



Supporting Information

© Wiley-VCH 2008

69451 Weinheim, Germany

Supporting Information

A General Palladium-Catalyzed Suzuki-Miyaura Coupling of Aryl Mesylates**

Chau Ming So, Chak Po Lau, Albert S. C. Chan and Fuk Yee Kwong*

Open Laboratory of Chirotechnology of the Institute of Molecular Technology for Drug Discovery and Synthesis, Department of Applied Biology and Chemical Technology, The Hong Kong Polytechnic University, Hung Hom, Hong Kong.
E-mail: bcfyk@inet.polyu.edu.hk

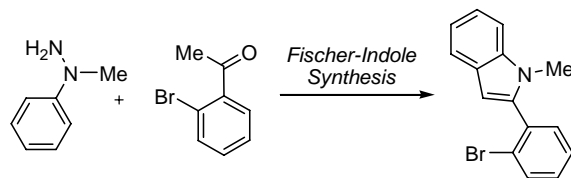
Table of Contents

1. General considerations.....	S2
2. Preparation of new indolyl phosphine ligands L1-L3	S3
3. Preparation of aryl mesylate substrates	S8
4. Procedures for preparation arylboronic acid substrates	S10
5. General procedures/data for initial ligand and reaction condition screenings.....	S12
6. General procedures for Suzuki-Miyaura coupling of aryl mesylates.....	S14
7. Characterization data for coupling products.....	S15
8. Preparation of single-component complex 5	S22
9. Data from aryl tosylate couplings (for comparison).....	S23
10. ¹ H, ¹³ C, ³¹ P NMR, MS, HRMS and IR spectra.....	S24
11. References.....	S122

1. General considerations.

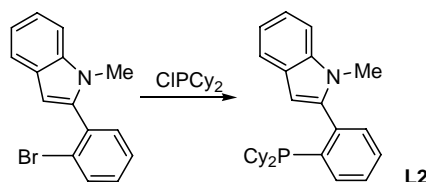
Unless otherwise noted, all reagents were purchased from commercial suppliers and used without purification. All Suzuki-Miyaura reactions were performed in Rotaflo[®] (England) resealable screw cap Schlenk flask (approx. 20 mL volume) in the presence of Teflon coated magnetic stirrer bar (3 mm × 10 mm). Toluene and tetrahydrofuran (THF) were distilled from sodium and sodium benzophenone ketyl under nitrogen, respectively.^[1] *tert*-Butanol was distilled from sodium under nitrogen.^[1] Chlorodiphenylphosphine (Tech grade from Aldrich) was distilled under vacuum prior to use. Most commercially available arylboronic acids were used as received. Some arylboronic acids may require further recrystallization depending on the received conditions. 1-Naphthylboronic acid and 2,4-di-*tert*-butyl-6-methoxyphenylboronic acid were synthesized according to literature methods. New bottle of *n*-butyllithium was used (*Note*: since the concentration of *n*-BuLi from old bottle may vary, we recommend to perform a titration prior to use). K₃PO₄•H₂O and K₃PO₄ were purchased from Fluka. Thin layer chromatography was performed on Merck precoated silica gel 60 F₂₅₄ plates. Silica gel (Merck, 70-230 and 230-400 mesh) was used for column chromatography. Melting points were recorded on an uncorrected Büchi Melting Point B-545 instrument. ¹H NMR spectra were recorded on a Bruker (400 MHz) spectrometer. Spectra were referenced internally to the residual proton resonance in CDCl₃ (δ 7.26 ppm), or with tetramethylsilane (TMS, δ 0.00 ppm) as the internal standard. Chemical shifts (δ) were reported as part per million (ppm) in δ scale downfield from TMS. ¹³C NMR spectra were referenced to CDCl₃ (δ 77.0 ppm, the middle peak). ³¹P NMR spectra were referenced to 85% H₃PO₄ externally. Coupling constants (*J*) were reported in Hertz (Hz). Mass spectra (EI-MS and ES-MS) were recorded on a HP 5989B Mass Spectrometer. High-resolution mass spectra (HRMS) were obtained on a Brüker APEX 47e FT-ICR mass spectrometer (ESIMS). GC-MS analysis was conducted on a HP 5973 GCD system using a HP5MS column (30 m × 0.25 mm). The products described in GC yield were accorded to the authentic samples/dodecane calibration standard from HP 6890 GC-FID system. All yields reported refer to isolated yield of compounds estimated to be greater than 95% purity as determined by capillary gas chromatography (GC) or ¹H NMR. Compounds described in the literature were characterized by comparison of their ¹H, and/or ¹³C NMR spectra to the previously reported data. The procedures in this section are representative, and thus the yields may differ from those reported in tables.

2. Preparation of new indolyl phosphine ligands L1-L4



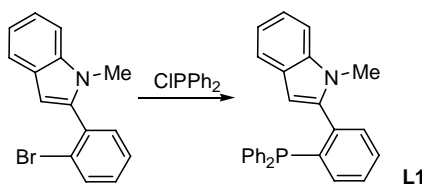
General procedure for Fischer-indole synthesis: 2'-Bromoacetophenone (1.31 mL, 10 mmol) was mixed with *N*-methylphenylhydrazine (1.3 mL, 11 mmol) in phosphoric acid (5 mL) and stirred at room temperature for 30 min. PPA (25-30 g) was added to the mixture and an exothermic reaction ensued whereupon the mixture was heated slowly to 120 °C and kept at this temperature for 1 h. The mixture was poured into ice water and then extracted with Et₂O (3 × ~150 mL). The organic phases were combined, dried over Na₂SO₄ and concentrated under reduced pressure. The crude product was filtered through a short silica pad (3 × ~10 cm) and washed with hexane then EA/Hexane (1:9). The solution was evaporated to yield a light yellow solid. Small amount of cold hexane was used to further wash the product. The product was then dried under vacuum to afford *N*-methyl-2-(2'-bromophenyl)indole (2.35 g, 95%) as a light yellow solid. Melting point. 85.5-87.5 °C; ¹H NMR (400 MHz, CDCl₃) δ 3.61 (s, 3H), 6.56 (s, 1H), 7.20-7.45 (m, 6H), 7.70-7.75 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 30.5, 102.0, 109.4, 119.7, 120.6, 121.6, 125.0, 127.1, 127.5 130.0, 132.6 132.7, 134.1, 137.1, 139.5; IR (cm⁻¹) 3049.43, 2933.63, 1536.35, 1457.76, 1431.08, 1382.94, 1337.55, 1309.58, 1164.78, 1061.55, 1023.56, 791.60, 749.44, 662.83, 578.63, 536.74, 454.92; MS (EI): *m/z* (relative intensity) 285 (M⁺, 100), 204 (75), 190 (10), 178 (20); HRMS: calcd. for C₁₅H₁₂BrNH⁺: 286.0231, found 286.0280.

N-Methyl-2-(2'- Dicyclohexylphosphinophenyl)indole (L2)



General procedure for ligand synthesis: *N*-methyl-2-(2'-bromophenyl)indole (2.2 g, 7.7 mmol) was dissolved in freshly distilled THF (25 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to -78 °C in dry ice/acetone bath. Titrated *n*-BuLi (8.47 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 30 min at -78 °C, chlorodicyclohexylphosphine (1.87 mL, 8.47 mmol) in THF (5 mL) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After the solvent was removed under vacuum, the product was successively washed with cold MeOH/EtOH mixture. The product was then dried under vacuum. White solid of *N*-methyl-2-(2'-dicyclohexylphosphinophenyl)indole (**L2**) (2.75g, 88%) were obtained. Melting point. 171.9-174.9 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.20-1.80 (m, 22H), 3.53 (s, 3H), 6.44 (s, 1H), 7.15(t, *J*=7.4 Hz, 1H), 7.24-7.27 (m, 1H), 7.36-7.50 (m, 4H), 7.66 (d, *J*=7.7 Hz, 2H); ¹³C NMR (100MHz, CDCl₃) δ 25.5, 26.2, 27.1, 28.9, 29.3, 29.5, 30.1, 30.7, 103.2, 109.3, 119.3, 120.2 121.0, 127.6, 127.8, 128.0, 128.2, 128.4, 131.8, 131.9, 132.7, 136.6 (unresolved complex C-P splittings were observed); ³¹P NMR (162 MHz, C₆D₆) δ -9.87; IR (cm⁻¹) 2422.67, 3050.84, 2924.05, 2846.58, 1445.74, 1384.98, 1338.55, 1309.68, 1264.17, 1123.17, 1001.01, 886.58, 848.87, 769.86, 746.53, 523.77; MS (EI): *m/z* (relative intensity) 403(M⁺, 25), 348 (5), 321 (30), 238 (100), 222 (30), 207 (20); HRMS: calcd. for C₂₇H₃₄NP: 403.2423, found 403.2414.

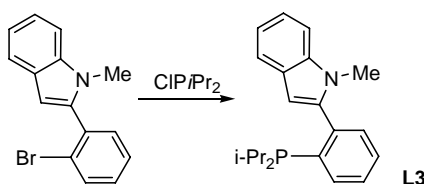
***N*-Methyl-2-(2'-Diphenylphosphinophenyl)indole (**L1**)**



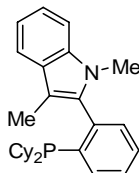
General procedures for the synthesis of ligand **L2** were followed. *N*-Methyl-2-(2'-bromophenyl)indole (1.89 g, 6.6 mmol), *n*-BuLi (6.9 mmol), chlorodiphenylphosphine (1.25 mL, 7.0 mmol) were used to afford *N*-methyl-2-(2'-diphenylphosphinophenyl)indole (**L1**) (1.65 g, 64%) as a white crystal. Melting point. 122.0-125.5 °C; ¹H NMR (400 MHz, C₆D₆) δ 3.18 (s, 3H), 6.55 (s, 1H), 7.08-7.66 (m, 18H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 30.5, 103.7, 103.8, 109.5, 119.5, 120.4, 121.5, 127.6, 128.5 (overlapped), 128.6, 128.7, 128.8, 131.6 (overlapped), 133.7, 133.8, 133.9, 137.2, 137.4, 137.5, 139.6 (overlapped) (unresolved complex C-P splittings were

observed); ^{31}P NMR (162 MHz, C_6D_6) δ -12.02; IR (cm^{-1}) 3446.06, 3048.19, 2931.48, 1582.52, 1461.53, 1429.92, 1377.43, 1359.35, 1336.25, 1308.54, 1235.94, 1176.72, 1123.94, 1093.80, 1025.26, 999.97, 924.13, 943.98, 769.18, 745.33, 695.27, 539.41, 495.26; MS (EI): m/z (relative intensity) 391 (M^+ , 100), 376 (50), 314 (30), 298 (27), 281 (5), 261 (5), 236 (50), 222 (40), 204 (25); HRMS: calcd. for $\text{C}_{27}\text{H}_{22}\text{NPH}^+$: 392.1568, found 392.1579.

***N*-Methyl-2-(2'-Diisopropylphosphinophenyl)indole (**L3**)**



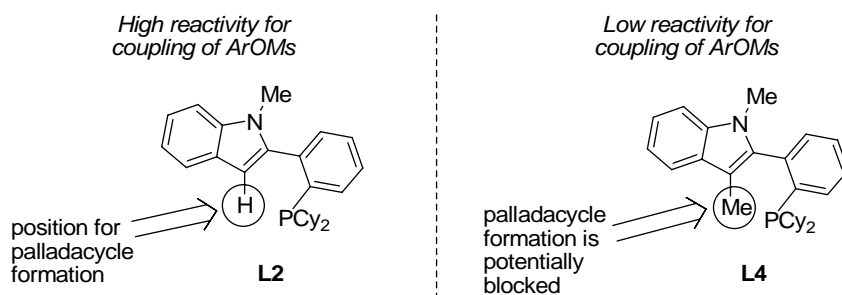
General procedures for the synthesis of ligand **L2** were followed. *N*-Methyl-2-(2'-bromophenyl)indole (1.66 g, 5.8 mmol), *n*-BuLi (6.38 mmol), chlorodiisopropylphosphine (1.03 mL, 6.4 mmol) were used to afford *N*-methyl-2-(2'-diisopropylphosphinophenyl)indole (**L3**) (1.41 g, 75%) as white crystal. Melting point. 146.1-147.8 °C; ^1H NMR (400 MHz, CD_2Cl_2) δ 1.06-1.15 (m, 12H), 1.80-2.80 (m, 2H), 3.61 (s, 3H), 6.52 (s, 1H), 7.23-7.77 (m, 8H); ^{13}C NMR (100 MHz, CD_2Cl_2) δ 20.0, 20.2, 23.3, 25.7 (overlapped), 30.8 (overlapped), 103.3 (overlapped), 109.4, 119.4, 120.1, 121.1, 127.8, 128.2, 128.4, 131.7, 131.8, 132.5, 132.6, 136.7, 137.9, 138.1, 140.6, 140.9, 141.5, 141.5 (overlapped) (unresolved complex C-P splitting was observed); ^{31}P NMR (162 MHz, CD_2Cl_2) δ -1.29; IR (cm^{-1}) 3445.89, 3050.50, 2941.77, 2860.55, 1542.40, 1454.90, 1420.35, 1380.67, 1336.26, 1309.58, 1237.42, 1119.75, 1005.64, 880.09, 778.80, 747.64, 676.90, 653.14, 606.16, 583.14, 531.39, 459.25; MS (EI): m/z (relative intensity) 323 (M^+ , 20), 280 (100), 236 (40), 222 (30), 204 (5); HRMS: calcd. for $\text{C}_{21}\text{H}_{26}\text{NP}$: 323.1797, found 323.1803.

Independent synthesis of ligand **L4** for a **L2/L4** ligand comparison study.**1,3-Dimethyl-2-(2'-dicyclohexylphosphinophenyl)indole (**L4**).**

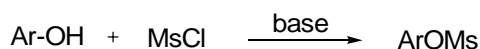
1,3-Dimethylindole (1.45 g, 10 mmol) was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to 0 °C in ice water bath. Titrated *t*-BuLi (10 mmol) was added dropwise by syringe. After the reaction mixture was stirred for 10 min at 0 °C, 2-bromochlorobenzene (1.2 mL, 10 mmol) was added dropwise by syringe and the solution was stirred for further 30 min. The solvent was removed under vacuum. Diethyl ether (150 mL) was added and washed with brine. The solution was concentrated and subjected to column chromatography and eluted with DCM:Hexane = 2:8. The fractions that are containing the 1,3-dimethylindole and 1,3-dimethyl 2-(2-bromophenyl)indole were concentrated and dried by vacuum. The amount of 1,3-dimethylindole and 2-(2-bromophenyl)indole in the mixture was judged by GC. The viscous mixture was dissolved in freshly distilled THF (20 mL) at room temperature under a nitrogen atmosphere. The solution was cooled to -78 °C in dry ice/acetone bath. Titrated *n*-BuLi (1.1 equiv.) was added dropwise by syringe. After the reaction mixture was stirred for 10 min at -78 °C, chlorodicyclohexylphosphine (1.1 equiv.) was added. The reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under reduced pressure. After the solvent was removed under vacuum, the product was washed with cold EtOH. The product was then dried under vacuum. White solid of 1,3-dimethyl-2-(2'-dicyclohexylphosphinophenyl)indole (20% overall yield in 2 steps) were obtained. Melting point. 199.8-201.9 °C; ¹H NMR (400 MHz, CD₂Cl₂) δ 1.22-2.12 (m, 22H), 2.23 (s, 3H), 3.54 (s, 3H), 7.20 (t, *J*=7.3, 1H), 7.28-7.57 (m, 5H), 7.65 (d, *J*=7.6 Hz, 1H), 7.74-7.76 (m, 1H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 10.5, 27.2, 27.3, 27.8, 27.9, 28.0, 28.2, 28.3, 28.4, 29.8, 30.7, 30.9, 31.1, 31.2, 31.7, 33.8, 34.0, 35.7, 35.8, 109.1, 109.7, 119.1, 119.3, 121.8, 128.6, 129.0, 129.1, 133.0, 133.1, 133.7, 137.1, 138.8, 139.0, 140.6, 140.9 (unresolved complex C-P splittings were observed); ³¹P NMR (162 MHz, CD₂Cl₂) δ -8.96; IR (cm⁻¹) 3046.64, 2923.29, 2846.57, 1573.98, 1459.63, 1445.76, 1421.75, 1381.21, 1357.91, 1327.96, 1265.92, 1243.64,

1178.72, 1156.81, 1129.37, 1080.41, 1001.15, 918.68, 881.88, 848.60, 820.68, 760.63, 738.96, 614.63, 552.33, 526.21, 484.54, 460.05; MS (EI): m/z (relative intensity) 417(M^+ ,15), 402 (100), 334 (25), 252 (30), 236 (29), 222 (10), 207 (3); HRMS: calcd. for $C_{27}H_{34}NPH^+$: 404.2507, found 404.2511.

The structural fine-tunings on ligand skeleton were reported to give significant impacts on the effectiveness of a catalytic system. In order to have a better insight on the relationship between the ligand reactivity and the structure of the indolyl ligand family, we performed additional probing experiments. Mechanistic studies showed that the substitution pattern on indole moiety is important to the reactivity. 1H NMR studies indicated that the signal of the C-H group (from 3-position of the indole ring, **L2**) rapidly disappeared before the catalysis starts. These spectroscopic data directly provide the evidence for the palladacycle formation. We are intrigued to investigate the importance of the indole 3-C-H position, we therefore independently synthesized a structurally similar ligand **L4** (Figure 2). Ligand **L4** contains all features of ligand **L2** except the C-H position from the indolyl scaffold is blocked by a methyl group. Surprisingly, a significantly lower reactivity (only 24% conv.) was observed when **L4** was subjected to the coupling of unactivated 4-*tert*-butylphenyl mesylate with 4-tolylboronic acid. From the ligand structural comparison, it seems to show that the ease of the palladacycle formation is important to accomplish mesylate catalysis.

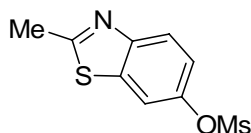


3. Preparation of aryl mesylates



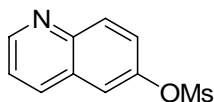
4-*tert*-Butylphenyl mesylate^[3], 4-methoxyphenyl mesylate^[4], 4-acetylphenyl mesylate^[4], 4-benzoylphenyl mesylate^[5], 4-acetyl-2-methoxyphenyl mesylate^[6], 1,3-Benzodioxol-5-ol mesylate^[7], 4-formyl-2-methoxyphenyl mesylate^[8], 3-pyridyl mesylate^[9], 4-cyanophenyl mesylate^[4], 4-(methoxycarbonyl)phenyl mesylate^[4] were prepared from their corresponding phenols with MsCl in the presence of triethylamine in CH₂Cl₂ according to the literature method.^[10] Unknown aryl mesylates were prepared according the the literature procedures without modification. Characterizations data of unknown aryl mesylates are shown below.^[10]

2-Methyl-5-benzothiazol mesylate

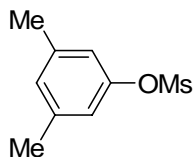


Obtained in 38% yield as white crystals: mp 119.5-120.8 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.76 (s, 3H), 3.12 (s, 3H), 7.24 (dd, *J*=2.3Hz, 8.7Hz, 1H), 7.74-7.78 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 20.0, 37.1, 115.4, 118.9, 122.1, 134.4, 147.4, 153.7, 169.6; MS (EI): *m/z* (relative intensity) 243 (M⁺, 35), 164 (100), 136 (28), 122 (15), 95 (18). HRMS: calcd. for C₉H₉N₁O₃S₂: 243.0018, found 243.0023.

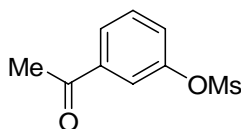
6-(Mesyloxy)quinoline



Obtained in 58% yield as deep purple crystals: mp 84.6-86.2 °C; ¹H NMR (400 MHz, CDCl₃) δ 3.15 (s, 3H), 7.32-7.35 (m, 1H), 7.52-7.55 (m, 1H), 7.67 (d, *J*=2.6Hz, 1H), 8.04-8.07 (m, 2H), 8.83-8.85 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 37.4, 119.2, 121.8, 124.0, 128.1, 131.6, 135.7, 146.4, 146.4, 150.7; MS (EI): *m/z* (relative intensity) 223 (M⁺, 63), 145 (60), 116 (100), 89 (30), 63 (10). HRMS: calcd. for C₁₀H₉N₁O₃S₁: 223.0298, found 223.0292.

3,5-Dimethylphenyl mesylate.

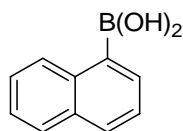
Obtained in 95% yield as white crystals: mp 41.7-43.4 °C; ^1H NMR (400 MHz, CDCl_3) δ 2.30 (s, 3H), 3.07 (s, 3H), 6.88 (s, 2H), 6.93 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.8, 36.8, 119.1, 128.7, 139.7, 149.0; MS (EI): m/z (relative intensity) 200 (M^+ , 65), 122 (100), 107 (20), 91 (30), 77 (30). HRMS: calcd. for $\text{C}_9\text{H}_{12}\text{O}_3\text{S}_1$: 200.0502, found 200.0507.

3-Acetylphenyl mesylate

Obtained in 83% yield as white crystals: mp 50.1-52.4 °C; ^1H NMR (400 MHz, CDCl_3) δ 2.56 (s, 3H), 3.14 (s, 3H), 7.44-7.48 (m, 2H), 7.78 (s, 1H), 7.86 (d, $J=7.1\text{Hz}$, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.5, 37.4, 121.4, 126.5, 127.0, 130.2, 138.7, 149.2, 196.3; MS (EI): m/z (relative intensity) 214 (M^+ , 28), 199 (100), 171 (5), 121 (35), 92 (10). HRMS: calcd. for $\text{C}_9\text{H}_{10}\text{O}_4\text{S}_1$: 214.2940, found 214.0304.

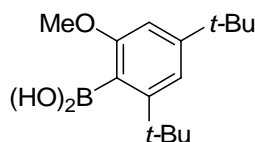
4. Procedures for preparation arylboronic acid substrates

1-Naphthylboronic acid^[11]



Anhydrous THF (20 mL) was added to 1-bromonaphthalene (5.56 mL, 40 mmol) under nitrogen at room temperature. *n*-Butyllithium (44 mmol) was added dropwise to the solution at -78 °C and stirred for 30 min. The reaction mixture was transferred to a solution of B(Oi-Pr)₃ (18.6 mL, 80 mmol) in THF (10 mL) at -78 °C *via* cannular under nitrogen. The solution was warm to room temperature and stirred overnight. Diluted HCl (10%, 20 mL) was added to the mixture and stirred for 30 min. The organic layer was separated and the aqueous phase was extracted by diethyl ether (50 mL × 3). The combined organic extracts were washed with brine and dried over MgSO₄. The solvent was removed and the residue was crystallized under ether/hexane to give colorless crystals (5.0 g, 72%). The product was used without further characterization.

2,4-Di-*tert*-butyl-6-methoxyphenylboronic acid^[12]



n-Butyllithium (1.6 M in hexane, 25 mL, 40 mmol) was added to a solution of 2-bromo-3,5-di-*tert*-butylanisole (12 g, 40 mmol) in THF (20 mL) at -78 °C under nitrogen. The mixture was stirred for 1 h and then transferred to a solution of distilled B(OMe)₃ (9.1 mL, 80 mmol) in THF (10 mL) at -78 °C under nitrogen. It was allowed to warm to room temperature and water was added. The organic layer was separated, and the aqueous phase was extracted with ether (3 × ~100 mL). The combined organic phases were washed with brine. After removing of solvent, the residue was redissolved in ethanol (60 mL). A solution of NaOH (2 M) was added and the mixture was stirred for 2 h. Solvent was rotary-evaporated, and the remaining aqueous layer was extracted with ether (3 × ~100 mL). The ethereal layer was then washed with brine and dried over MgSO₄. The solvent was removed, and the residue was recrystallized from ether/hexane to give

2,4-di-*tert*-butyl-6-methoxyphenylboronic acid as colorless crystals (7.8 g, 74%). The product was used without further characterization.

5. General procedures/data for initial ligand and reaction condition screenings

General procedure for screening: Pd(OAc)₂ (6.8 mg, 0.030 mmol) and ligand (Pd:L = 1:4) were loaded into a Schlenk tube equipped with a Teflon-coated magnetic stir bar. The tube was evacuated and flushed with nitrogen for three times. Precomplexation was applied by adding freshly distilled dichloromethane (0.5 mL) and Et₃N (50 μ L) into the tube. The palladium complex stock solution was stirred and warmed using hair drier for about 1 to 2 minutes until the solvent started boiling. The solvent was then evaporated under high vacuum. 4-*tert*-Butylphenyl mesylate (1.0 mmol), 4-methylphenylboronic acid (2.0 mmol) and base (3.0 mmol) were loaded into the tube, and the system was further evacuated and flushed with nitrogen for three times. The solvent (3.0 mL) was added with stirring at room temperature for several minutes. The tube was then placed into a preheated oil bath (110 °C) and stirred for the time as indicated. After completion of reaction, the reaction tube was allowed to cool to room temperature. Ethyl acetate (~10 mL), dodecane (227 μ L, internal standard) and water were added. The organic layer was subjected to GC analysis. The GC yield obtained was previously calibrated by authentic sample/dodecane calibration curve.

$t\text{-Bu-C}_6\text{H}_4\text{-OMs} + 4\text{-MeC}_6\text{H}_4\text{B(OH)}_2 \xrightarrow[\text{base, solvent, 110 }^\circ\text{C, 4 h}]{\text{2 mol\% Pd(OAc)}_2, \text{L1-L3}} t\text{-Bu-C}_6\text{H}_4\text{-tol}$				
entry	Ligand	base	Solvent	GC yield (%)
1	L1	K ₃ PO ₄ ·H ₂ O	^t BuOH	<1%
2	L3	K ₃ PO ₄ ·H ₂ O	^t BuOH	84%
3	L2	K ₃ PO ₄ ·H ₂ O	^t BuOH	89%
4	L2	K ₃ PO ₄ ·H ₂ O	Toluene	42%
5	L2	K ₃ PO ₄ ·H ₂ O	DMF	50%
6	L2	K ₃ PO ₄ ·H ₂ O	^t BuOH	89%
7	L2	K ₃ PO ₄ ·H ₂ O	THF	33%
8	L2	Cs ₂ CO ₃	^t BuOH	83%
9	L2	CsF	^t BuOH	93%

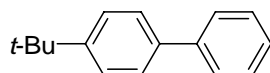
10	L2	K ₂ CO ₃	^t BuOH	79%
11	L2	^t BuOK	^t BuOH	<1%
12	L2	K ₃ PO ₄	^t BuOH	97%
13	L2	K ₃ PO ₄	DMF	81%

6. General procedures for Suzuki-Miyaura couplings of aryl mesylates

General procedure for Suzuki-Miyaura coupling of aryl mesylates: Pd(OAc)₂ (2.3 mg, 0.010 mmol) and ligand (Pd:L = 1:4) were loaded into a Schlenk tube equipped with a Teflon-coated magnetic stir bar. The tube was evacuated and flushed with nitrogen for several times. Precomplexation was applied by adding freshly distilled dichloromethane (0.5 mL) and Et₃N (50 μL) into the tube. The solution was stirred and warmed using hair drier for about 1 to 2 minutes until the solvent started boiling. The solvent was then evaporated under high vacuum. Aryl mesylate (1.0 mmol), arylboronic acid (2.0 mmol) and K₃PO₄ (3.0 mmol) were loaded into the tube, and the system was further evacuated and flushed with nitrogen for several times. The solvent *tert*-butanol (3.0 mL) was then added. The tube was stirred at room temperature for several minutes and then placed into a preheated oil bath (110 °C) for the time period as indicated in Table 2. After completion of reaction as judged by GC analysis, the reaction tube was allowed to cool to room temperature and quenched with water and diluted with EtOAc. The organic layer was separated and the aqueous layer was washed with EtOAc. The filtrate was concentrated under reduced pressure. The crude products were purified by flash column chromatography on silica gel (230-400 mesh) to afford the desired product.

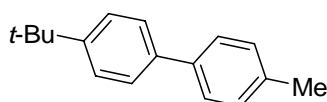
7. Characterization data for coupling products

4-*tert*-Butylbiphenyl (Table 2, entries 1 and 2, Scheme 2).^[14]



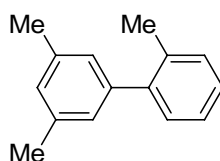
Hexane, $R_f=0.55$; ^1H NMR (400 MHz, CDCl_3) δ 1.72 (s, 9H), 7.64 (t, $J=7.4$ Hz, 1H), 7.72-7.82 (m, 4H), 7.89-7.96 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.3, 34.4, 125.6, 126.7, 126.9, 128.6, 138.2, 140.9, 150.0; MS (EI): m/z (relative intensity) 210 (M^+ , 35), 195 (100), 178 (20), 167 (30).

4-Methyl-4'-*tert*-butylbiphenyl (Table 2, entry 3).^[13]

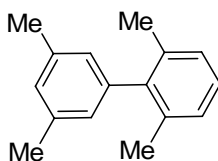


Hexane, $R_f=0.5$; ^1H NMR (400 MHz, CDCl_3) δ 1.65 (s, 9H), 2.65 (s, 3H), 7.49 (d, $J=8.0$ Hz, 2H), 7.73 ($J=8.4$ Hz, 2H), 7.78 ($J=8.0$ Hz, 2H), 7.82 ($J=8.3$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 21.0, 31.3, 34.4, 125.6, 126.5, 126.7, 129.4, 136.5, 138.1, 138.2, 149.7; MS (EI): m/z (relative intensity) 224 (M^+ , 33), 209 (100), 193 (5), 181 (14), 165 (10).

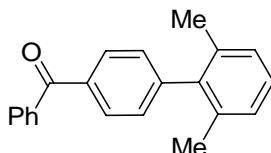
3, 5-Dimethyl-2'-methylbiphenyl (Table 2, entry 4).^[15]



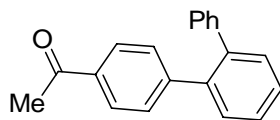
Hexane, $R_f=0.55$; ^1H NMR (400 MHz, CDCl_3) δ 2.59 (s, 3H), 2.65 (s, 6H), 7.27 (m, 3H), 7.50-7.54 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.4, 21.2, 125.6, 126.9, 128.3, 129.6, 130.1, 135.1, 137.3, 141.9, 142.1; MS (EI): m/z (relative intensity) 196 (M^+ , 70), 181 (100), 165 (45), 152 (10).

3, 5-Dimethyl-2',6'-dimethylbiphenyl (Table 2, entry 5).^[16]

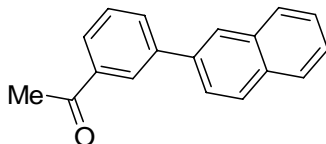
Hexane, $R_f=0.55$; ^1H NMR (400 MHz, CDCl_3) δ 2.29 (s, 6H), 2.57 (s, 6H), 7.01 (s, 2H), 7.20 (s, 1H), 7.30-7.40 (m, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.8, 21.3, 126.6, 127.1, 128.1, 135.8, 137.6, 11.0, 142.0; MS (EI): m/z (relative intensity) 210 (M^+ , 65), 195 (100), 180 (30), 152 (25).

4-(2,6-Dimethylphenyl)benzophenone (Table 2, entry 6).

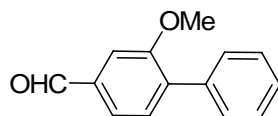
EA:Hexane = 1:20, $R_f=0.4$; white solid, m.p. = 76.5-78.9 °C; ^1H NMR (400 MHz, CDCl_3) δ 2.13 (s, 6H), 7.18-7.25 (m, 3H), 7.33 (d, $J=8.2$ Hz, 2H), 7.54 (t, $J=7.7$ Hz, 2H), 7.62-7.63 (m, 1H), 7.93-7.98 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.5, 127.1, 127.2, 127.9, 128.7, 129.6, 130.0, 131.9, 135.1, 135.5, 137.3, 140.3, 145.4, 195.8; MS (EI): m/z (relative intensity) 286 (M^+ , 80), 209 (100), 181 (25), 165 (50), 105 (45). HRMS: calcd. for $\text{C}_{21}\text{H}_{18}\text{O}$: 287.1436, found 287.1442.

4-(2-Biphenyl)acetophenone (Table 2, entry 7).^[18]

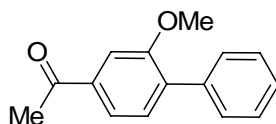
EA:Hexane = 1:20, $R_f=0.4$; ^1H NMR (400 MHz, CDCl_3) δ 2.56 (s, 3H), 7.20-7.31 (m, 7H), 7.46-7.49 (m, 4H), 7.86 (d, $J=8.3$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 126.5, 127.4, 127.7, 127.8, 127.9, 129.0, 129.8, 130.1, 130.5, 134.8, 139.0, 140.3, 140.7, 146.3; MS (EI): m/z (relative intensity) 272 (M^+ , 70), 257 (100), 228 (50), 202 (20).

3-(2-Naphthyl)acetophenone (Table 2, entry 8).

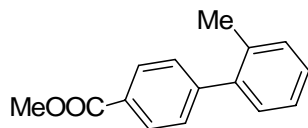
EA:Hexane = 1:9, R_f =0.35; White solid, m.p. = 69.5-71.8 °C; ^1H NMR (400 MHz, CDCl_3) δ 2.60 (s, 3H), 7.45-7.50 (m, 3H), 7.68 (dd, J =1.6, 8.5 Hz, 1H), 7.78-7.89 (m, 5H), 8.00 (bs, 1H), 8.30 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.2, 124.7, 125.5, 125.8, 126.0, 126.5, 126.8, 127.2, 127.8, 128.2, 128.6, 131.3, 132.4, 133.2, 136.8, 137.2, 140.9, 197.5; MS (EI): m/z (relative intensity) 246 (M^+ , 100), 231 (90), 202 (95). HRMS: calcd. for $\text{C}_{18}\text{H}_{14}\text{O}$: 247.1123, found 247.1106.

3'-Methoxy-4'-phenylbenzaldehyde (Table 2, entry 9).^[24]

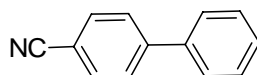
EA:Hexane = 1:9, R_f =0.35; ^1H NMR (400 MHz, CDCl_3) δ 3.86 (s, 3H), 7.39-7.59 (m, 8H), 10.00 (s, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.3, 109.4, 124.0, 127.6, 127.8, 129.1, 130.9, 136.4, 136.8, 136.9, 156.7, 191.5; MS (EI): m/z (relative intensity) 212 (M^+ , 100), 169 (20), 152 (10), 139 (20), 115(15).

3'-Methoxy-4'-phenylacetophenone (Table 2, entry 10).^[19]

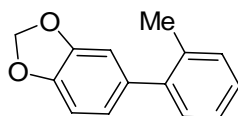
EA:Hexane = 1:9, R_f =0.3; ^1H NMR (400 MHz, CDCl_3) δ 2.61 (s, 3H), 3.82 (s, 3H), 7.37-7.46 (m, 4H), 7.57-7.62 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 26.1, 55.1, 109.5, 121.3, 127.2, 129.0, 130.3, 135.1, 136.8, 137.0, 156.2, 197.0; MS (EI): m/z (relative intensity) 226 (M^+ , 80), 211 (100), 183 (10), 168 (30).

Methyl 4-(2'-tolyl)benzoate (Table 2, entry 11).

EA:Hexane = 1:9, $R_f=0.4$; ^1H NMR (400 MHz, CDCl_3) δ 2.3 (s, 3H), 3.96 (s, 3H), 7.24-7.32 (m, 4H), 7.42(d, J = 8.0 Hz, 2H), 8.14 (d, J = 8.3 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.1, 125.6, 127.5, 128.3, 129.0, 129.1, 129.2, 130.2, 134.8, 140.5, 146.4, 166.6; MS (EI): m/z (relative intensity) 226 (M^+ , 100), 195 (98), 168 (60), 152 (50), 139 (7).

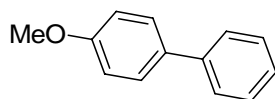
4-Cyanobiphenyl (Table 2, entry 12).^[28]

DCM:Hexane = 1:20, $R_f=0.35$; ^1H NMR (400 MHz, CDCl_3) δ 7.43-7.50 (m, 3H), 7.57-7.59 (m, 2H), 7.64-7.70 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 110.6, 118.6, 126.9, 127.4, 128.4, 128.8, 132.3, 138.8, 145.2; MS (EI): m/z (relative intensity) 179 (M^+ , 100), 151 (20).

2'-Methyl- 3,4-methylenedioxy biphenyl (Table 2, entry 13).^[23]

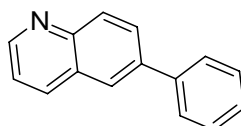
EA:Hexane = 1:4, $R_f=0.7$; ^1H NMR (400 MHz, CDCl_3) δ 2.44 (s, 3H), 6.07 (s, 2H), 6.91-7.01 (m, 3H), 7.37-7.39 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.3, 100.8, 107.8, 109.6, 122.3, 125.6, 127.0, 129.6, 130.1, 135.2, 135.7, 141.4, 146.3, 147.1; MS (EI): m/z (relative intensity) 212 (M^+ , 100), 181 (20), 153 (30).

4-Phenylanisole (Table 2, entry 14).^[17]



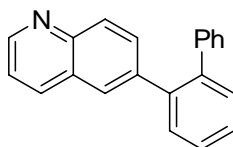
EA:Hexane = 1:20, R_f =0.4; ^1H NMR (400 MHz, CDCl_3) δ 3.97 (s, 3H), 7.17 (d, J =8.7 Hz, 2H), 7.51-7.54 (m, 1H), 7.62 (t, J =7.8 Hz, 2H), 7.73-7.79 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 55.0, 114.1, 126.5, 127.9, 128.3, 128.6, 133.5, 140.6, 159.0; MS (EI): m/z (relative intensity) 184 (M^+ , 100), 169 (50), 141 (50), 115 (40).

6-Phenylquinoline (Table 3, entry 1).^[22]



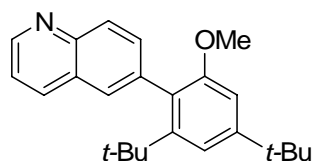
EA:Hexane = 1:4, R_f =0.25; ^1H NMR (400 MHz, CDCl_3) δ 7.21-7.33 (m, 2H), 7.37-7.71 (m, 2H), 7.58-7.60 (m, 2H), 7.82-7.87 (m, 2H), 7.96-8.15 (m, 2H), 8.83 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 121.0, 125.0, 127.0, 127.3, 128.0, 128.5, 128.7, 129.4, 135.7, 138.7, 139.7, 147.2, 149.9; MS (EI): m/z (relative intensity) 205 (M^+ , 100), 176 (10).

5-(2-Phenyl)quinoline (Table 3, entry 2).



