



Supporting Information

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Branched-Regioselective Hydroformylation with Catalytic Amounts of a Reversibly Bound Directing Group

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Supporting Information

I General

All reactions were carried out in oven-dried glassware under an atmosphere of argon (Argon 5.0 from Sauerstoffwerke Friedrichshafen). All reagents were obtained commercially unless otherwise noted. 3-Buten-1-ol was purchased from Aldrich, cis-3-Hexen-1-ol and trans-3-Hexen-1-ol from ABCR. All solvents were dried and distilled by standard procedures. Chromatographic purification of products was accomplished using flash chromatography^[1] on Machery-Nagel silica gel 60[®] (230-400 mesh). Hydroformylation reactions were performed in an Endeavor parallel autoclave with 8 reaction vessels from Argonaut Technologies. Gases: Carbon monoxide 3.7, hydrogen 4.3 (1:1, Messer-Griesheim).

Achiral GC analyses were performed on an Agilent 6890N apparatus equipped with a Supelcowax 10 column (30 m x 0.25 mm ID x 0.25 µm film).

Nuclear magnetic resonance spectra were acquired on a Bruker Avance 400 spectrometer (400 MHz and 100 MHz for ¹H and ¹³C respectively), on a Bruker Avance DRX 250 spectrometer and on a Varian Mercury 300 spectrometer. NMR-spectra are referenced internally according to TMS [0.00 ppm (¹H)] or residual protio solvent signals [77.10 ppm for CDCl₃ (¹³C)].^[2] Data for ¹H-NMR are reported as follows: chemical shift (δ in ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; quint, quintet; sext, sextet; hept, heptet; oct, octet; m, multiplet; m_c, symmetrical multiplet; br, broad

signal; *p*, pseudo), integration, coupling constant (Hz). Data for ^{13}C -NMR are reported in terms of chemical shift, integration, coupling constant (Hz). High-resolution mass spectra were obtained on a Finnigan MAT 8200 instrument (EI: 70 eV; CI/NH₃: 110 eV).

II Experimental procedures and characterizations

General procedure (g. p. 1) for the synthesis of (*E*)-alk-3-enoic acids or (*E*)-alk-3-enoic acid-methylesters from aldehydes by a deconjugative Knoevenagel reaction.^[3]

Piperidine (40 μl , 34 mg, 0.40 mmol, 0.02 eq.) and acetic acid (23 μl , 0.40 mmol, 0.02 eq.) was stirred for 5 min. in DMSO (1ml). Afterwards DMSO (20 ml), Malonic acid (4.16 g, 40.0 mmol, 2.0 eq.) [or Malonic acid monomethylester (4.18 ml, 4.72 g, 40.0 mmol) to yield the (*E*)-alk-3-enoic acid-methylesters] and the aldehyde (20 mmol) were added and the reaction mixture was stirred under argon at RT for 20 min. Then the solution was heated to 100 °C for 4-12 h. The solution was cooled to RT and water (40 ml) and diethyl ether (40 ml) were added. The phases were separated and the aqueous phase was extracted with diethyl ether (3 x 40 ml). The combined organic phases were washed with water (3 x 30 ml), dried over Na₂SO₄, filtrated and the solvent was evaporated under reduced pressure. The crude product was purified by column chromatography (ethyl acetate/ cyclohexane).

General procedure (g. p. 2) for the synthesis of homoallylic alcohols from (*E*)-alk-3-enoic acids.

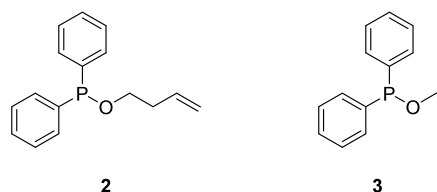
The β,γ -unsaturated acid (9.0 mmol, 1.0 eq.) was dissolved in THF (5 ml), and this solution was added to a suspension of LiAlH₄ (9.0 mmol, 1.0 eq.) in THF (5 ml) at 0 °C. The reaction mixture was allowed to warm to RT and was stirred for further 2 h. The reaction was quenched by addition of aqueous saturated Na₂SO₄ (until the reaction mixture turned white) and additional Na₂SO₄ (solid) was added. The precipitate was filtered off and washed with diethyl ether. The solvent of the filtrate was removed under reduced pressure. The crude product was purified by column chromatography (ethyl acetate/ cyclohexane).

General procedure (g. p. 3) for the synthesis of γ -lactones from homoallylic alcohols (Hydroformylation and PCC oxidation).

The oven dried glass inlet of the autoclave was equipped with molecular sieves (4 Å) and washed with argon. Homoallylic alcohol (0.6 mmol) was added under argon to the glass inlet and dried over the molecular sieves for 30 min. During this time a solution of the ligand $\text{Ph}_2\text{P}(\text{OMe})$ (13.0 mg, 0.06 mmol) in THF (3 ml) was prepared. To this solution $\text{Rh}(\text{CO})_2\text{acac}$ (1.5 mg, 0.006 mmol) was added and the mixture was stirred for 5 min. This mixture was transferred into the glass inlet equipped with the homoallylic alcohol and the molecular sieves. The glass inlet was transferred into the autoclave. The autoclave was sealed and purged three times with 5 bar of the CO/H_2 gas-mixture (1:1). The autoclave was pressurized with 5 bar of CO/H_2 (1:1), heated to 40 °C and pressurized with 20 bar of CO/H_2 (1:1). The reaction mixture was stirred at 40 °C for 12 h.

After pressure release, a sample was taken for GC-analysis. The solvent of the crude reaction mixture was evaporated. The residue was dissolved in CH_2Cl_2 (5 ml) and Al_2O_3 (1.2 g), PCC (256 mg, 1.2 mmol, 2.0 eq.) and NaOAc (25 mg, 0.3 mmol) were added. The reaction mixture was stirred at RT for 12 h, and subsequently filtered through a plug of silica gel. The silica gel was washed with CH_2Cl_2 (70 ml) and for more polar substrates with additional EtOAc (70 ml). The solvent of the filtrate was evaporated and the γ -lactone could be obtained analytically pure without further purification.

Synthesis of Homoallyl-diphenylphosphinite (**2**) and Methyl-diphenylphosphinite (**3**)



The alcohol (5.0 mmol) was dissolved in Et_2O (30 ml). The reaction mixture was cooled to 0 °C and NEt_3 (0.69 ml, 0.51 g, 5.0 mmol) was added. Afterwards ClPPh_2 (0.90 ml, 1.1 g, 5.0 mmol) was added over a period of 10 min. The reaction mixture was allowed to warm up to RT and was filtrated under argon over a plug of Al_2O_3 (neutral). The precipitate was washed with Et_2O (80 ml) and the solvent of combined filtrates was removed in vacuo to give the analytically pure title compounds (1.16 g, 91% for **2**, 919mg, 85% for **3**).

Analytical data for **2**:

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 2.43 (m_c , 2H, 2-H), 3.89 (dt, J = 9.4, 6.8 Hz, 2H, 1-H), 5.05 (m_c , 2H, 4-H), 5.81 (ddt, J = 17.1, 10.3, 6.8 Hz, 3-H), 7.26-7.36 (m, 6H, Ar-H), 7.45-7.52 (m, 4H, Ar-H).

$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 36.0 (d, J = 7.5 Hz, 1C), 69.5 (d, J = 19.6 Hz, 1C), 117.0 (s, 1C), 128.3 (d, J = 7.0 Hz, 4C), 129.3 (s, 2C), 130.4 (d, J = 21.5 Hz, 4C), 134.6 (s, 1C), 142.1 (d, J = 17.9 Hz, 2C).

$^{31}\text{P-NMR}$ (161.976 MHz, CDCl_3): δ = 112.4.

HRMS (CI, NH_3): calculated $(\text{M}+\text{H})^+$: 257.109528, found: 257.109701.

Analytical data for **3**:

$^1\text{H-NMR}$ (250.130 MHz, C_6D_6): δ = 3.39 (d, J = 13.8 Hz, 3H, OMe), 6.98-7.18 (m, 6H, Ar-H), 7.52-7.62 (m, 4H, Ar-H).

$^{13}\text{C-NMR}$ (62.895 MHz, C_6D_6): δ = 56.4 (d, J = 19.9 Hz, 1C, OMe), 128.6 (d, J = 6.4 Hz, 4C), 129.4 (2C), 130.7 (d, J = 22.0 Hz, 4C), 142.6 (d, J = 19.3 Hz, 2C).

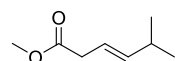
$^{31}\text{P-NMR}$ (101.253 MHz, C_6D_6): δ = 117.0.

Analytical data for **3** were identical to those reported previously.^[4]

Substrate synthesis

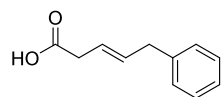
Synthesis of (*E*)-Alk-3-enoic acids

Synthesis of (*E*)-5-Methyl-hex-3-enoic acid methyl ester



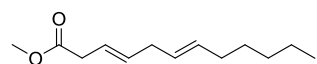
According to general procedure 1, (*E*)-5-Methyl-hex-3-enoic acid methyl ester (1.65 g, 58%) was obtained from commercially available Isovaleraldehyde and Malonic acid monomethylester. The analytical data were identical to those reported previously.^[5]

Synthesis of (*E*)-5-Phenyl-pent-3-enoic acid



According to general procedure 1, (*E*)-5-Phenyl-pent-3-enoic acid (2.60 g, 74%) was obtained from commercially available 3-Phenylpropionaldehyde and Malonic acid. The analytical data were identical to those reported previously.^[6]

Synthesis of (*E,E*)-Dodeca-3,6-dienoic acid methyl ester

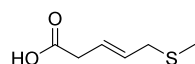


According to general procedure 1, (*E,E*)-Dodeca-3,6-dienoic acid methyl ester (3.36 g, 80%) was obtained from commercially available *trans*-4-Decenal and Malonic acid monomethylester.

¹H-NMR (400.130 MHz, CDCl₃): δ = 0.88 (t, *J* = 7.1 Hz, 3H, CH₃), 1.18-1.42 (m, 6H), 1.9-2.5 (m, 2H), 2.72 (m_c, 2H), 3.02-3.07 (m, 2H), 3.68 (s, 3H, OCH₃), 5.32-5.62 (m, 4H).

¹³C-NMR (100.613 MHz, CDCl₃): δ = 14.1, 22.6, 29.2, 31.5, 32.6, 35.6, 38.0, 51.8, 122.1, 127.6, 131.9, 133.4, 172.6.

Synthesis of (*E*)-5-Methylsulfanyl-pent-3-enoic acid

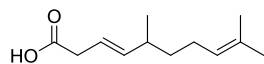


According to general procedure 1, (*E*)-5-Methylsulfanyl-pent-3-enoic acid (1.81 g, 62%) was obtained from commercially available 3-(Methylthio)-propanal and Malonic acid.

¹H-NMR (400.130 MHz, CDCl₃): δ = 2.02 (s, 3H, SCH₃), 3.08-3.12 (m, 2H), 3.13-3.17 (m, 2H), 5.52-5.70 (m, 2H).

¹³C-NMR (100.613 MHz, CDCl₃): δ = 14.4, 35.7, 37.3, 123.9, 130.7, 177.7.

Synthesis of (*E*)-5,9-Dimethyl-deca-3,8-dienoic acid

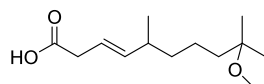


According to general procedure 1, (*E*)-5,9-Dimethyl-deca-3,8-dienoic acid (3.18 g, 81%) was obtained from commercially available Citronellal and Malonic acid.

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 0.98 (d, J = 6.7 Hz, 3H), 1.18-1.34 (m, 2H), 1.58 (d, J = 0.9 Hz, 3H), 1.68 (d, J = 1.3 Hz, 3H), 1.90-2.03 (m, 2H), 2.09-2.20 (m, 1H), 3.07 (d, J = 5.7 Hz, 2H), 5.08 (tqq, J = 7.2, 1.4, 1.4 Hz, 1H), 5.45-5.50 (m, 2H).

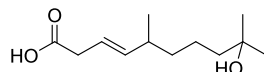
$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 17.7, 20.4, 25.8, 25.8, 36.3, 37.0, 37.8, 119.3, 124.6, 131.4, 141.2, 177.7.

Synthesis of (*E*)-9-Methoxy-5,9-dimethyl-dec-3-enoic acid



According to general procedure 1, (*E*)-9-Methoxy-5,9-dimethyl-dec-3-enoic acid (4.10 g, 90%) was obtained from commercially available 7-Methoxy-3,7-dimethyloctanal and Malonic acid. The analytical data were identical to those reported previously.^[7]

Synthesis of (*E*)-9-Hydroxy-5,9-dimethyl-dec-3-enoic acid



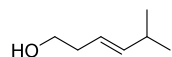
According to general procedure 1, (*E*)-9-Hydroxy-5,9-dimethyl-dec-3-enoic acid (3.64 g, 85%) was obtained from commercially available 3,7-Dimethyl-7-hydroxyoctanal and Malonic acid.

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 0.98 (d, J = 6.7 Hz, 3H), 1.20 (s, 6H), 1.22-1.52 (m, 6H), 2.15 (qdt, J = 6.8, 6.8, 6.6 Hz, 1H), 3.05 (d, J = 6.1 Hz, 2H), 5.38-5.54 (m, 2H), 5.87 (sbr, 2H, COOH, OH).

$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 20.6, 21.9, 29.0, 29.2, 36.7, 37.2, 37.9, 43.7, 71.5, 119.7, 140.9, 177.3.

