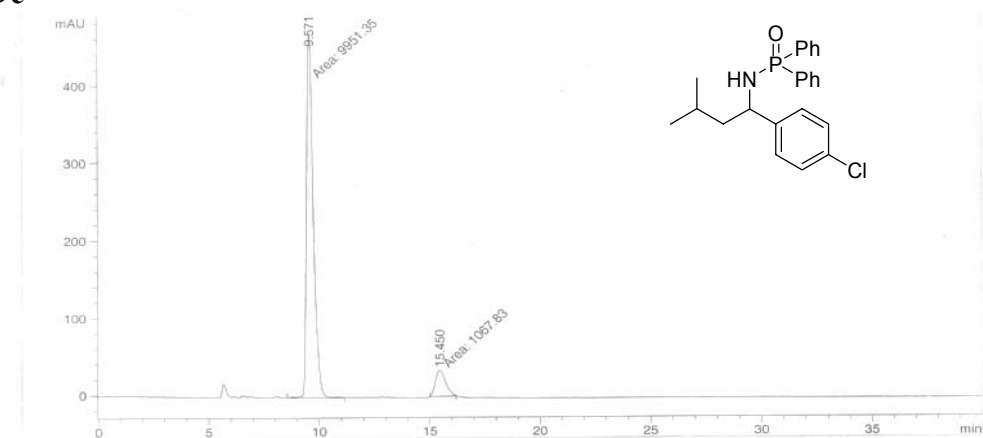
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.408	MM	0.5226	2.79947e4	892.86774	93.1222
2	18.537	MM	0.7076	2067.61182	48.70267	6.8778

Totals : 3.00623e4 941.57040

9c



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Area Percent Report
=====

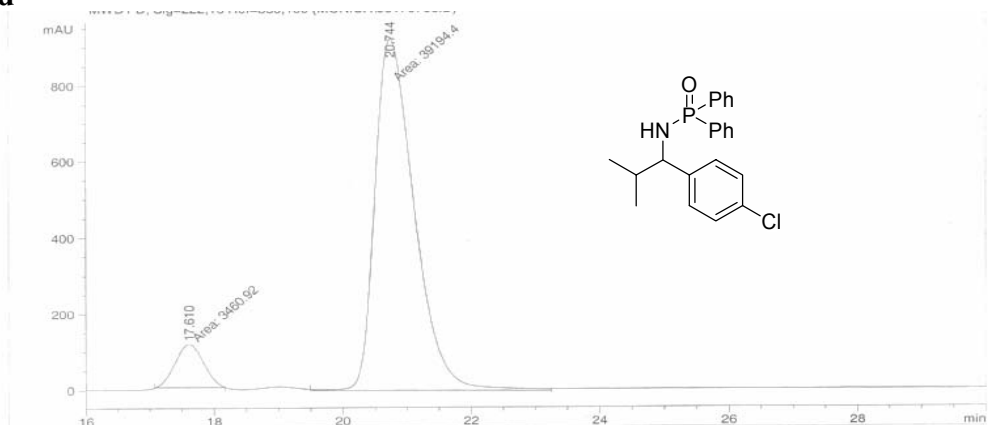
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.571	MM	0.3525	9951.35352	470.49469	90.3094
2	15.450	MM	0.5330	1067.82703	33.39040	9.6906

Totals : 1.10192e4 503.88509

9d



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Area Percent Report
=====

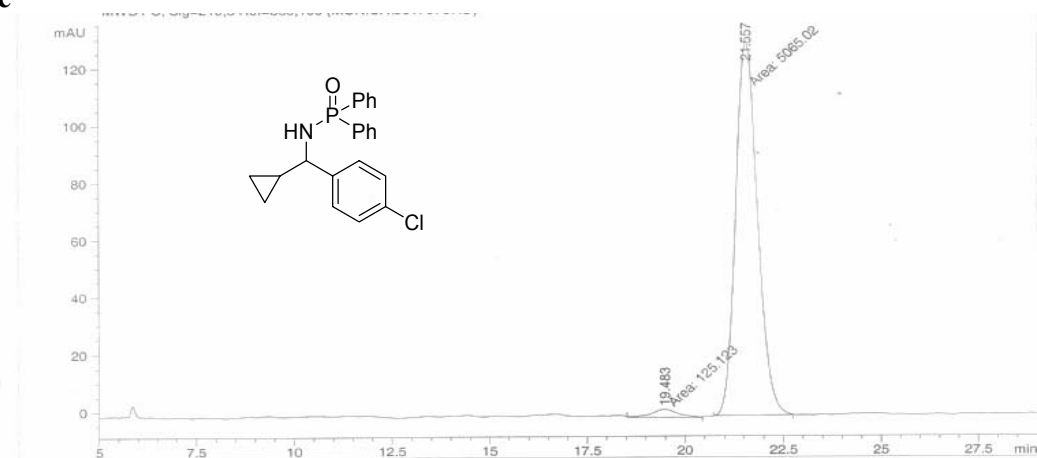
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=222,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.610	MM	0.5099	3460.92041	113.13480	8.1137
2	20.744	MM	0.7076	3.91944e4	923.11694	91.8863