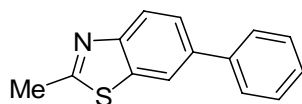
DCM, R_f =0.2; light yellow viscous liquid; ^1H NMR (400 MHz, CDCl_3) δ 7.18 (bs, 5H), 7.25-7.54 (m, 6H), 7.69 (s, 1H), 7.94-7.98 (m, 2H), 8.85 (d, J = 4.0 Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 120.8, 126.4, 127.4, 127.7, 127.8, 127.9, 128.3, 129.6, 130.4, 130.5, 131.7, 135.6, 139.2, 139.7, 140.4, 140.8, 146.8, 149.9; MS (EI): m/z (relative intensity) 281 (M^+ , 100), 266 (30), 252 (20); HRMS: calcd. for $\text{C}_{21}\text{H}_{15}\text{N}$: 281.1199, found 281.1203.

6-(2,4-Di-*tert*-butyl-6-methoxyphenyl)quinoline (Table 3, entry 3).



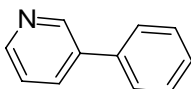
EA:Hexane = 1:9, R_f =0.15; white solid, m.p. = 136.4-138.1 °C. ^1H NMR (400 MHz, CDCl_3) δ 1.19 (s, 9H), 1.42 (s, 9H), 3.56 (s, 3H), 6.92 (s, 1H); 7.24-7.31 (m, 4H), 8.02 (d, J =7.9 Hz, 1H), 8.14 (d, J =8.5 Hz, 1H), 8.87 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 31.1, 32.5, 34.8, 36.6, 55.5, 105.7, 116.0, 120.5, 127.0, 127.2, 127.5, 129.2, 133.9, 135.5, 138.3, 147.0, 148.4, 149.6, 150.6, 157.3; MS (EI): m/z (relative intensity) 347 (M^+ , 100), 332 (75), 317 (5), 302 (3), 290 (5), 276(75), 261(30); HRMS: calcd. for $\text{C}_{24}\text{H}_{29}\text{NO}$: 347.2244, found 347.2253.

2-Methyl-5-phenylbenzothiazole (Table 3, entry 4).^[20]



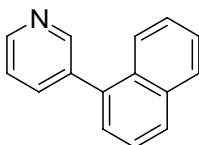
EA:Hexane = 1:4, R_f =0.4; ^1H NMR (400 MHz, CDCl_3) δ 2.73 (s, 3H), 7.22-7.24 (m, 1H), 7.40 (t, J =8.3Hz, 2H), 7.62 (d, J =7.3Hz, 1H), 7.69 (d, J =8.3Hz, 2H), 8.21 (d, J =1.1Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 19.5, 120.1, 121.0, 123.5, 126.8, 126.9, 128.4, 134.1, 138.9, 140.2, 153.6, 167.0; MS (EI): m/z (relative intensity) 225 (M^+ , 100), 184 (10), 152 (10).

3-Phenyl pyridine (Table 3, entry 5).^[25]



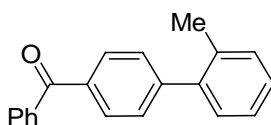
EA:Hexane = 3:7, R_f =0.5; ^1H NMR (400 MHz, CDCl_3) δ 7.31-7.83 (m, 7H), 8.5-8.58 (m, 1H), 8.85 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 123.3, 126.8, 127.9, 128.8, 134.2, 136.4, 137.4, 147.8, 147.9; MS (EI): m/z (relative intensity) 155 (M^+ , 100), 127 (10), 102 (7).

3-(1-Naphthalenyl) pyridine (Table 3, entry 6).^[29]



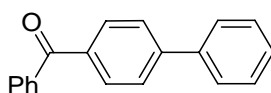
EA:Hexane = 1:4, $R_f=0.3$; ^1H NMR (400 MHz, CDCl_3) δ 7.34-7.48 (m, 5H), 7.50-7.86 (m, 4H), (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 122.7, 124.8, 125.0, 125.7, 126.1, 127.0, 128.1, 131.0, 133.4, 135.8, 135.9, 136.9, 148.1, 150.1; MS (EI): m/z (relative intensity) 205 (M^+ , 90), 204 (100), 176 (18), 151 (10).

4-(2'-Tolyl)benzophenone (Scheme 2).^[21]



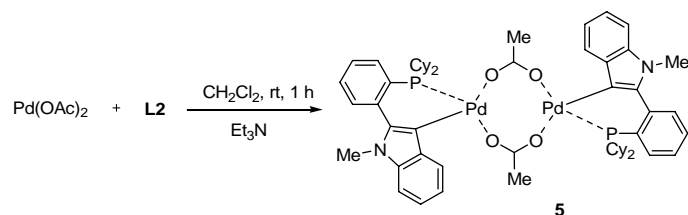
EA:Hexane = 1:20, $R_f=0.4$; ^1H NMR (400 MHz, CDCl_3) δ 2.36 (s, 3H), 7.30-7.33 (m, 4H), 7.46-7.61 (m, 5H), 7.90-7.93 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 20.2, 125.7, 127.6, 128.0, 128.9, 129.3, 129.7, 130.3, 132.0, 135.6, 137.4, 140.4, 146.0, 195.9; MS (EI): m/z (relative intensity) 272 (M^+ , 70), 195 (100), 164 (40), 152 (40), 105 (40).

4-Phenylbenzophenone (Scheme 2).^[26]

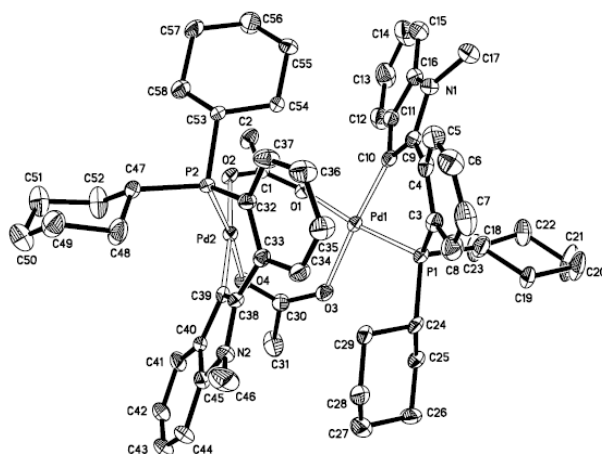


EA:Hexane = 1:20, $R_f=0.4$; ^1H NMR (400 MHz, CDCl_3) δ 7.38-7.42 (m, 1H), 7.46-7.51 (m, 4H), 7.57-7.61 (m, 1H), 7.65-7.71 (m, 4H), 7.82-7.92 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3) δ 126.6, 126.9, 127.9, 128.0, 128.6, 129.6, 130.4, 132.0, 135.8, 137.4, 139.5, 144.7, 195.7; MS (EI): m/z (relative intensity) 258 (M^+ , 75), 181 (100), 152 (40), 105 (20), 77 (19).

8. Preparation of single-component complex **5**

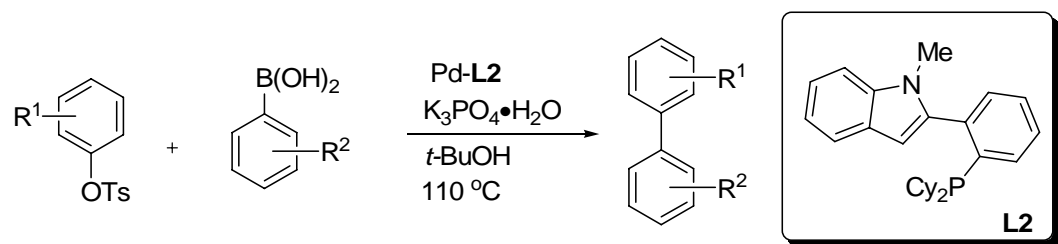
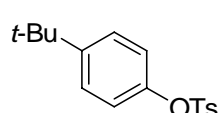
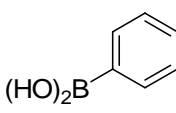
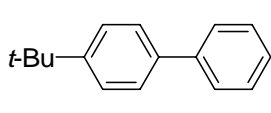
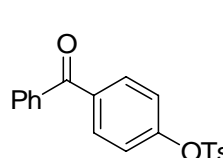
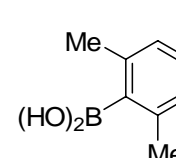
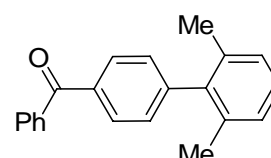
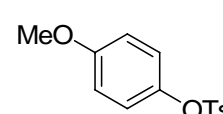
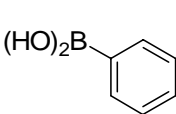
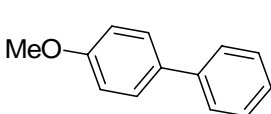


Palladium(II) acetate (224 mg, 1.0 mmol) and **L2** (1.0 mmol) were dissolved in freshly distilled dichloromethane (10 mL) and anhydrous Et₃N (2.0 mmol) under nitrogen at room temperature. The red solution was stirred for one hour. Solvent was partially removed under reduced pressure. Anhydrous diethyl ether was then slowly added and reddish brown precipitates were obtained (70%). Crystals for X-ray diffraction were obtained by recrystallization of the pure product from dichloromethane/hexane layers.

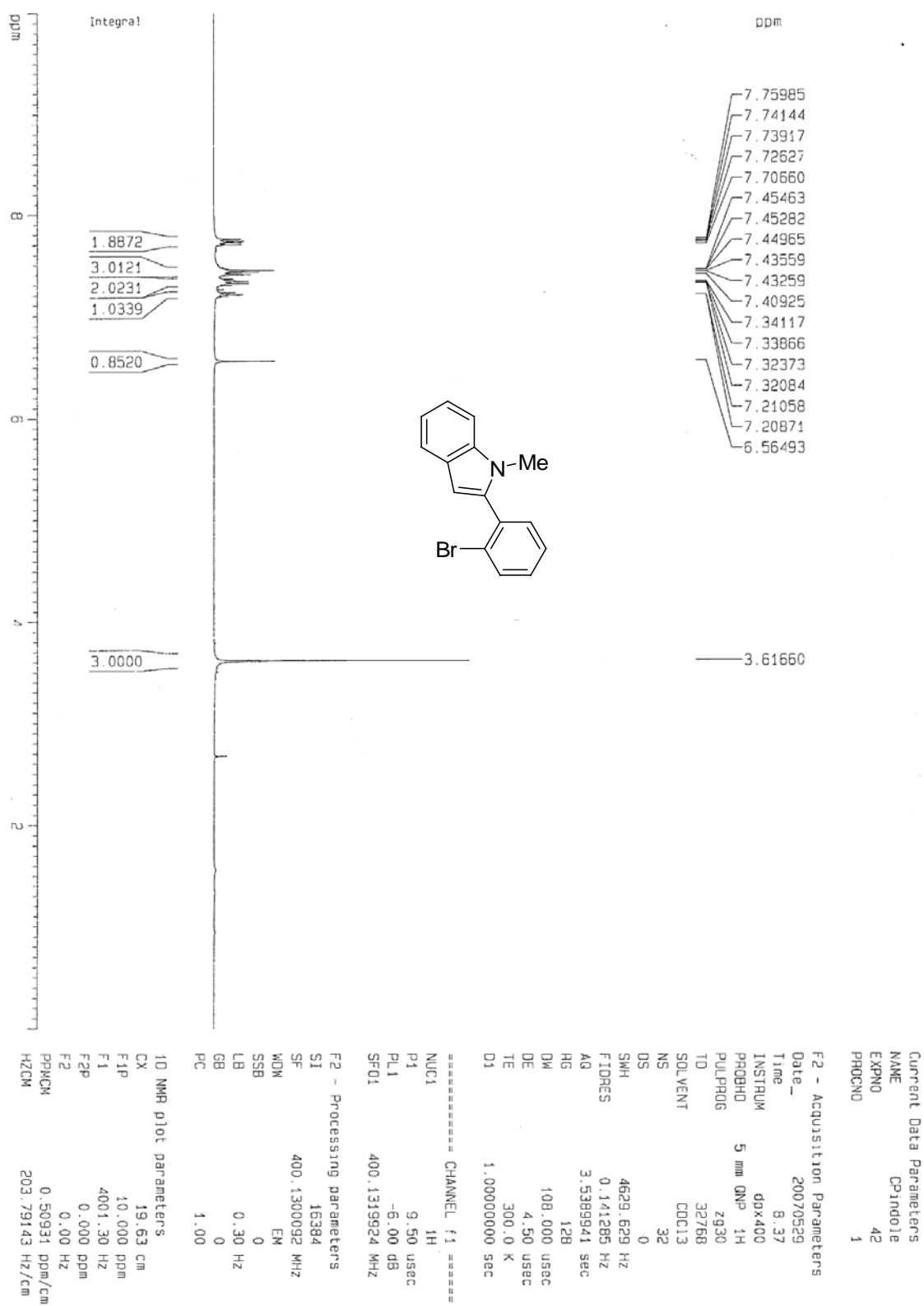


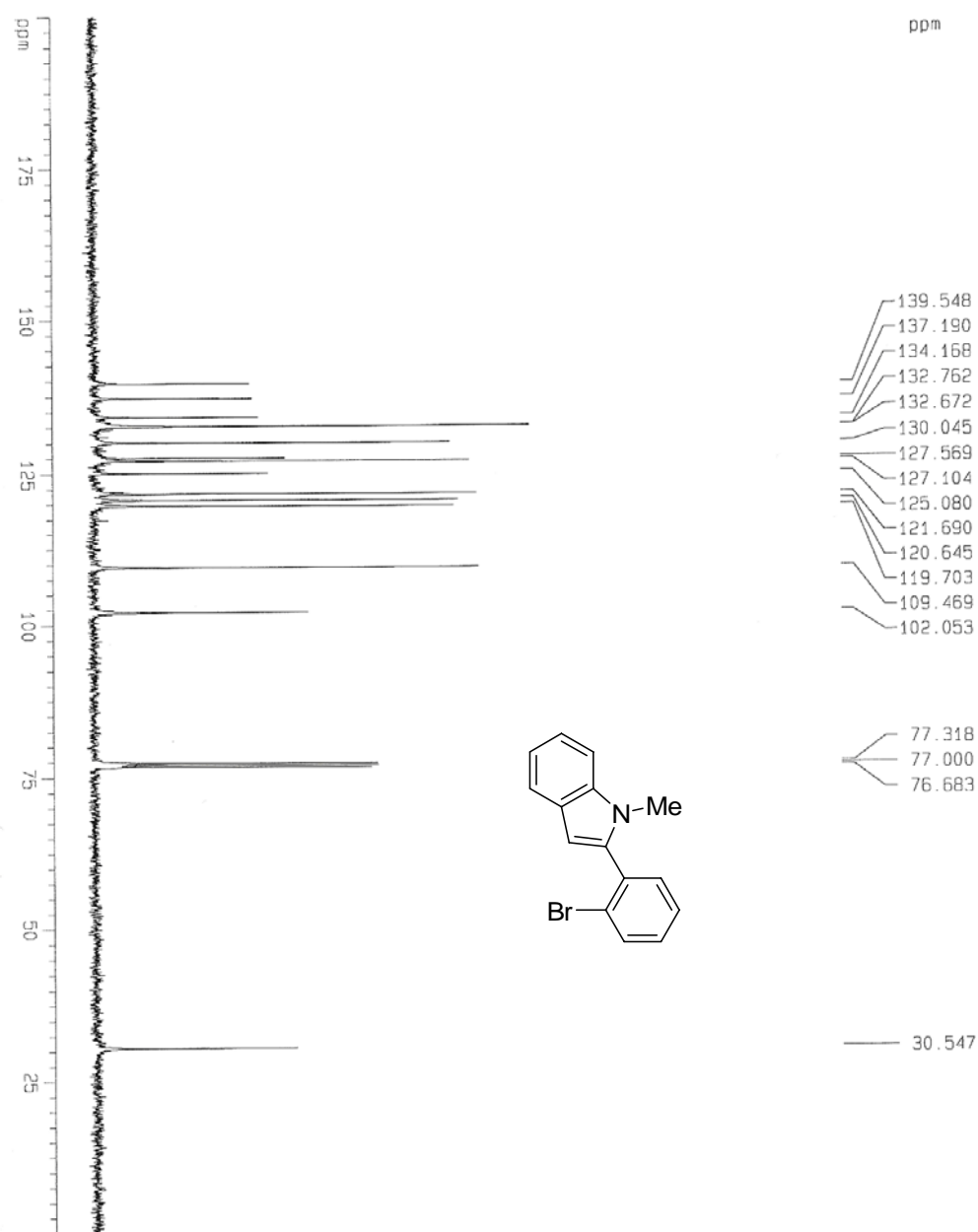
¹H NMR (400 MHz, CD₂Cl₂) δ 8.01 (d, *J* = 7.0 Hz, 1 H), 7.54-7.62 (m, 2 H), 7.45-7.49 (m, 1 H), 7.20-7.34 (m, 3 H), 7.07 (t, *J* = 7.2 Hz, 1 H), 3.83 (s, 3H), 1.11-2.07 (m, 30H); ¹³C NMR (100 MHz, CD₂Cl₂) δ 143.1, 141.2 (d, *J*_{CP} = 10.6 Hz), 137.6 (d, *J*_{CP} = 26.8 Hz), 134.2, 133.7, 131.4, 128.6, 126.4, 126.3, 124.9, 123.1, 122.4, 119.9, 110.2, 104.0 (d, *J*_{CP} = 14.5 Hz), 35.6 (d, *J*_{CP} = 28.1 Hz), 34.5, 29.3, 28.2, 27.7, 26.5, 24.6; ³¹P NMR (162 MHz, CD₂Cl₂) δ +44.0. Full X-ray data were reported in supplied cif file.

9. Data from aryl tosylate couplings (for comparison)

					
entry	ArOTs	Ar'B(OH) ₂	product	mol% Pd	% yield
1				0.2%, 12.5 h	85
2				0.5%, 2 h	90
3				3%, 4 h	90

Reaction conditions: ArOTs (1.0 mmol), Ar'B(OH)₂ (2.0 mmol), K₃PO₄ (3.0 mmol), Pd(OAc)₂/L2 (mol% as indicated), *t*-BuOH (3.0 mL), at 110 °C under N₂ for indicated period of time.

10. ^1H , ^{13}C , ^{31}P NMR, MS, HRMS and IR spectra



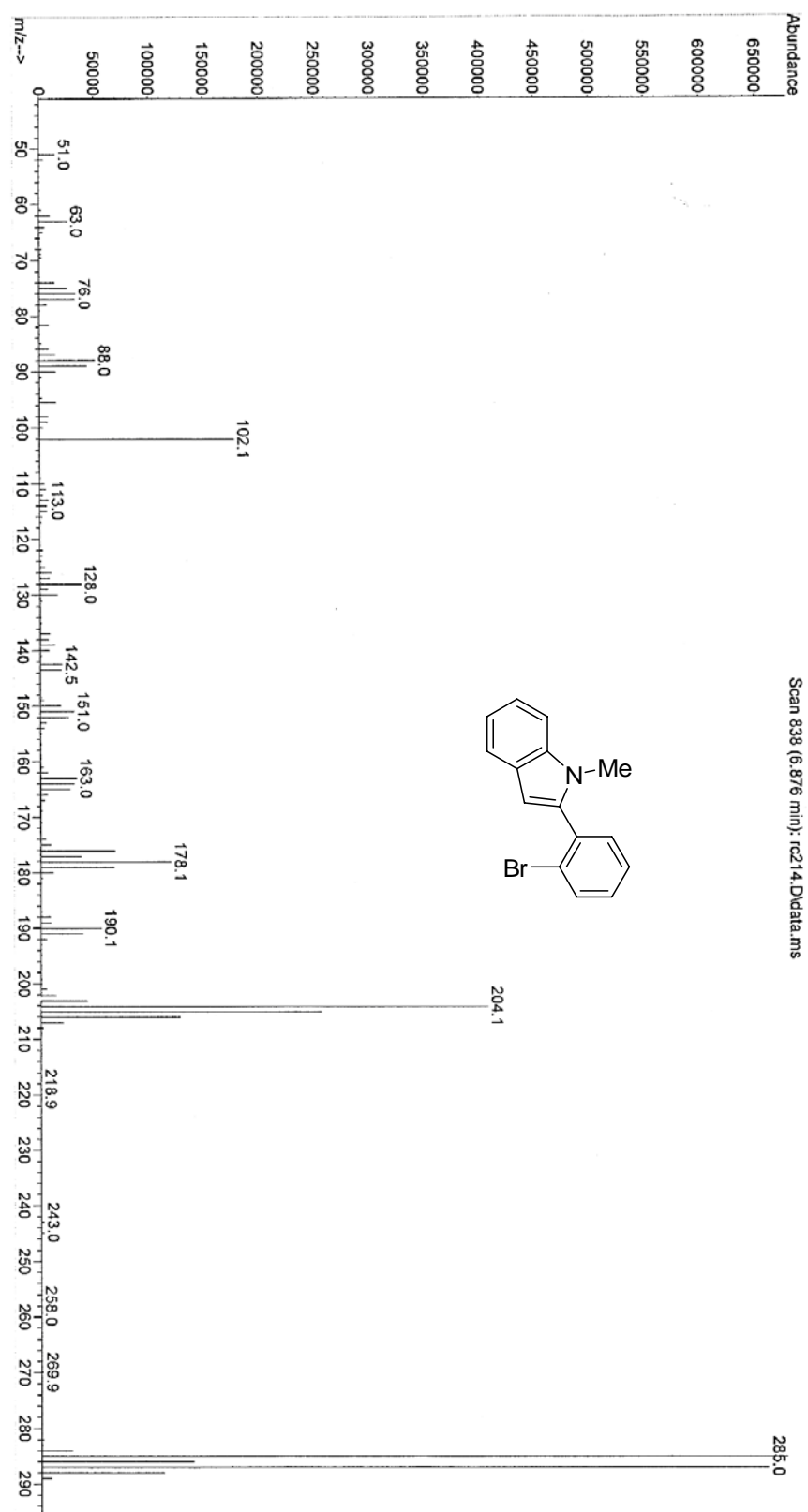
F2 - Acquisition Parameters
 Date_ 20070528
 Time 19.00
 INSTRUM dx400
 PROBRD 5 mm QNP 1H
 PULPROG zgpg30
 TD 131072
 SOLVENT CDCl₃
 NS 128
 DS 0
 SWH 25125.629 Hz
 FIDRES 0.191693 Hz
 AQ 2.6083827 sec
 RG 9195.2
 DW 19.900 usec
 DE 4.50 usec
 TE 300.0 K
 D1 3.00000000 sec
 d11 0.03000000 sec

===== CHANNEL f1 =====
 NUC1 ¹³C
 P1 3.30 usec
 PL1 -6.00 dB
 SFO1 100.6231263 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 ¹H
 PCPD2 71.00 usec
 PL2 120.00 dB
 PL12 17.00 dB
 SFO2 400.1326008 MHz

F2 - Processing parameters
 SI 65536
 SF 100.6127947 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 19.50 cm
 F1P 200.000 ppm
 F1 20122.56 Hz
 F2P -0.000 ppm
 F2 -0.00 Hz
 PPMCM 10.25641 ppm/cm
 HZCM 1031.92615 Hz/cm



Elemental Composition Report

Single Mass Analysis

Tolerance = 100.0 PPM / DBE: min = -1.5, max = 60.0

Selected filters: None

Monoisotopic Mass, Even Electron Ions

7 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-15 H: 0-1000 N: 0-1 Br: 0-1

So Chau Ming, N-methyl 2(2Bromophenyl)indole

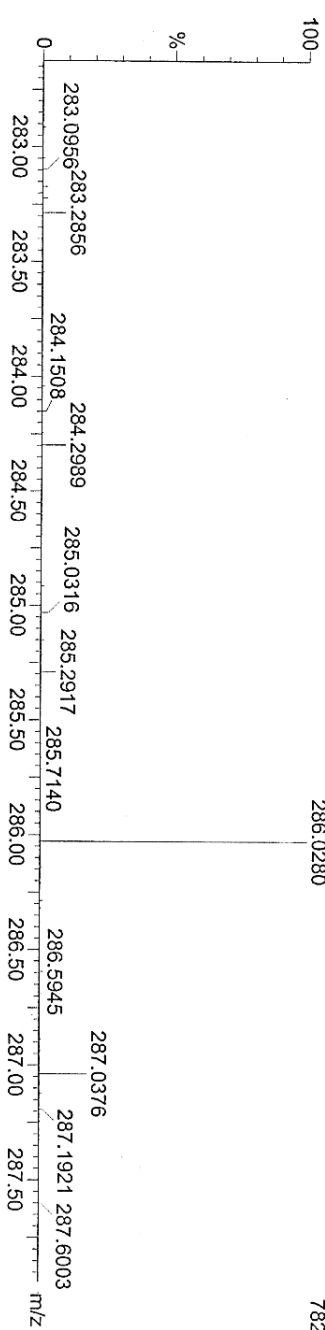
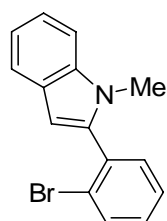
HR07_0615_4B 41 (0.778) AM (Cen.5, 80.00, Ht, 10000.0, 0.00, 0.70); Sm (SG, 3x2.00); Cm (41.59)

286.0280

15-Jun-2007

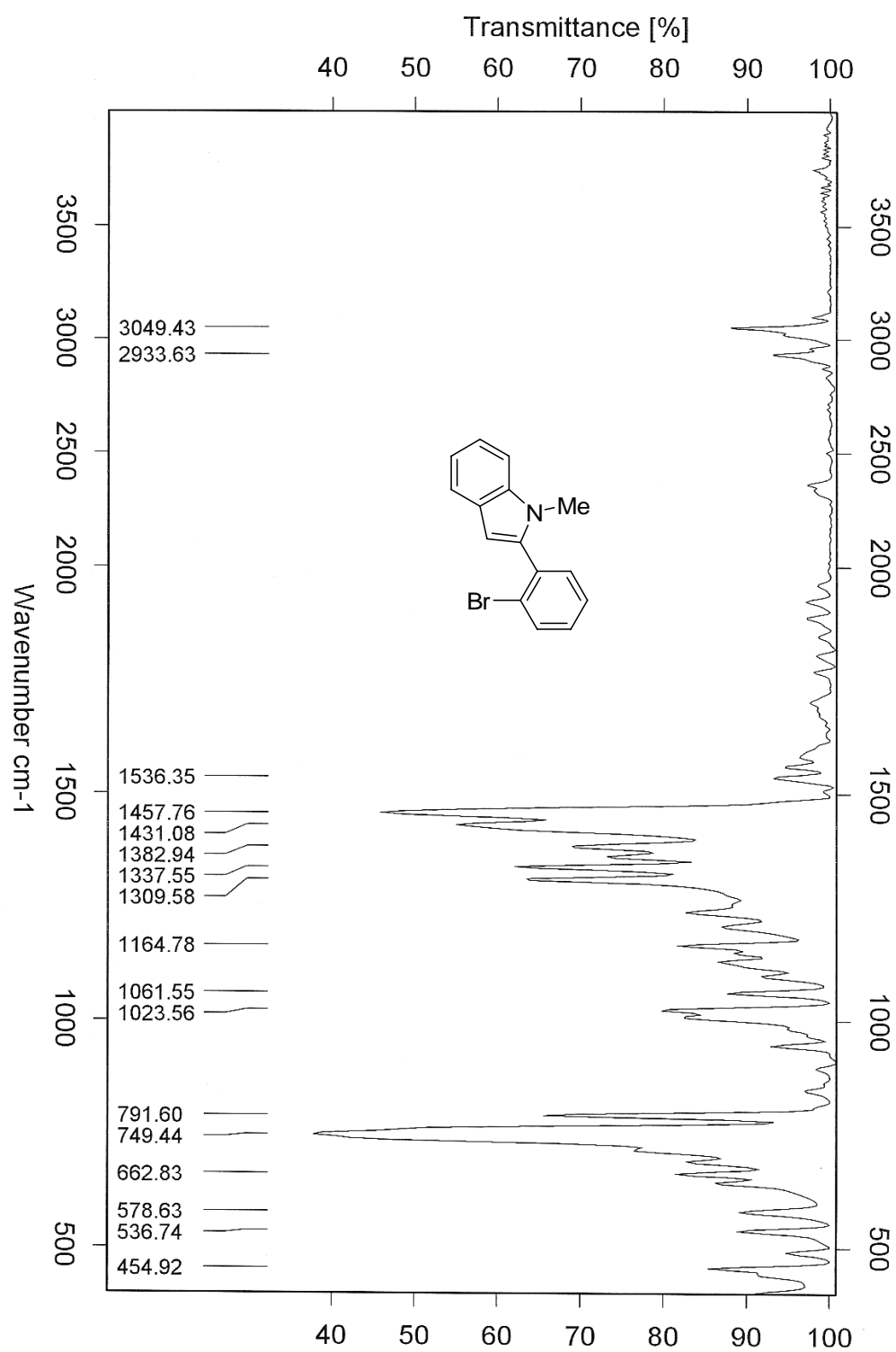
TOF MS ES+

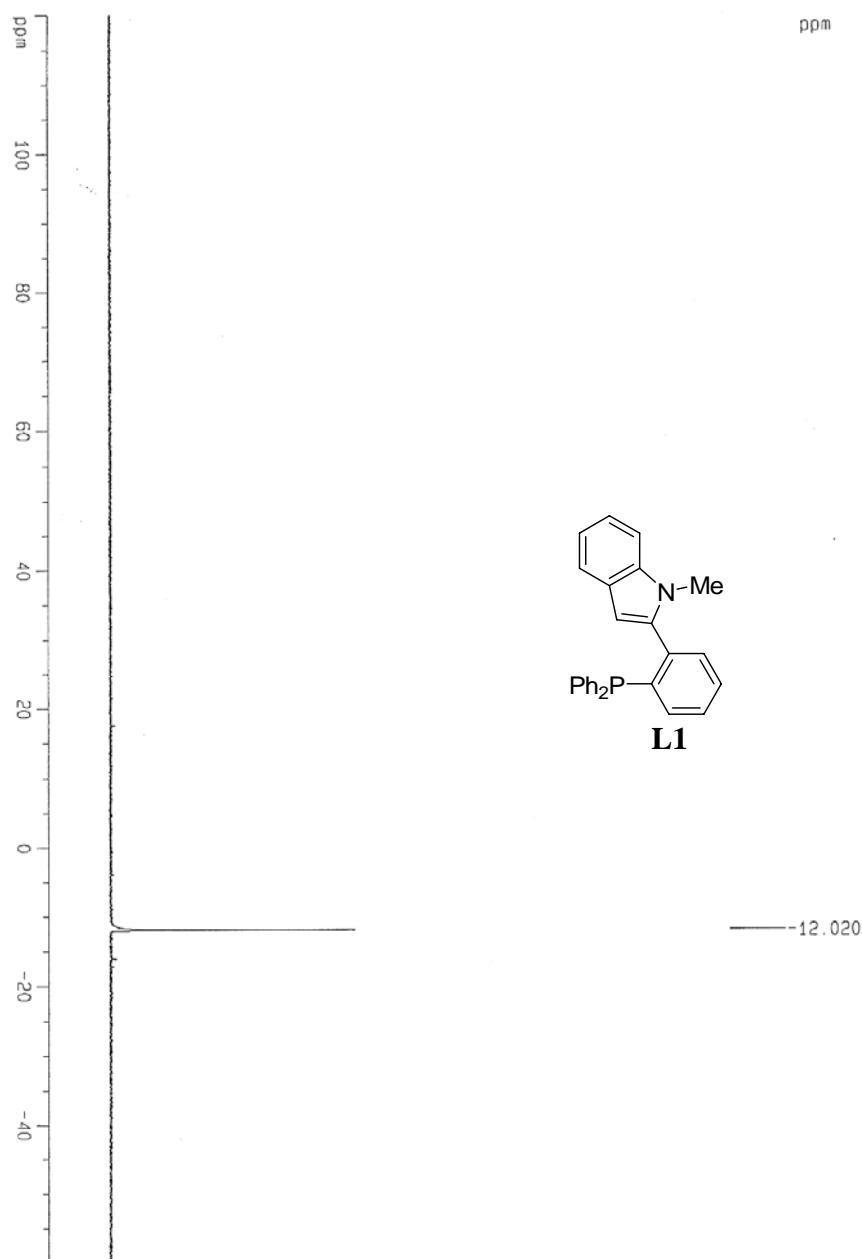
782



Minimum: 5.0
Maximum: 100.0
-1.5
60.0

Mass	Calc. Mass	mDa	PPM	DBE	I-FIT	Formula
286.0280	286.0231	4.9	17.1	9.5	463.5	C15 H13 N Br





Current Data Parameters
 NAME PCPindole
 EXPNO 40
 PROCNO 1

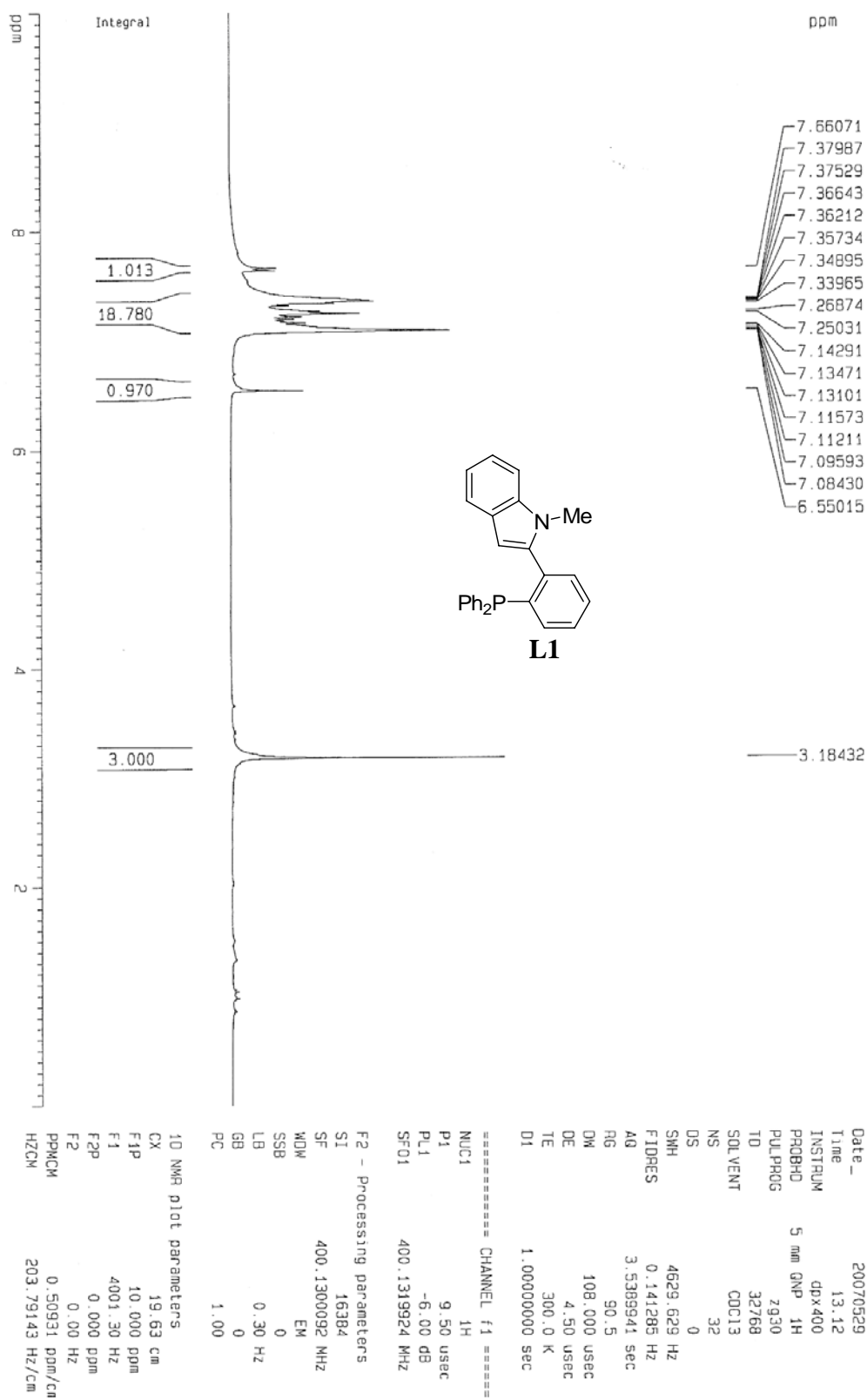
F2 - Acquisition Parameters
 Date_ 20070529
 Time 13.06
 INSTRUM dp400
 PROBRD 5 mm QNP 1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl₃
 NS 32
 DS 0
 SWH 48651.901 Hz
 FIDRES 0.742520 Hz
 AQ 0.6734324 sec
 RG 13004
 DM 10.275 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec

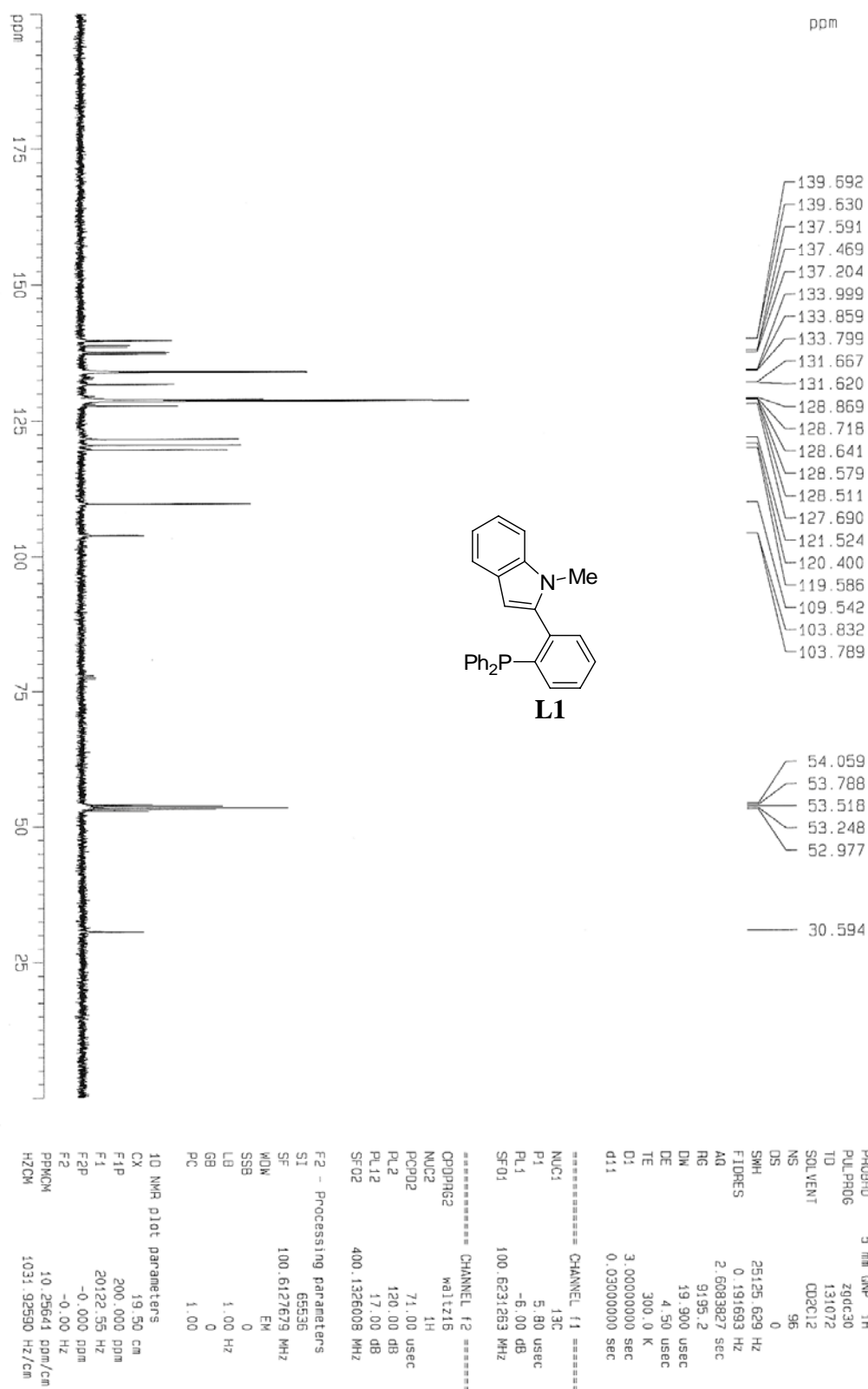
===== CHANNEL f1 =====
 NUC1 31P
 P1 7.20 usec
 PL1 -5.00 dB
 SF01 161.985115 MHz