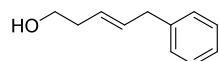
Synthesis of the substrates for hydroformylation

Synthesis of (*E*)-5-Methyl-hex-3-en-1-ol



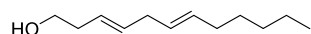
According to general procedure 2, (*E*)-5-Methyl-hex-3-en-1-ol (739 mg, 72%) was obtained by LiAlH_4 reduction of (*E*)-5-Methyl-hex-3-enoic acid methyl ester. The analytical data were identical to those reported previously.^[8]

Synthesis of (*E*)-5-Phenyl-pent-3-en-1-ol



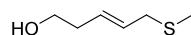
According to general procedure 2, (*E*)-5-Phenyl-pent-3-en-1-ol (1.21 g, 83%) was obtained by LiAlH_4 reduction of (*E*)-5-Phenyl-pent-3-enoic acid. The analytical data were identical to those reported previously.^[9]

Synthesis of (*E,E*)-Dodeca-3,6-dien-1-ol



According to general procedure 2, (*E,E*)-Dodeca-3,6-dien-1-ol (1.19 g, 73%) was obtained by LiAlH_4 reduction of (*E,E*)-Dodeca-3,6-dienoic acid methyl ester. The analytical data were identical to those reported previously.^[10]

Synthesis of (*E*)-5-Methylsulfanyl-pent-3-en-1-ol



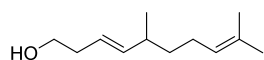
According to general procedure 2, (*E*)-5-Methylsulfanyl-pent-3-en-1-ol (939 mg, 79%) was obtained by LiAlH_4 reduction of (*E*)-5-Methylsulfanyl-pent-3-enoic acid.

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 1.42 (s_{br} , 1H, OH), 2.02 (s, 3H, S- CH_3), 2.30-2.37 (m, 2H, 2-H), 3.07-3.11 (m, 2H, 5-H), 3.63-3.72 (m, 2H, 1-H), 5.47-5.60 (m, 2H, 3-H and 4-H).

$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 14.5, 35.7, 36.0, 62.1, 129.0, 129.3.

HRMS (EI): calculated: 132.060885, found: 132.061103.

Synthesis of (*E*)-5,9-Dimethyl-deca-3,8-dien-1-ol



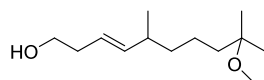
According to general procedure 2, (*E*)-5,9-Dimethyl-deca-3,8-dien-1-ol (1.39 g, 85%) was obtained by LiAlH₄ reduction of (*E*)-5,9-Dimethyl-deca-3,8-dienoic acid.

¹H-NMR (400.132 MHz, CDCl₃): δ = 0.97 (d, *J* = 6.7 Hz, 3H, 5-CH₃), 1.26-1.33 (m, 2H), 1.38-1.45 (m, 1H, OH), 1.59 (d, *J* = 0.9 Hz, 3H, 9-CH₃), 1.68 (d, *J* = 1.3 Hz, 3H, 10-H), 1.90-2.02 (m, 2H), 2.11 (qdt, *J* = 6.9, 6.9, 6.9 Hz, 1H, 5-H), 2.26 (dt, *J* = 6.3, 6.3 Hz, 2H), 3.62 (dt, *J* = 6.1, 6.0 Hz, 2H, 1-H), 5.09 (tqq, *J* = 7.1, 1.4, 1.4 Hz, 1H, 8-H), 5.29-5.38 (m, 1H, 3-H), 5.43 (ddt, *J* = 15.4, 7.3, 1.0 Hz, 1H, 4-H).

¹³C-NMR (100.613 MHz, CDCl₃): δ = 17.8, 20.8, 25.8, 25.9, 36.1, 36.5, 37.2, 62.2, 124.1, 124.7, 131.4, 140.3.

HRMS (CI, NH₃): calculated (M+NH₄)⁺: 200.201439, found: 200.201497.

Synthesis of (*E*)-9-Methoxy-5,9-dimethyl-dec-3-en-1-ol



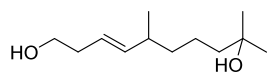
According to general procedure 2, (*E*)-9-Methoxy-5,9-dimethyl-dec-3-en-1-ol (1.52 g, 79%) was obtained by LiAlH₄ reduction of (*E*)-9-Methoxy-5,9-dimethyl-dec-3-enoic acid.

¹H-NMR (400.130 MHz, CDCl₃): δ = 0.96 (d, *J* = 6.7 Hz, 3H, 5-CH₃), 1.11 (s, 6H, 9-CH₃, 10-H), 1.20-1.49 (m, 6H), 1.64 (s_{br}, 1H, OH), 2.11 (dtq, *J* = 6.8, 6.7, 6.7 Hz, 1H, 5-H), 2.25 (dt, *J* = 6.4, 6.3, 2H), 3.15 (s, 3H, OCH₃), 3.60 (t, *J* = 6.3 Hz, 2H, 1-H), 5.28-5.44 (m, 2H, 3-H, 4-H).

¹³C-NMR (100.613 MHz, CDCl₃): δ = 20.9, 21.6, 24.9, 25.0, 36.1, 36.8, 37.5, 40.0, 49.1, 62.2, 74.7, 124.2, 140.1.

HRMS (CI, NH₃): calculated (M+NH₄)⁺: 232.227654, found: 232.227100.

Synthesis of (*E*)-5,9-Dimethyl-dec-3-ene-1,9-diol



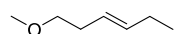
According to general procedure 2, (*E*)-5,9-Dimethyl-dec-3-ene-1,9-diol (1.44 g, 80%) was obtained by LiAlH_4 reduction (in this case 1.5 eq. LiAlH_4 was used) of (*E*)-9-Hydroxy-5,9-dimethyl-dec-3-enoic acid.

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 0.98 (d, J = 6.7 Hz, 3H, 5- CH_3), 1.20 (s, 6H, 9- CH_3 , 10-H), 1.23-1.51 (m, 7H, including OH), 1.67 (s_{br}, 1H, OH), 2.13 (dtq, J = 6.8, 6.8, 6.8 Hz, 1H, 5-H), 2.26 (td, J = 6.2, 6.1 Hz, 2H, 2-H), 3.62 (t, J = 6.3 Hz, 2H, 1-H), 5.30-5.45 (m, 2H, 3-H, 4-H).

$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 20.9, 22.0, 29.2, 29.5, 36.1, 36.8, 37.3, 43.8, 62.2, 71.1, 124.4, 140.0.

HRMS (CI, NH_3): calculated ($\text{M}+\text{NH}_4$)⁺: 218.212004, found: 218.212398.

Synthesis of (*E*)-1-Methoxy-hex-3-ene



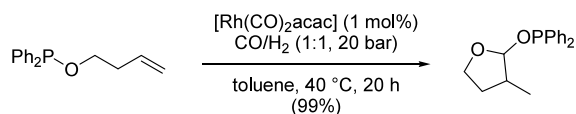
NaH (60% suspension in oil, 300 mg) was washed with THF. The remaining pure NaH (180 mg, 7.50 mmol, 1.5 eq.) was suspended in THF (6 ml). At 0 °C MeI (934 μl , 2.13 g, 15.0 mmol, 3.0 eq.) and *trans*-3-Hexen-1-ol (613 μl , 501 mg, 5.00 mmol) were added. The reaction mixture was allowed to warm up to RT and was stirred for further 16 h. The reaction was quenched with water (5 ml) and the mixture was extracted with Et_2O (3 x 20 ml). The combined organic phases were dried over MgSO_4 and the solvents were removed under reduced pressure. The crude product was purified by column chromatography (pentane: Et_2O = 70:30) to give the title compound (353 mg, 62 %).

$^1\text{H-NMR}$ (400.130 MHz, CDCl_3): δ = 0.97 (t, J = 7.5 Hz, 3H), 2.02 (m_c, 2H), 2.27 (m_c, 2H), 3.34 (s, 3H), 3.39 (t, J = 6.8 Hz, 2H), 5.35-5.45 (m, 1H), 5.50-5.59 (m, 1H).

$^{13}\text{C-NMR}$ (100.613 MHz, CDCl_3): δ = 13.8, 25.7, 33.0, 58.7, 72.8, 125.3, 134.3.

Hydroformylation reactions

Hydroformylation of Homoallyl-diphenylphosphinite (2)



Homoallyl-diphenylphosphinite (**2**) (174 mg, 0.680 mmol) was dissolved in toluene (3.4 ml). $\text{Rh}(\text{CO})_2\text{acac}$ (1.8 mg, 0.0068 mmol) was added and the reaction mixture was transferred to the parallel autoclave. The reaction was heated to 40 °C and pressurized with CO/H_2 (1:1, 20 bar). The reaction mixture was stirred under these conditions for 20 h. After the pressure was released, the reaction mixture was filtrated over Alox N. The solvent was evaporated to yield the title compound (192 mg, 99%). One part of the product was analyzed by NMR. The other part was dissolved in MeOH (3 ml) and Tetrazole (5 mg) was added to liberate the free lactol. This sample was analyzed by GC. Method: 135 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: γ -lactol: 5.03 min, δ -lactol: 6.72 min.

Regioselectivity: 99 : 1.

NMR-analysis of the lactol phosphinites (2 diastereomers) [signals of the other diastereomer, if separated]:

^1H -NMR (400.130 MHz, C_6D_6): δ = 0.67 (d, J = 7.2 Hz, 3H), [0.97 (d, J = 6.7 Hz)], 1.01-2.32 (m, 3H), 3.49-3.96 (m, 2H), 4.90-5.66 (m, 1H), 7.00-7.18 (m, 6H), 7.56-7.70 (m, 4H).

^{13}C -NMR (100.613 MHz, C_6D_6) [doubled signal set due to 2 diastereomers]: δ = 13.3, 17.1, 30.8, 31.2, 40.1 (d, J = 6.3 Hz), 41.4 (d, J = 6.0 Hz), 67.3, 67.6, 106.2 (d, J = 18.1 Hz), 110.6 (d, J = 18.6 Hz), 128.4 (d, J = 6.5 Hz, 2C), 128.4 (d, J = 7.2 Hz, 2C), 128.5 (d, J = 7.0 Hz, 2C), 128.6 (d, J = 11.1 Hz, 2C), 129.1 (2C), 129.2, 129.2, 130.7 (d, J = 21.7 Hz, 2C), 130.8 (d, J = 21.5 Hz, 2C), 130.9 (d, J = 22.2 Hz), 131.0 (d, J = 22.2 Hz), 131.0 (d, J = 22.0 Hz, 2C), 143.1 (d, J = 15.5 Hz), 143.2 (d, J = 15.0 Hz), 143.6 (d, J = 19.3 Hz), 144.0 (d, J = 20.0 Hz).

^{31}P -NMR (121.468 MHz, C_6D_6): δ = 104.1, 104.9.

Synthesis of 3-Methyl-dihydro-furan-2-one



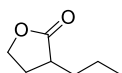
Following general procedure 3, 3-Methyl-dihydro-furan-2-one (54.6 mg, 91%) was obtained on hydroformylation of But-3-ene-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 135 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: But-3-en-1-ol: 2.53 min., γ -lactol: 5.03 min, δ -lactol: 6.72 min.

Conversion: 100%, Regioselectivity: 99 : 1.

The analytical data of the title compound were identical to those reported previously.^[11]

Synthesis of 3-Propyl-dihydro-furan-2-one from *trans*-Hex-3-ene-1-ol



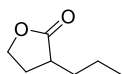
Following general procedure 3, 3-Propyl-dihydro-furan-2-one (65.3 mg, 85%) was obtained on hydroformylation of *trans*-Hex-3-ene-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 155 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: *trans*-Hex-3-ene-1-ol: 2.68 min., γ -lactol: 5.55 min., δ -lactol: 6.08 min.

Conversion: 100%, Regioselectivity: >99 : <1.

The analytical data of the title compound were identical to those reported previously.^[12]

Synthesis of 3-Propyl-dihydro-furan-2-one from *cis*-Hex-3-ene-1-ol



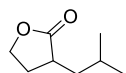
Following general procedure 3, 3-Propyl-dihydro-furan-2-one (67.6 mg, 88%) was obtained on hydroformylation of *cis*-Hex-3-ene-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 155 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: *cis*-Hex-3-ene-1-ol: 2.77 min., γ -lactol: 5.55 min., δ -lactol: 6.08 min.

Conversion: 100%, Regioselectivity: >99 : <1.

The analytical data of the title compound were identical to those reported previously.^[12]

Synthesis of 3-Isobutyl-dihydro-furan-2-one



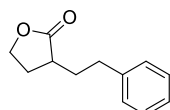
Following general procedure 3, 3-Isobutyl-dihydro-furan-2-one (70.1 mg, 82%, 84% based on conversion) was obtained on hydroformylation of (*E*)-5-Methyl-hex-3-en-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 135 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-5-Methyl-hex-3-en-1-ol: 3.57 min., γ -lactol: 10.73 min., δ -lactol: 12.03 min.

Conversion: 98%, Regioselectivity: 97 : 3.

The analytical data of the title compound were identical to those reported previously.^[13]

Synthesis of 3-Phenethyl-dihydro-furan-2-one



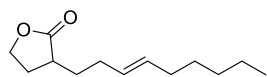
Following general procedure 3, 3-Phenethyl-dihydro-furan-2-one (87.8 mg, 77 %, 92% based on conversion) was obtained on hydroformylation of (*E*)-5-Phenyl-pent-3-en-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 200 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-5-Phenyl-pent-3-en-1-ol: 7.31 min., γ -lactols (the 2 diastereomers are separated): 14.22, 15.09 min., δ -lactols (the 2 diastereomers are separated): 14.02, 15.45 min.