Totals : 4.26554e4 1036.25174

9e



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Area Percent Report
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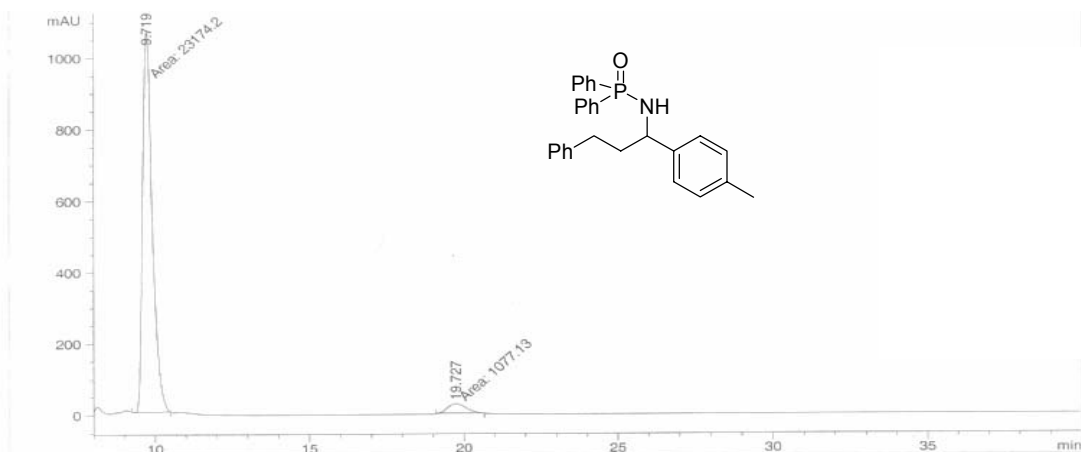
Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.483	MM	0.7622	125.12321	2.73602	2.4108
2	21.557	MM	0.6444	5065.02148	131.00134	97.5892

Totals : 5190.14470 133.73736

9f



Area Percent Report

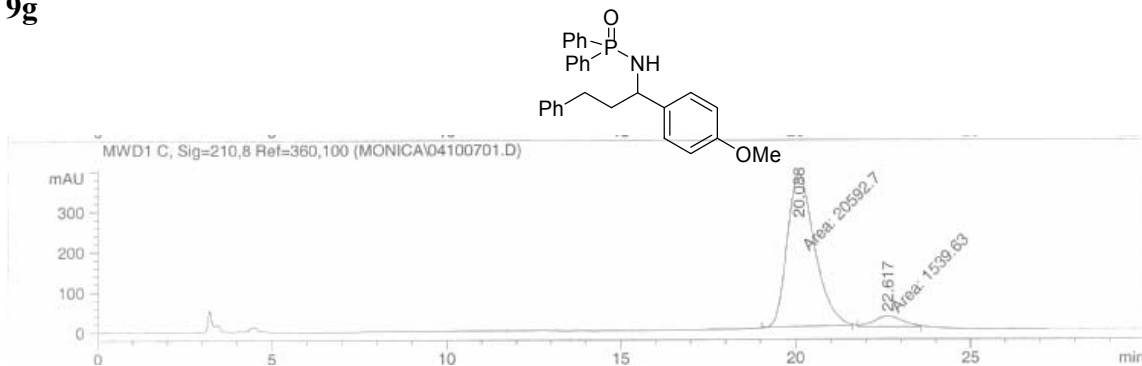
Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.719	MM	0.3629	2.31742e4	1064.24548	95.5585
2	19.727	MM	0.6791	1077.13159	26.43612	4.4415

Totals : 2.42514e4 1090.68160

9g

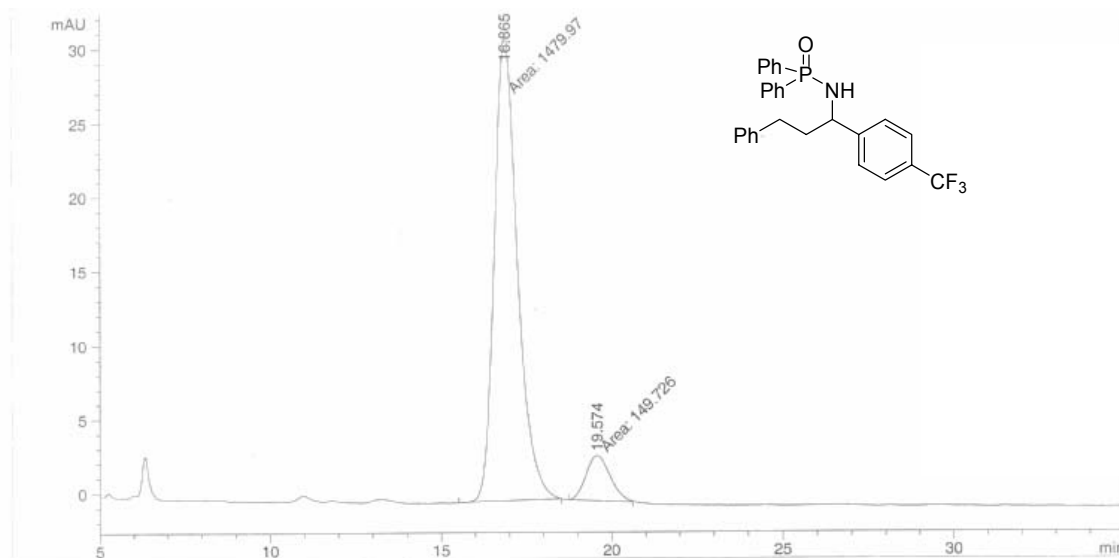


Signal 3: MWD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.088	MM	0.9089	2.05927e4	377.60992	93.0435
2	22.617	MM	0.9724	1539.63428	26.38808	6.9565

Totals : 2.21323e4 403.99801

9h



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Area Percent Report
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Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Signal 1: MWD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.865	MM	0.7873	1479.97034	31.32814	90.8126
2	19.574	MM	0.8181	149.72626	3.05014	9.1874

Totals : 1629.69659 34.37828