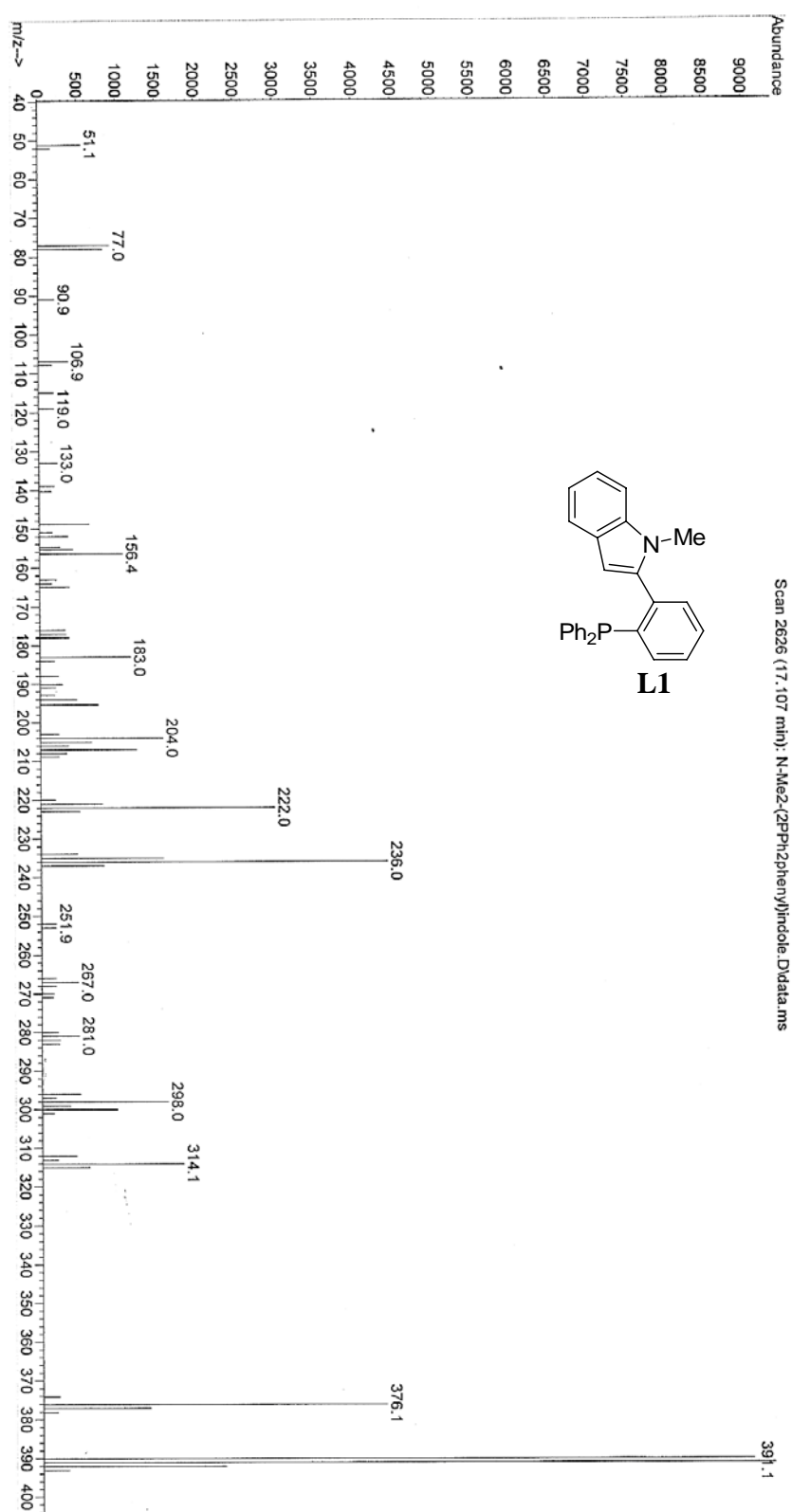
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 71.00 usec
 PL2 120.00 dB
 PL12 17.00 dB
 SF02 400.1324009 MHz

F2 - Processing parameters
 SI 32768
 SF 161.975931 MHz
 WDM EN
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.40

1D NMR Plot parameters
 CX 20.00 cm
 F1P 120.000 ppm
 F1 19437.07 Hz
 F2P -60.000 ppm
 F2 -9718.54 Hz
 PPMCH 9.00000 ppm/cf
 HZCM 1457.78027 Hz/cm







Elemental Composition Report

Single Mass Analysis

Tolerance = 100.0 PPM / DBE: min = -1.5, max = 60.0

Selected filters: None

Monoisotopic Mass, Even Electron Ions

9 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

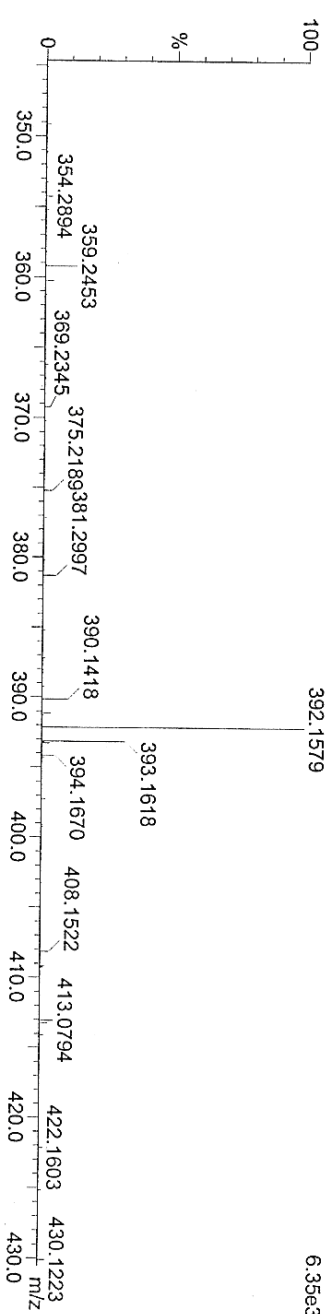
C: 0-27 H: 0-1000 N: 0-1 P: 0-1

So Chau Ming, 2-(N-methylindolyl)phenylPh₂phosphine

HR07_0615_766 (1.249) AM (Cen.5, 80.00, Ar,10000.0,0.00,0.70); Cm (66.77)

392.1579

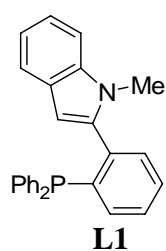
15-Jun-2007
TOF MS ES+
6.35e3

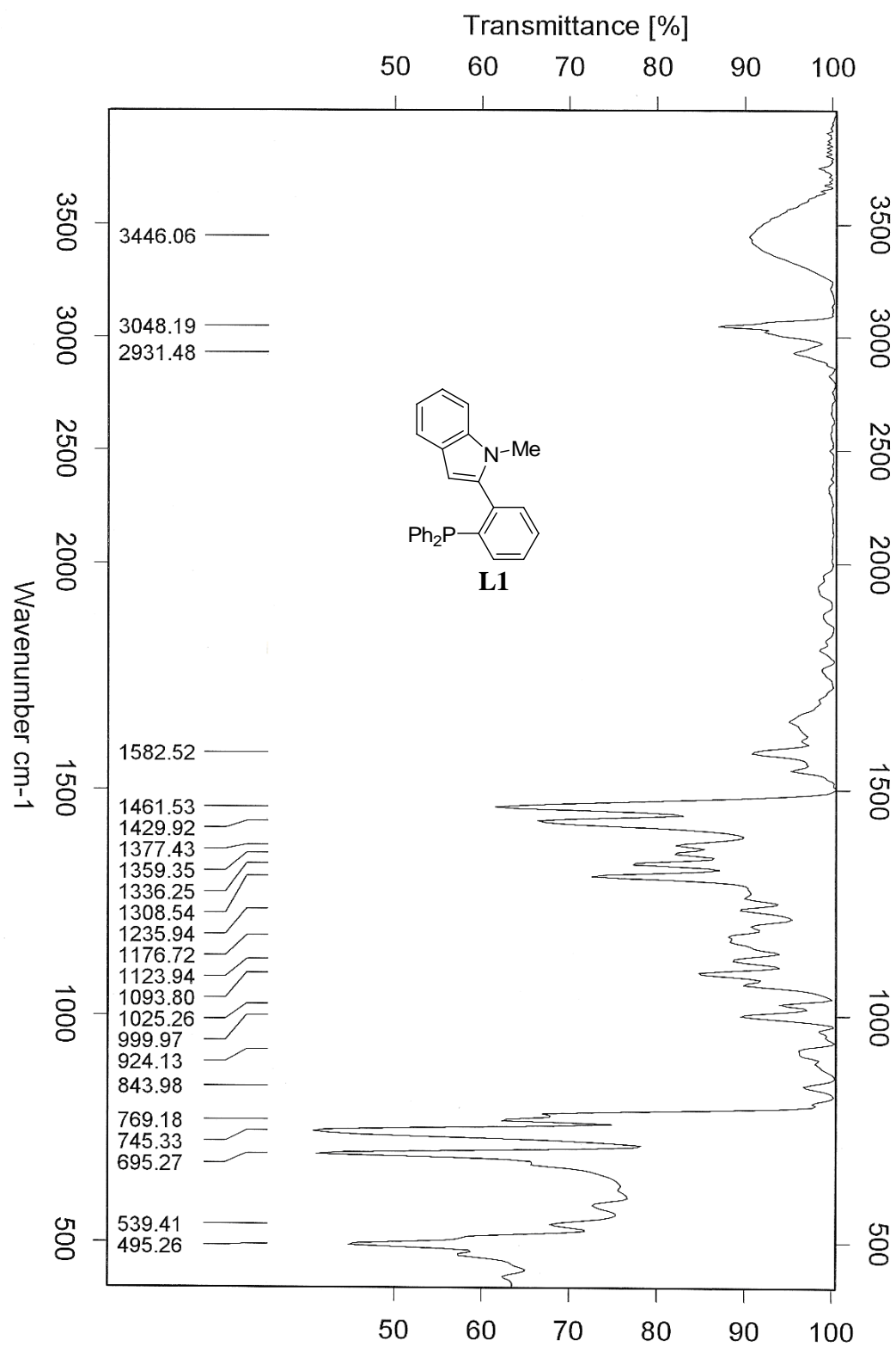


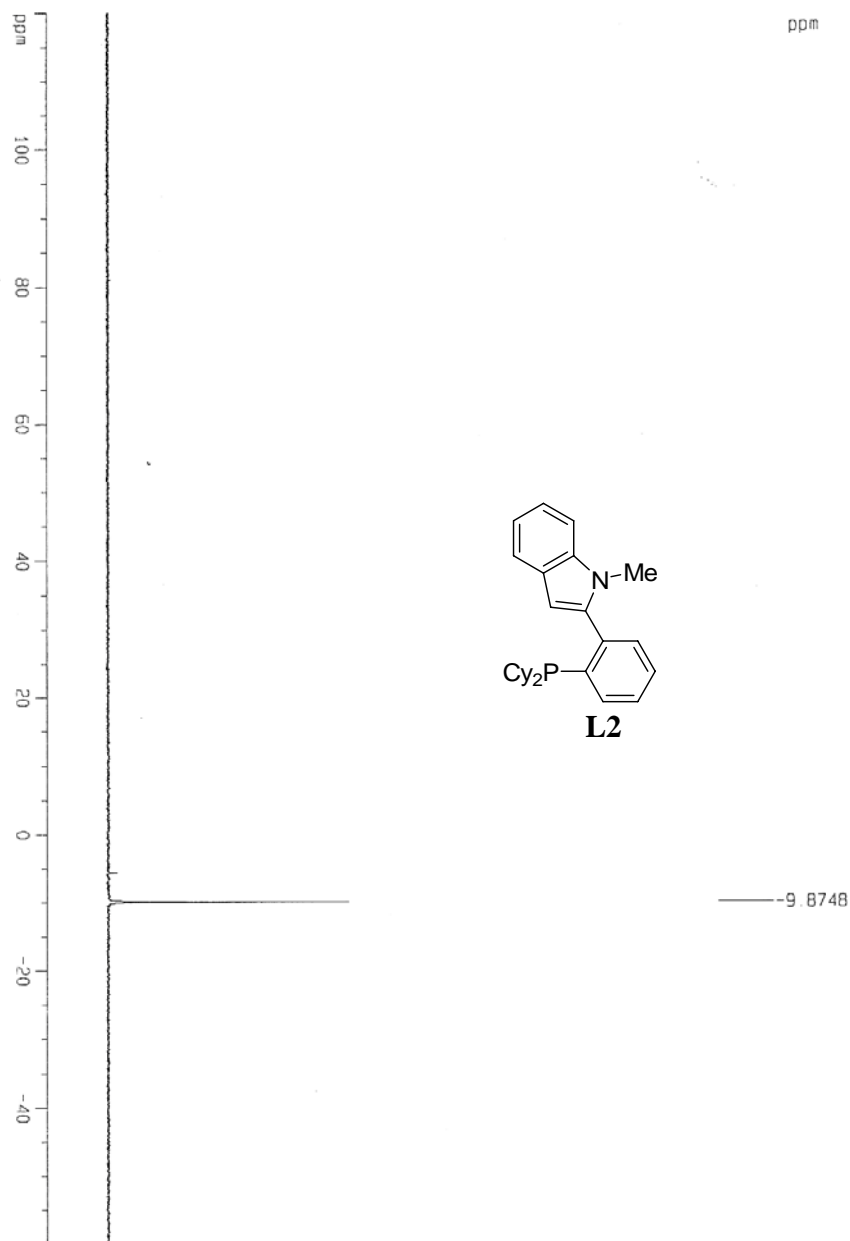
Minimum:
Maximum:

5.0 100.0 -1.5
60.0

Mass	Calc. Mass	mda	PPM	DBE	I-FIT	Formula
392.1579	392.1568	1.1	2.8	17.5	1.2	C ₂₇ H ₂₃ NP







Current Data Parameters
 NAME PCPindole
 EXPNO 31
 PROCNO 1

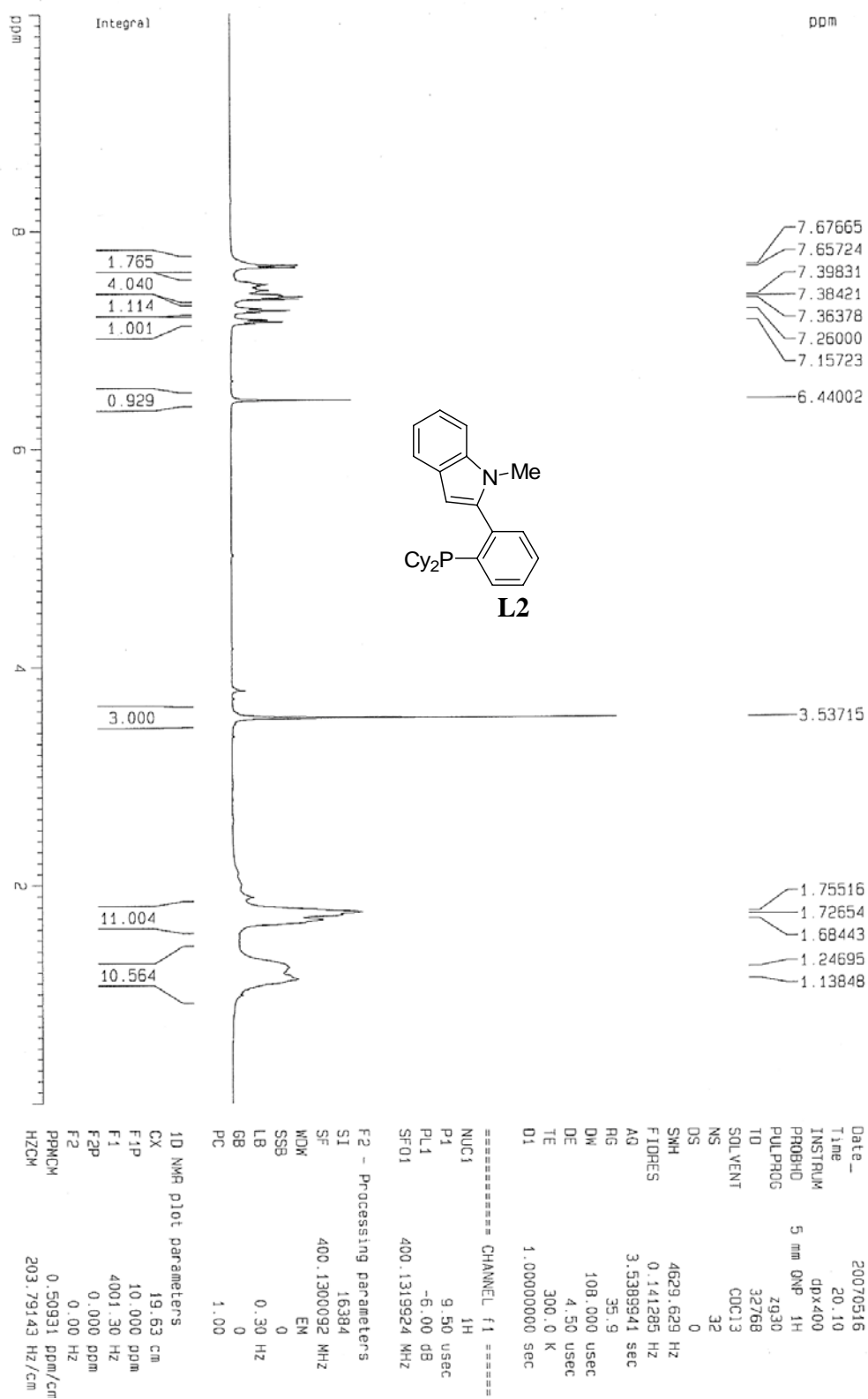
F2 - Acquisition Parameters
 Date_ 20070516
 Time 20.50
 INSTRUM dx400
 PROBHD 5 mm QNP 1H
 PULPROG zgpg30
 TO 65536
 SOLVENT CDCl₃
 NS 16
 DS 0
 SHH 48661.801 Hz
 FIDRES 0.742520 Hz
 AQ 0.6734324 sec
 RG 13004
 DM 10.275 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec

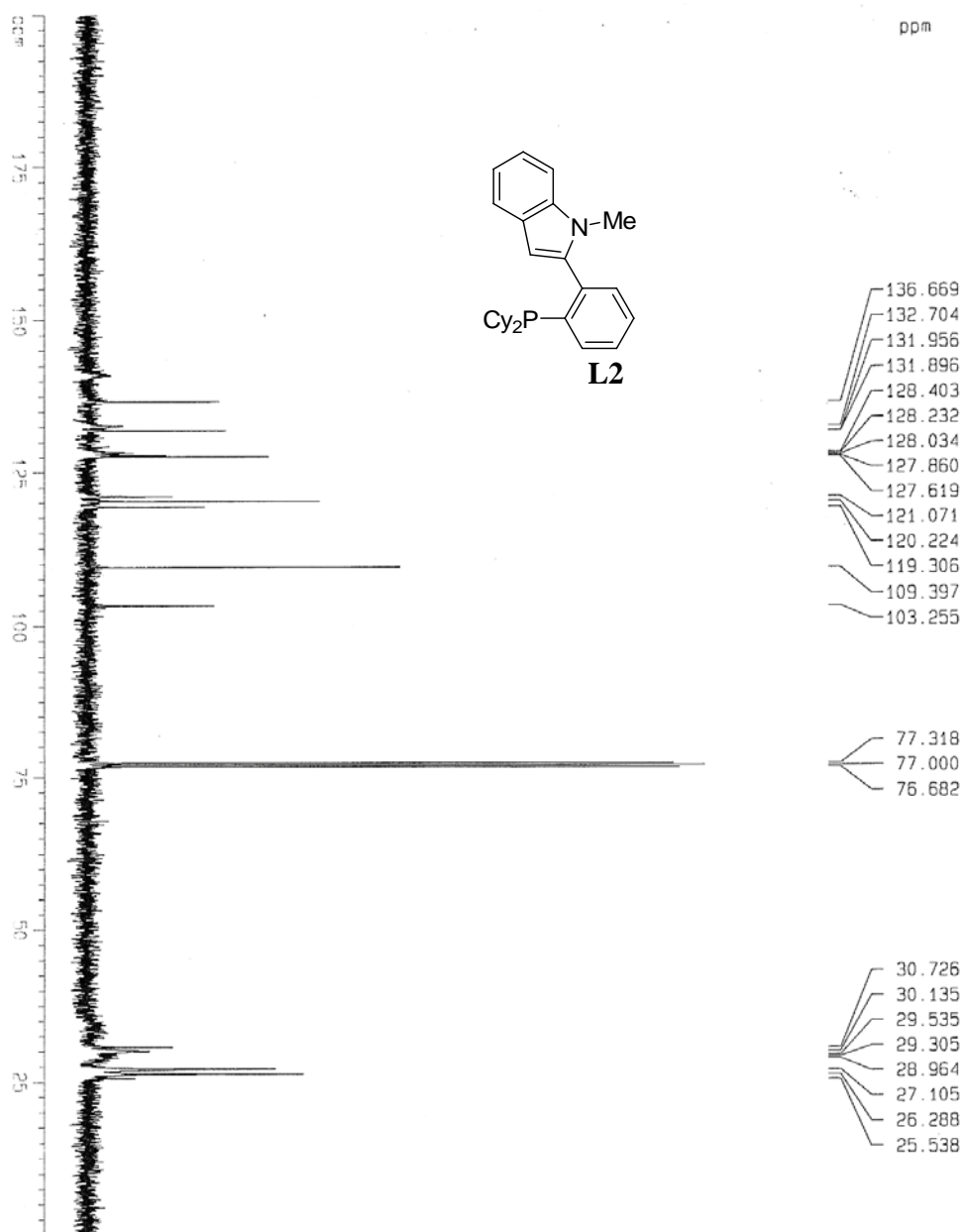
===== CHANNEL f1 =====
 NUC1 31P
 P1 7.20 usec
 PL1 -5.00 dB
 SF01 161.985115 MHz

===== CHANNEL f2 =====
 CPOPRG2 waltz16
 NUC2 1H
 PCPD2 71.00 usec
 PL2 120.00 dB
 PL12 17.00 dB
 SF02 400.1324008 MHz

F2 - Processing parameters
 SI 32768
 SF 161.9756831 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 120.000 ppm
 F1 19437.07 Hz
 F2P -60.000 ppm
 F2 -9718.54 Hz
 PPMCM 9.00000 ppm/cm
 HZCM 1457.78027 Hz/cm





Current Data Parameters

NAME	CCPModule
EXRNO	21
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070516	20.14

INSTRUM 5 mm QNP 1H
PROBHD zgpg30
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 128
DS 0
SWH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.6083827 sec
RG 5195.2
DM 19.900 usec
DE 4.50 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

OPPRG2	wait216
NUC2 <td>1H</td>	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

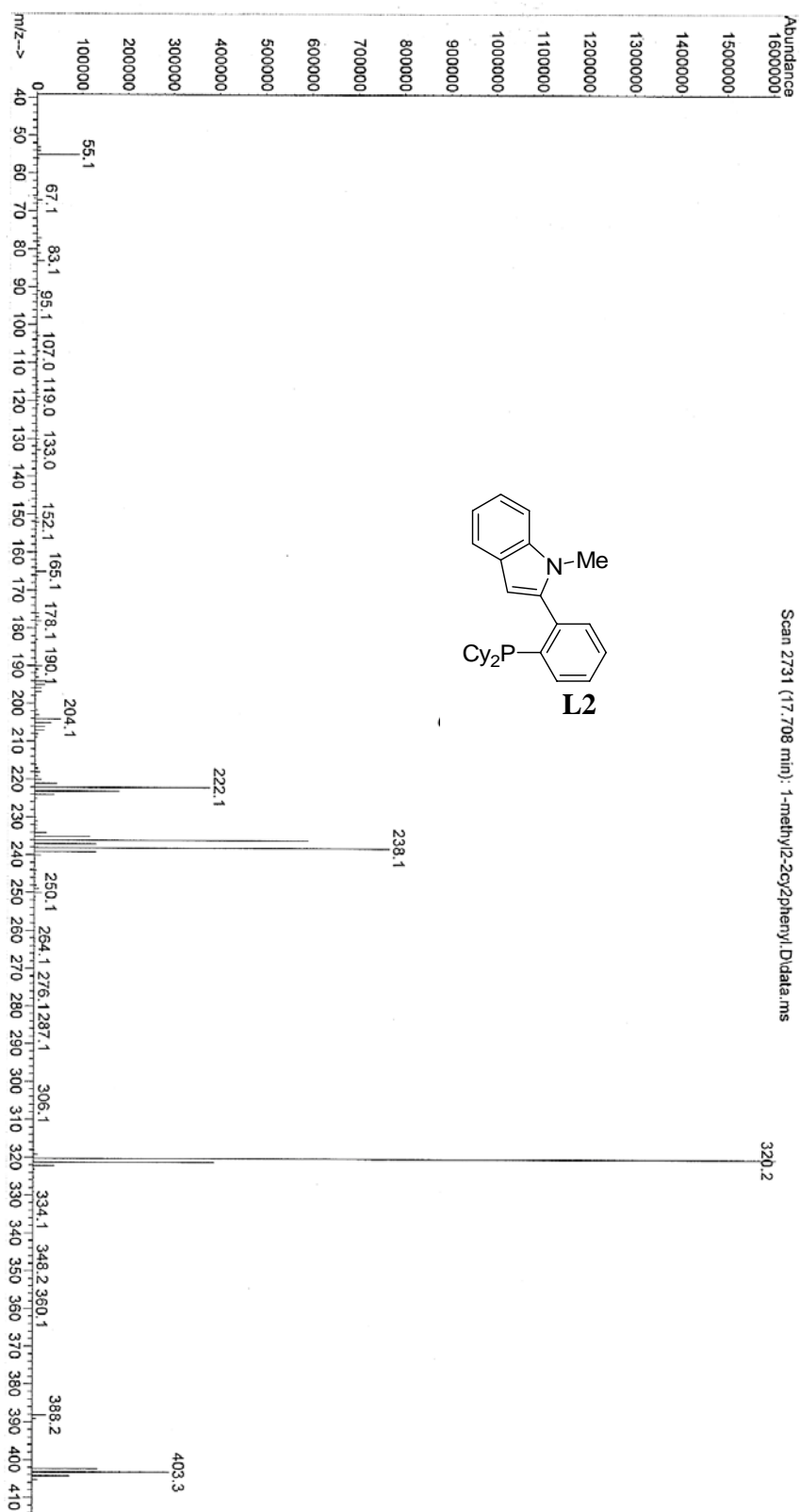
F2 - Processing parameters

SI	65536
SF	100.6127828 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.00

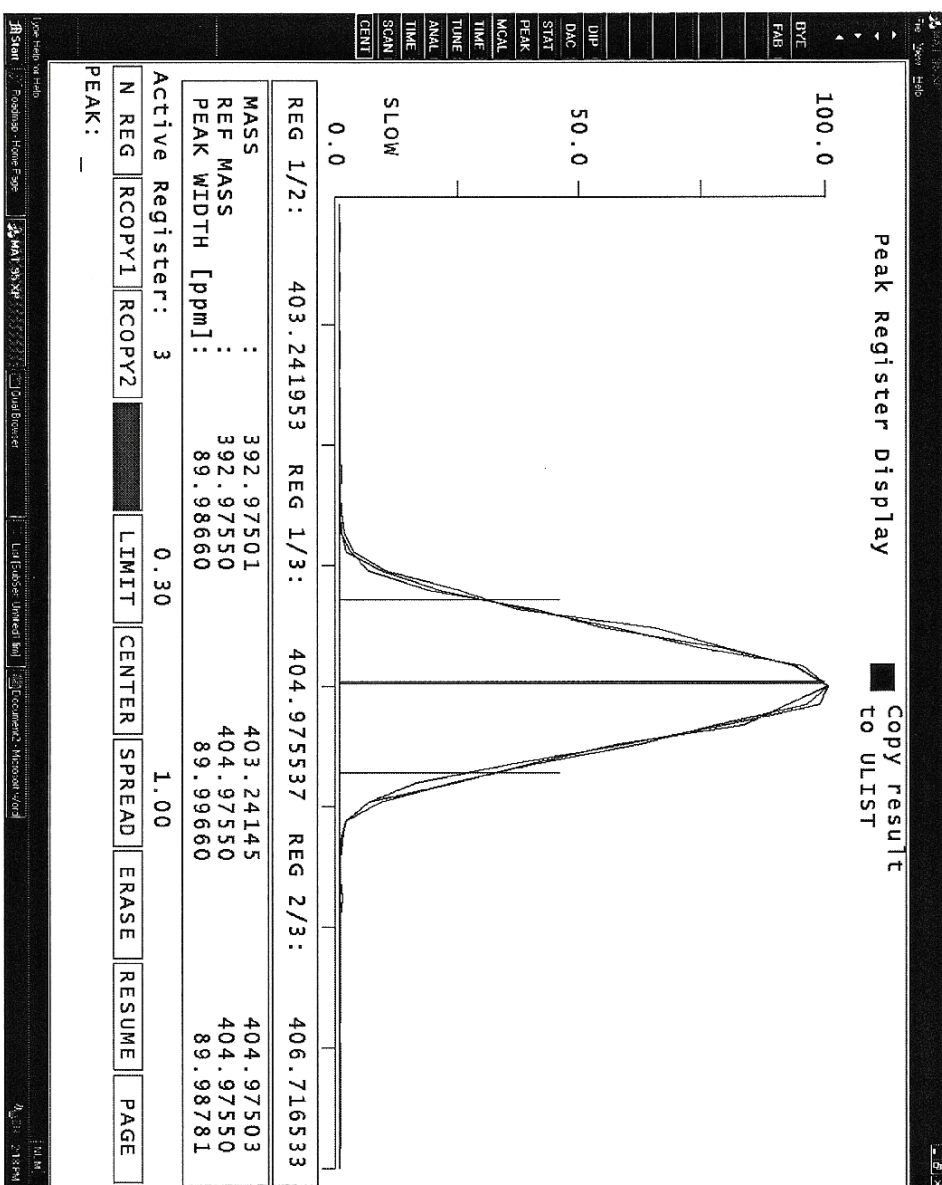
1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCK	10.25641 ppm/cm
HZCM	1031.92603 Hz/cm

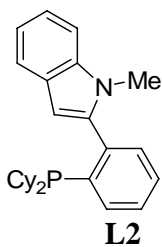
File : C:\MSDCHEM\1\DATA\CMO\C-PHOS\Snapshot\1-methyl2-2cy2phenyl
Operator :
Instrument : 5973N
Acquired : 16 May 2007 16:45 using AcqMethod METHOD2.M
Sample Name :
Misc Info :

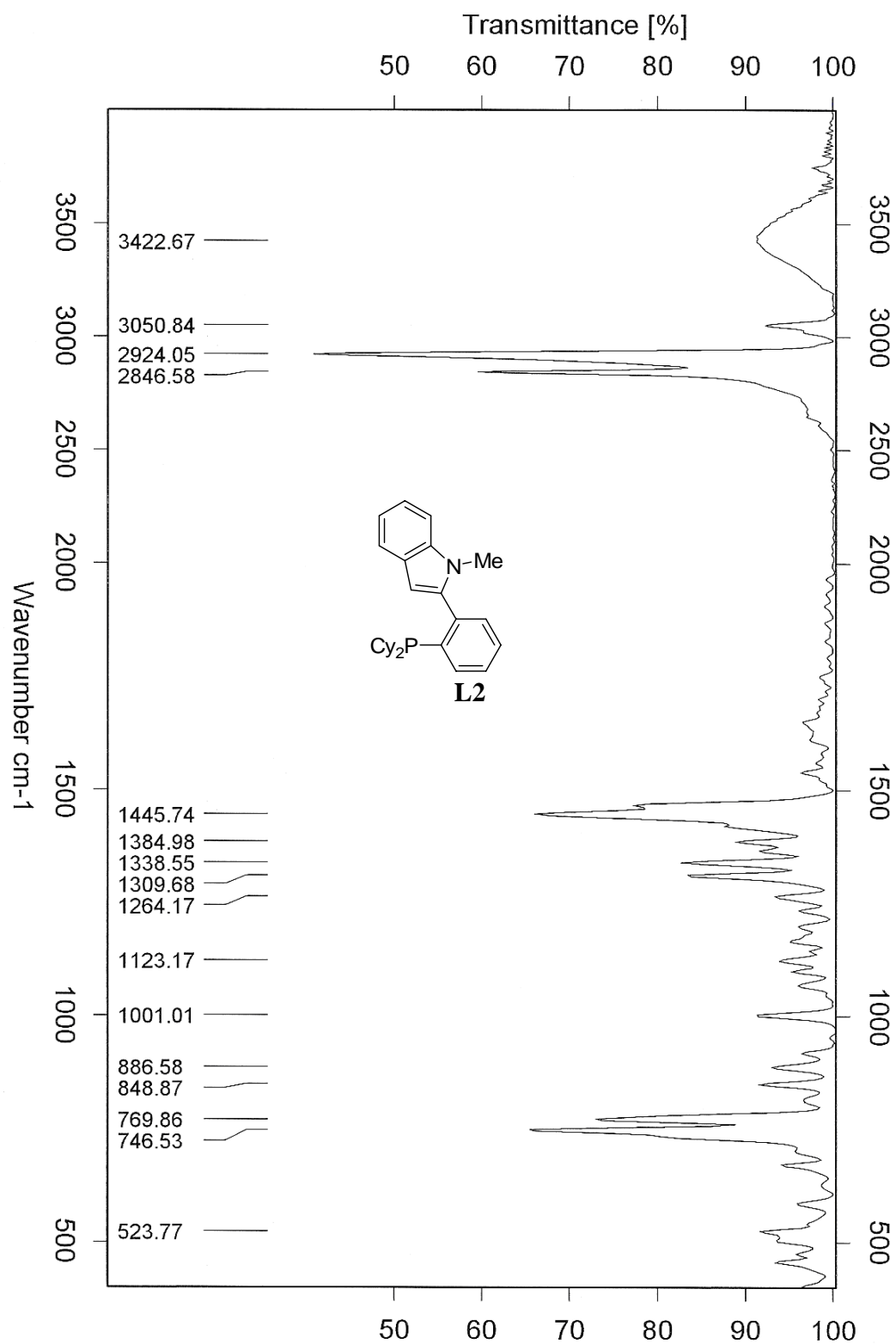


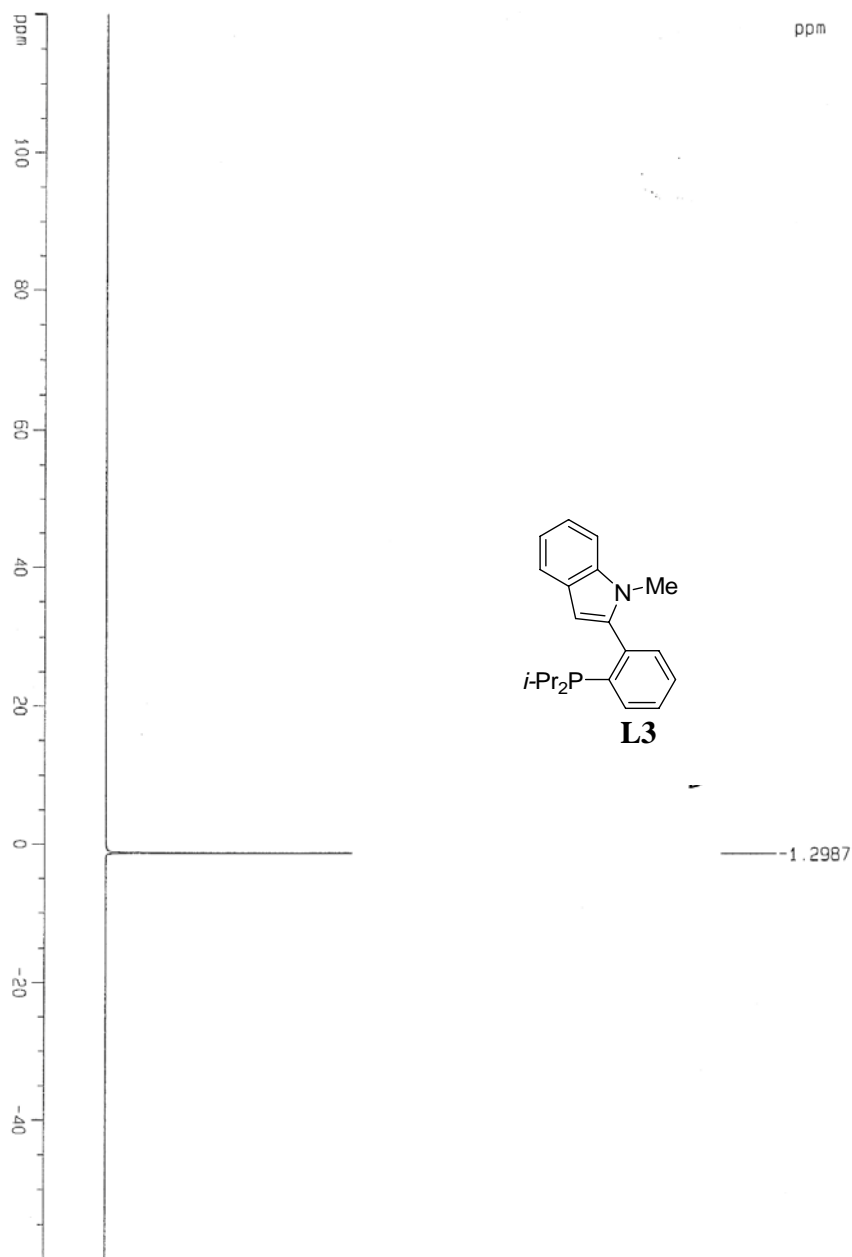
Accurate Mass Measurement:



Molecular formula:
 $C_{27}H_{34}N_1P_1$
 $[M]^+$ (theoretical)
 = 403.2423







Current Data Parameters

NAME	PCPndole
EXPNO	42
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070531	11.06

INSTRUM gp400
PROBHD 5 mm QNP 1H
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 32
DS 0
SMH 49661.801 Hz
FIDRES 0.742520 Hz
AQ 0.6734324 sec
RG 6502
DM 10.275 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	31P
P1	7.20 usec
PL1	-5.00 dB
SFO1	161.9851115 MHz

===== CHANNEL f2 =====

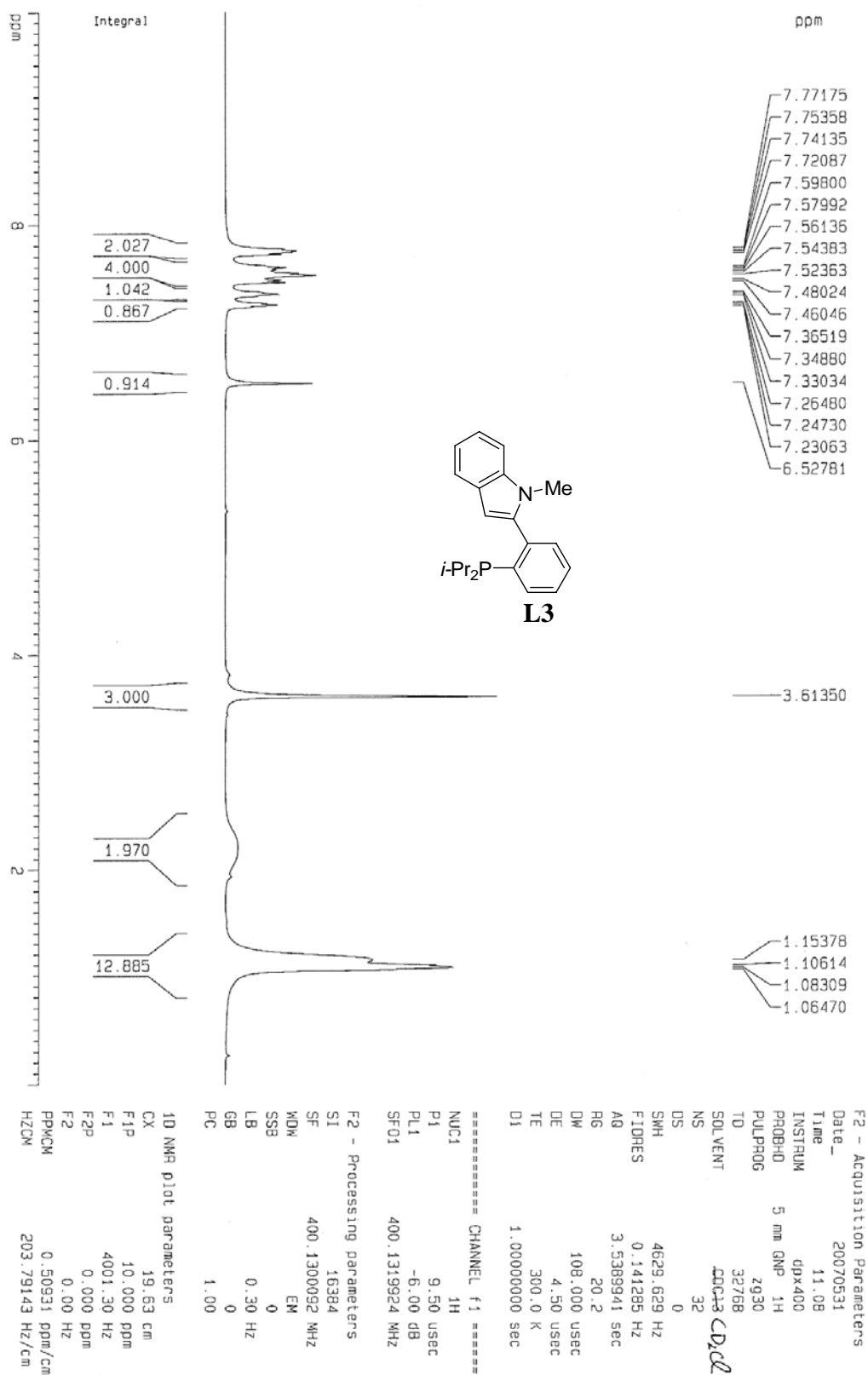
CPDPRG2	waltz15
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1324008 MHz

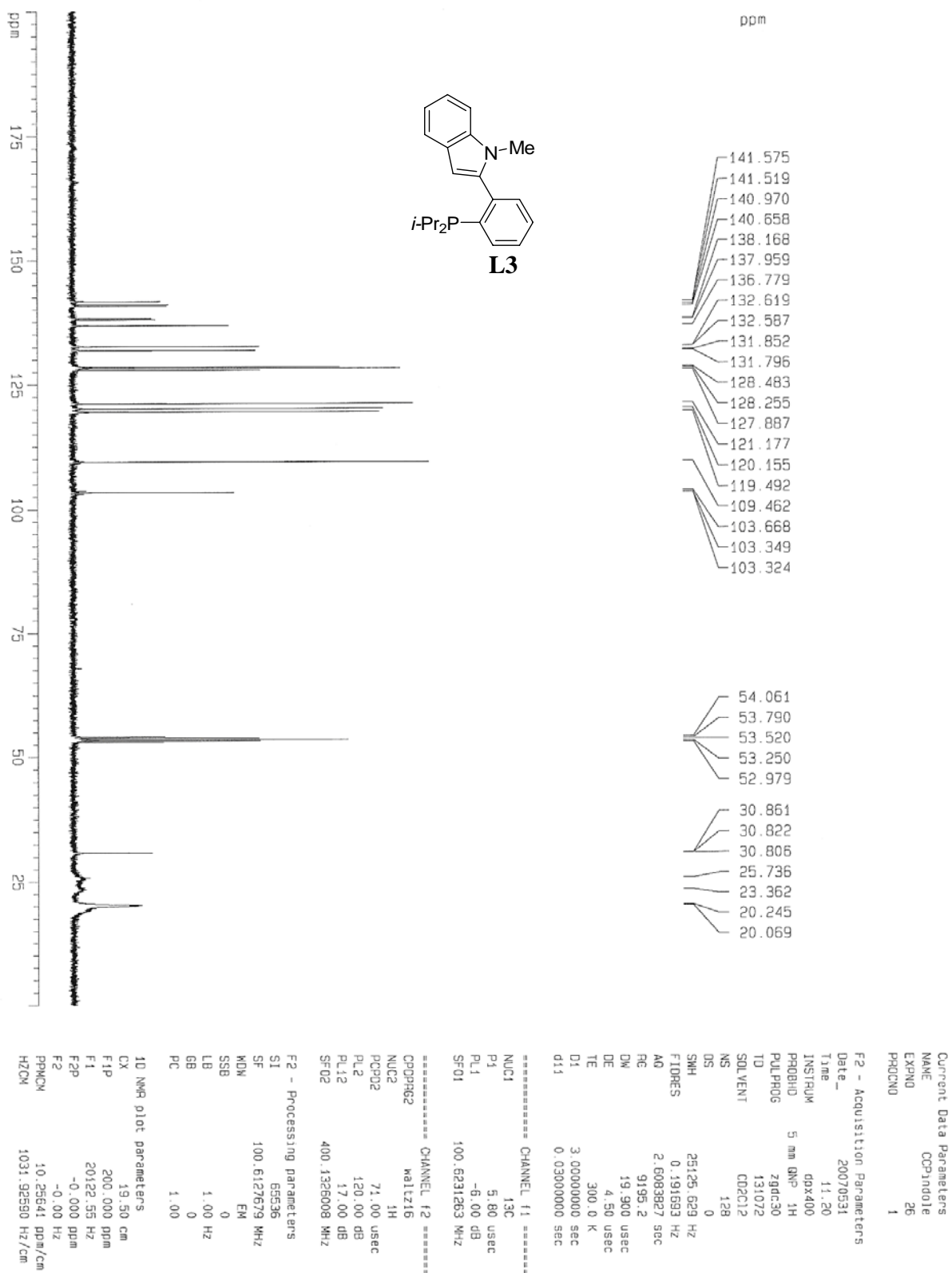
F2 - Processing parameters

SF	32768
SF	161.9755831 MHz
KOM	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.40

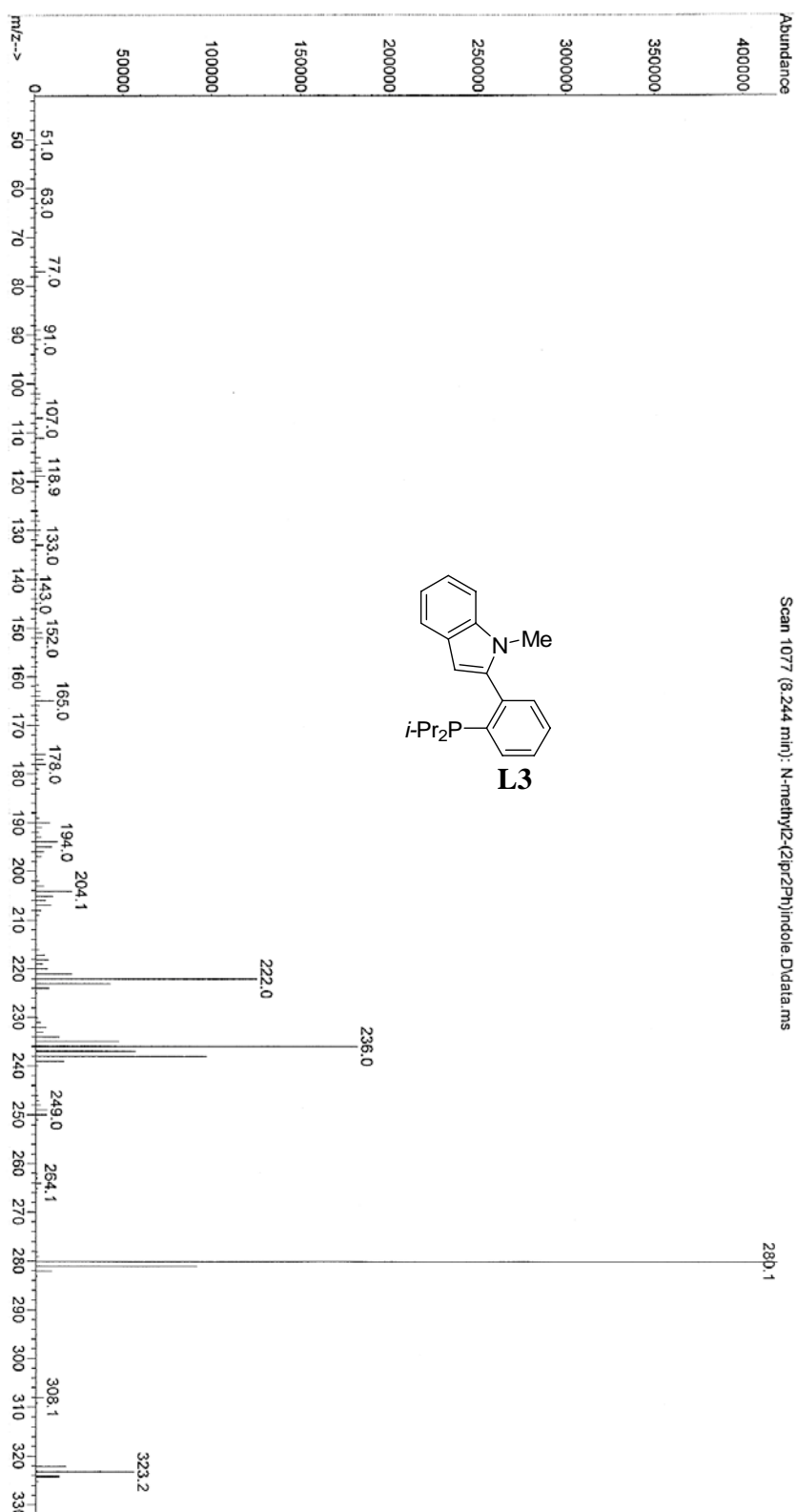
1D NMR Plot Parameters

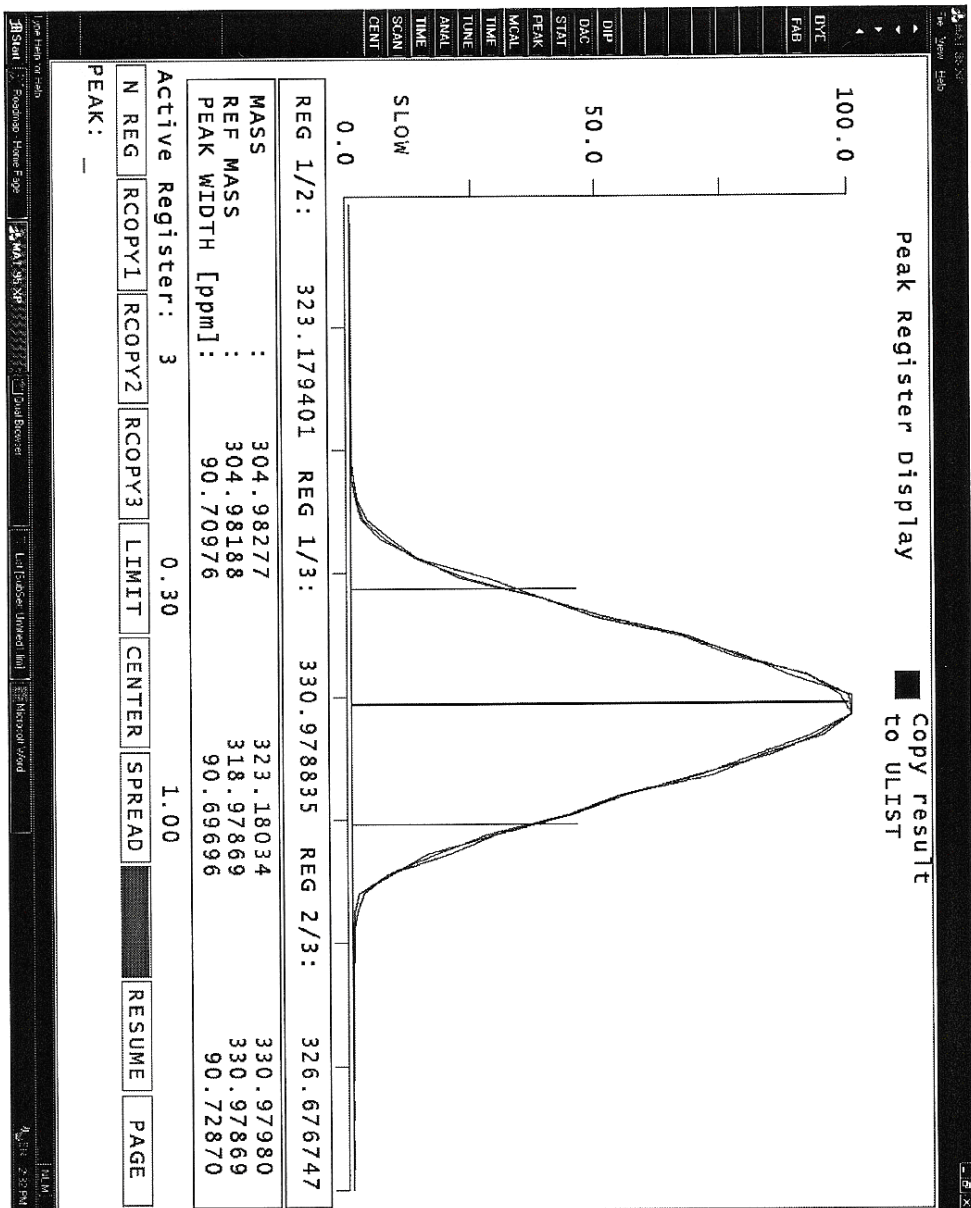
CX	20.00 cm
F1P	120.000 ppm
F1	19437.07 Hz
F2P	-60.000 ppm
F2	-9718.54 Hz
PPMCK	9.00000 ppm/cm
HZCM	1457.79027 Hz/cm



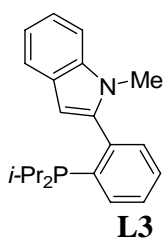


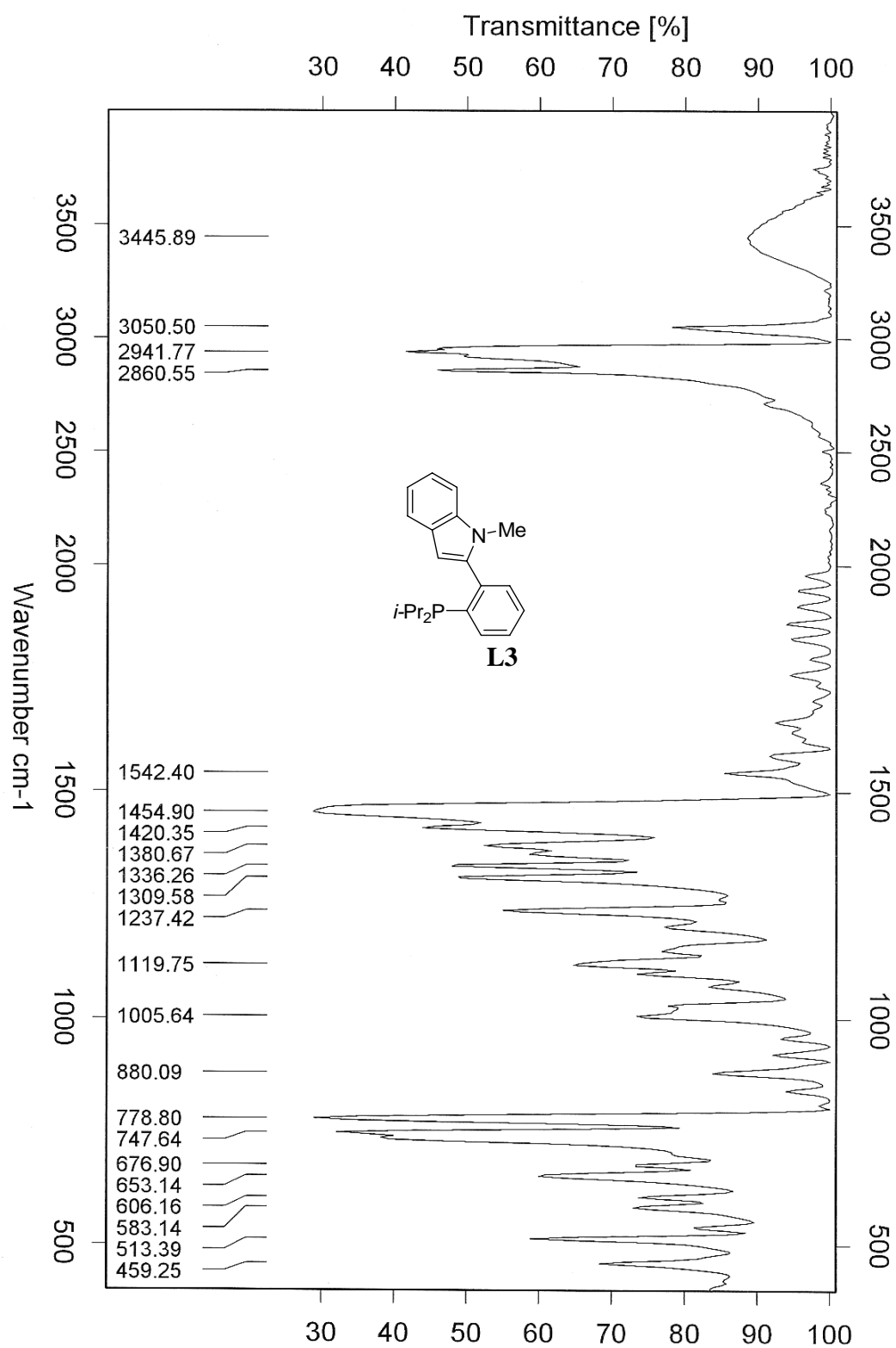
File : C:\msdchem\1\DATA\cmso\C-Pphos\N-methyl2-(2ipr2Ph)indole.D
Operator :
Acquired : 30 May 2007 20:57 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 8

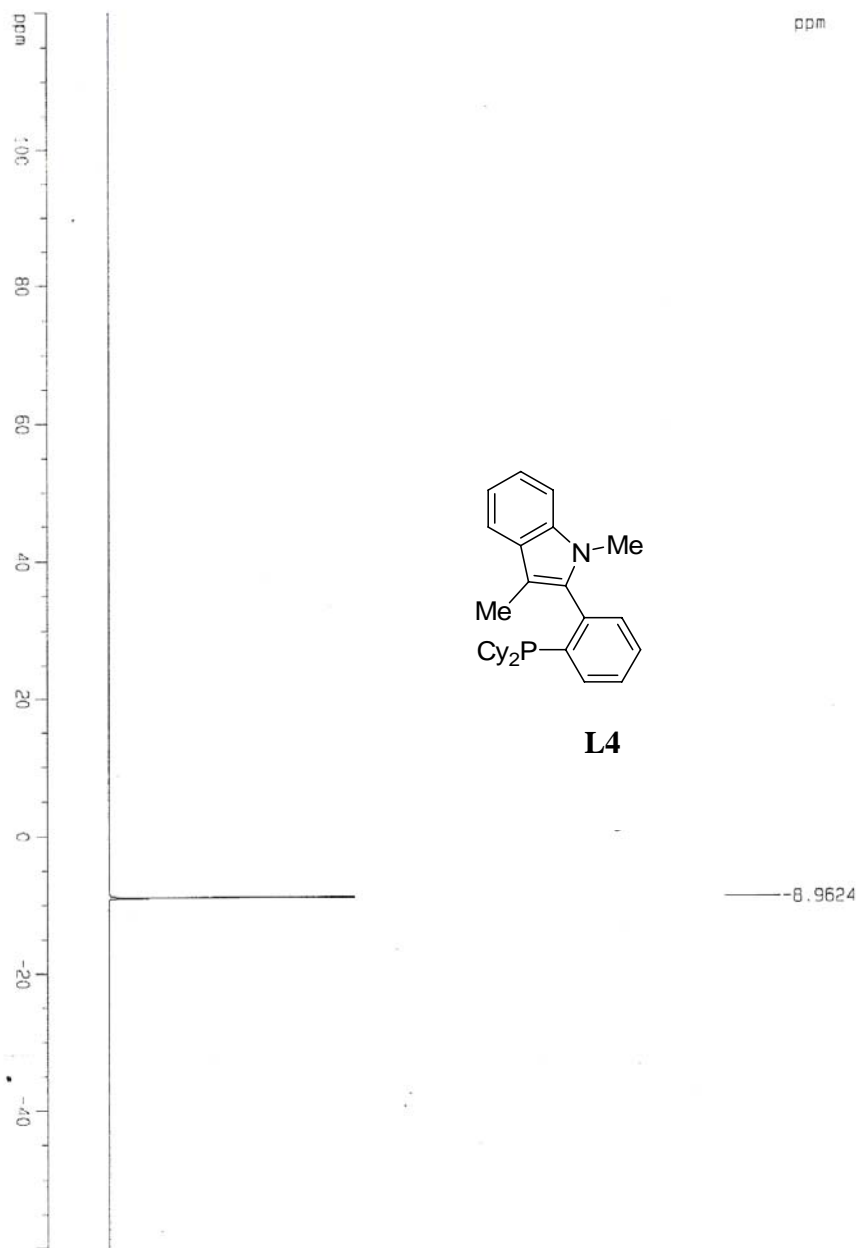


Accurate Mass Measurement:

Molecular formula:
 $C_{21}H_{26}N_1P_1$
 $[M]^+$ (theoretical)
 = 323.1797







Current Data Parameters
 NAME PCP.indole
 EXPNO 80
 PROCNO 1

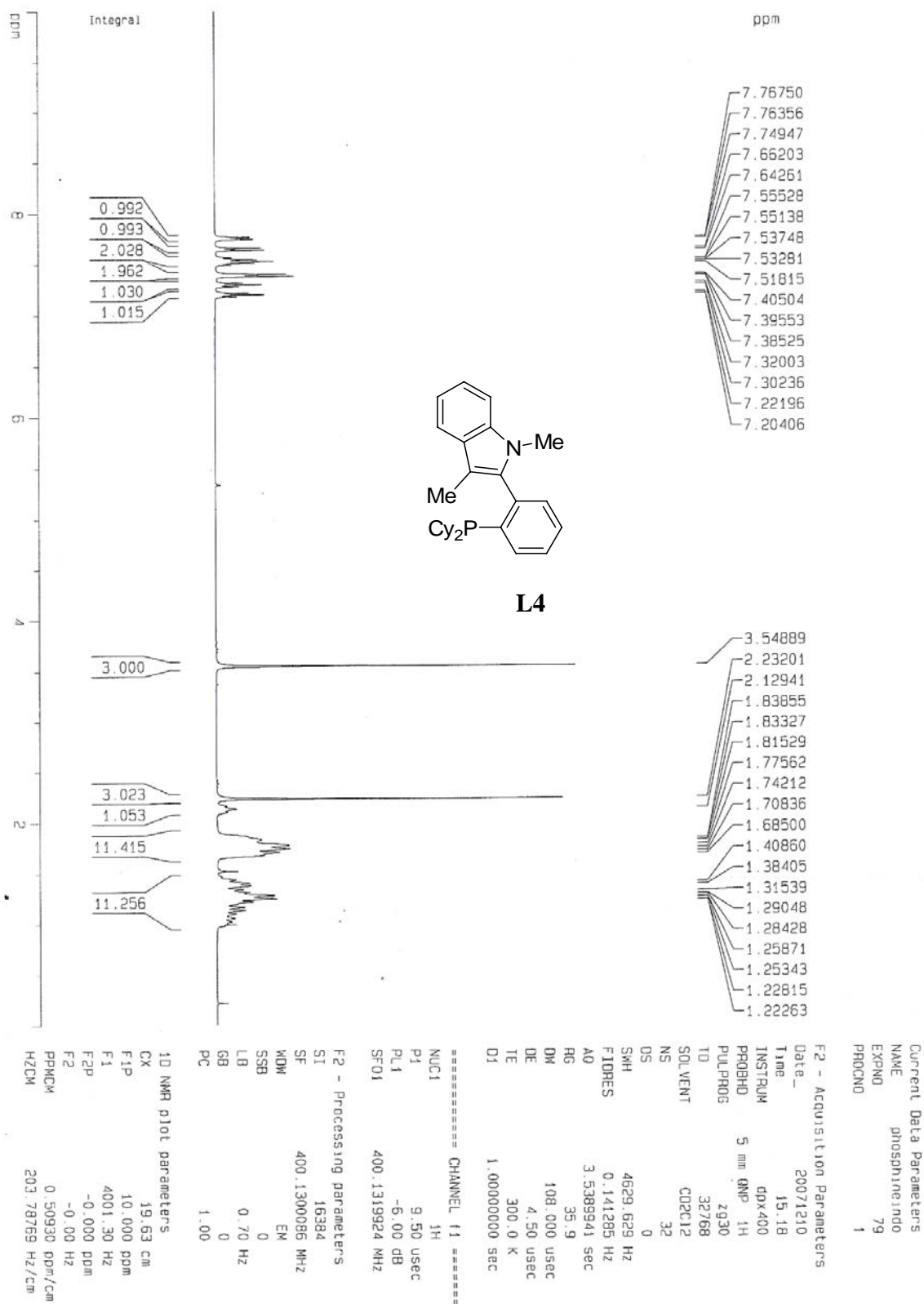
F2 - Acquisition Parameters
 Date_ 20071210
 Time 15.16
 INSTRUM spect
 PROBHD 5 mm QNP 1H
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 0
 SWH 48661.801 Hz
 FIDRES 0.742520 Hz
 AQ 0.6734324 sec
 RG 6502
 DM 10.275 usec
 DE 4.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec

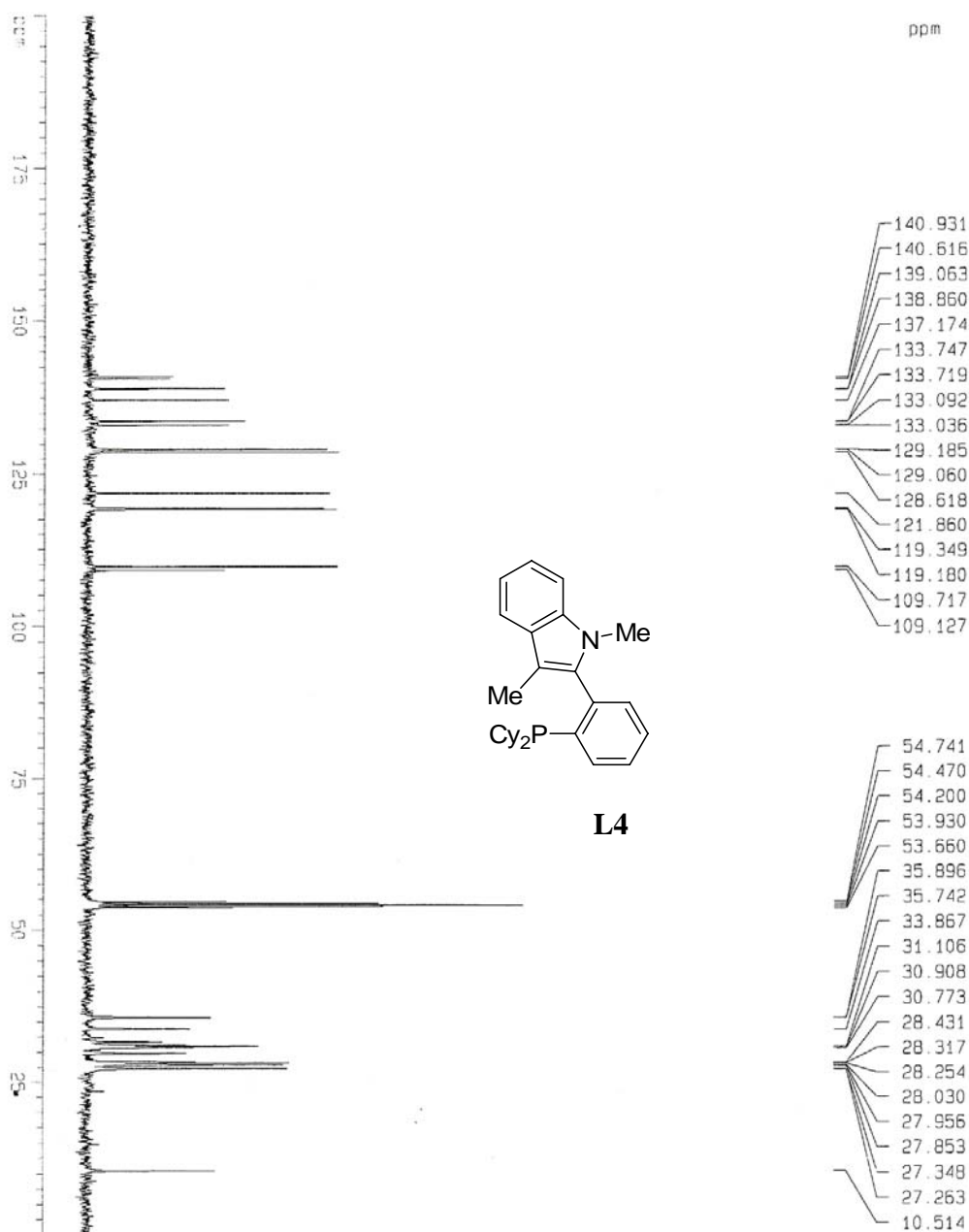
===== CHANNEL f1 =====
 NUC1 13C
 P1 7.20 usec
 PL1 -5.00 dB
 SF01 161.9851115 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 71.00 usec
 PL2 120.00 dB
 PL12 17.00 dB
 SF02 400.1324008 MHz

F2 - Processing parameters
 SI 32768
 SF 161.9755831 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.40

1D NMR parameters
 CX 20.00 cm
 F1P 120.000 DDM
 F1 19437.07 Hz
 F2P -60.000 DDM
 F2 -9718.54 Hz
 PPMCM 9.00000 DDM/cm
 HZCM 1457.78027 Hz/cm





Current Data Parameters
 NAME: CPlndole
 EXPNO: 34
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20071210
 Time: 15.21
 INSTRUM: gp400
 PULPROG: zgpg30
 TO: 131072
 SOLVENT: CDCl3
 NS: 115
 DS: 0
 SM: 25125.629 Hz
 FIDRES: 0.151693 Hz
 AQ: 2.6083827 sec
 RG: 5792.6
 DW: 19.900 usec
 DE: 4.50 usec
 TE: 300.0 K
 D1: 3.00000000 sec
 d11: 0.03000000 sec

===== CHANNEL f1 =====
 NUC1: 13C
 P1: 5.80 usec
 PL1: -6.00 dB
 SFO1: 100.6231263 MHz

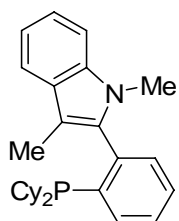
===== CHANNEL f2 =====
 CPOPRG2: waltz16
 NUC2: 1H
 PCPD2: 71.00 usec
 PL2: 120.00 dB
 PL12: 17.00 dB
 SFO2: 400.1326008 MHz

F2 - Processing parameters
 SI: 65536
 SF: 100.6126960 MHz
 MDW: EM
 SSB: 0
 LB: 3.00 Hz
 GB: 0
 PC: 1.00

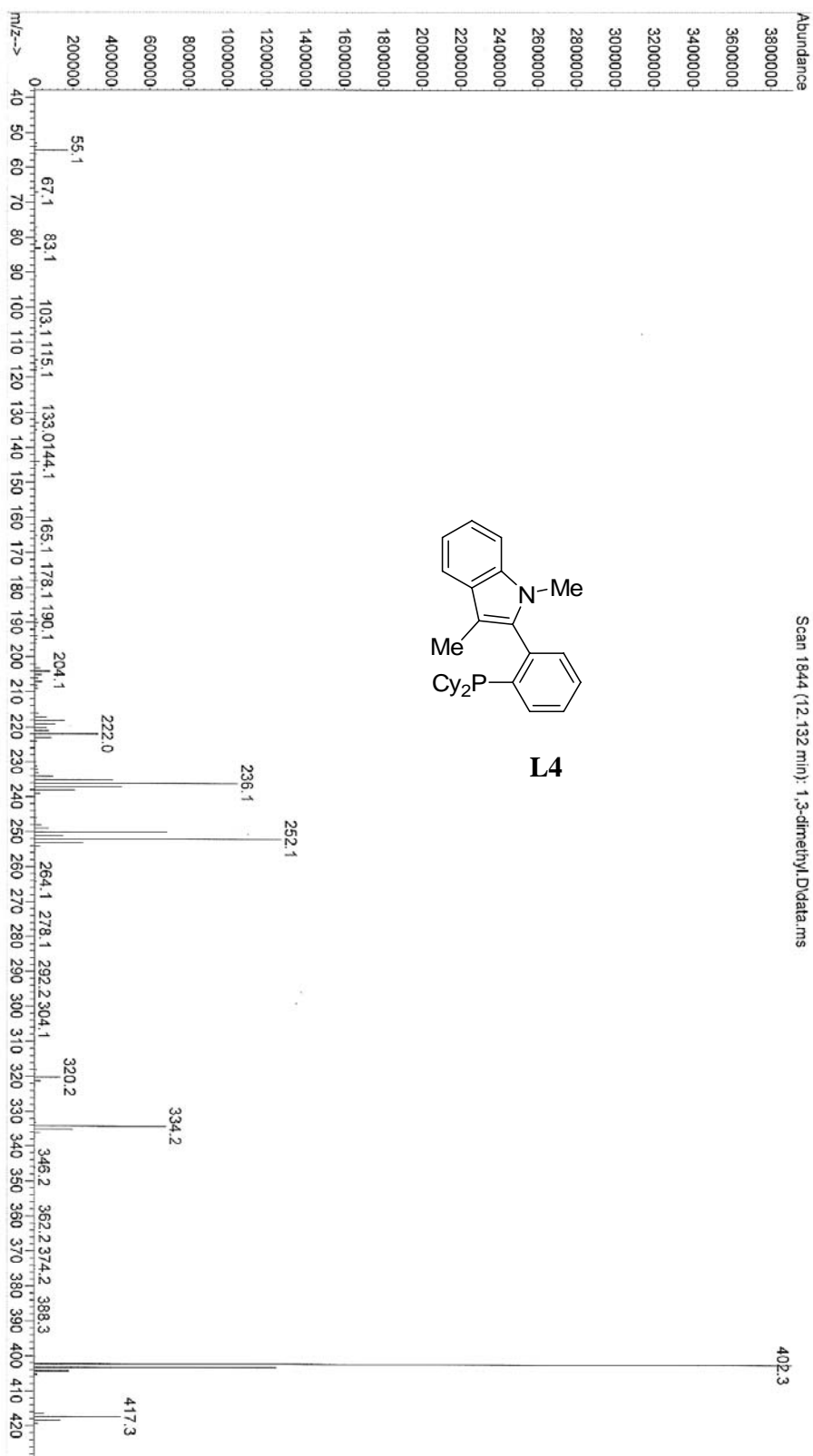
1D NMR plot parameters
 CX: 19.50 cm
 FIP: 200.000 ppm
 F1: 20122.54 Hz
 F2P: -0.000 ppm
 F2: -0.00 Hz
 PPMQV: 10.25641 ppm/cm
 HZCM: 1031.92505 Hz/cm

File : C:\msdchem\1\DATA\cmso\C-Pphos\1,3-dimethyl.D
Operator :
Acquired : 8 Dec 2007 11:20 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 8

Scan 1844 (12.132 min): 1,3-dimethyl.D\data.ms



L4



Single Mass Analysis

Selected filters: None

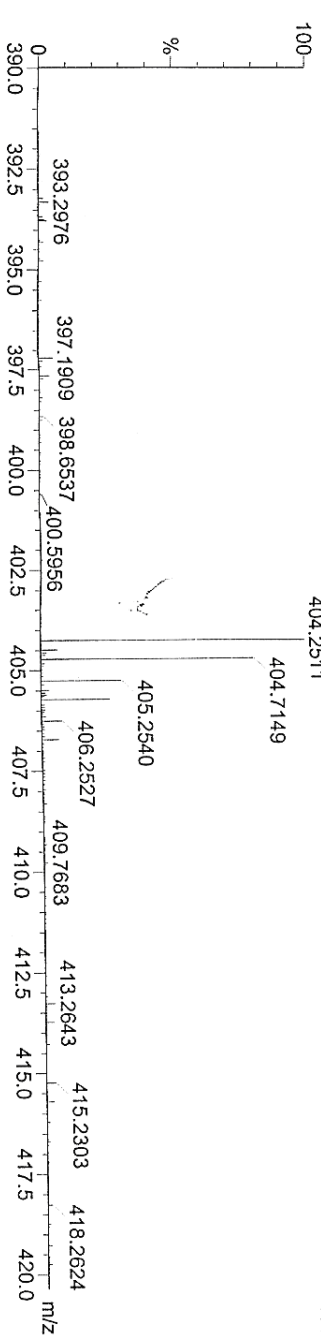
24 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