Conversion: 84%, Regioselectivity: 99 : 1.

The analytical data of the title compound were identical to those of the literature.^[14]

Synthesis of 3-Non-3-enyl-dihydro-furan-2-one



Following general procedure 3, 3-Non-3-enyl-dihydro-furan-2-one (84.4 mg, 67%, 83% based on conversion) was obtained on hydroformylation of (*E,E*)-Dodeca-3,6-dien-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 200 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E,E*)-Dodeca-3,6-dien-1-ol: 4.08 min., γ -lactols (the 2 diastereomers are separated): 10.20, 11.38 min., other products including double hydroformylated products: 10.44, 11.17, 11.49 min.

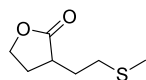
Conversion: 81%, Regioselectivity (γ -lactol vs. all other products): 97 : 3.

¹H-NMR (400.130 MHz, CDCl₃): δ = 0.88 (t, *J* = 7.0 Hz, 3H), 1.20-1.42 (m, 6H), 1.44-1.56 (m, 1H), 1.86-2.24 (m, 6H), 2.34-2.45 (m, 1H), 2.48-2.59 (m, 1H), 4.18 (ddd, *J* = 9.4, 9.3, 6.7 Hz, 1H), 4.34 (ddd, *J* = 8.8, 8.8, 2.9 Hz, 1H), 5.30-5.52 (m, 2H).

¹³C-NMR (100.613 MHz, CDCl₃): δ = 14.1, 22.6, 28.7, 29.2, 30.3 (2C), 31.5, 32.6, 38.6, 66.5, 128.4, 132.2, 179.6.

HRMS (EI): calculated: 210.161980, found: 210.162005

Synthesis of 3-(2-Methylsulfanyl-ethyl)-dihydro-furan-2-one



Following general procedure 3, 3-(2-Methylsulfanyl-ethyl)-dihydro-furan-2-one (30.3 mg, 32%, 42% based on conversion) was obtained on hydroformylation of (*E*)-5-Methylsulfanyl-pent-3-en-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion and regioselectivity: Method: 155 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-5-Methylsulfanyl-pent-3-en-1-ol: 11.00 min., γ -lactols (the 2 diastereomers are separated): 15.33, 15.62 min., δ -lactols (the 2 diastereomers are separated): 15.15, 16.28 min.

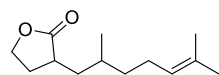
Conversion: 75%, Regioselectivity: 97 : 3.

^1H -NMR (400.130 MHz, CDCl_3): δ = 1.67 (dddd, J = 14.1, 8.6, 7.7, 5.8 Hz, 1H), 1.89 (dddd, J = 12.5, 10.4, 10.3, 8.7 Hz, 1H), 2.05 (s, 3H, SCH_3), 2.06-2.18 (m, 1H), 2.37 (dddd, J = 12.6, 8.8, 6.5, 2.5 Hz, 1H), 2.52 (ddd, J = 13.3, 7.6, 7.6 Hz, 1H), 2.62 (ddd, J = 13.3, 7.8, 5.7 Hz, 1H), 2.70 (dddd, J = 10.7, 8.7, 8.7, 5.3 Hz, 1H), 4.14 (ddd, J = 10.0, 9.1, 6.7 Hz, 1H), 4.30 (ddd, J = 8.8, 8.8, 2.6 Hz, 1H).

^{13}C -NMR (100.613 MHz, CDCl_3): δ = 14.2, 27.7, 28.5, 30.7, 36.9, 65.4, 178.1.

HRMS (CI, NH_3): calculated $(\text{M}+\text{H})^+$: 161.063625, found: 161.063404.

Synthesis of 3-(2,6-Dimethyl-hept-5-enyl)-dihydro-furan-2-one



Following general procedure 3, 3-(2,6-Dimethyl-hept-5-enyl)-dihydro-furan-2-one (116 mg, 92%, 99% based on conversion) was obtained on hydroformylation of (*E*)-5,9-Dimethyl-deca-3,8-dien-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion: Method: 200 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-5,9-Dimethyl-deca-3,8-dien-1-ol: 3.46 min., γ -lactols (3 peaks for 4 diastereomers): 8.54, 9.96, 10.20 min., δ -lactols not detected due to low conversion of PPh_3 hydroformylation. Determination of regioselectivity by ^1H - and ^{13}C -NMR analysis after oxidation to the corresponding lactones, where no δ -lactone was detected.

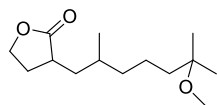
Conversion: 93%, Regioselectivity: 99 : 1.

^1H -NMR (400.130 MHz, CDCl_3) [signals of the other diastereomer, if separated]: δ = 0.92 (d, J = 6.6 Hz, 3H), [0.95 (d, J = 6.7 Hz], 1.08-1.57 (m, 4H), 1.60 (s, 3H), 1.68 (s, 3H), 1.70-2.10 (m, 4H), 2.34-2.46 (m, 1H), 2.51-2.64 (m, 1H), 4.14-4.23 (m, 1H), 4.31-4.39 (m, 1H), 5.08 (ddqq, J = 8.5, 5.7, 1.4, 1.4 Hz, 1H).

^{13}C -NMR (100.613 MHz, CDCl_3) [doubled signal set due to 2 diastereomers]: 17.7 (2C), 18.6, 20.0, 25.3, 25.5, 25.8 (2C), 29.1, 29.6, 30.6, 30.7, 36.0, 37.4 (2C), 37.5, 37.6, 38.1, 66.4, 66.5, 124.5, 124.5, 131.5, 131.6, 179.9, 180.1.

HRMS (EI): calculated: 210.161980, found: 210.161499.

Synthesis of 3-(6-Methoxy-2,6-dimethyl-heptyl)-dihydro-furan-2-one



Following general procedure 3, 3-(6-Methoxy-2,6-dimethyl-heptyl)-dihydro-furan-2-one (126 mg, 86%, 88% based on conversion) was obtained on hydroformylation of (*E*)-9-Methoxy-5,9-dimethyl-dec-3-en-1-ol and PCC-oxidation of the lactol.

GC-analysis for the determination of conversion: Method: 200 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-9-Methoxy-5,9-dimethyl-dec-3-en-1-ol: 5.06 min., γ -lactols (3 peaks for 4 diastereomers): 12.58, 13.30, 13.43 min., δ -lactols not detected due to low conversion of PPh_3 hydroformylation. Determination of regioselectivity by ^1H - and ^{13}C -NMR analysis after oxidation to the corresponding lactones, where no δ -lactone was detected.

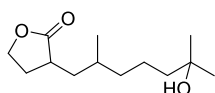
Conversion: 98%, Regioselectivity: 99 : 1.

^1H -NMR (400.130 MHz, CDCl_3) [signals of the other diastereomer, if separated]: δ = 0.85 (d, J = 6.6 Hz, 3H), [0.88 (d, J = 6.6 Hz)], 1.07 (s, 6H), [1.07 (s)], 1.10-1.52 (m, 8H), 1.60-1.69 (m, 1H), 1.76-1.92 (m, 1H), 2.27-2.39 (m, 1H), 2.42-2.58 (m, 1H), 3.10 (s, 3H), [3.10 (s)], 4.08-4.16 (m, 1H), 4.24-4.32 (m, 1H).

^{13}C -NMR (100.613 MHz, CDCl_3) [doubled signal set due to 2 diastereomers]: δ = 17.6, 18.9, 20.0, 20.2, 23.9 (2C), 23.9 (2C), 28.0, 28.6, 30.0, 30.0, 35.6, 36.3 (2C), 36.7, 37.0, 37.1, 39.1, 39.3, 48.1 (2C), 65.4, 65.4, 73.5 (2C), 178.8, 179.0.

HRMS (CI, NH_3): calculated $(\text{M}+\text{H})^+$: 243.196020, found: 243.196102.

Synthesis of 3-(6-Hydroxy-2,6-dimethyl-heptyl)-dihydro-furan-2-one



Following general procedure 3, 3-(6-Hydroxy-2,6-dimethyl-heptyl)-dihydro-furan-2-one (115 mg, 84%, 99% based on conversion) was obtained on hydroformylation of (*E*)-5,9-Dimethyl-dec-3-ene-1,9-diol and PCC-oxidation of the lactol.

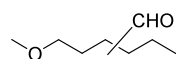
GC-analysis for the determination of conversion: Method: 200 °C, hold 10 min.; 20 °C/min. to 250 °C, hold 15 min. Retention times: (*E*)-5,9-Dimethyl-dec-3-ene-1,9-diol: 8.16 min., γ -lactols (3 peaks for 4 diastereomers): 15.33, 16.41, 16.62 min., δ -lactols not detected due to low conversion of PPh₃ hydroformylation. Determination of regioselectivity by ¹H- and ¹³C-NMR analysis after oxidation to the corresponding lactones, where no δ -lactone was detected. Conversion: 85%, Regioselectivity: 99 : 1.

¹H-NMR (400.130 MHz, CDCl₃) [signals of the other diastereomer, if separated]: δ = 0.92 (d, *J* = 6.4 Hz, 3H), [0.95 (d, *J* = 6.7 Hz)], 1.21 (s, 6H), 1.22-1.79 (m, 10H), 1.82-1.99 (m, 1H), 2.34-2.47 (m, 1H), 2.51-2.65 (m, 1H), 4.14-4.24 (m, 1H), 4.31-4.39 (m, 1H).

¹³C-NMR (100.613 MHz, CDCl₃) [doubled signal set due to 2 diastereomers]: δ = 18.7, 20.0, 21.5, 21.7, 29.1, 29.3, 29.4, 29.4, 29.4, 29.7, 31.0, 31.0, 36.6, 37.3 (2C), 37.7, 38.0, 38.2, 44.1, 44.1, 66.5, 66.5, 71.1 (2C), 179.9, 180.1.

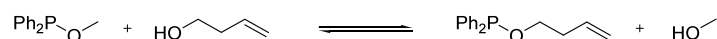
HRMS (CI): calculated (M-CH₃): 213.149070, found: 213.148997.

Hydroformylation of (*E*)-1-Methoxy-hex-3-ene



Following general procedure 3, (*E*)-1-Methoxy-hex-3-ene was hydroformylated and conversion and regioselectivity were determined by GC-analysis. Method: 135 °C, hold 10 min. Retention times: (*E*)-1-Methoxy-hex-3-ene: 2.25 min, hydroformylated products: 3.88, 4.14 min. Conversion: 6%, Regioselectivity: 53 : 47.

Transesterification experiments



Three different experiments were launched to determine the transesterification catalyst. Three flasks were charged each with Methyl-diphenylphosphinite (**3**) (108 mg, 0.50 mmol) in THF (25 ml). To each reaction mixture was added But-3-en-1-ol (434 μ l, 361 mg, 5.00 mmol). To flask 1 Rh(CO)₂acac (12.9 mg, 0.05 mmol) was added. To flask 2 molecular sieves (4 Å) were added. To flask 3 Rh(CO)₂acac (12.9 mg, 0.05 mmol) and molecular sieves (4 Å) were added. The reaction mixtures were heated at 40 °C for 2 ½ h. After cooling to RT the reaction mixtures containing Rh(CO)₂acac were filtrated over Al₂O₃ (neutral) to remove the Rhodium catalyst. The solvents of all reaction mixtures were evaporated and the residue was

analyzed by NMR spectroscopy. The samples (1 and 3) which contained Rh(CO)₂acac showed both phosphinites [Homoallyl-diphenylphosphinite (**2**) as well as Methyl-diphenylphosphinite (**3**)] in a ratio of about 1 : 1. Sample 2 which contained only the molecular sieves showed only the Methyl-diphenylphosphinite (**3**) as the starting material.

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CM306-4010, Gruenanger, C9-273, CDCl3



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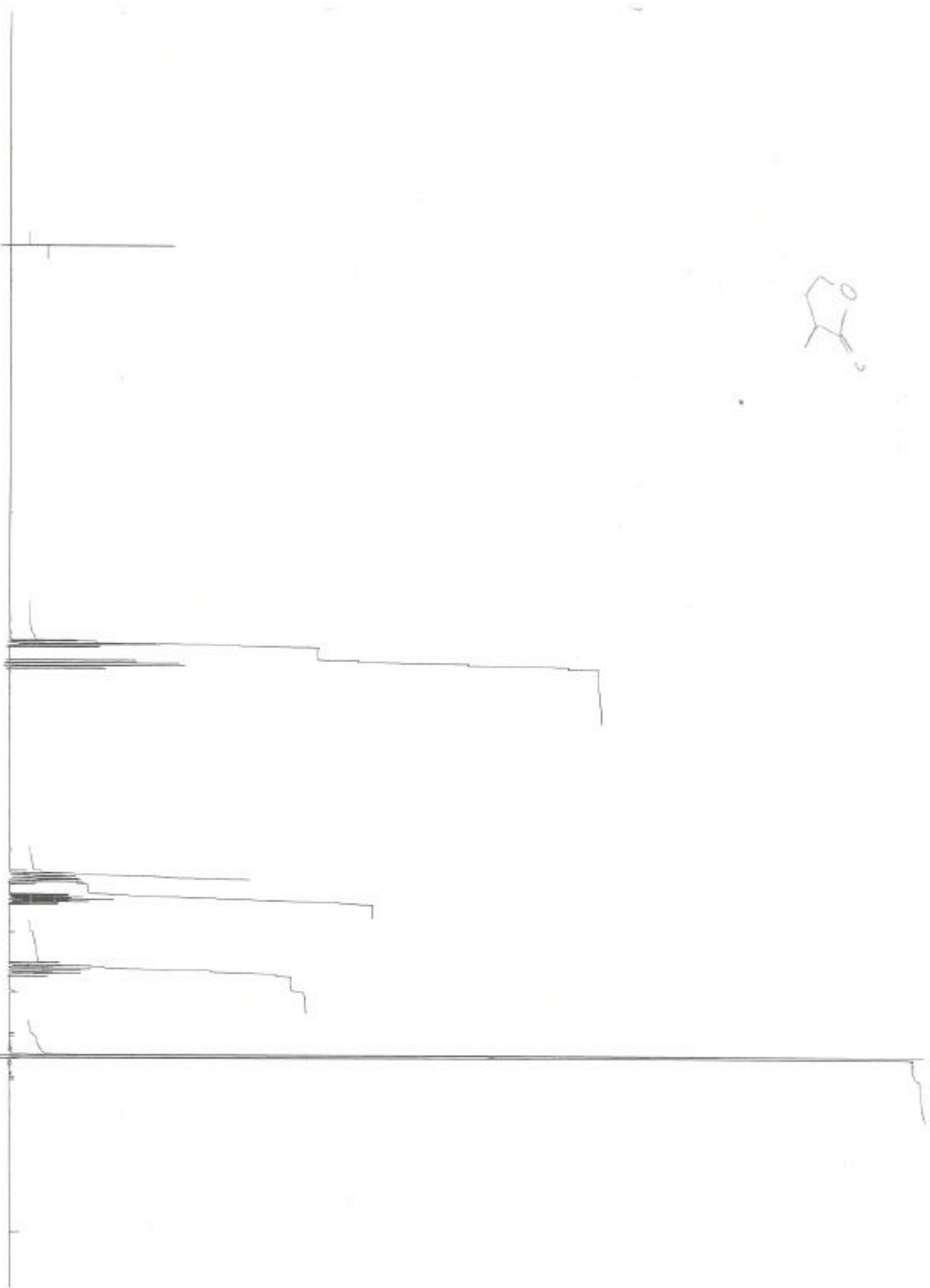
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DS 0.10000000 sec
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WDW EM
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RR

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1	TDO

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SP01 100.6258487 8887

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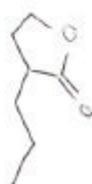
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GPBCTM107-4010, Gruenanger, cg-275, CDCl₃, TMS



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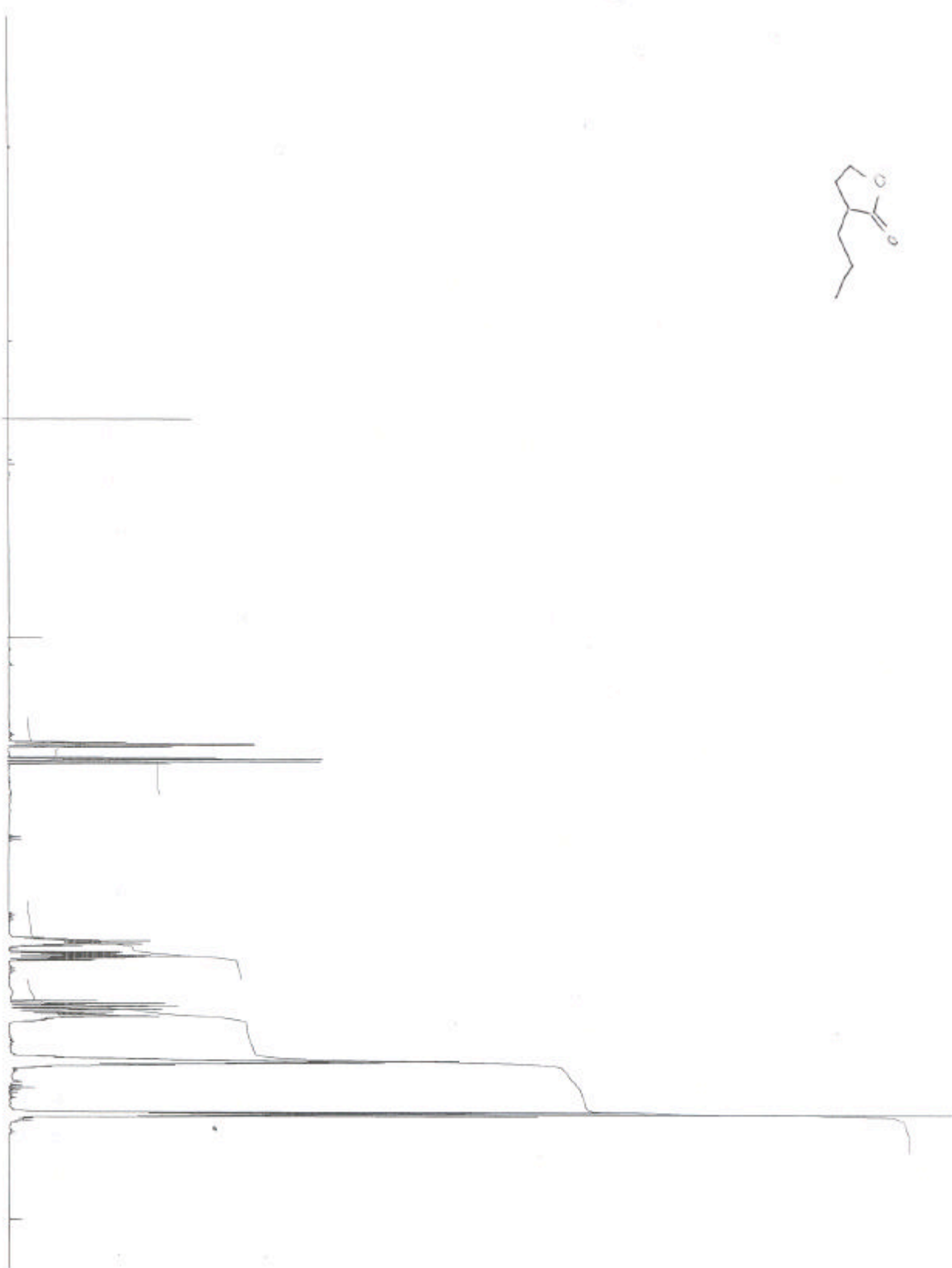
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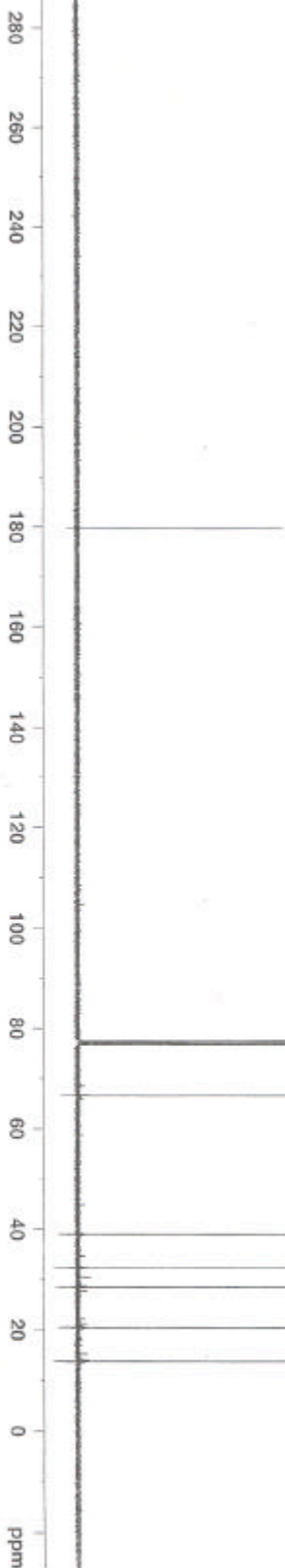
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[illegible]

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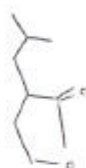
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Wt		15.00 db
Wt		15.00 db
Wt		15.00 db
SP02		400.1320007 kHz
F3 - Processing parameters		
ST		111072
SR		100.6137669 kHz
MDW		CM
SEB		CM
GB		-0.50 Hz
GB		0.1
FC		1.40
ER		-2.05 Hz

02HFM07-4060, Gruenanger, c9-275-2, CDCl3, TM

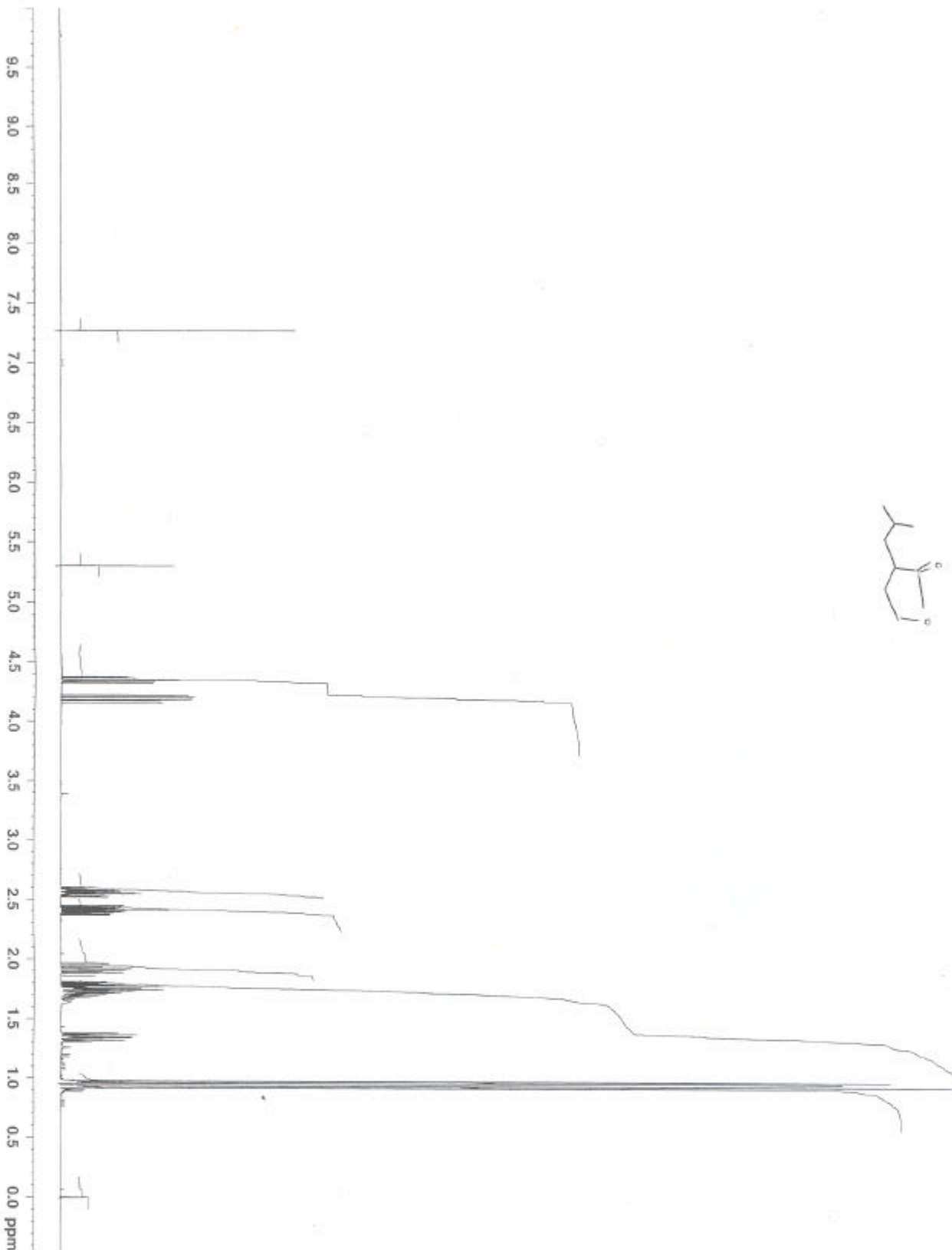


Current Data Parameters
NAME 20080507
EXTRNO 60
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080507
Time 15.03
INSTRUM spect
PROBHD 5 mm BBOBO MM-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 2
DS 2
SWH 8378.146 Hz
FIDRES 0.126114 Hz
AQ 3.9584243 sec
RG 203.2
WDW 50.400 uSec
DE 6.00 uSec
TE 300.0 K
D1 0.10000000 sec
TD0 1