C:0-28 H:0-1000 N:0-1 O:0-1 P:0-1

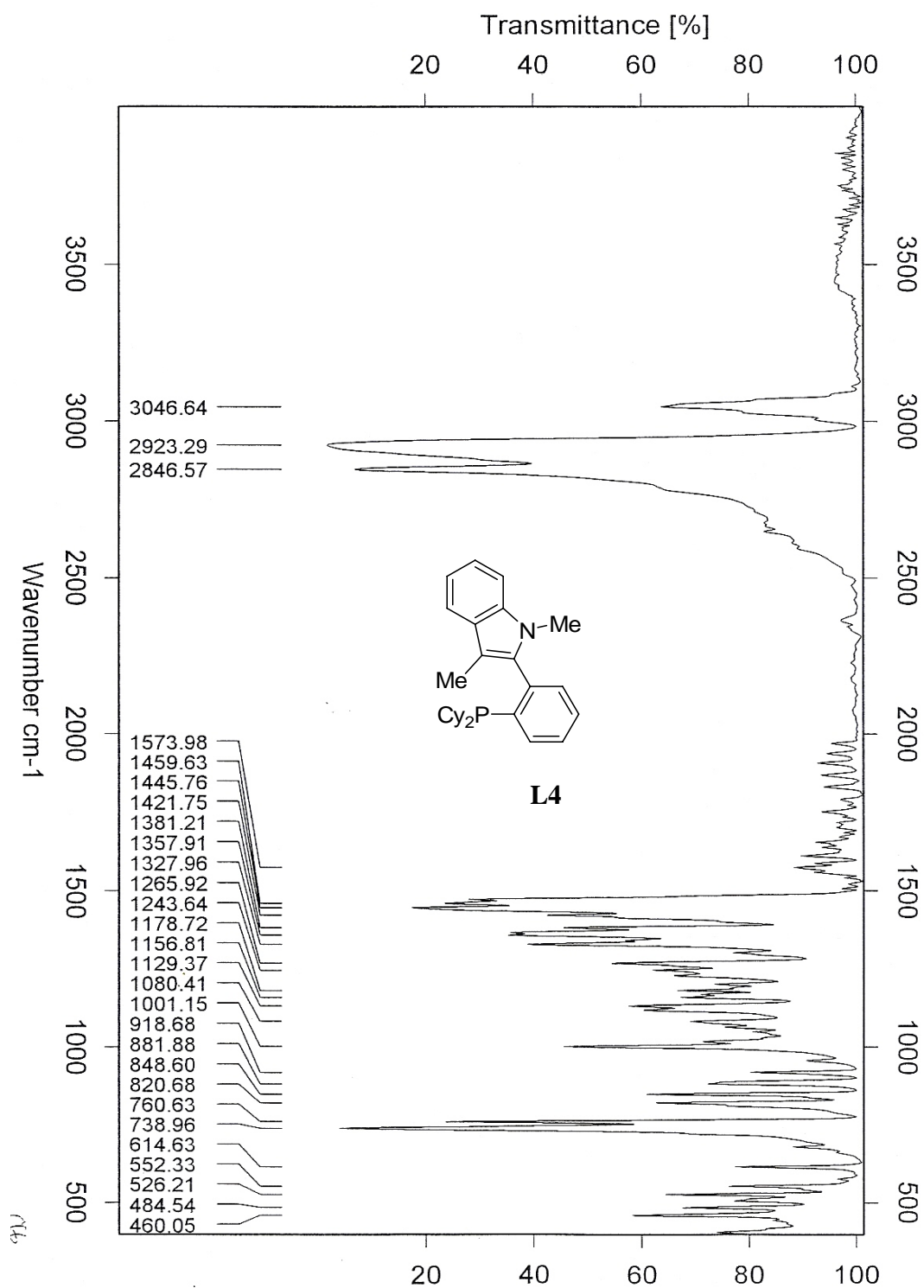
HR07_0517_1 14 (0.271) AM (Cen, 4, 80.00, Ht, 10000.0, 0.00, 1.00); Sm (SG, 2x3.00); Sb (15, 10.00); Cm (13:19)

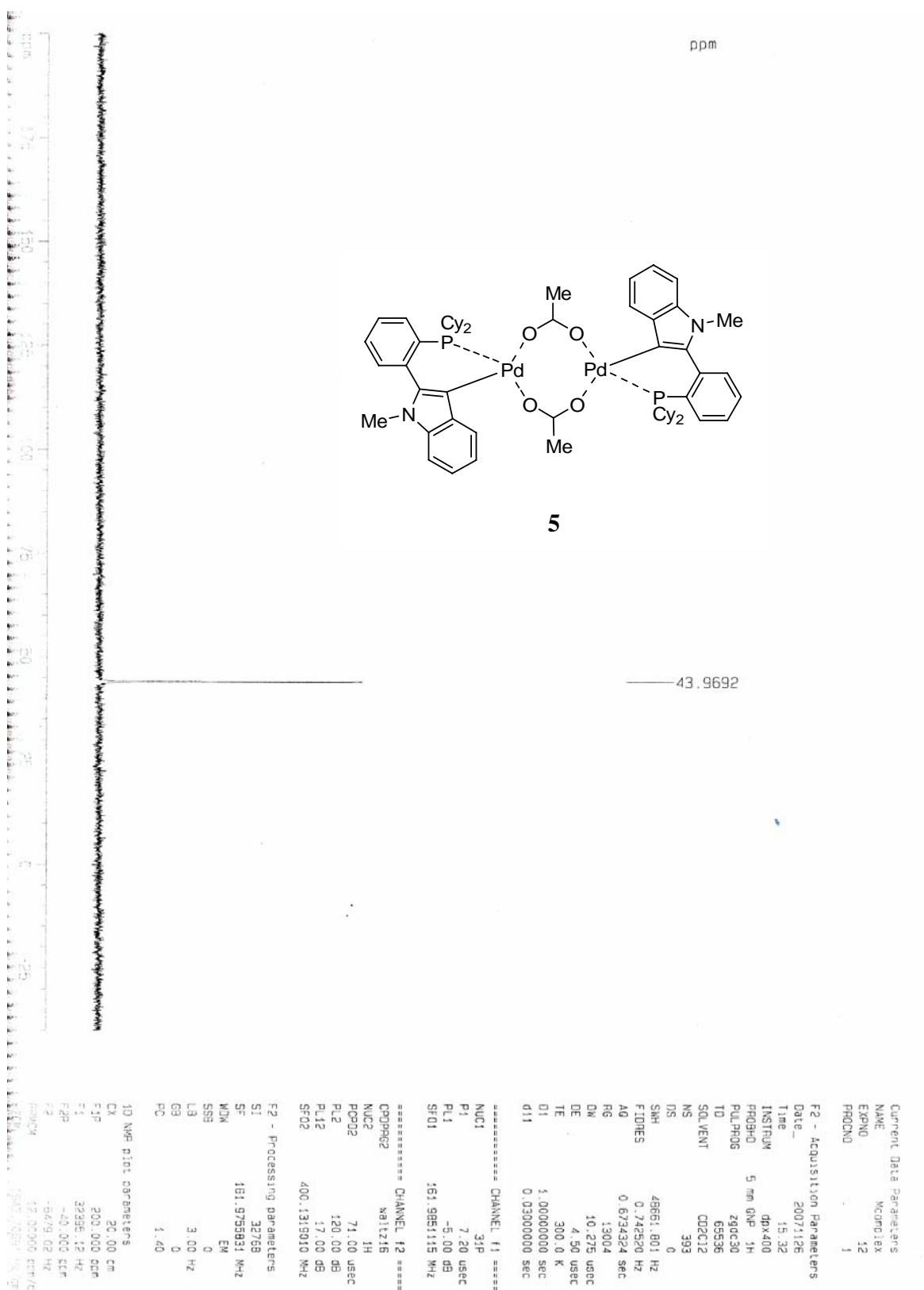
404.7149

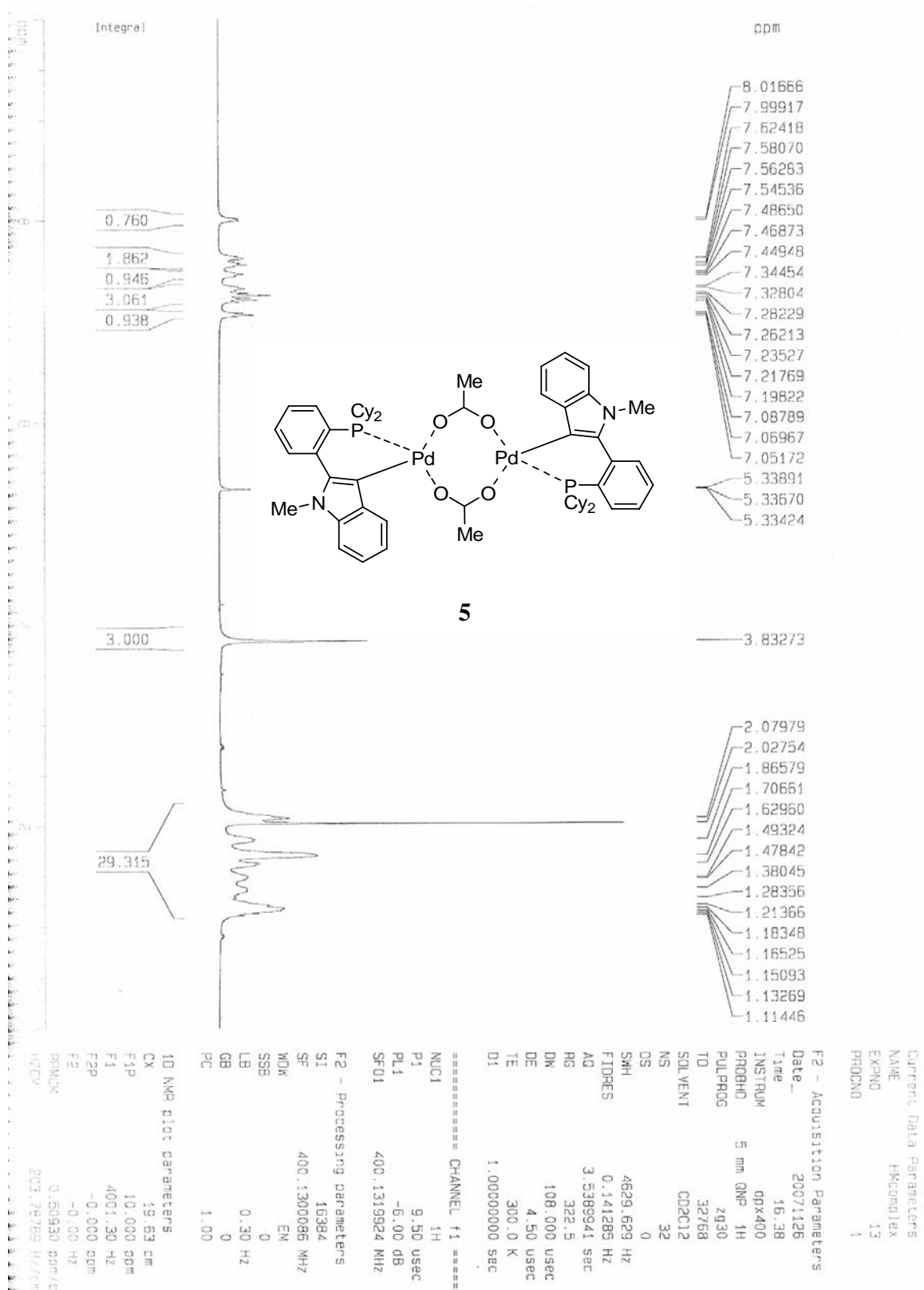
L4

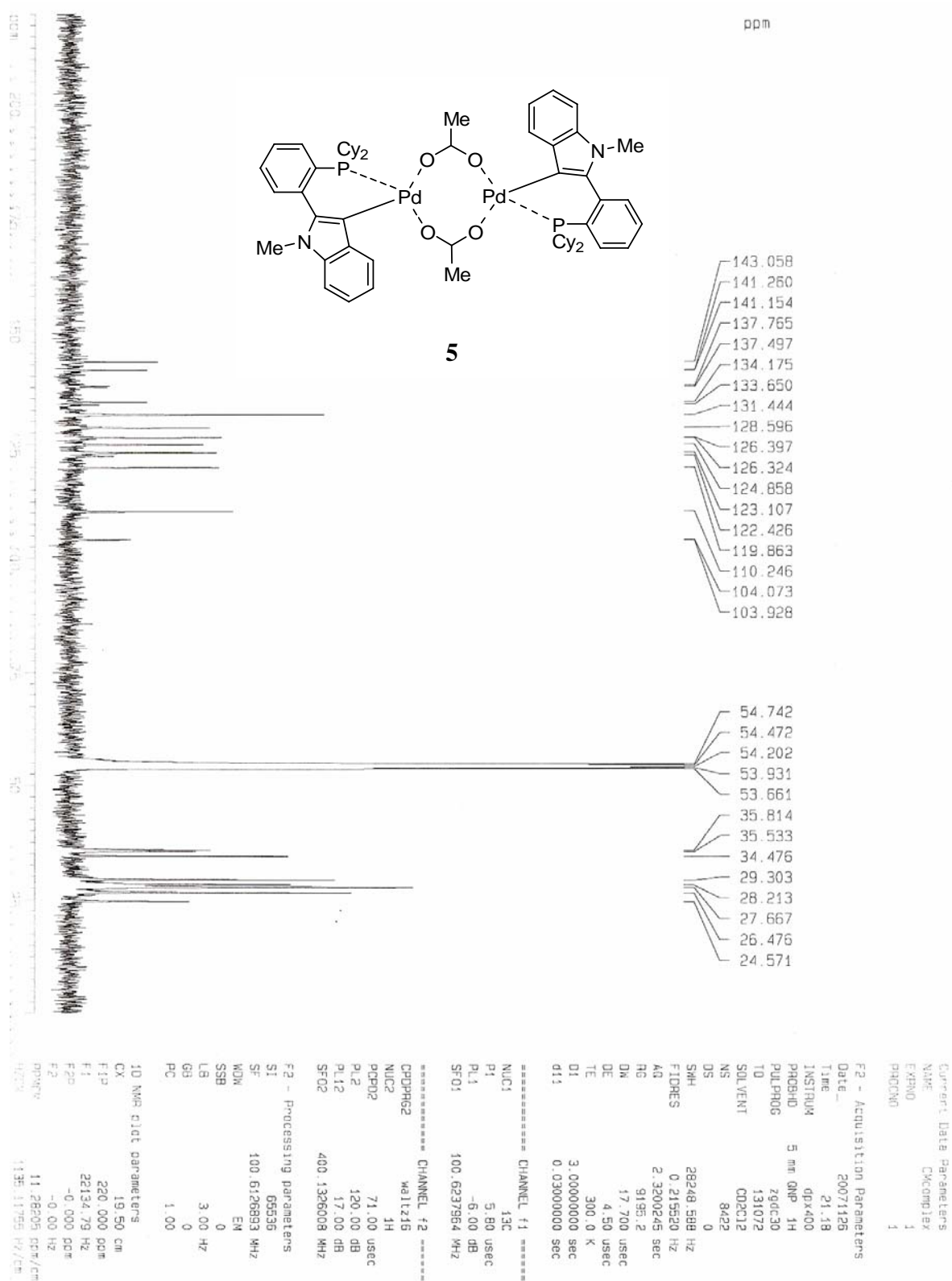


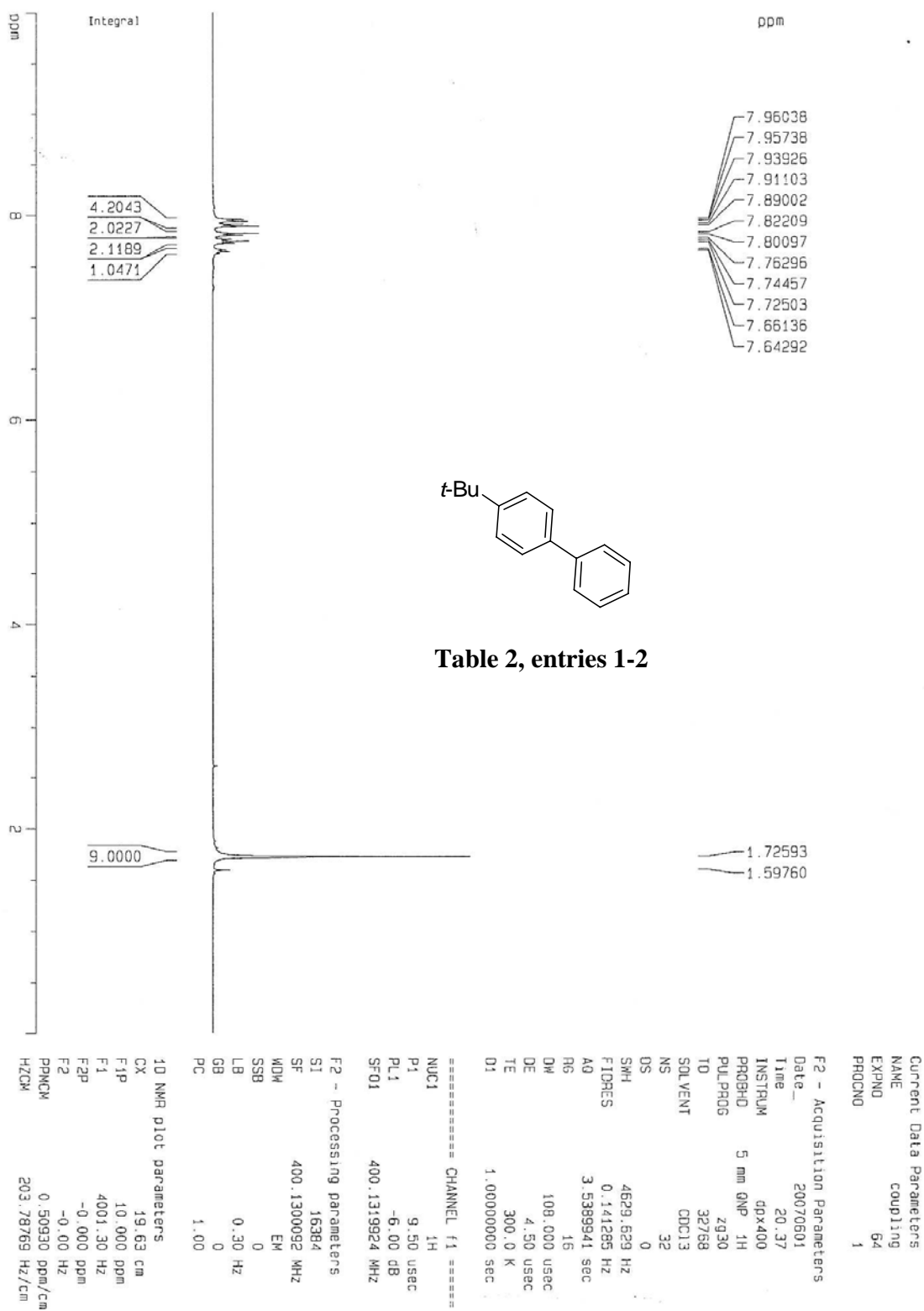
Minimum:				-1.5		
Maximum:			5.0	5.0	60.0	
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	Formula
404.2511	404.2507	0.4	1.0	11.5	1.1	C27 H35 N P











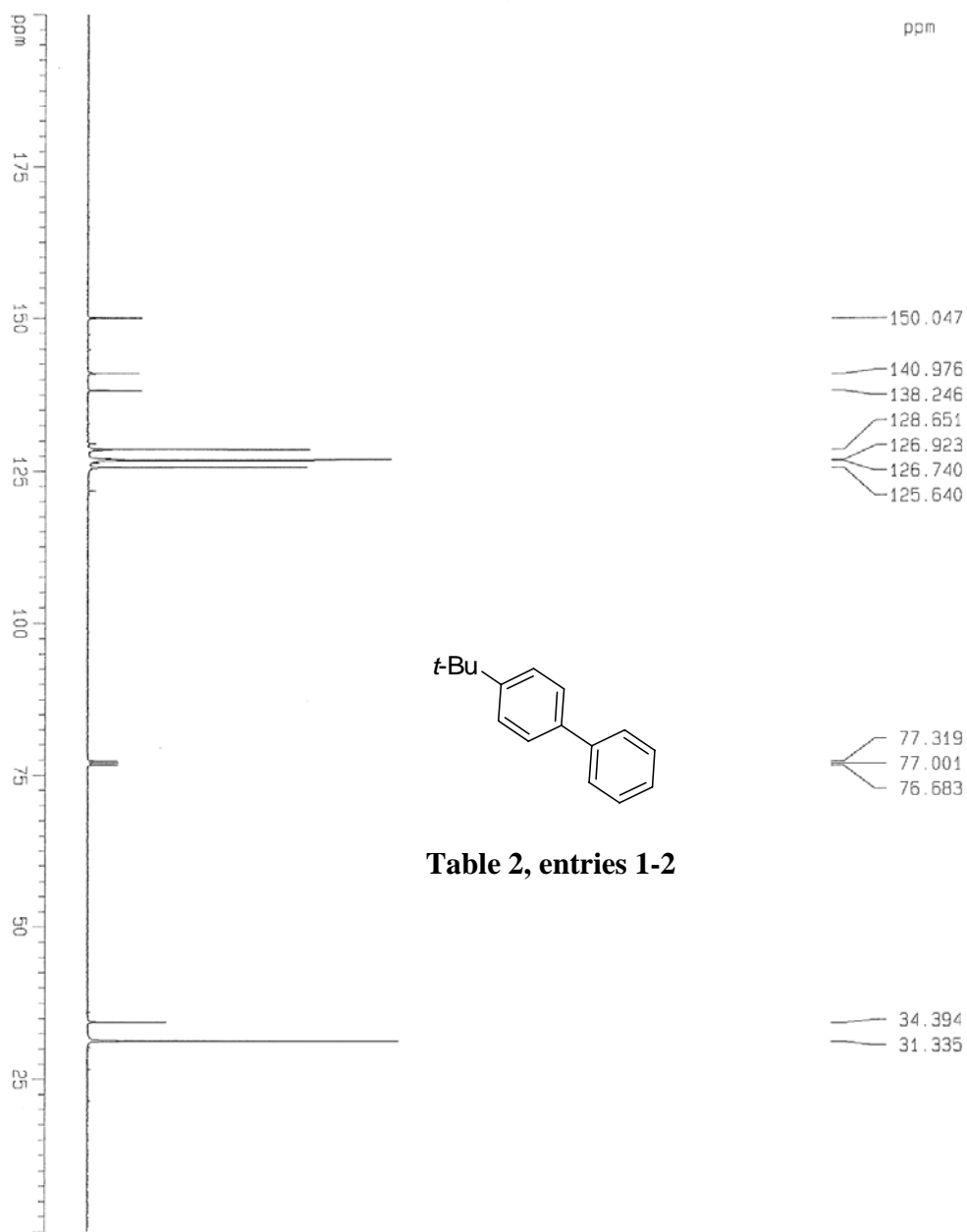


Table 2, entries 1-2

```

Current Data Parameters
NAME      Coupling
EXPNO     30
PROCNO    1

F2 - Acquisition Parameters
Date_     20070601
Time      20.41
INSTRUM   dpx400
PROBHD    5 mm QNP 1H
PULPROG   zgpg30
TO        131072
SOLVENT   CDCl3
NS         64
DS         0
SWH        25125.629 Hz
FIDRES     0.191693 Hz
AQ         2.6083827 sec
RG         8192
DM         19.500 usec
DE         4.50 usec
TE         300.0 K
D1         3.00000000 sec
d11        0.03000000 sec

===== CHANNEL f1 =====
NUC1       13C
P1         5.80 usec
PL1        -6.00 dB
SFO1       100.6231263 MHz

===== CHANNEL f2 =====
CHPROG2    waltz16
NUC2        1H
PCPD2       71.00 usec
PL2         120.00 dB
PL12        17.00 dB
SFO2        400.1326008 MHz

F2 - Processing parameters
SI          65536
SF          100.6128150 MHz
WDW         EM
SSB         0
LB          3.00 Hz
GB          0
PC          1.00

1D NMR plot parameters
CX          19.50 cm
F1P         200.000 ppm
F1          20122.56 Hz
F2P         -0.000 ppm
F2          -0.00 Hz
PPMCM       10.25641 ppm/cm
HZCM        1031.92639 Hz/cm

```

File : C:\msdchem\1\DATA\cmso\CT\c289-2.D
Operator :
Acquired : 1 Jun 2007 19:37 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

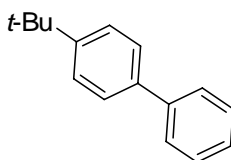
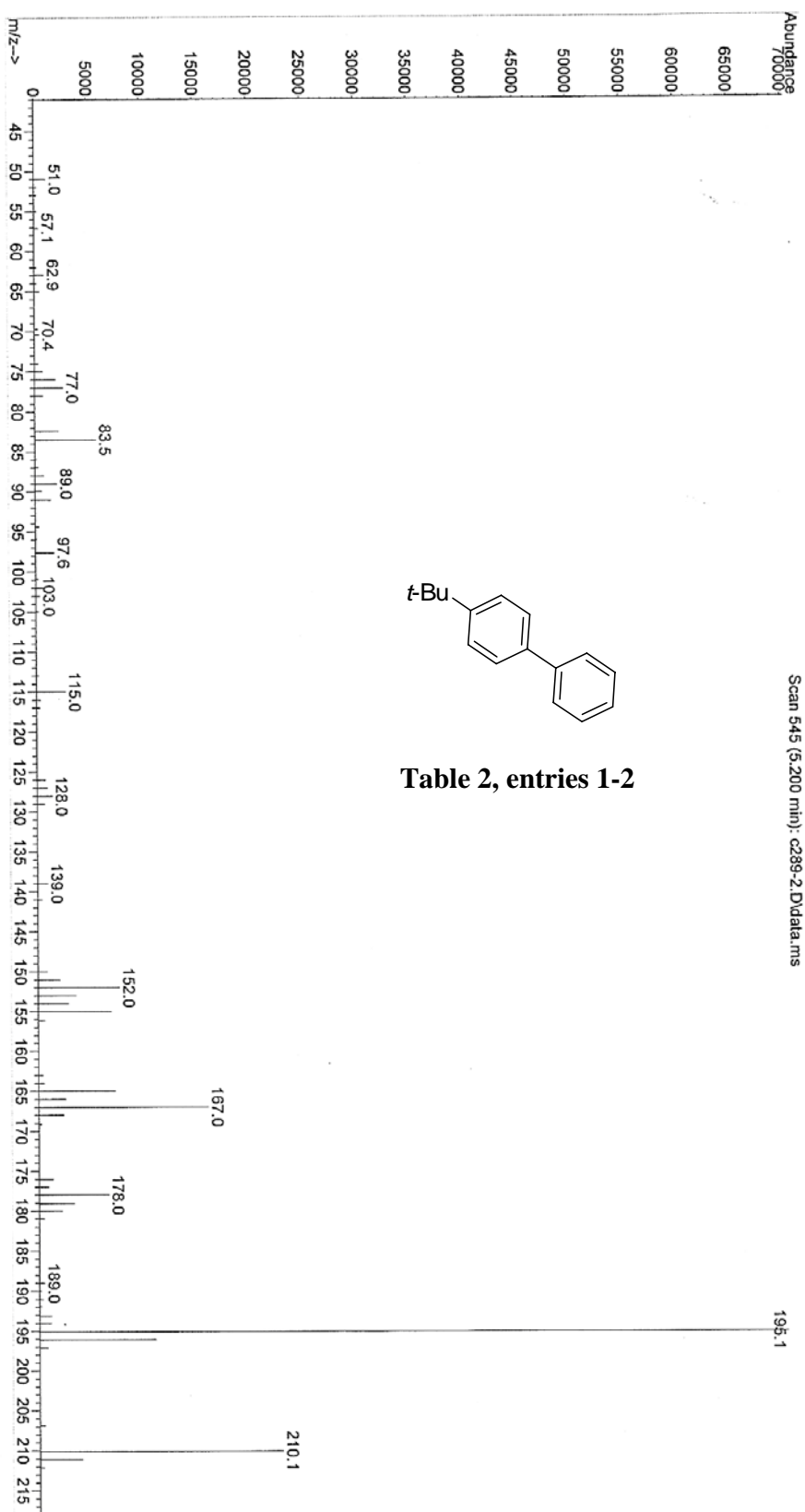
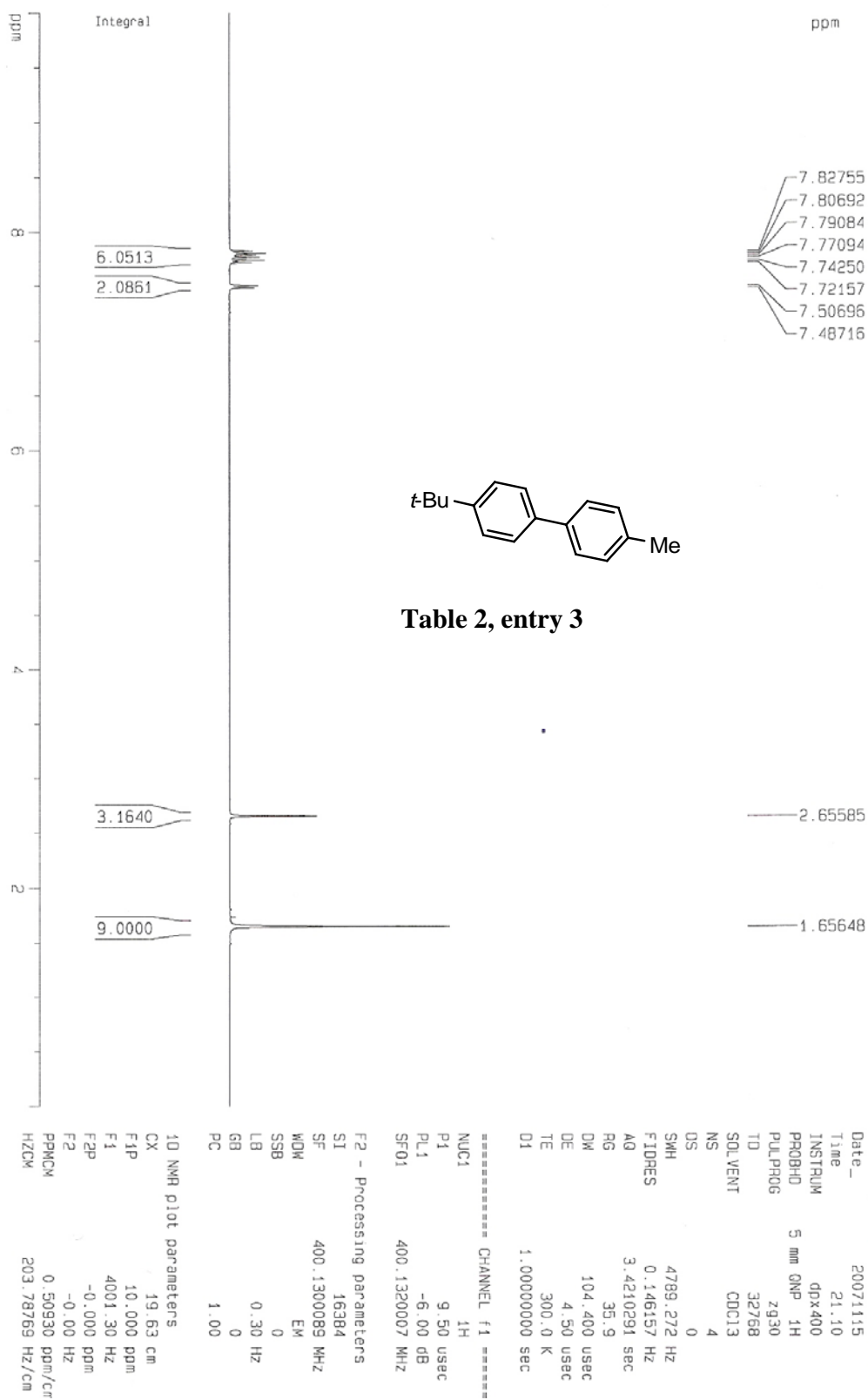


Table 2, entries 1-2





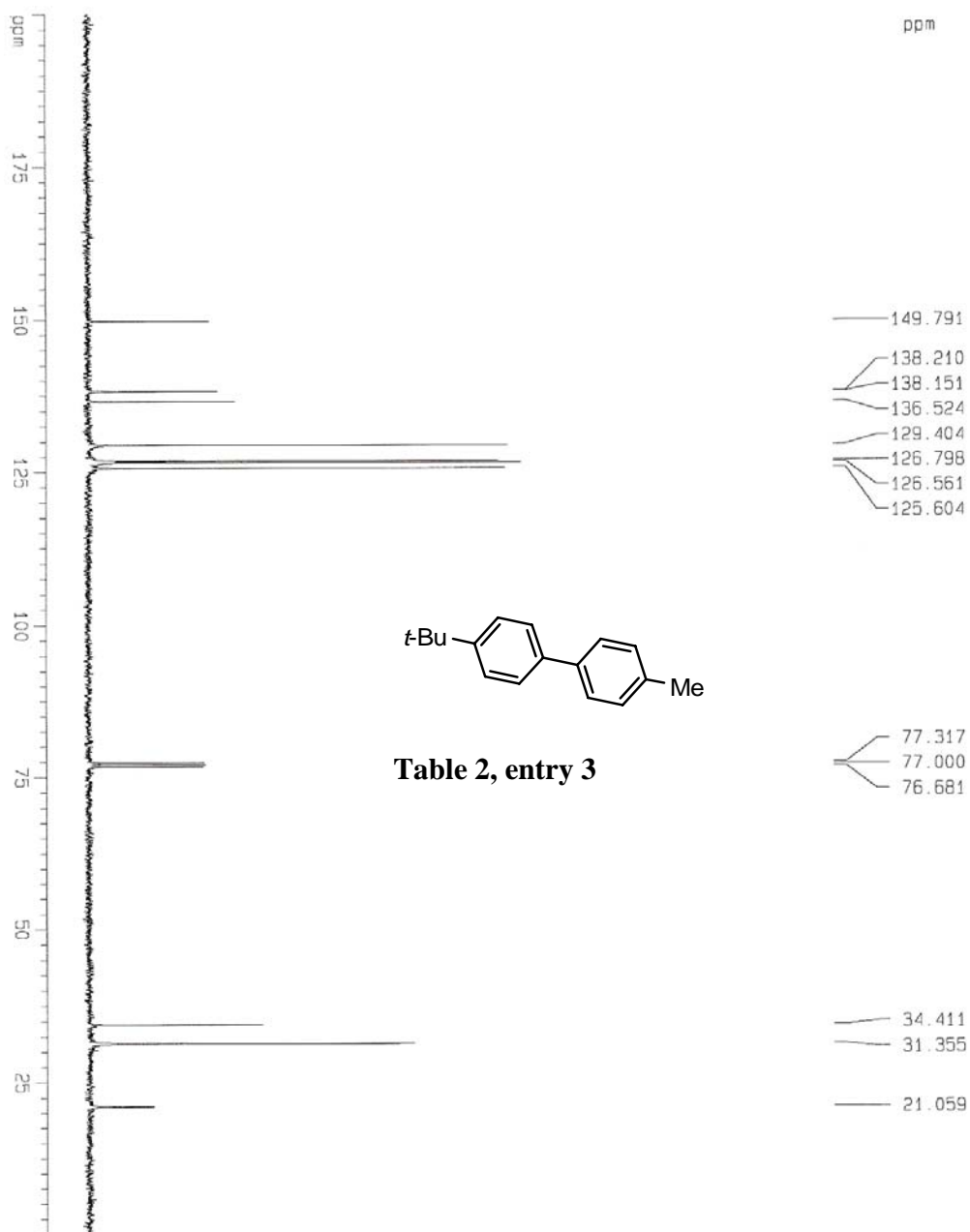


Table 2, entry 3

Current Data Parameters

NAME	Coupling
EXPNO	99
PROCNO	1

F2 - Acquisition Parameters

Date_	20071115
Time	21.11
INSTRUM	gpc400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	16
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DW	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	¹³ C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	¹ H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128035 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm/c
HZCM	1031.92615 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c518.D
Operator :
Acquired : 15 Nov 2007 14:21 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