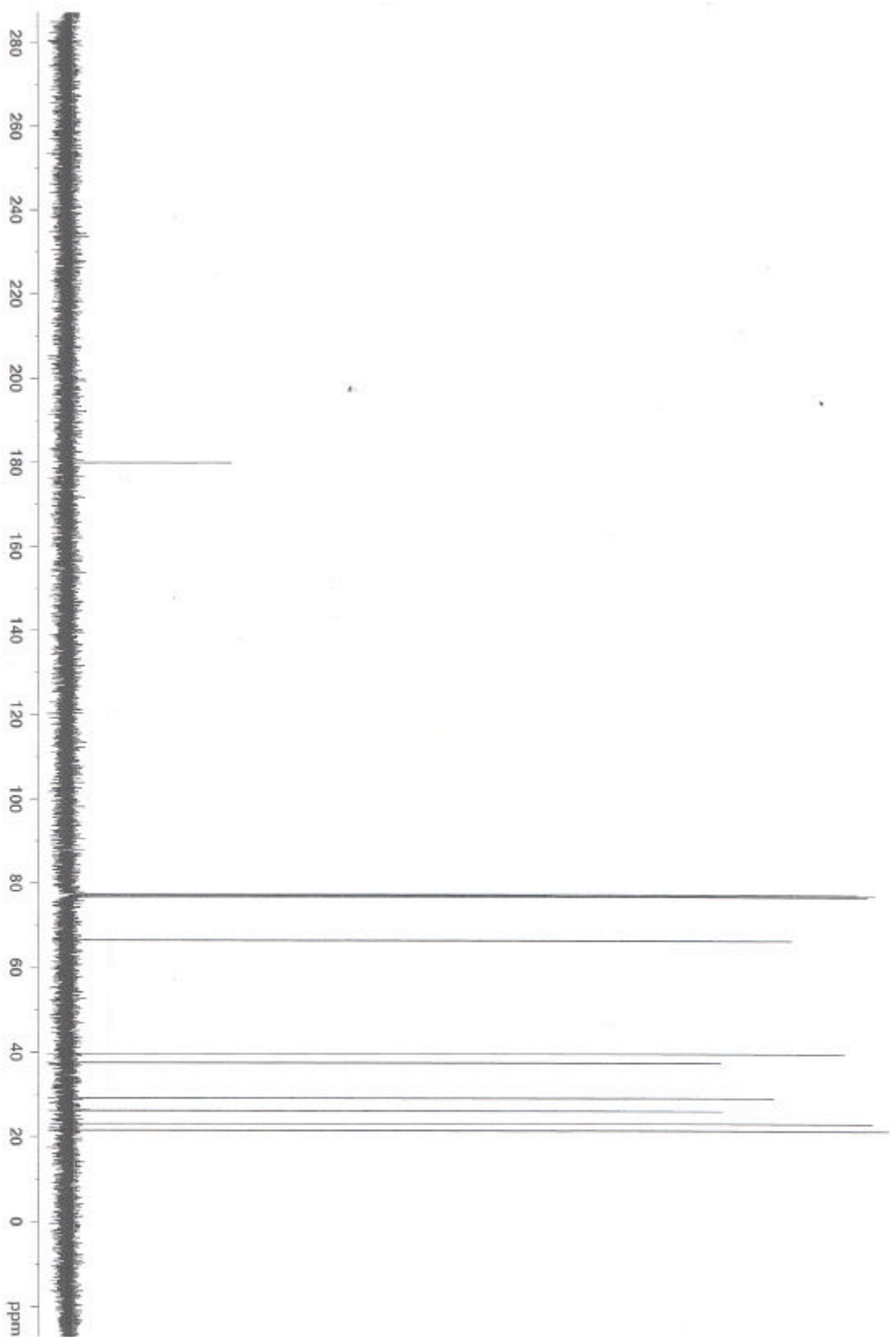
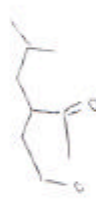
===== CHANNEL f1 =====
NUC1 1H
P1 9.30 uSec
PL1 -3.00 dB
SFO1 400.1324710 MHz

F2 - Processing Parameters
SI 65536
WDW EM
SSB 0
LB -0.30 Hz
GB 0.2
PC 1.00
SC 0.05 Hz



4.07-4.062, Grunanger, c9-276-2, CDCl3, TMS

Channel	Offset	Frequency	Intensity
1	4.062	100.625	100
2	4.062	100.625	100
3	4.062	100.625	100
4	4.062	100.625	100
5	4.062	100.625	100
6	4.062	100.625	100
7	4.062	100.625	100
8	4.062	100.625	100
9	4.062	100.625	100
10	4.062	100.625	100
11	4.062	100.625	100
12	4.062	100.625	100



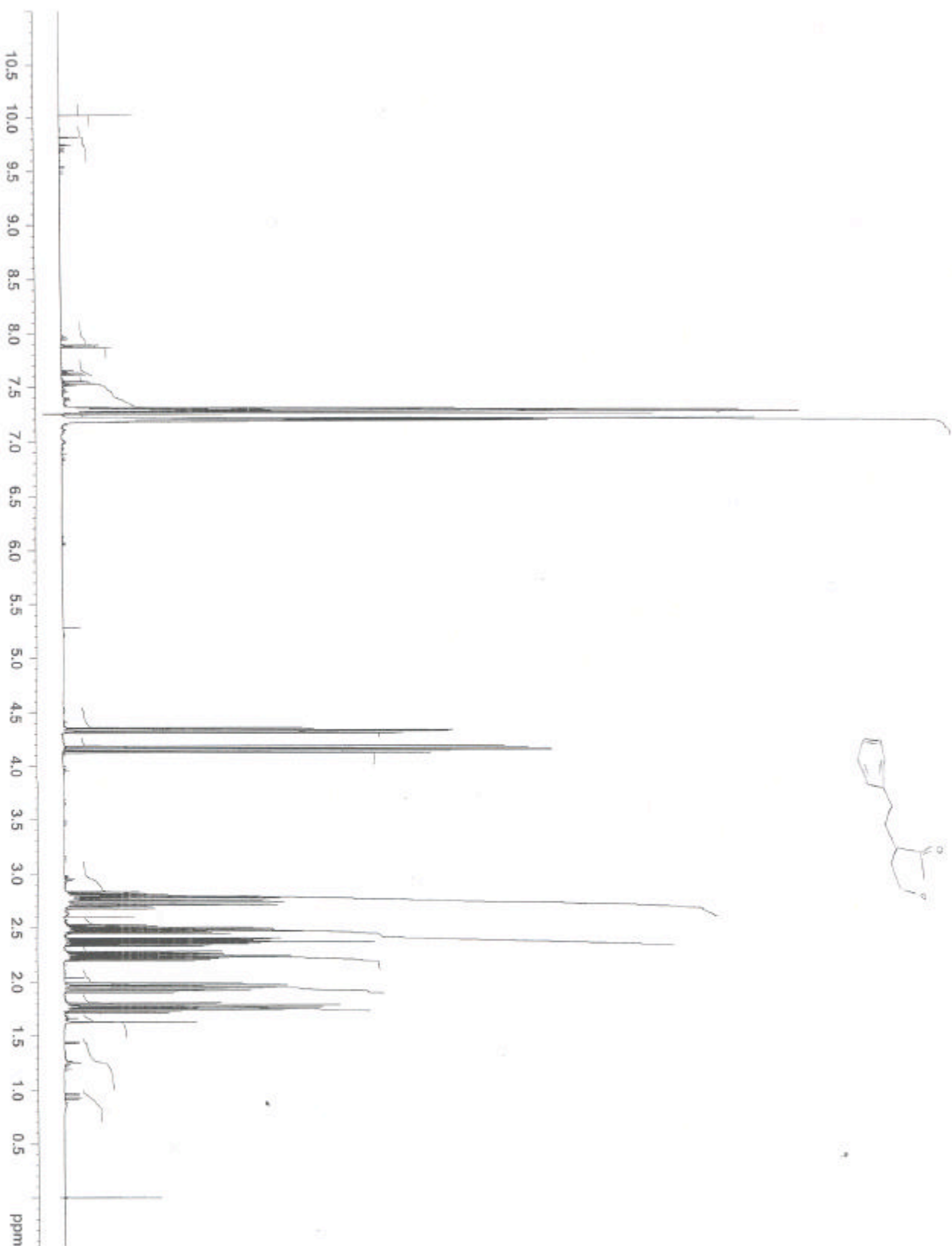
Current Data Parameters
NAME 20080507
EXPNO 52
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080507
Time 15.29
INSTRUM spect
PROBHD 5 mm RABBO HS-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 320
DS 2
SWH 3145.570 Hz
FIDRES 0.482872 Hz
AQ 1.035188 sec
RG 645.1
IN 15.800 usec
DE 6.00 usec
TE 300.0 K
D1 1.0000000 sec
d11 0.0300000 sec
DELTA 0.6999998 sec
TD0 1

===== CHANNEL F1 =====
NUC1 13C
P1 7.90 usec
PL1 -1.00 dB
SFO1 100.625487 MHz

===== CHANNEL F2 =====
CPDPRG22 waltz16
NUC2 1H
PCPD2 90.00 usec
PL2 3.00 dB
PL3 15.00 dB
PL4 11.00 dB
SFO2 400.126087 MHz

F2 - Processing parameters
SI 131072
SF 100.627613 MHz
WDW EM
SSB 0
LB -0.50 Hz
GB 0.1
PC 1.40
SR -7.75 Hz



Current Data Parameters
NAME 200805070
EXNO 70
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080502
Time_ 2:05
INSTRUM spect
PROBHD 5 mm DABO-BB-
PULPROG zg30
TD 65536
ID
SOLVENT CDCl₃
NS 64
DS 2
SWH 0.278, 346 Hz
FIDRES 0.120114 Hz
AQ 3.3584243 sec
RG 181
BO 60.400 usec
DE 38.00 usec
TE 300.2 K
D1 0.1000000 sec
TP0 1

===== CHANNEL f1 =====
NUC1 ¹H
P1 5.00 usec
PL -3.00 dB
SFO1 400.1384710 MHz

F2 - Processing parameters
SI 65536
SF 400.138079 MHz
WDW EM
SSB 0
GB 0
PC 1.00
SR 7.83 MHz

107-4072, Gruenanger, cg-276-5, CDCl3, TMS

Channel	F2 - Processing Parameters	F1 - Acquisition Parameters
1	100.626 MHz	500.135 MHz
2	100.626 MHz	500.135 MHz
3	100.626 MHz	500.135 MHz
4	100.626 MHz	500.135 MHz
5	100.626 MHz	500.135 MHz
6	100.626 MHz	500.135 MHz
7	100.626 MHz	500.135 MHz
8	100.626 MHz	500.135 MHz
9	100.626 MHz	500.135 MHz
10	100.626 MHz	500.135 MHz
11	100.626 MHz	500.135 MHz
12	100.626 MHz	500.135 MHz
13	100.626 MHz	500.135 MHz
14	100.626 MHz	500.135 MHz
15	100.626 MHz	500.135 MHz
16	100.626 MHz	500.135 MHz
17	100.626 MHz	500.135 MHz
18	100.626 MHz	500.135 MHz
19	100.626 MHz	500.135 MHz
20	100.626 MHz	500.135 MHz
21	100.626 MHz	500.135 MHz
22	100.626 MHz	500.135 MHz
23	100.626 MHz	500.135 MHz



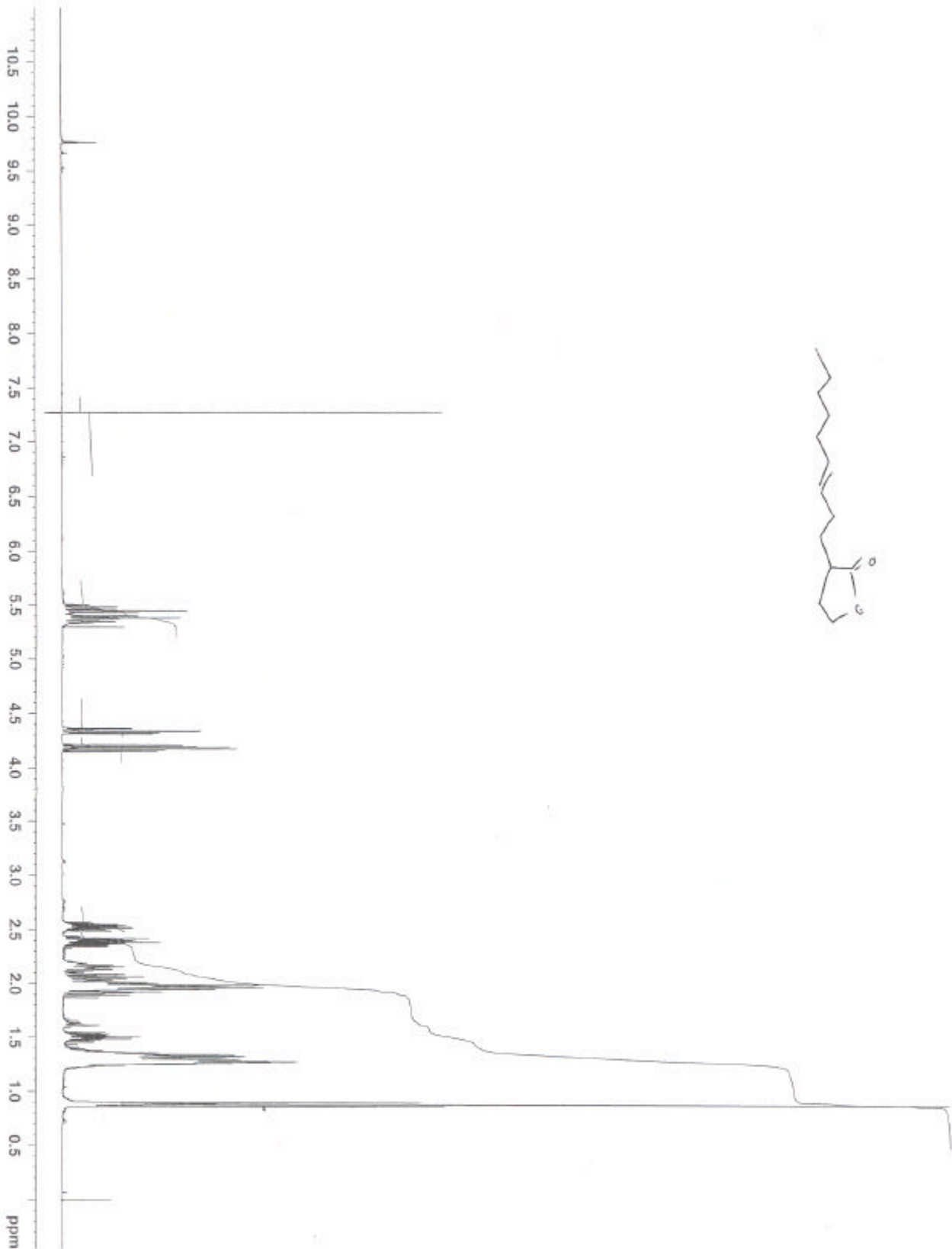
Current Data Parameters
NAME: 100605072
EXPNO: 2
PROCNO: 1