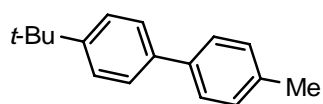
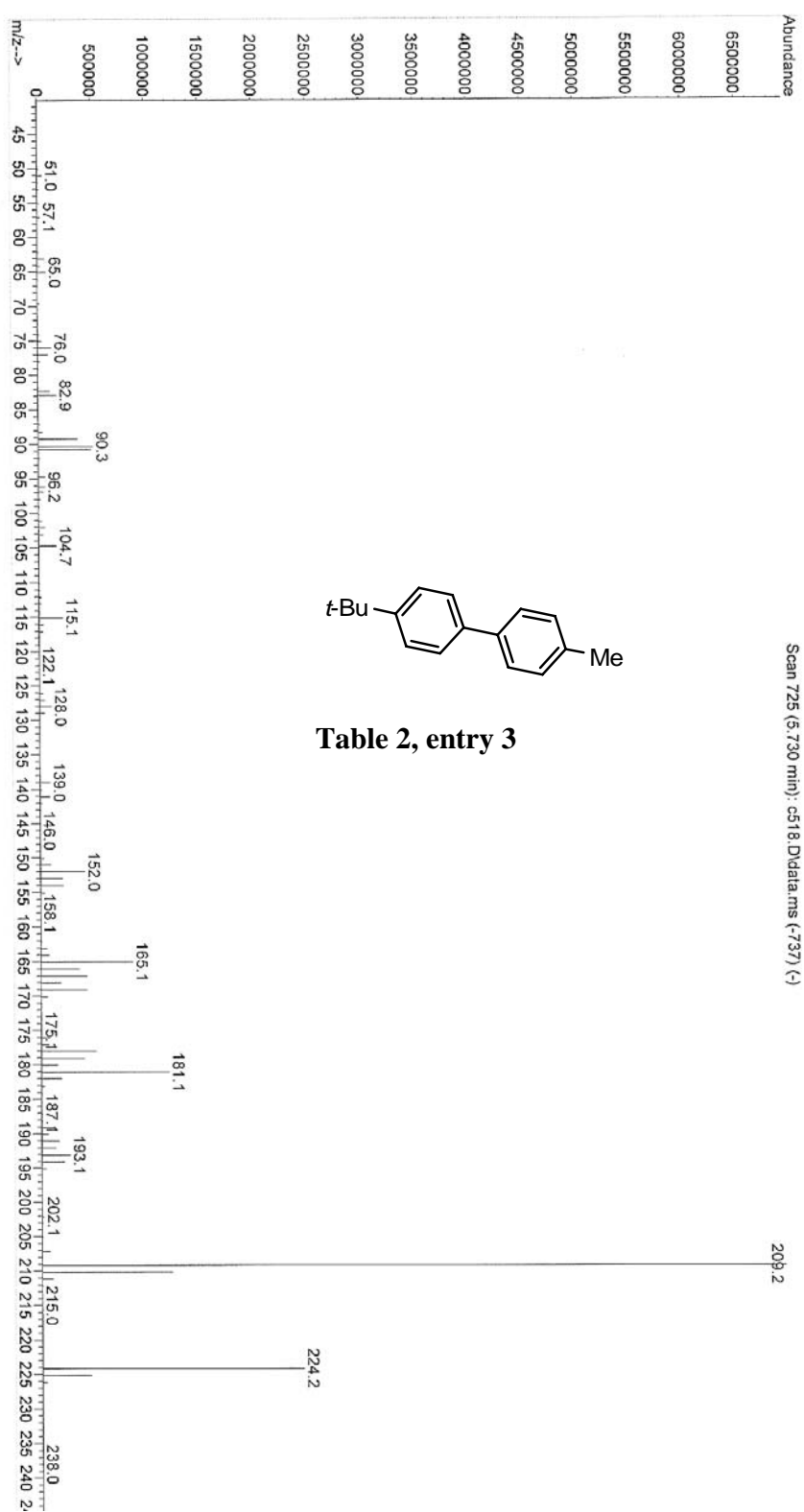
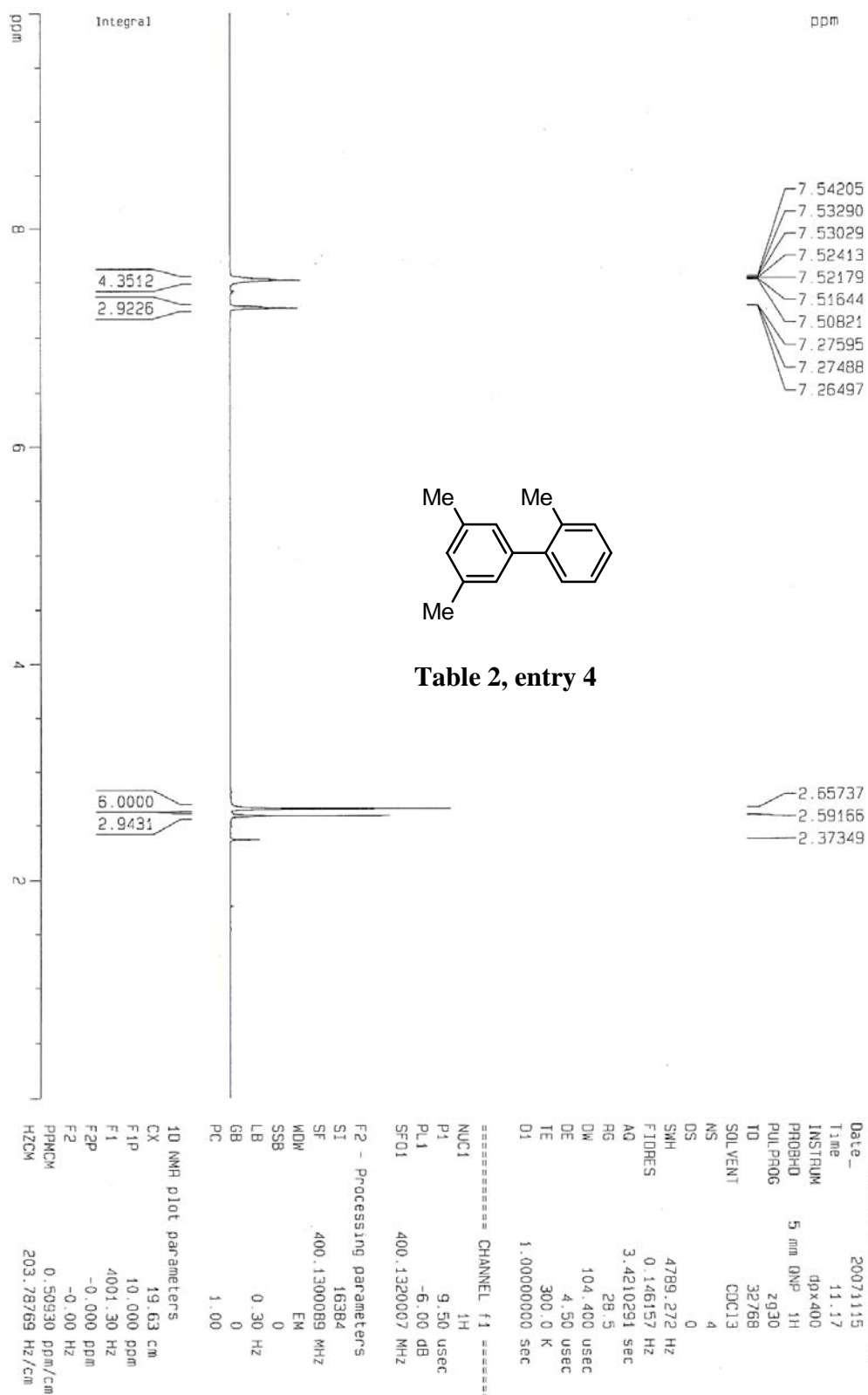


Table 2, entry 3





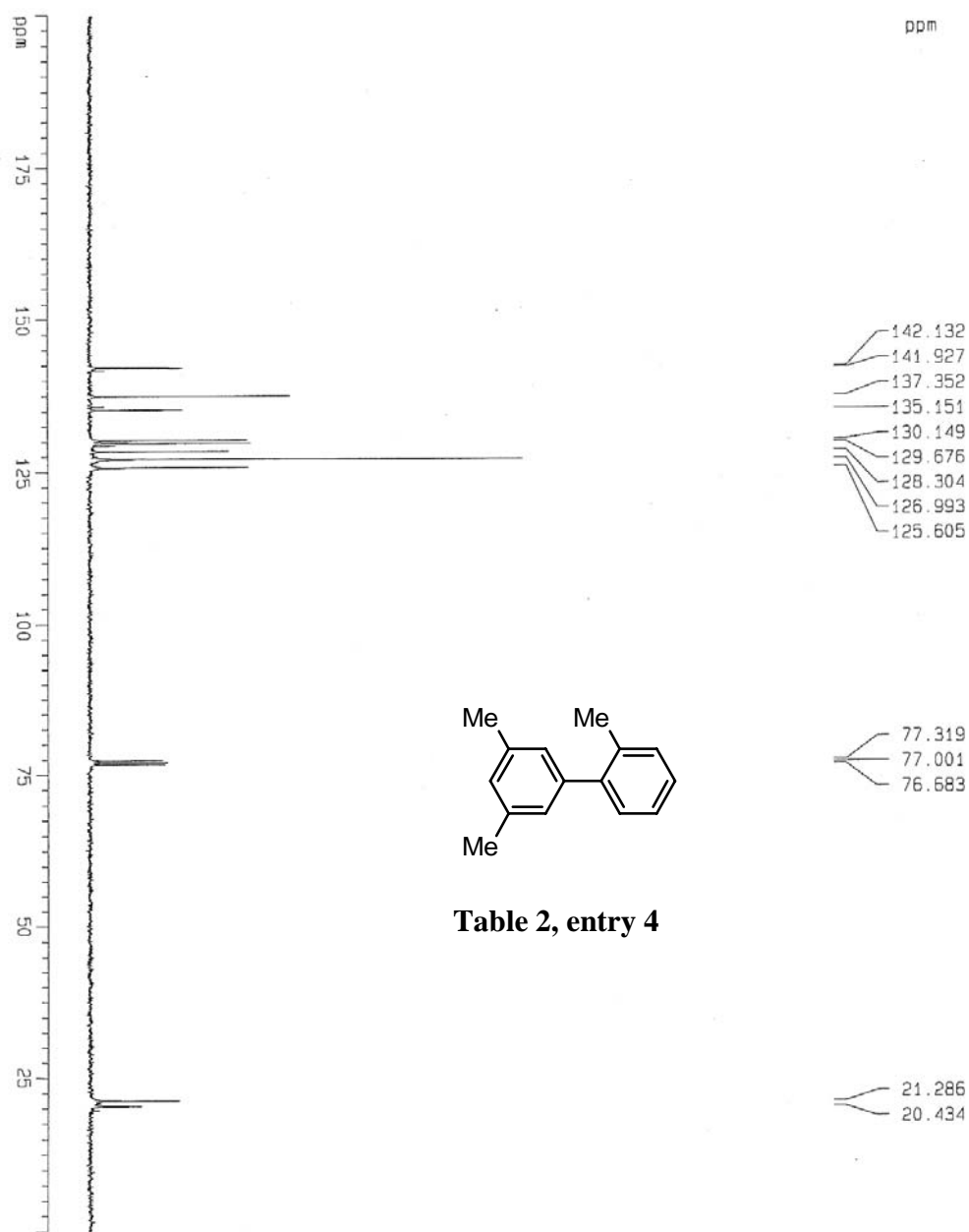


Table 2, entry 4

Current Data Parameters

NAME	Value	Unit
EXPNO	94	
PROCNO	1	

F2 - Acquisition Parameters

Parameter	Value	Unit
Date_	20071115	
Time	11.19	
INSTRUM	dp400	
PROBHD	5 mm QNP 1H	
PULPROG	zgpg30	
TD	131072	
SOLVENT	CDCl3	
NS	16	
DS	0	
SWH	25125.629	Hz
FIDRES	0.191693	Hz
AQ	2.6083827	sec
RG	8192	
DW	19.500	usec
DE	4.50	usec
TE	300.0	K
D1	3.00000000	sec
d11	0.03000000	sec

===== CHANNEL f1 =====

Parameter	Value	Unit
NUC1	13C	
P1	5.80	usec
PL1	-6.00	dB
SFO1	100.6231263	MHz

===== CHANNEL f2 =====

Parameter	Value	Unit
CPDPRG2	waltz16	
NUC2	1H	
PCPD2	71.00	usec
PL2	120.00	dB
PL12	17.00	dB
SFO2	400.1326008	MHz

F2 - Processing parameters

Parameter	Value	Unit
SI	65536	
SF	100.6128074	MHz
WDW	EM	
SSB	0	
LB	3.50	Hz
GB	0	
PC	1.00	

1D NMR plot parameters

Parameter	Value	Unit
CX	19.50	cm
F1P	200.000	ppm
F1	20122.56	Hz
F2P	-0.000	ppm
F2	-0.00	Hz
PPMCM	10.25641	ppm/cm
HZCM	1031.92627	Hz/cm

Supporting Information

File : C:\msdchem\1\DATA\cms0\CT\c514.D
 Operator :
 Acquired : 14 Nov 2007 19:27 using AcqMethod METHOD2.M
 Instrument : 5973N
 Sample Name :
 Misc Info :
 Vial Number: 1

Scan 571 (4.848 min): c514.D\data.ms

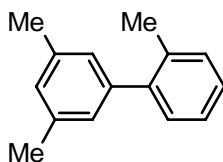
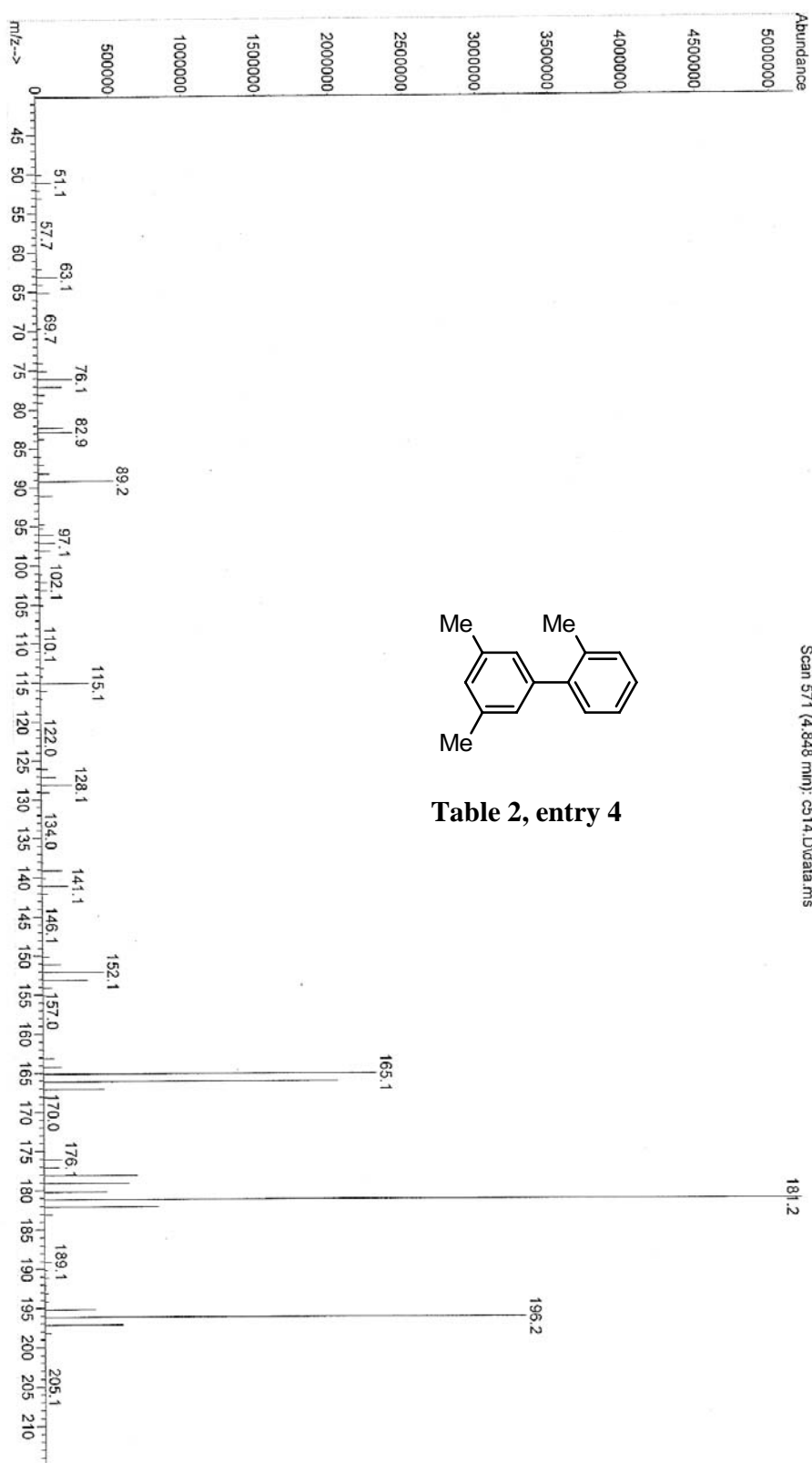
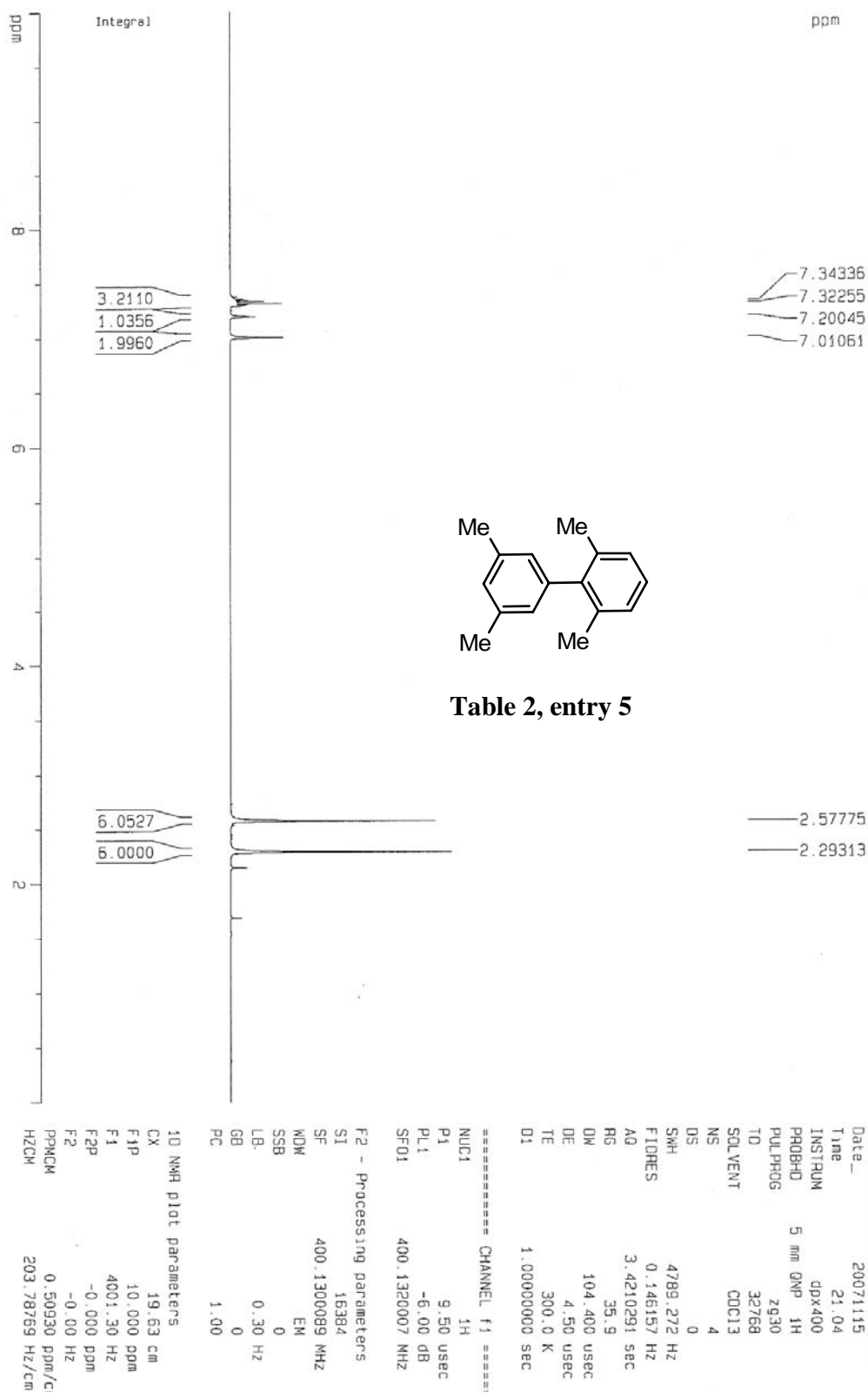


Table 2, entry 4





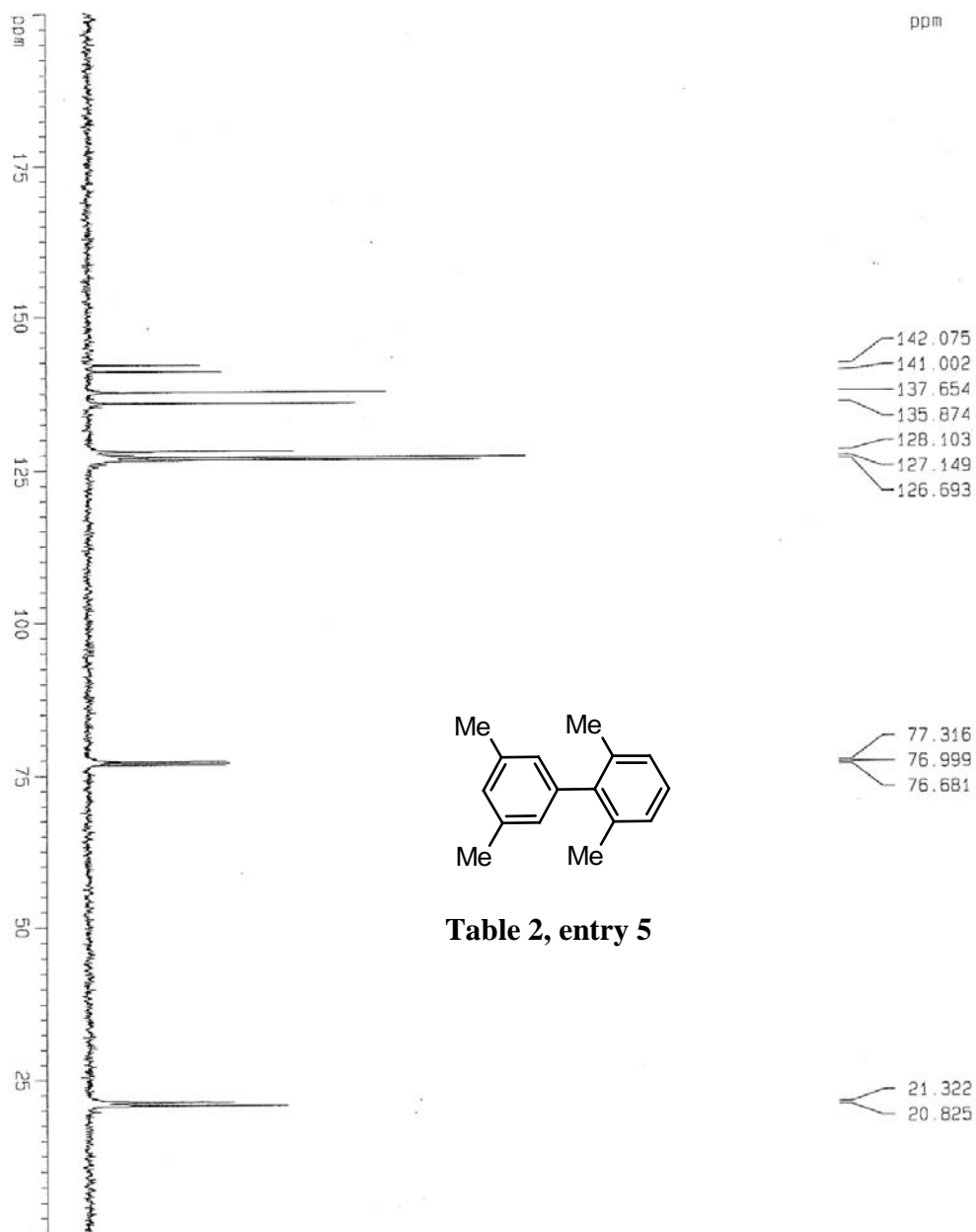


Table 2, entry 5

Current Data Parameters

NAME	Coupling
EXPNO	98
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071115	21.05

INSTRUM dpx400
PROBHD 5 mm QNP 1H
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 16
DS 0
SMH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.6083827 sec
RG 8192
DW 19.900 usec
DE 4.50 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	wa1tz16
NUC2 <td>1H</td>	1H
PCPD2 <td>71.00 usec</td>	71.00 usec
PL2 <td>120.00 dB</td>	120.00 dB
PL12 <td>17.00 dB</td>	17.00 dB
SFO2 <td>400.1326008 MHz</td>	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6127985 MHz
MDM	EW
SSB	0
LB	5.00 Hz
GB	0
PC	1.00

10 NMR plot parameters

CX	19.50 cm
F1P <td>200.000 ppm</td>	200.000 ppm
F1 <td>20122.56 Hz</td>	20122.56 Hz
F2P <td>-0.000 ppm</td>	-0.000 ppm
F2 <td>-0.00 Hz</td>	-0.00 Hz
PPMCM <td>10.25641 ppm/cm</td>	10.25641 ppm/cm
HZCM <td>1031.92615 Hz/cm</td>	1031.92615 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c515.D
Operator :
Acquired : 14 Nov 2007 19:41 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 2

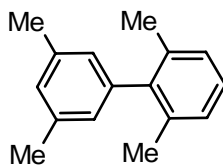
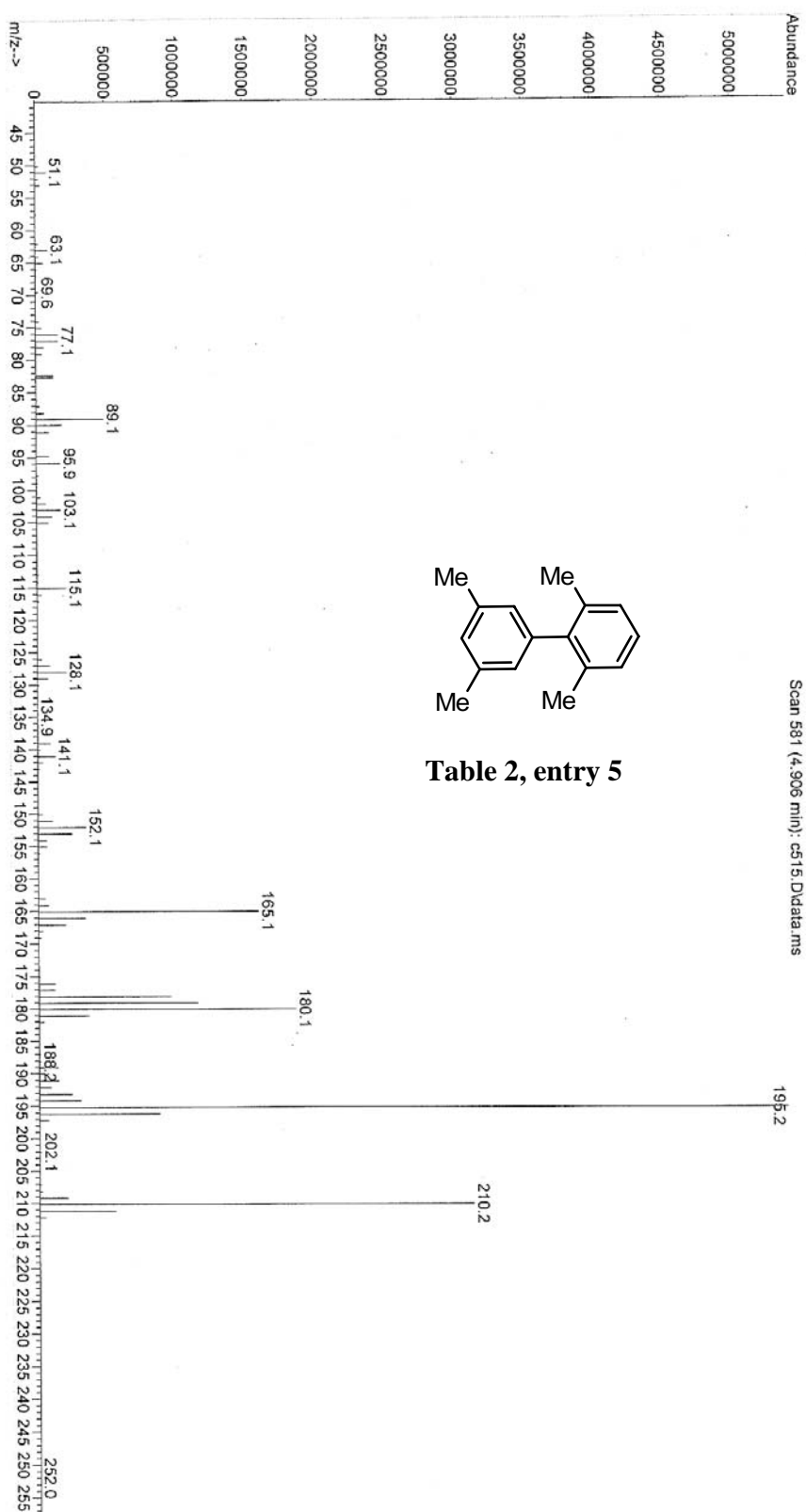
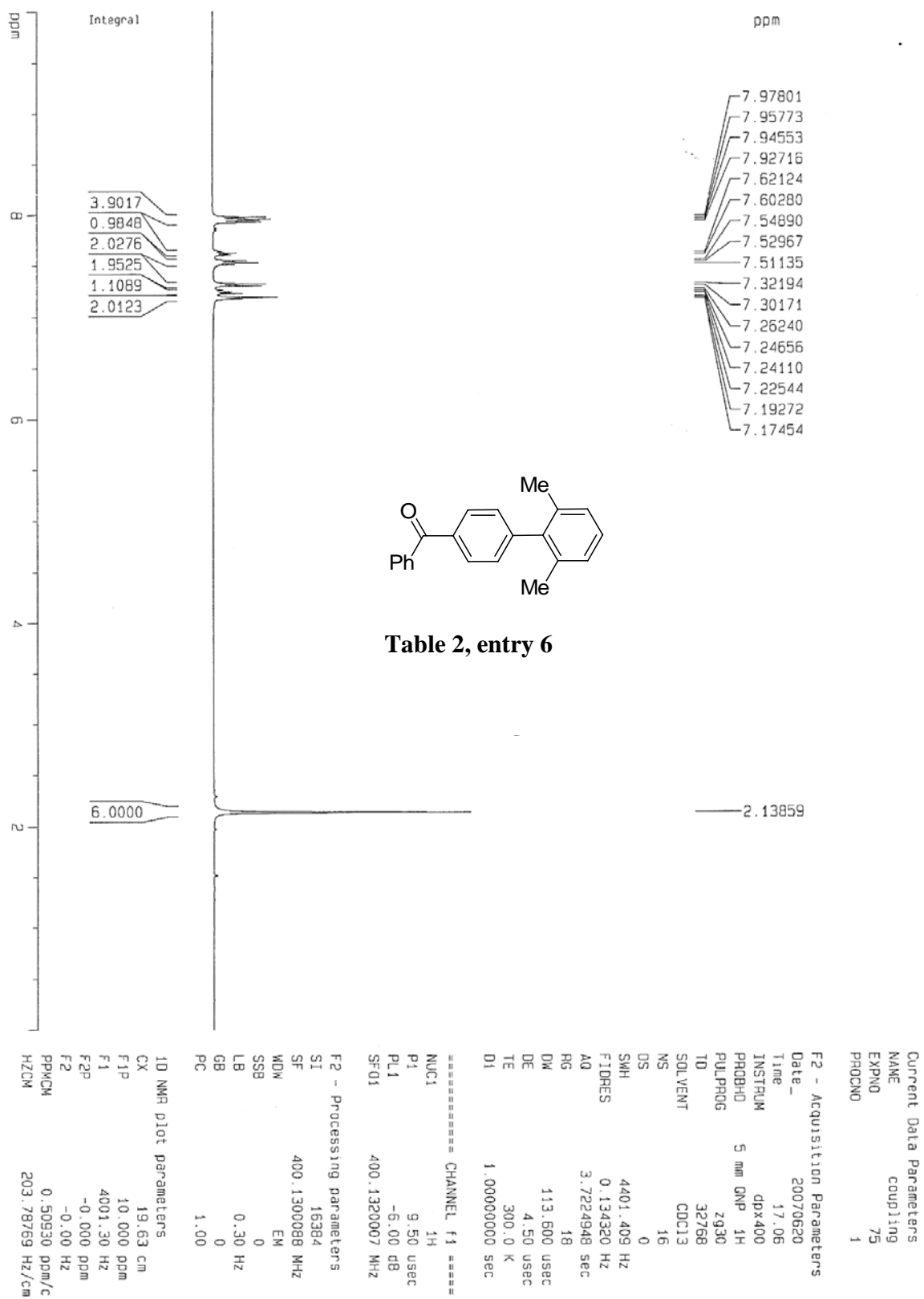


Table 2, entry 5





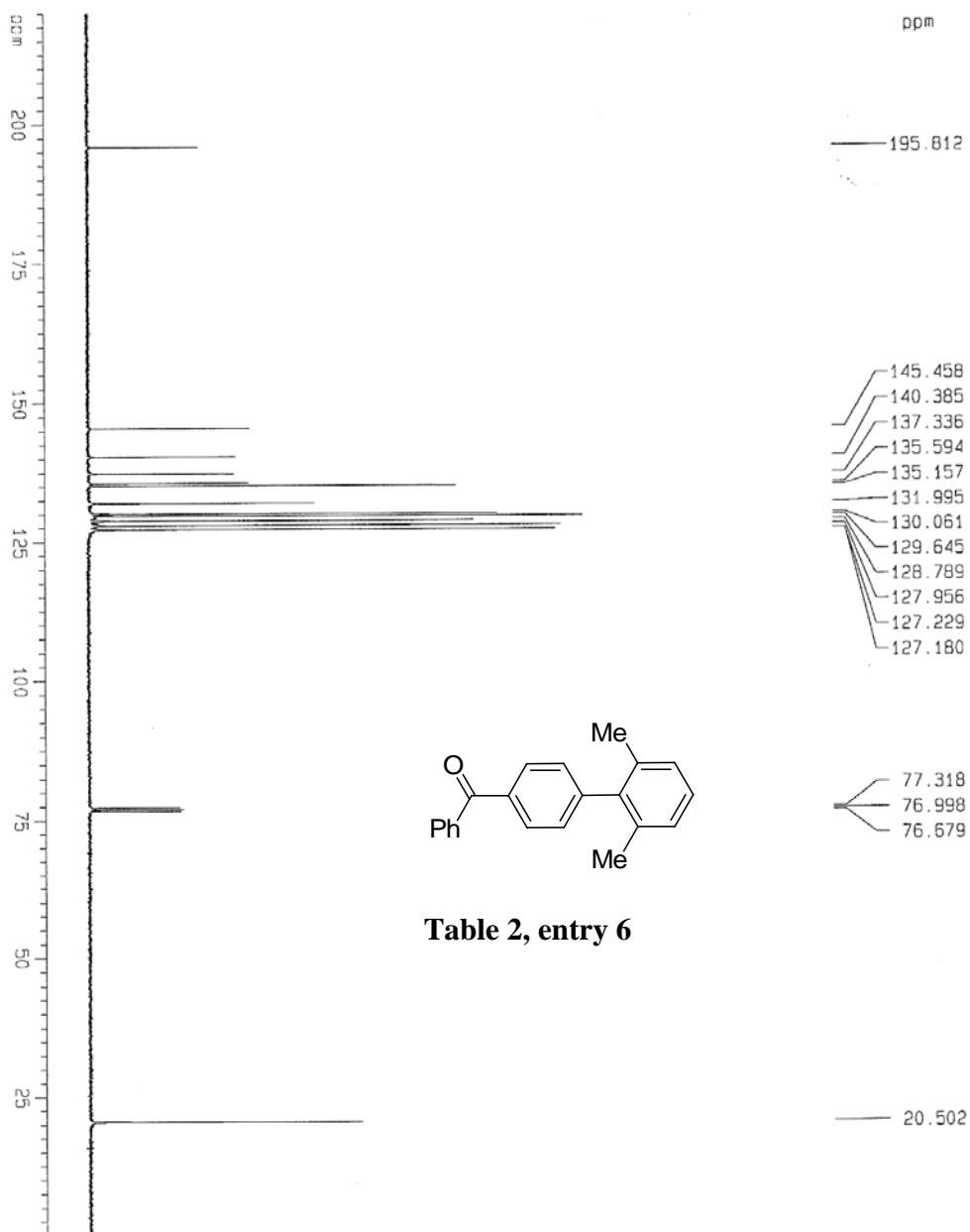


Table 2, entry 6

Current Data Parameters

NAME	Coupling
EXPNO	40
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070620	17.08
INSTRUM	dp400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	64
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.603827 sec
RG	8192
DM	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

OPPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SF	100.6128177 MHz
MDW	EX
SSB	0
LB	1.50 Hz
GB	0
PC	1.00

10 NMR plot parameters

CX	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	11.26205 ppm/cm
HZCM	1135.11902 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c348.D
Operator :
Acquired : 20 Jun 2007 13:17 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

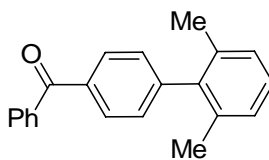
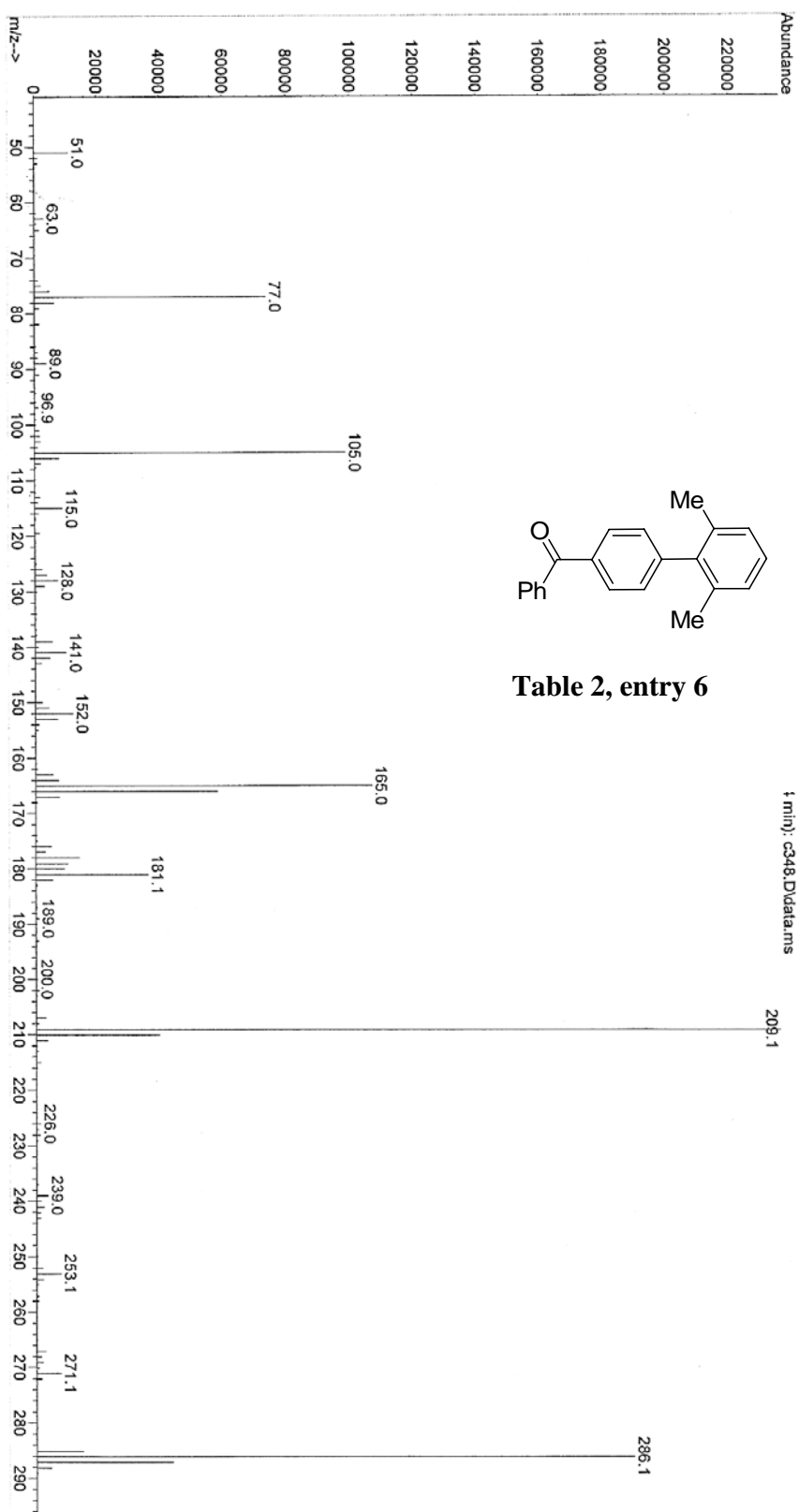