F2 - Acquisition Parameters
Date_: 20080507
Time: 21.36
INSTRUM: spect
PROBHD: 5 mm PABBO-BB-
PULPROG: zgpg30
TD: 65536
SOLVENT: CDCl3
NS: 2000
DS: 2
SWH: 31645.540 MHz
FIDRES: 0.482873 MHz
AQ: 1.035188 sec
RG: 1024
DM: 15.800 usec
DE: 29.8 dB
TE: 300.2 K
D1: 1.0000000 sec
d11: 0.0500000 sec
DELTA: 0.8999998 sec
CPO: 1

===== CHANNEL F1 =====
NUC1: 13C
P1: 7.90 usec
PL1: -1.00 dB
SFO1: 100.6258487 MHz

===== CHANNEL F2 =====
CPDPRG2: waltz16
NUC2: 1H
PCPD2: 60.00 usec
PL2: -3.00 dB
PL12: 15.00 dB
PL13: 15.00 dB
SFO2: 400.1320007 MHz
F2 - Processing Parameters
SI: 131072
SF: 100.6127648 MHz
WDW: EM
SSB: 0
LB: -0.50 Hz
GB: 0
DS: 1.40
PC: -4.25 dB
SR:

GBB108-4060, Gruenanger, cg-277-8, CDCl₃, TM

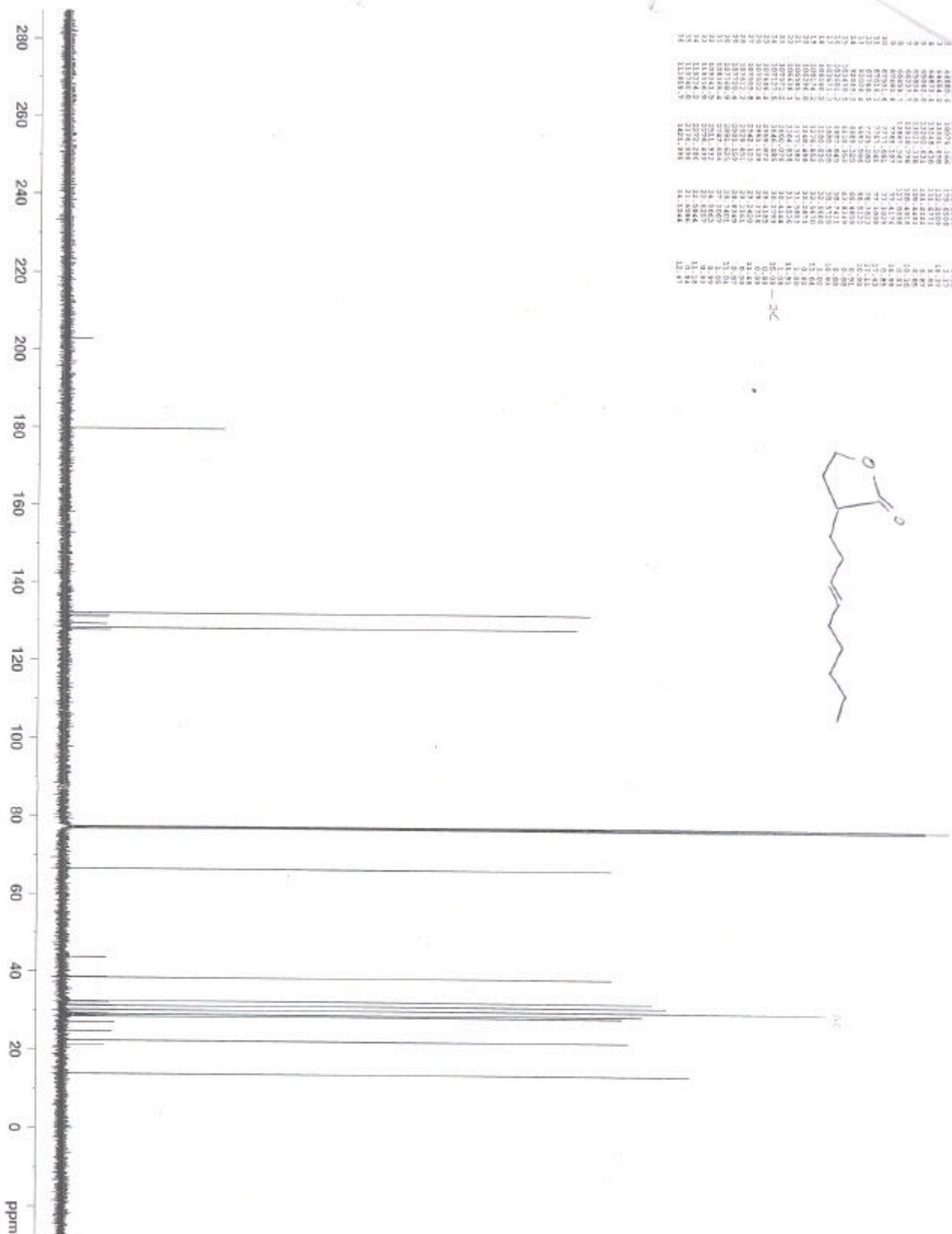


Current Data Parameters
NAME 20080508
EXPNO 60
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080508
Time 14.30
INSTRUM spect
PROBHD 5 mm PABBO BBI
PULPROG zgpg30
TD 65536
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 161.3
IN 60.400 usec
TE 300.0 K
DE 0.10000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.90 usec
PL1 -3.00 dB
SFO1 400.1324710 MHz

F2 - Processing Parameters
SI 65536
SF 400.130018 MHz
WDW EM
SSB 0
GB 0
PC 1.00
DS 1.00
SR 1.77 Hz

[illegible]

G-BTNC1314010, Gruenanger, c9-282-3, CDCl3, TM



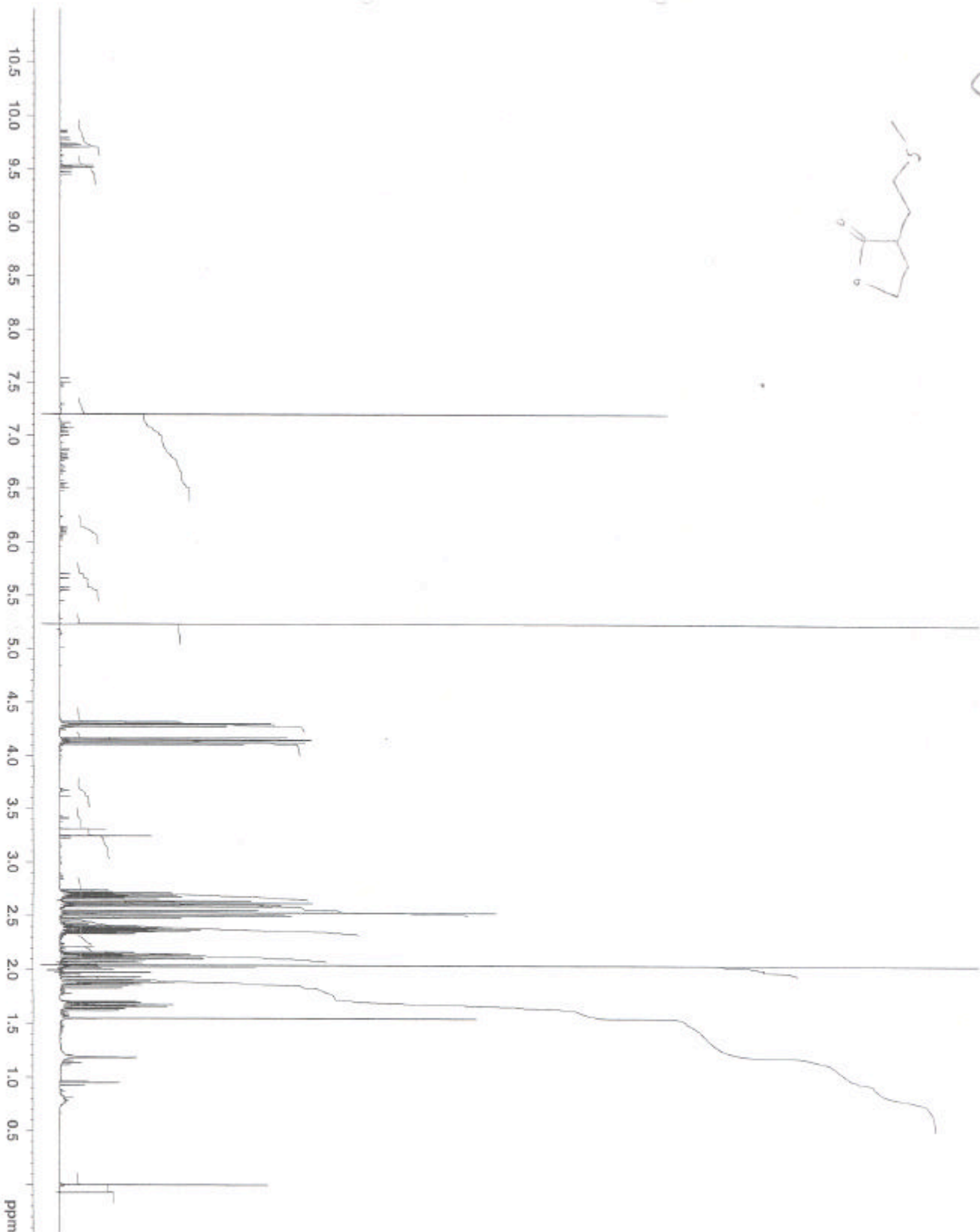
Current Data Parameters
NAME 200805138
EXTNO 10
PROCNO 1

F2 - Acquisition Parameters

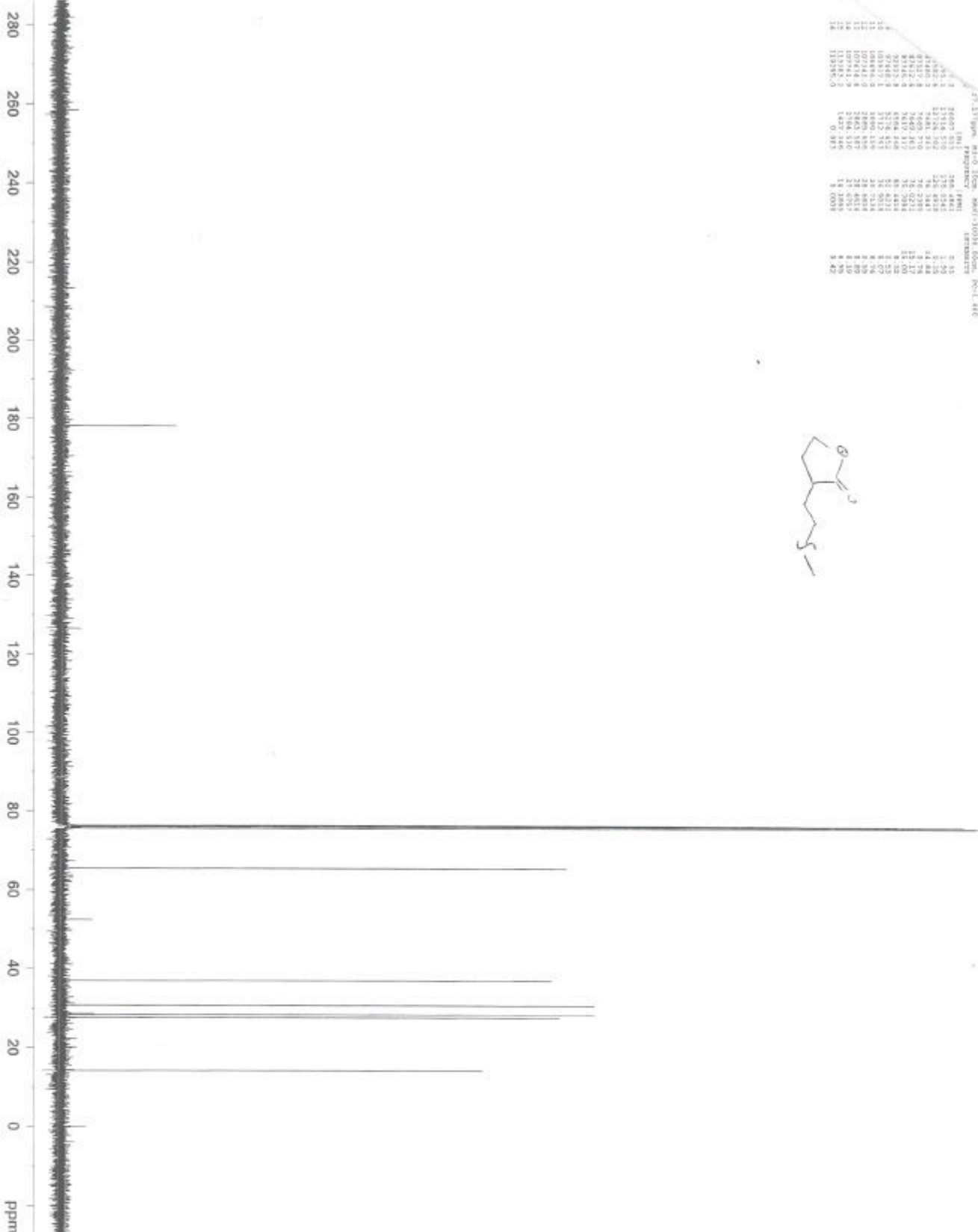
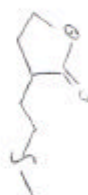
Date_ 20080513
Time 7.40
INSTRUM spect
PROBHD 5 mm PABBO BBO
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 64
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 287.4
IN 60.400 uSec
DE 6.00 uSec
TE 300.0 K
D1 0.10000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.90 uSec
PL1 -3.00 dB
SFO1 400.1324710 MHz