Table 2, entry 6



Elemental Composition Report

Single Mass Analysis

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 50.0

Selected filters: None

Monoisotopic Mass, Even Electron Ions

5 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-21 H: 0-1000 O: 0-1

So chau Ming, 4-(2,6-dimethylphenyl)benzophenone

HR07_0724_4 47 (0.891) AM (Cen:5, 80.00; Ht:10000.0, 0.00, 1.00); Sm (SG, 2x3.00); Cm (31:57)

287.1442

TOF MS ES+
2.73e3

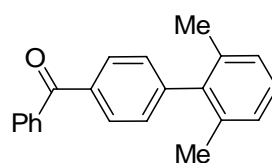
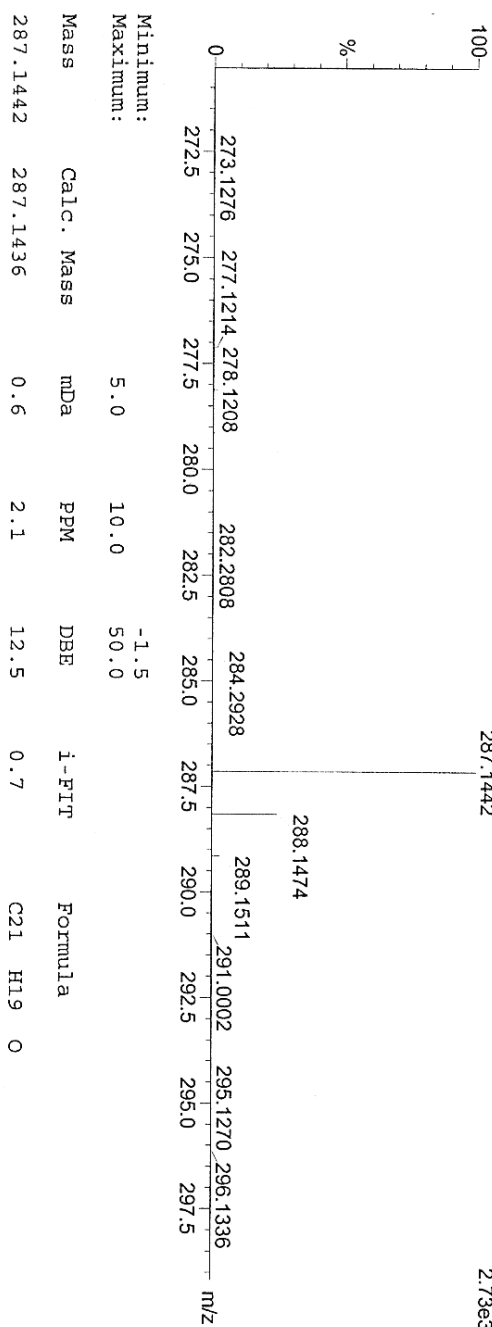
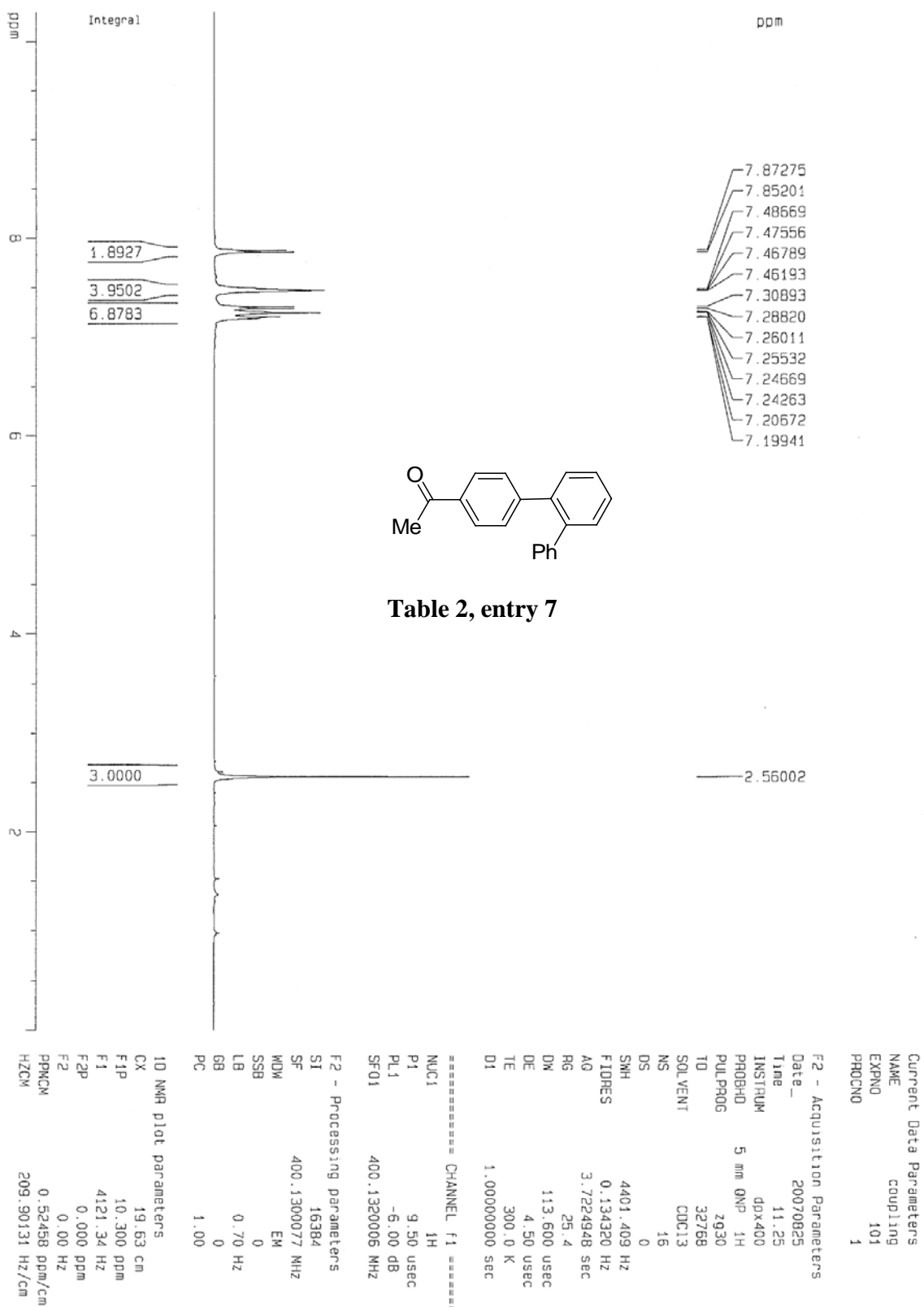


Table 2, entry 6





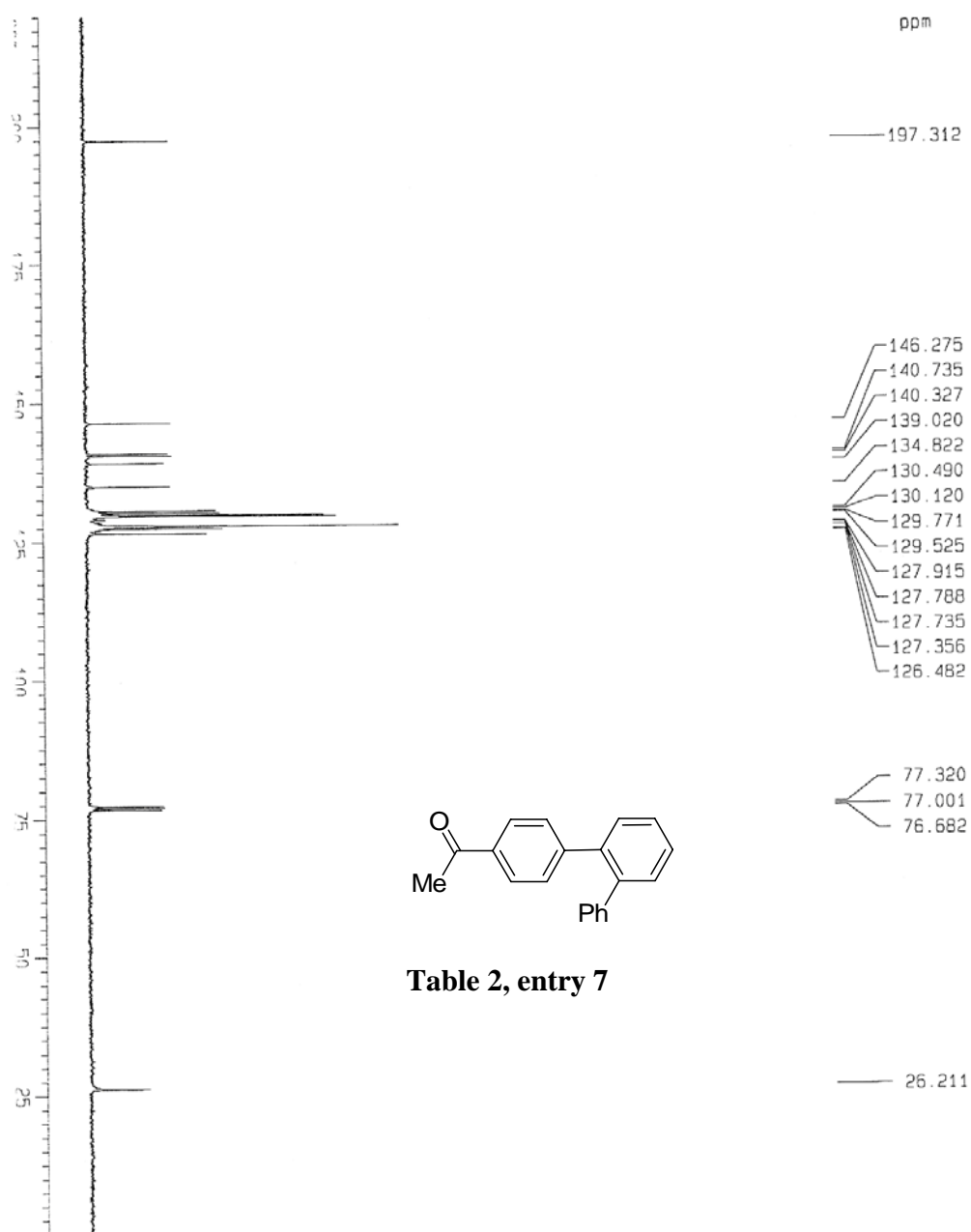


Table 2, entry 7

Current Data Parameters

NAME	COUPLING
EXPNO	58
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070825	11.27

INSTRUM: dp400
PROBHD: 5 mm QNP 1H
PULPROG: zgpg30
TD: 131072
SOLVENT: CDCl3
NS: 32
DS: 0
SWH: 25125.629 Hz
FIDRES: 0.191693 Hz
AQ: 2.6083827 sec
RG: 8192
DM: 19.900 usec
DE: 4.50 usec
TE: 300.0 K
D1: 3.00000000 sec
d11: 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231253 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128116 MHz
WDW	EM
SSB	0
LB	4.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CK	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	11.28205 ppm/cm
HZCM	1135.11850 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c381.D
Operator :
Acquired : 24 Aug 2007 17:44 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

Scan 1008 (7.349 min): c381.D\data.ms

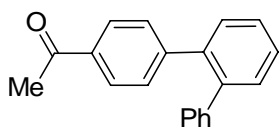
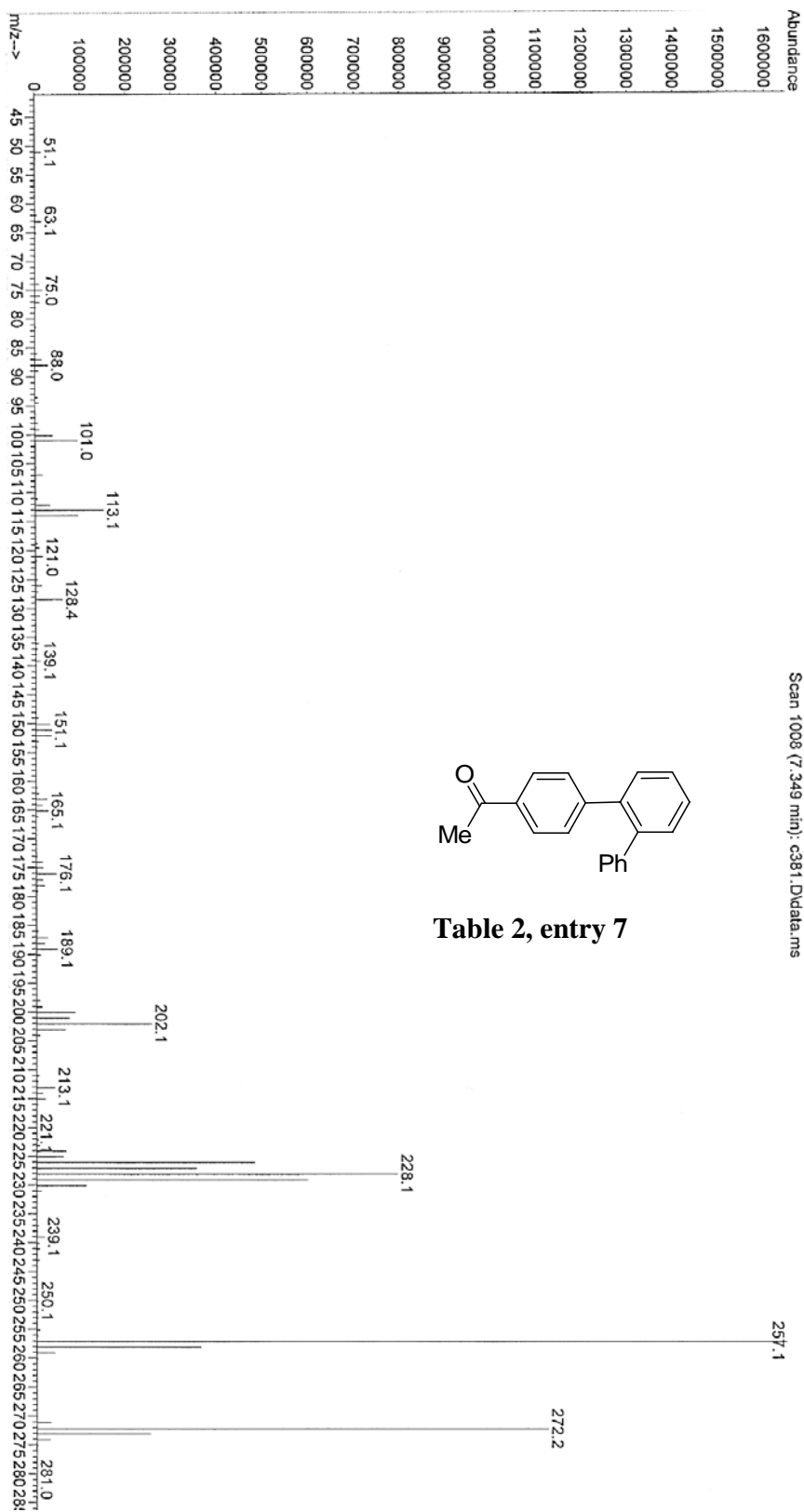
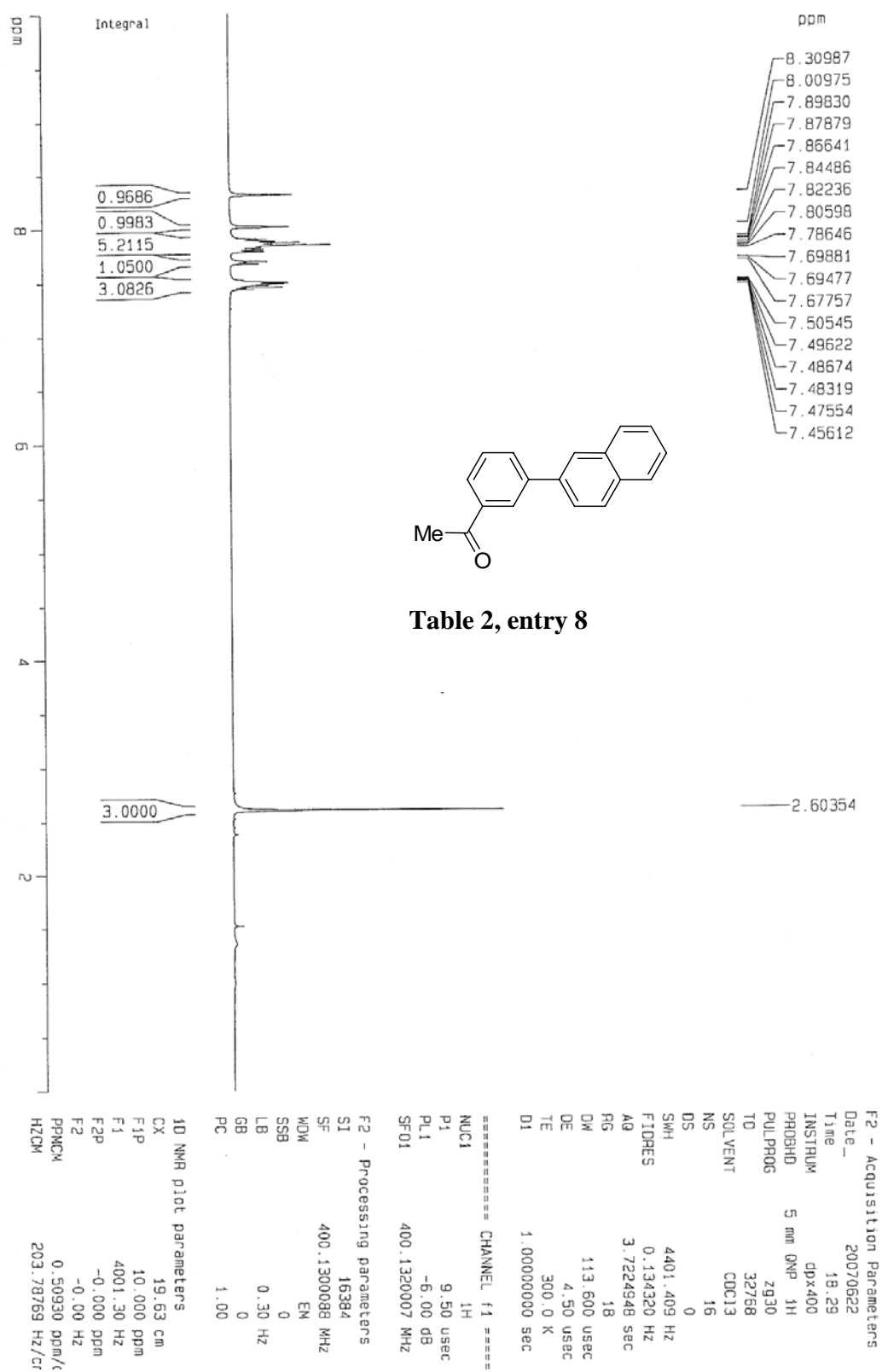


Table 2, entry 7





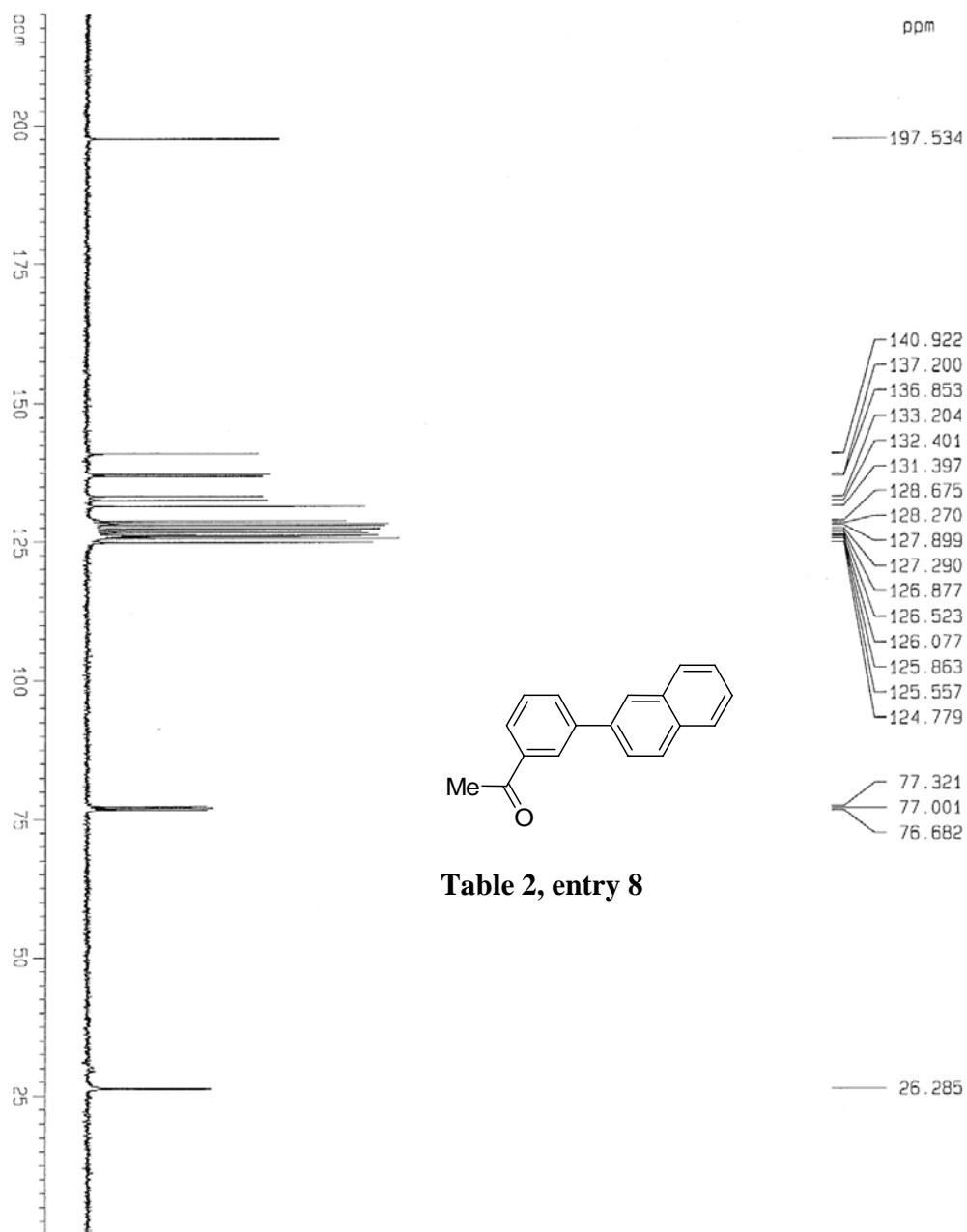


Table 2, entry 8

Current Data Parameters

NAME	COUPLING
EXPNO	43
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070622	18.31
INSTRUM	ddx400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	32
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DM	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2 <td>1H</td>	1H
PCPD2 <td>71.00 usec</td>	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128181 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	11.28205 ppm/c
HZCM	1135.11902 Hz/cm

File : C:\msdchem\1\DATA\cmso\CT\c353.D
Operator :
Acquired : 22 Jun 2007 14:35 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

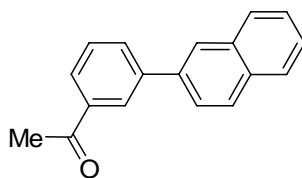
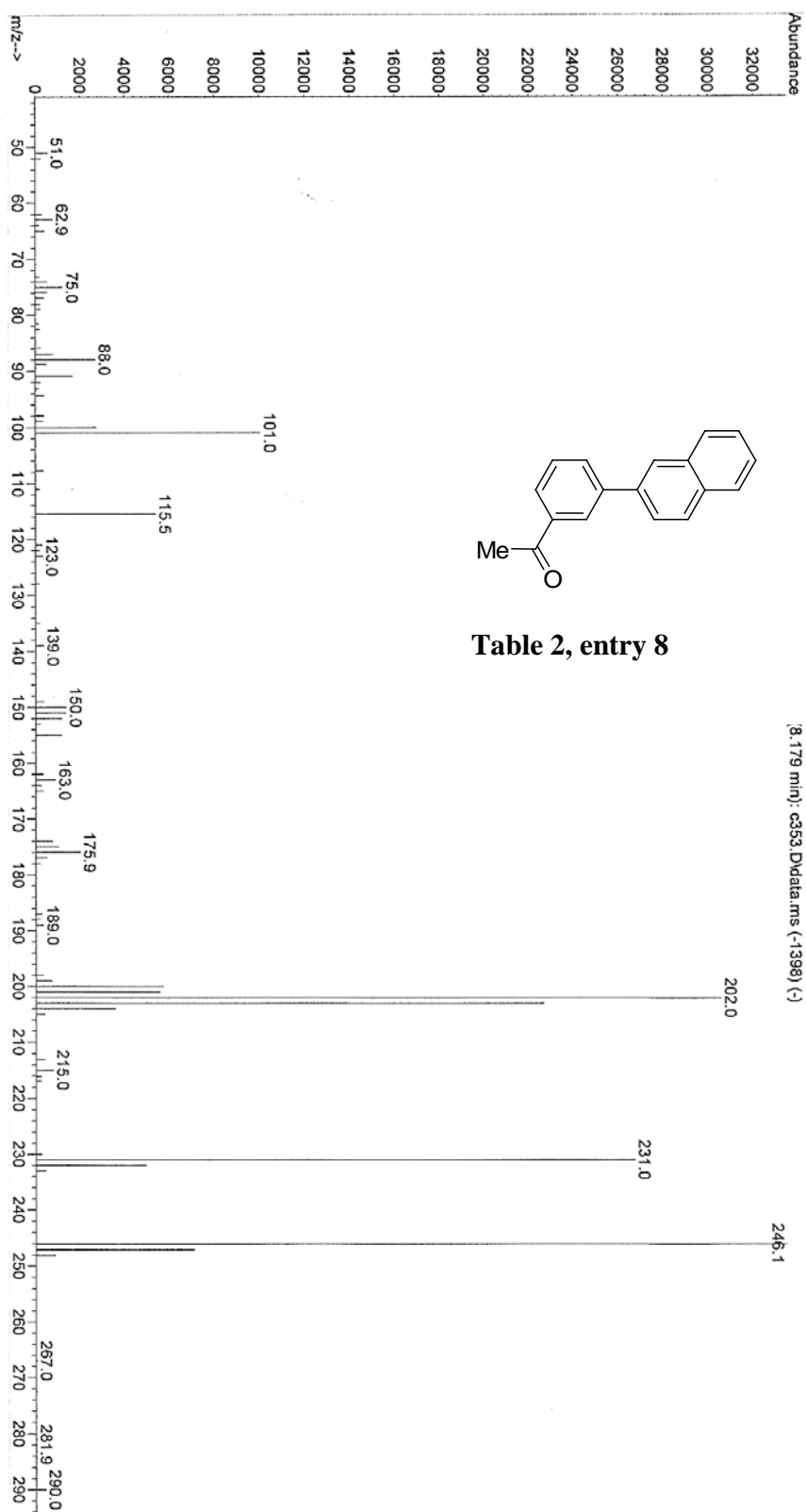


Table 2, entry 8



Elemental Composition Report

Single Mass Analysis

Tolerance = 10.0 PPM / DBE: min = -1.5, max = 50.0

Selected filters: None

Monoisotopic Mass, Even Electron Ions

8 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-21 H: 0-1000 O: 0-1

So Chau Ming, 3-naphthylacetophenone

HR07_0724_5 48 (0.910) AM (Cen:5, 80.00, Ht:10000.0,0.00,1.00); Sm (SG, 2x3.00); Cm (23:53)

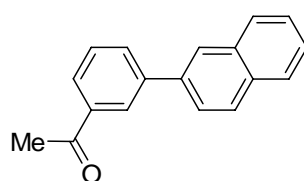
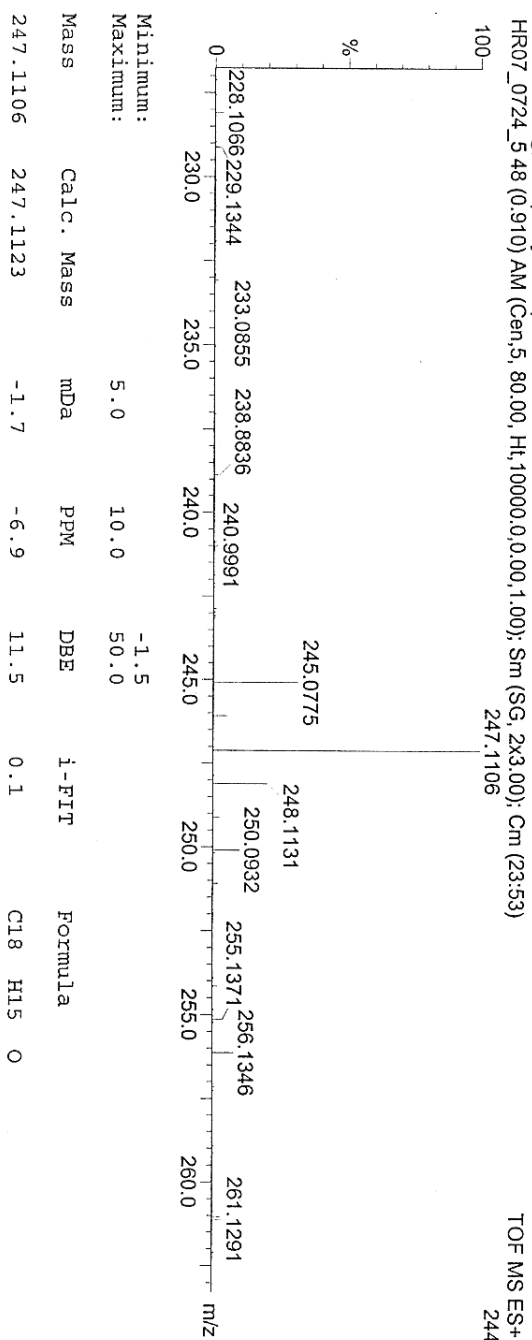


Table 2, entry 8



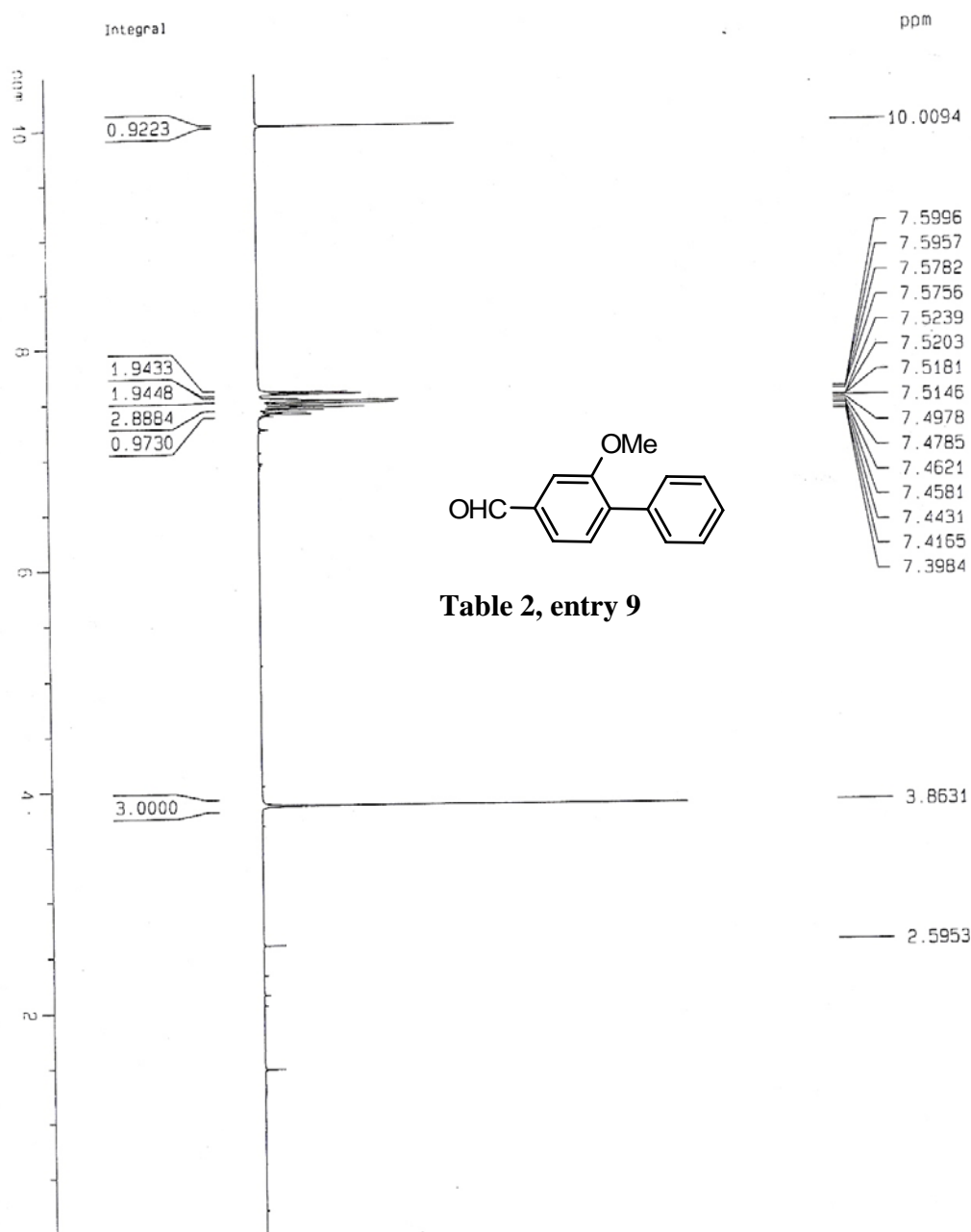


Table 2, entry 9

Current Data Parameters

NAME	COUPLING
EXPNO	129
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071106	19.12

INSTRUM: gpc400
PROBHD: 5 mm GNP 1H
PULPROG: zg30
TD: 32768
SOLVENT: CDCl3
NS: 16
DS: 0
SWH: 4401.409 Hz
FIDRES: 0.13420 Hz
AQ: 3.7224948 sec
RG: 32
DW: 113.600 usec
DE: 4.50 usec
TE: 300.0 K
D1: 1.00000000 sec

===== CHANNEL f1 =====

NUC1: 1H
P1: 9.50 usec
PL1: -6.00 dB
SF01: 400.132006 MHz

F2 - Processing parameters

SI	SI
16384	16384
SF	400.130085 MHz
WDW	EM
SSB	0
LB	0.30 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.63 cm
F1P	11.000 ppm
F1	4401.43 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PCNCH	0.56023 ppm/cr
HZCM	224.16646 Hz/cm

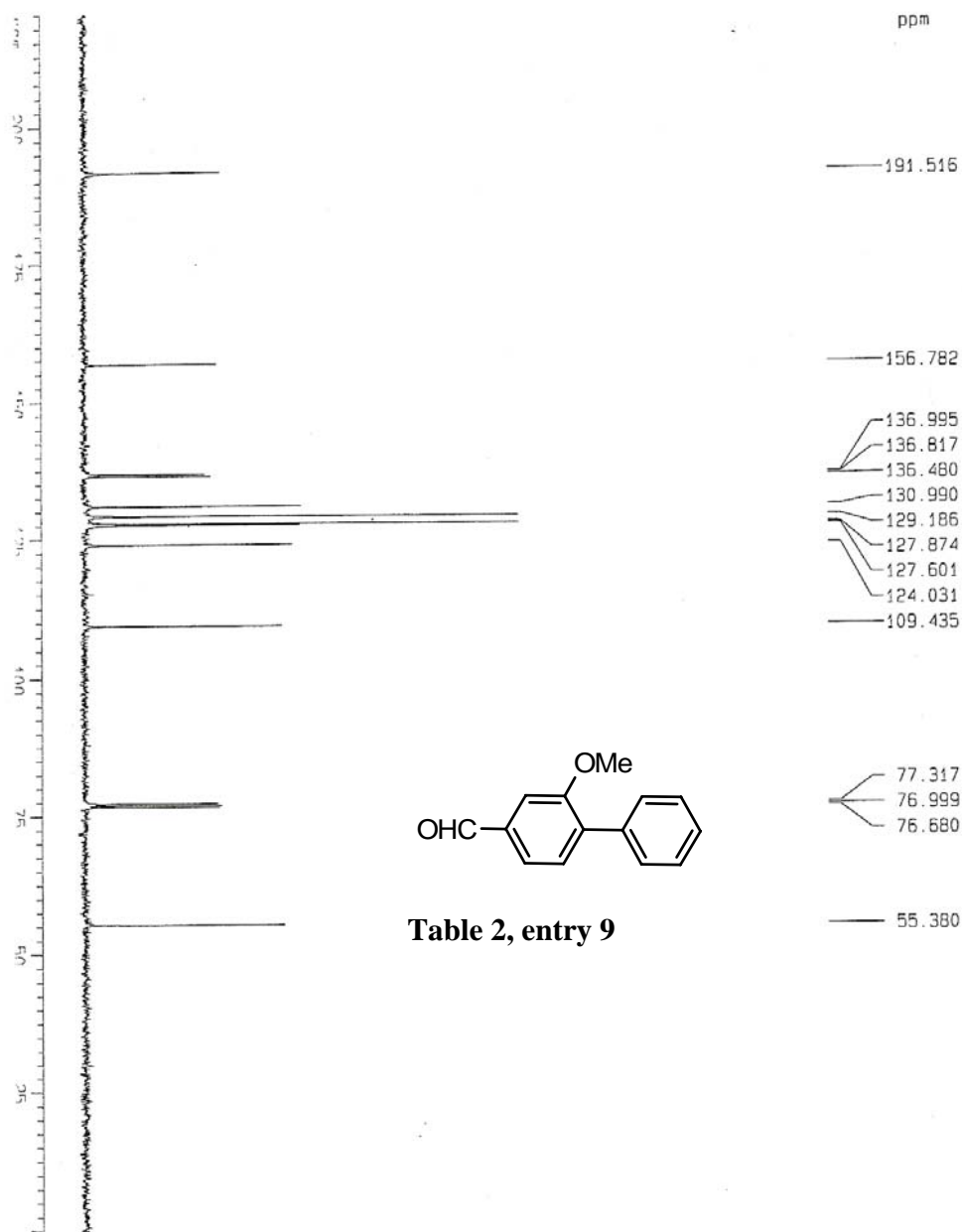


Table 2, entry 9

Current Data Parameters

NAME	COUPLING
EXPNO	B3
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071106	19.16
INSTRUM	dp400
PROBHD	5 mm GNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	16
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DW	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	wa1tz16
NUC2 <td>1H</td>	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128047 MHz
WDW	EM
SSB	0
LB	4.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PRFCKM	11.28205 ppm/cm
HZCM	1135.11850 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c481.D
Operator :
Acquired : 6 Nov 2007 15:14 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 8