F2 - processing parameters
SI 65536
SF 400.1300288 MHz
WDW GM
SSB 0
LB -0.30 Hz
GB 0
PC 1.00
SC 28.80 Hz
ZM



5

[illegible]

```
Current Data Parameters
NAME          20080513M
EXPNO         12
PROCNO        1
```

```

F2 - Acquisition Parameters
Date_ 20080513
Time 8.22

```

INSTRUM	spec
THORND	5 mm HABO 88-
#10, PRC	29pg30
TD	65536
SOLVENT	CDCl3
NOE	1.024

DO	2
EMR	31645.570 Hz
FIDREF	0.482073 Hz

NO	1.0355188	sec
MC	1625.5	
LM	15.800	use

```

DEC      6.00 used
TDC      300.0 M
D1       1.00000000 dec

```

ALL	0.0300000	sec
DELTA	0.0000000	sec
TDO	1	

```

***** CHANNEL F1 *****
NOCT 13C
BT 7 30 1100

```

PL1	-3.00 dB
SFO1	100.6258487 MHz

```

***** CHANNEL F2 *****
CWDPRG      woltz16
MUC2        1H

```

INCPD2	60.00 u00
PL2	-3.00 dR
PL12	15.00 dR

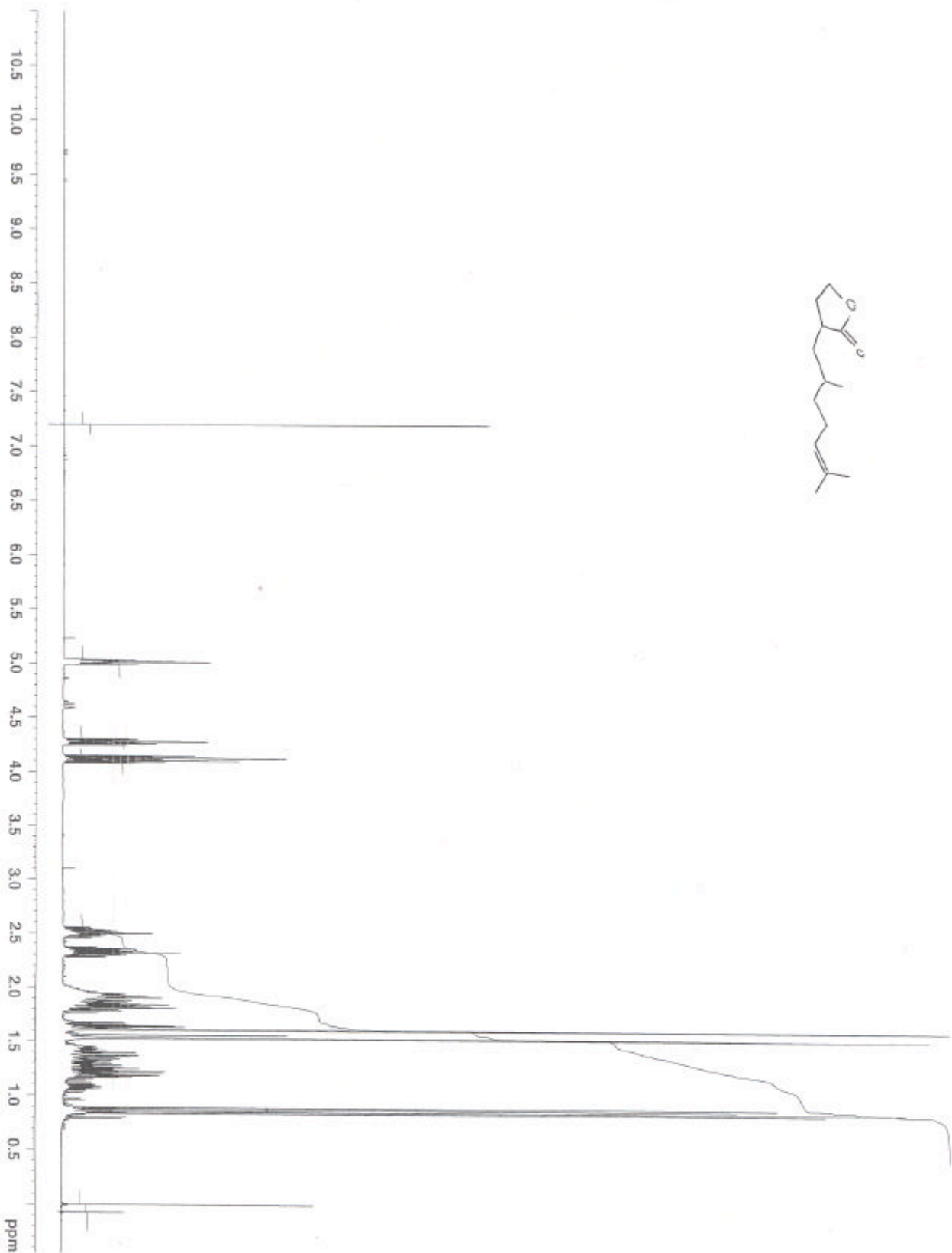
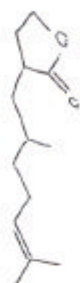
PL13	15.00 GHz
SFO2	400.1320007 MHz

Parameter	Value
Processing parameters	
BT	131072
SR	100,612869 MHz
NTSC	CM

0
-0.50 Hz
0.1

EC	1.40
EN	100.26 Hz

GRBEN 1,3-4020, Gruenanger, CG-281-4, CDCl3, TM



Nutrient Data Parameters
 NAME: 20080513
 EXPNO: 20
 PROCNO: 1
 F2 - Acquisition Parameters
 Date_: 20080513
 Time: 8.55
 INSTRUM: spect
 PROBRD: 5 mm PABBO BB-
 PULPROG: zg30
 VD: 65536
 SOLVENT: CDCl3
 NS: 64
 DS: 2
 SWH: 6278.146 Hz
 FWHM: 0.11148 Hz
 AQ: 3.958443 sec
 RG: 141.7
 INW: 60.400 usec
 DS: 5.00 usec
 GE: 300.0 K
 D1: 0.10000000 sec
 TDO: 1
 ===== CHANNEL f1 =====
 NUCL1: 1H
 P1: 9.30 usec
 PL1: -2.00 dB
 SFO1: 400.1324710 MHz
 F2 - Processing parameters
 SI: 65536
 SF: 400.1300291 MHz
 WDW: EM
 SSF: 0
 LB: -0.30 Hz
 GB: 0.3
 PC: 1.00
 SR: 28.14 Hz

GrbM13-4040, Gruenanger, cg-281-2, CDCl₃, TM

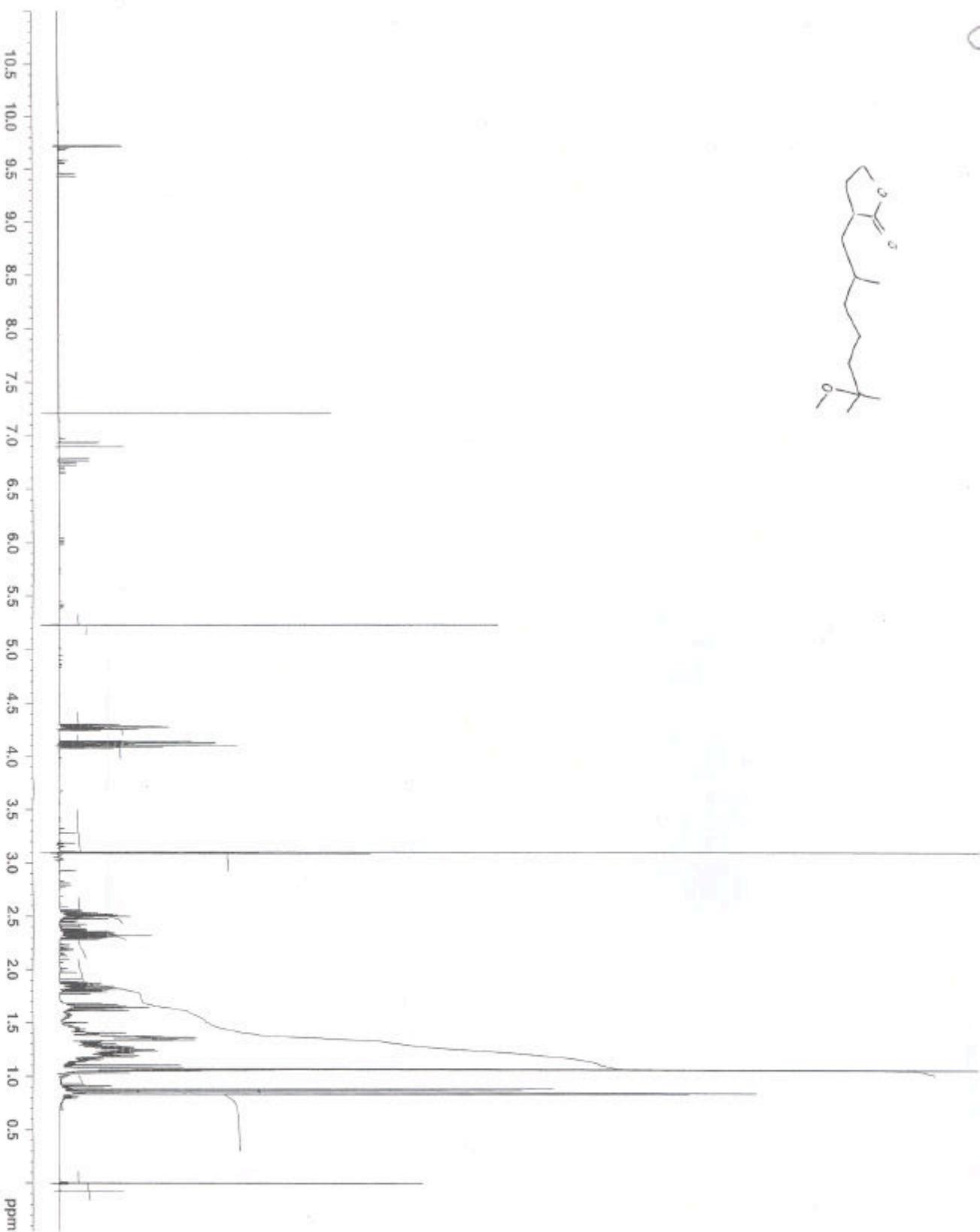


Current Data Parameters
NAME 200805138
EXPNO 40
PROCNO 1

F2 - Acquisition Parameters
Date_ 20080513
Time 10.51
INSTRUM spect
PROBHD 5 mm PABBO BBO
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 64
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 128
W 60.400 usec
DS 4.00 usec
TE 300.0 K
DE 0.10000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.90 usec
PL -3.00 dB
SFO1 400.1324710 MHz

F2 - Processing parameters
SI 65536
SF 400.1300260 MHz
WDW EM
SSB 0
LB -0.30 Hz
GB 0.3
PC 1.00
TC 28.99 Hz



3-4042, Gruenanger, eg-281-2, CDCl3, TMS

NAME: 3-4042, Gruenanger, eg-281-2, CDCl3, TMS
EXPNO: 1
PROCNO: 1
F2 - Acquisition Parameters
Date_ : 20080513
Time : 11.48
INSTRUM : spect
PROBHD : 5 mm PABBO BB-
PULPROG : zgpg30
TD : 65536
SOLVENT : CDCl3
NS : 1024
DS : 2
SWH : 31645.570 Hz
AQ : 0.482972 sec
RG : 1.015108 sec
DE : 11.8200 usec
TE : 300.0 K
FIDRES : 0.0000000 sec
DELTA : 0.8999998 sec
TDO : 1



Current Data Parameters
NAME : 20080513
EXPNO : 42
PROCNO : 1

CH1	F1	13C	=====
1	125.00	125.00	125.00
2	125.00	125.00	125.00
3	125.00	125.00	125.00
4	125.00	125.00	125.00
5	125.00	125.00	125.00
6	125.00	125.00	125.00
7	125.00	125.00	125.00
8	125.00	125.00	125.00
9	125.00	125.00	125.00
10	125.00	125.00	125.00
11	125.00	125.00	125.00
12	125.00	125.00	125.00
13	125.00	125.00	125.00
14	125.00	125.00	125.00
15	125.00	125.00	125.00
16	125.00	125.00	125.00
17	125.00	125.00	125.00
18	125.00	125.00	125.00
19	125.00	125.00	125.00
20	125.00	125.00	125.00
21	125.00	125.00	125.00
22	125.00	125.00	125.00
23	125.00	125.00	125.00
24	125.00	125.00	125.00
25	125.00	125.00	125.00
26	125.00	125.00	125.00
27	125.00	125.00	125.00
28	125.00	125.00	125.00
29	125.00	125.00	125.00
30	125.00	125.00	125.00
31	125.00	125.00	125.00
32	125.00	125.00	125.00
33	125.00	125.00	125.00
34	125.00	125.00	125.00
35	125.00	125.00	125.00
36	125.00	125.00	125.00
37	125.00	125.00	125.00
38	125.00	125.00	125.00
39	125.00	125.00	125.00
40	125.00	125.00	125.00
41	125.00	125.00	125.00
42	125.00	125.00	125.00
43	125.00	125.00	125.00
44	125.00	125.00	125.00
45	125.00	125.00	125.00
46	125.00	125.00	125.00
47	125.00	125.00	125.00
48	125.00	125.00	125.00
49	125.00	125.00	125.00
50	125.00	125.00	125.00
51	125.00	125.00	125.00
52	125.00	125.00	125.00

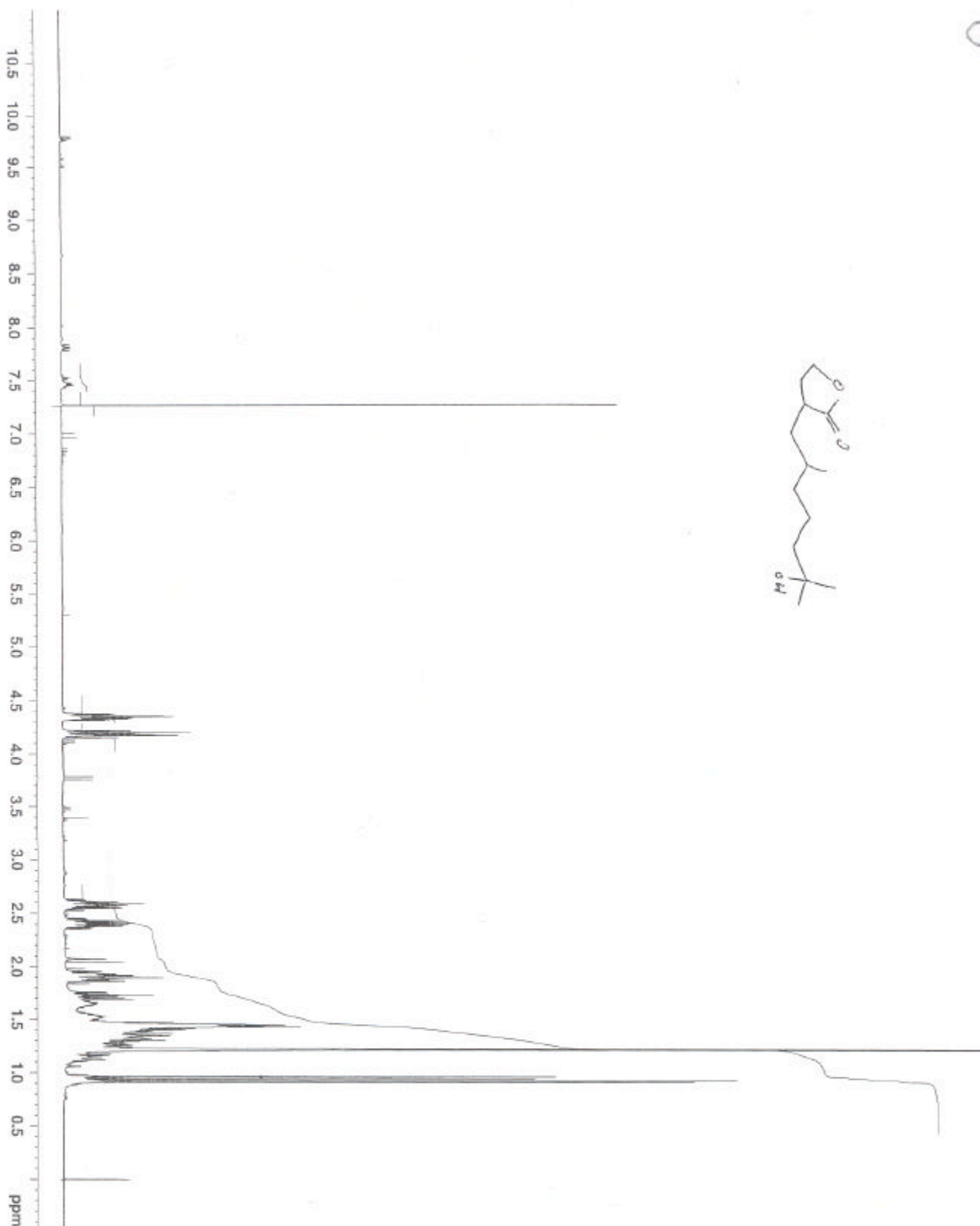


F2 - Processing Parameters
SI : 131072
SF : 100.625881 MHz
WDW : EM
SSB : 0
GB : -0.50 Hz
PC : 1.40
SR : 99.07 Kz

===== CHANNEL F1 =====
NUC1 : 13C
P1 : 7.50 usec
PL : -3.00 dB
SFO1 : 100.625881 MHz

===== CHANNEL F2 =====
CPDPRG2 : waltz16
NUC2 : 1H
PCPD2 : 80.00 usec
PL2 : -3.00 dB
PL12 : 15.00 dB
PL13 : 15.00 dB
SFO2 : 400.132007 MHz

GfBtM14-4030, Gruenanger, cg-283-8, CDCl3, TM

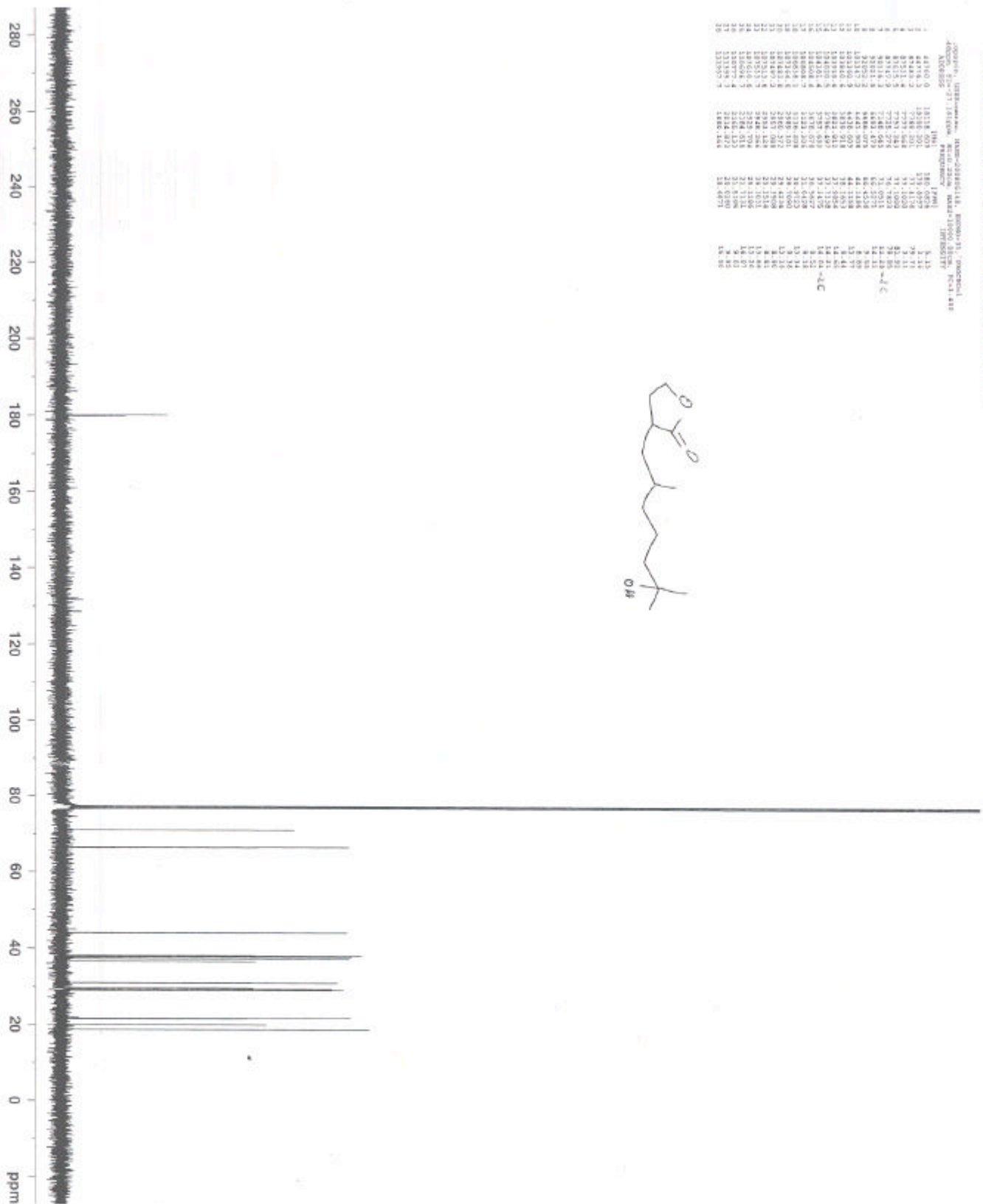


```

Current Data Parameters
NAME      20005148
EXPNO     30
PROCNO    1
F2 - Acquisition Parameters
Date_     20080531
Time      10.20
INSTRUM   spect
PROBHD    5 mm DABO BB-
PULPROG   zgpg30
TD        65536
SOLVENT   CDCl3
NS         64
DS         2
SWH        6270.146 Hz
FIDRES     0.126314 Hz
AQ         3.9584243 sec
RG         60.400 usec
TM         4.000 sec
DE         100.0 K
TE         300.0 K
D1         0.10000000 sec
TDO        1
===== CHANNEL f1 =====
NUC1       1H
P1         9.90 usec
PL         -9.00 dB
SFO1       400.1326710 MHz
F2 - Processing parameters
SI         65536
SF          400.1300910 MHz
WDW         EM
SSB         0
LB          0.3
GB          0
PC          1.00
SR          1.02 Hz
  
```

[illegible]

	1961	1962	1963	1964	1965	1966	1967	1968	1969	1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100	2101	2102	2103	2104	2105	2106	2107	2108	2109	2110	2111	2112	2113	2114	2115	2116	2117	2118	2119	2120	2121	2122	2123	2124	2125	2126	2127	2128	2129	2130	2131	2132	2133	2134	2135	2136	2137	2138	2139	2140	2141	2142	2143	2144	2145	2146	2147	2148	2149	2150	2151	2152	2153	2154	2155	2156	2157	2158	2159	2160	2161	2162	2163	2164	2165	2166	2167	2168	2169	2170	2171	2172	2173	2174	2175	2176	2177	2178	2179	2180	2181	2182	2183	2184	2185	2186	2187	2188	2189	2190	2191	2192	2193	2194	2195	2196	2197	2198	2199	2200	2201	2202	2203	2204	2205	2206	2207	2208	2209	2210	2211	2212	2213	2214	2215	2216	2217	2218	2219	2220	2221	2222	2223	2224	2225	2226	2227	2228	2229	2230	2231	2232	2233	2234	2235	2236	2237	2238	2239	2240	2241	2242	2243	2244	2245	2246	2247	2248	2249	2250	2251	2252	2253	2254	2255	2256	2257	2258	2259	2260	2261	2262	2263	2264	2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275	2276	2277	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306	2307	2308	2309	2310	2311	2312	2313	2314	2315	2316	2317	2318	2319	2320	2321	2322	2323	2324	2325	2326	2327	2328	2329	2330	2331	2332	2333	2334	2335	2336	2337	2338	2339	2340	2341	2342	2343	2344	2345	2346	2347	2348	2349	2350	2351	2352	2353	2354	2355	2356	2357	2358	2359	2360	2361	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390	2391	2392	2393	2394	2395	2396	2397	2398	2399	2400	2401	2402	2403	2404	2405	2406	2407	2408	2409	2410	2411	2412	2413	2
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CURRENT DATE	20080514
NAME	
EXPNO	31
PROCNO	1

#2 - Acquisition Parameters	
Date_____	20080514
Time_____	10.11
INSTRUM_____	
PROBHD_____	5 mm BBOBO BB-
NUMSCA_____	exp230
TD_____	65536
SOLVENT_____	CDCl3
DOS_____	12000
DS_____	2
DEPH_____	33.45 520 Hz
NUC1_____	13C
NUC2_____	400.1300 MHz
AC_____	1.0333168 sec
RG_____	1024
RG_____	15.8000 usec
TS_____	5.00 usec
TE_____	300.0 K
DL1_____	1.00000000 sec
DL1_____	0.01000000 sec
DELTA_____	0.65999998 sec
TD0_____	1

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----- CHANNEL 11 -----
MUC1      13C
P1         7.90 usec
PIL        -3.00 dB
SF01      100.6256487 MHz

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===== CHANNEL #2 =====  
CEDEFB03  
RUCZ2  
WPCP2      80.00 USBC  
PIA2       -3.00 dB  
PLI2       15.00 dB  
PLI3       15.00 dB  
SPC2      600.1370007 MHz
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W1	-	precipitating potential	mm
S1		3.81072	
EP		10.6237605	mm
W2		0	mm
S2		0	
L2		-0.50	mm
C2		0.1	
FC		1.40	
S3		-8.39	mm