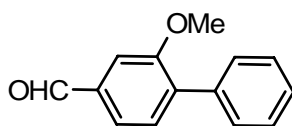
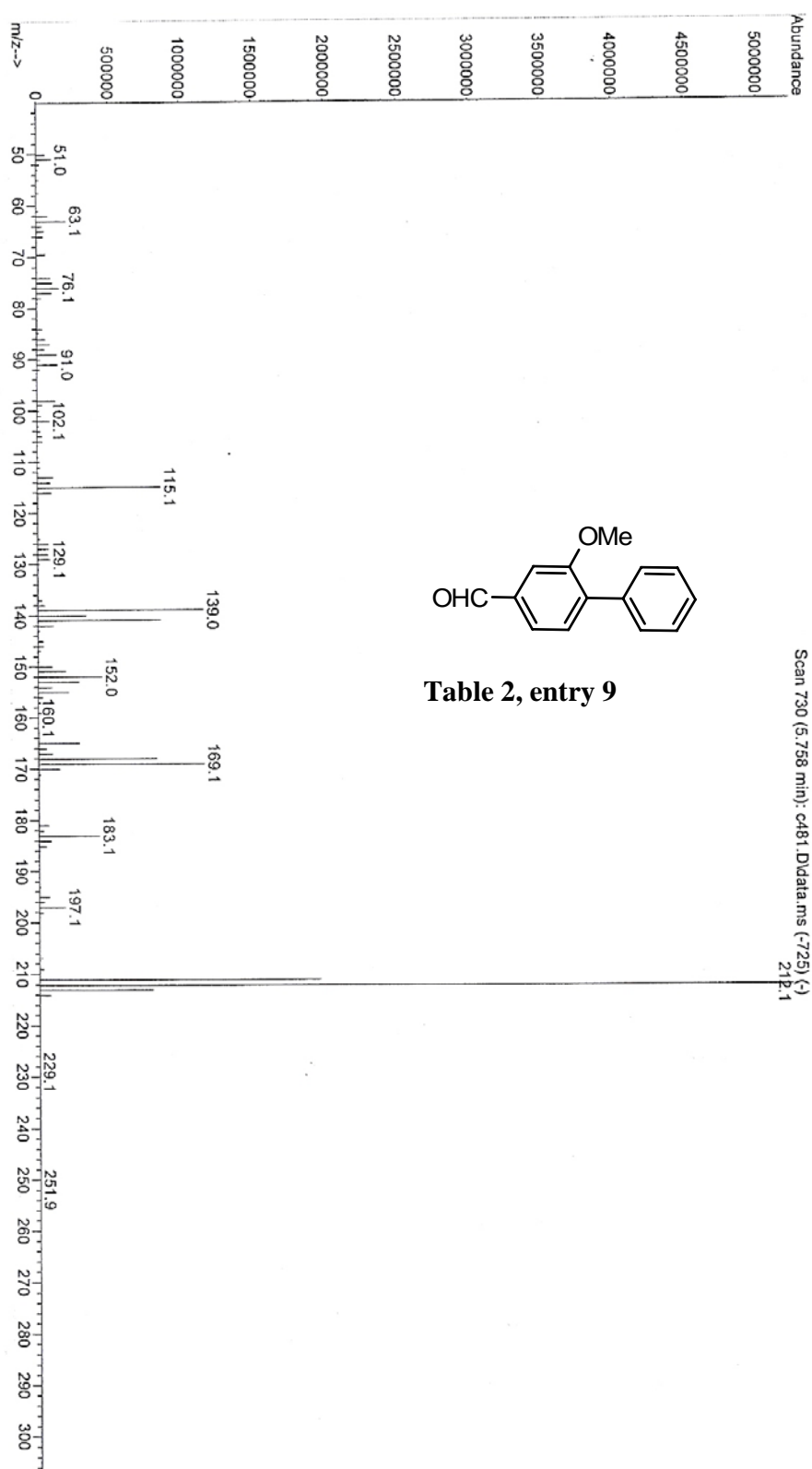
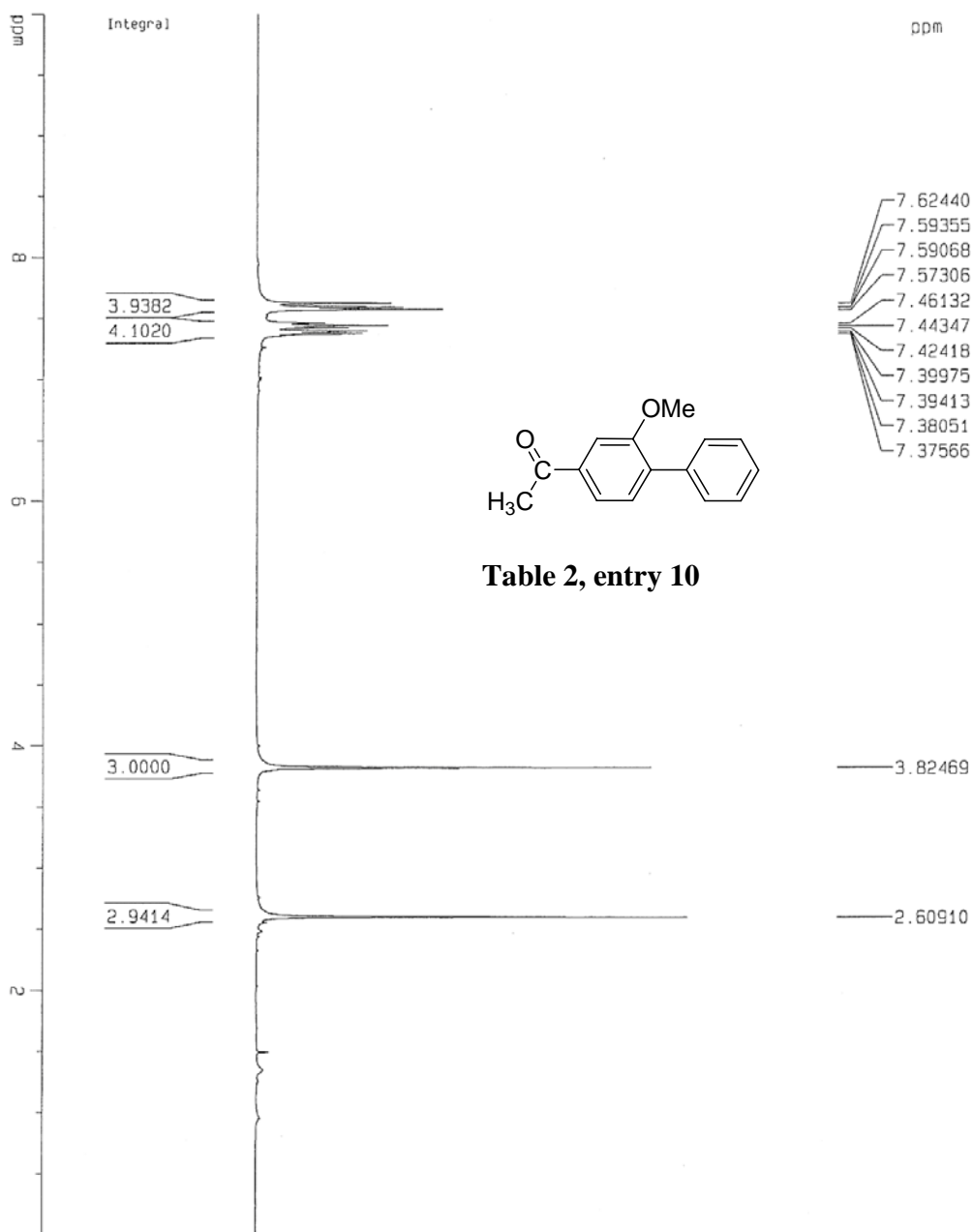


Table 2, entry 9





Current Data Parameters

NAME	coupling
EXPNO	94
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070818	11.31
INSTRUM	dp400
PROBHD	5 mm QNP 1H
PULPROG	zg30
TD	32768
SOLVENT	CDCl3
NS	16
DS	0
SWH	4401.409 Hz
FIDRES	0.134320 Hz
AQ	3.7224948 sec
R6	16
DM	113.600 usec
DE	4.50 usec
TE	300.0 K
D1	1.00000000 sec

===== CHANNEL f1 =====

NUC1	1H
P1	9.50 usec
PL1	-6.00 dB
SFO1	400.132006 MHz

F2 - Processing parameters

SI	16384
SF	400.130077 MHz
WDW	EM
SSB	0
LB	0.70 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.63 cm
F1P	10.000 ppm
F1	4001.30 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	0.50930 ppm/c
HZCM	203.78769 Hz/cm

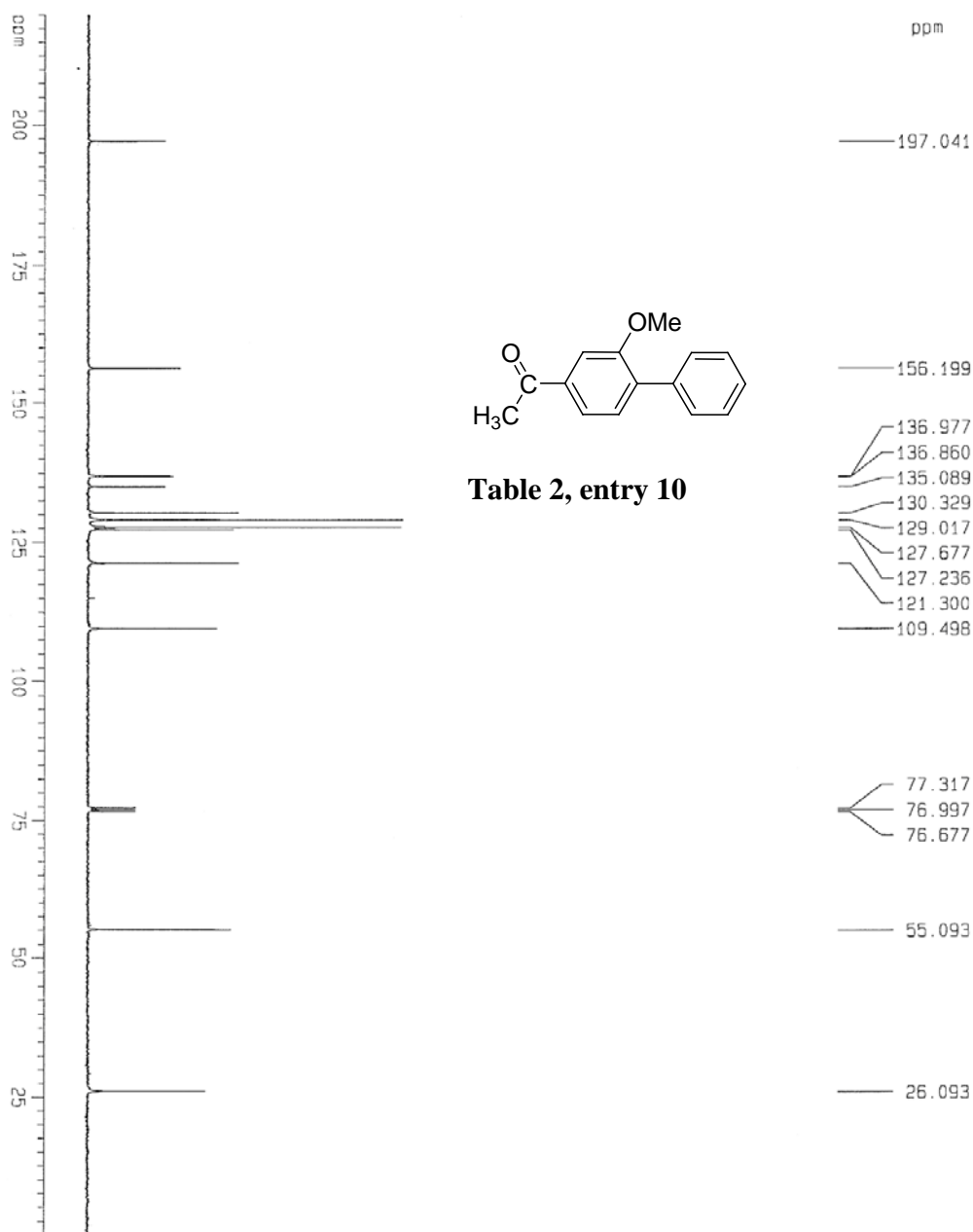


Table 2, entry 10

Current Data Parameters
NAME Coupling
EXPNO 54
PROCNO 1

F2 - Acquisition Parameters
Date_ 20070818
Time 11.33
INSTRUM dpx400
PROBHD 5 mm QNP 1H
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 32
DS 0
SWH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.5083827 sec
RG 8192
DM 19.900 usec
DE 4.50 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 5.80 usec
PL1 -6.00 dB
SFO1 100.6231263 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 71.00 usec
PL2 120.00 dB
PL12 17.00 dB
SFO2 400.1326008 MHz

F2 - Processing parameters
SI 65536
SF 100.6128219 MHz
WDW EM
SSB 0
LB 3.00 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 19.50 cm
F1P 220.000 ppm
F1 22134.82 Hz
F2P -0.000 ppm
F2 -0.00 Hz
PPMCM 11.28205 ppm/cm
HZCM 1135.11902 Hz/cm

File : C:\msdchem\1\DATA\cmso\CT\c377.D
Operator :
Acquired : 17 Aug 2007 18:08 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 8

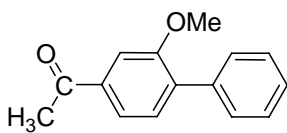
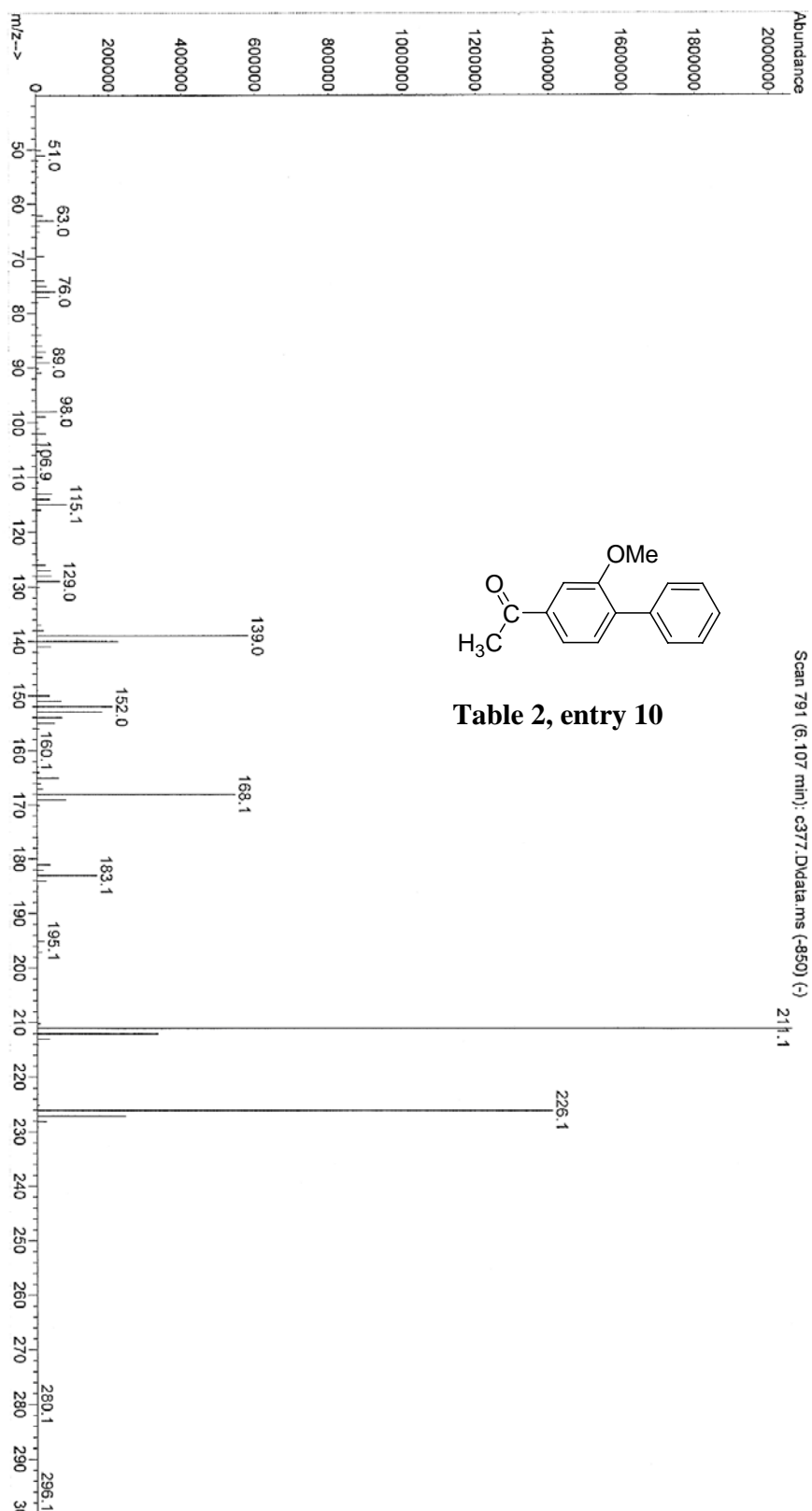
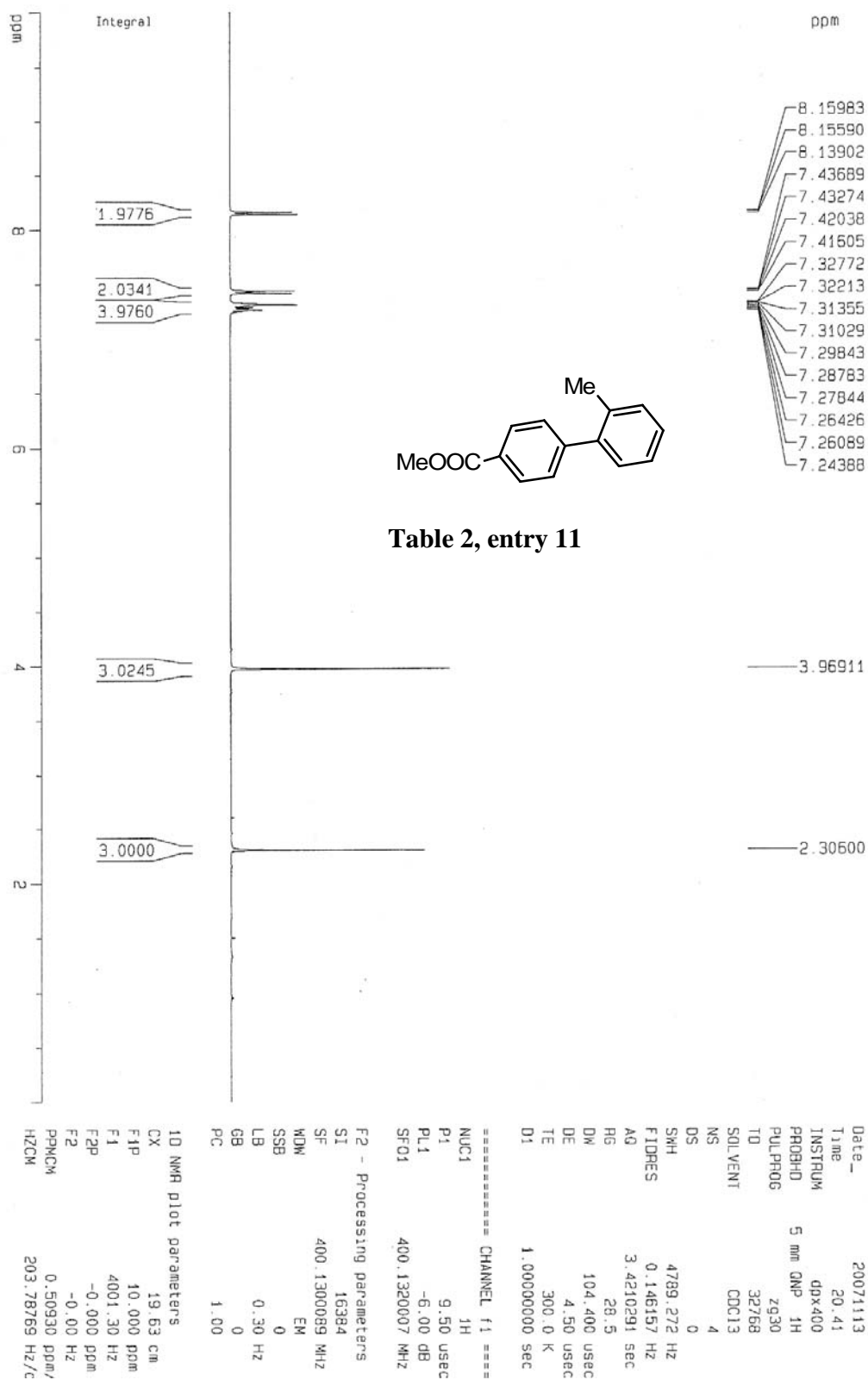


Table 2, entry 10





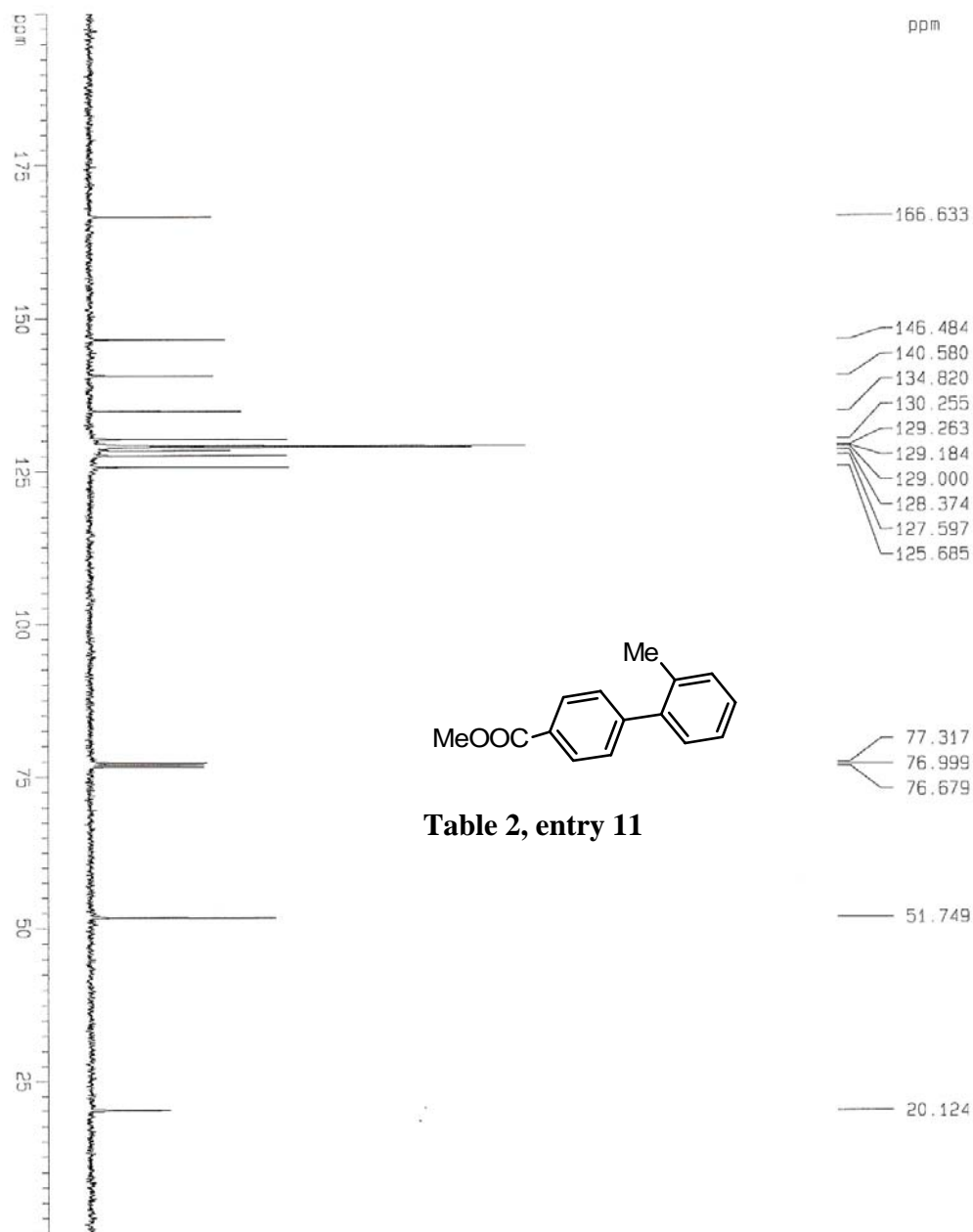


Table 2, entry 11

```

Current Data Parameters
NAME      Coupling
EXPNO     93
PROCNO    1

F2 - Acquisition Parameters
Date_     20071113
Time      20.43
INSTRUM   spect
PROBHD    5 mm GNP 1H
PULPROG   zgpg30
TD         131072
SOLVENT   CDCl3
NS         16
DS         0
SWH         25125.629 Hz
FIDRES     0.191693 Hz
AQ          2.6063627 sec
RG          6192
DE          19.900 usec
TE          300.0 K
D1          3.00000000 sec
d11         0.03000000 sec

===== CHANNEL f1 =====
NUC1       13C
P1          5.80 usec
PL1         -6.00 dB
SFO1       100.6231263 MHz

===== CHANNEL f2 =====
CPOPRG2    waltz16
NUC2        1H
PCPD2       71.00 usec
PL2         120.00 dB
PL12        17.00 dB
SFO2        400.1326008 MHz

F2 - Processing parameters
SI          65536
SF          100.6128020 MHz
WDW         EM
SSB         0
LB          3.50 Hz
GB          0
PC          1.00

1D NMR plot parameters
CX          19.50 cm
F1P         200.000 ppm
F1          20122.56 Hz
F2P         -0.000 ppm
F2          -0.00 Hz
PPMCK       10.25641 ppm/cm
HZCK        1031.92615 Hz/cm

```

File : C:\msdchem\1\DATA\cms0\CT\c511.D
Operator :
Acquired : 13 Nov 2007 18:32 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

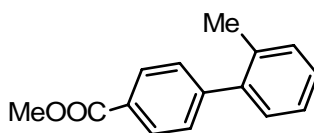
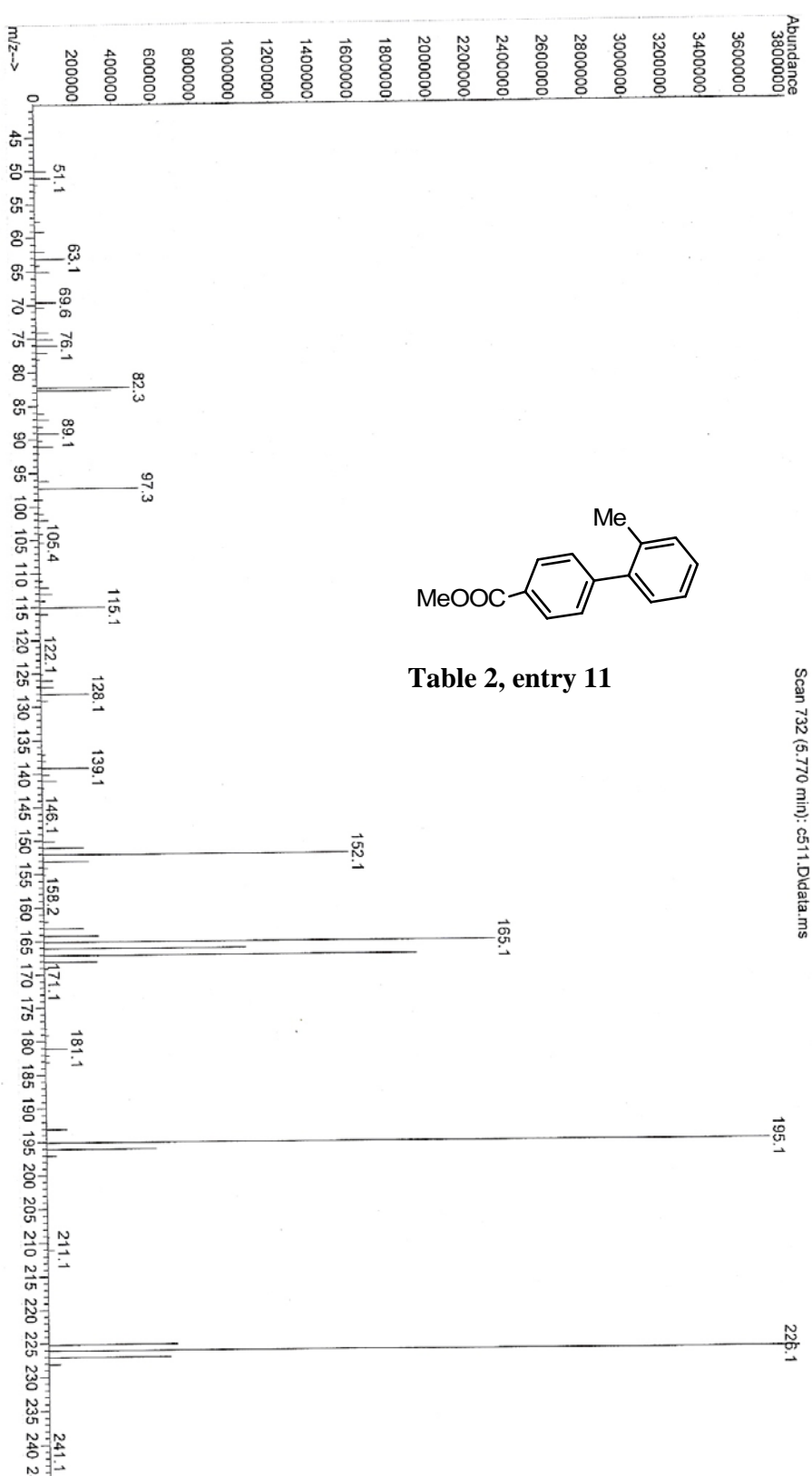


Table 2, entry 11



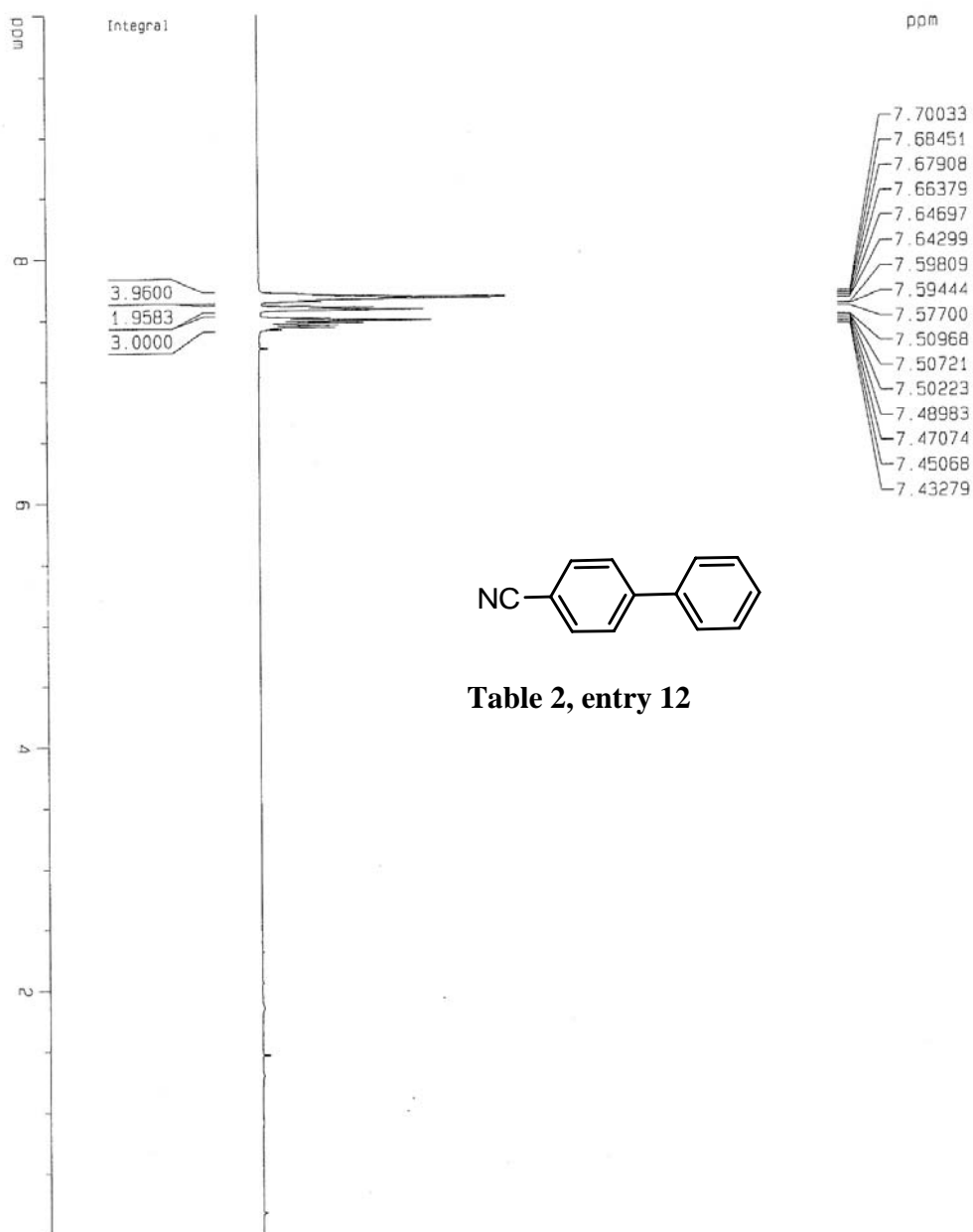


Table 2, entry 12

Current Data Parameters
NAME Coupling
EXPNO 137
PROCNO 1

F2 - Acquisition Parameters
Date_ 20071112
Time 20.25
INSTRUM gpc400
PROBHD 5 mm QNP 1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 4
DS 0
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4210291 sec
RG 57
DM 104.400 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.50 usec
PL1 -6.00 dB
SFO1 400.132007 MHz

F2 - Processing parameters
SI 16384
SF 400.130089 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters
CX 19.63 cm
F1P 10.000 ppm
F1 4001.30 Hz
F2P -0.000 ppm
F2 -0.00 Hz
PPMCM 0.50930 ppm/cm
HZCM 203.78759 Hz/cm

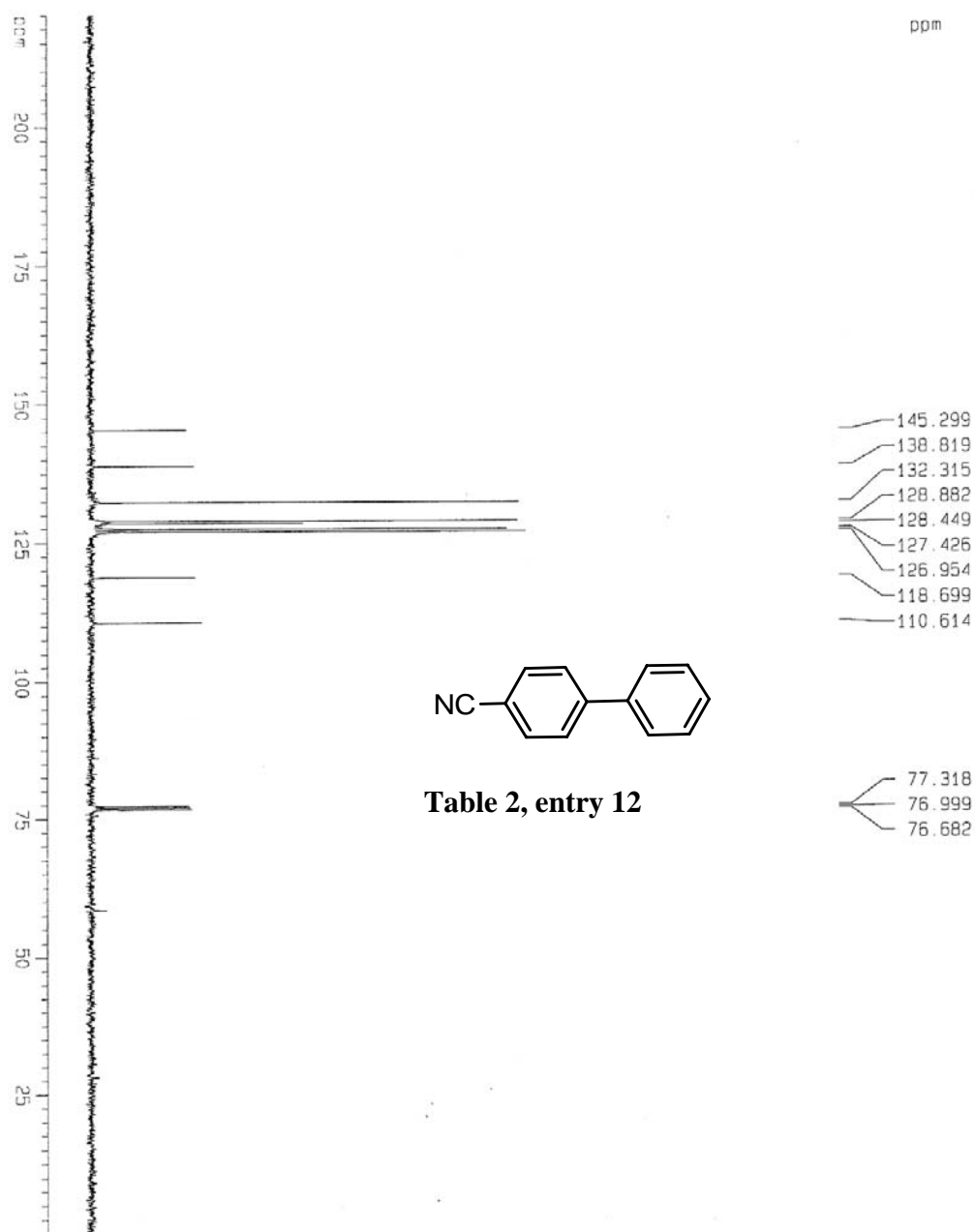


Table 2, entry 12

Current Data Parameters

NAME	Grouping
EXPNO	91
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071112	20.28
INSTRUM	cpd400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	32
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DM	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

MUCL	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
MUCL	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	55536
SF	100.6127993 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CK	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCK	11.28205 ppm/cm
HZCK	1135.11877 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c504.D
Operator :
Acquired : 12 Nov 2007 14:43 using AcqMethod METHOD2.M
Instrument : S973N
Sample Name:
Misc Info :
Vial Number: 4

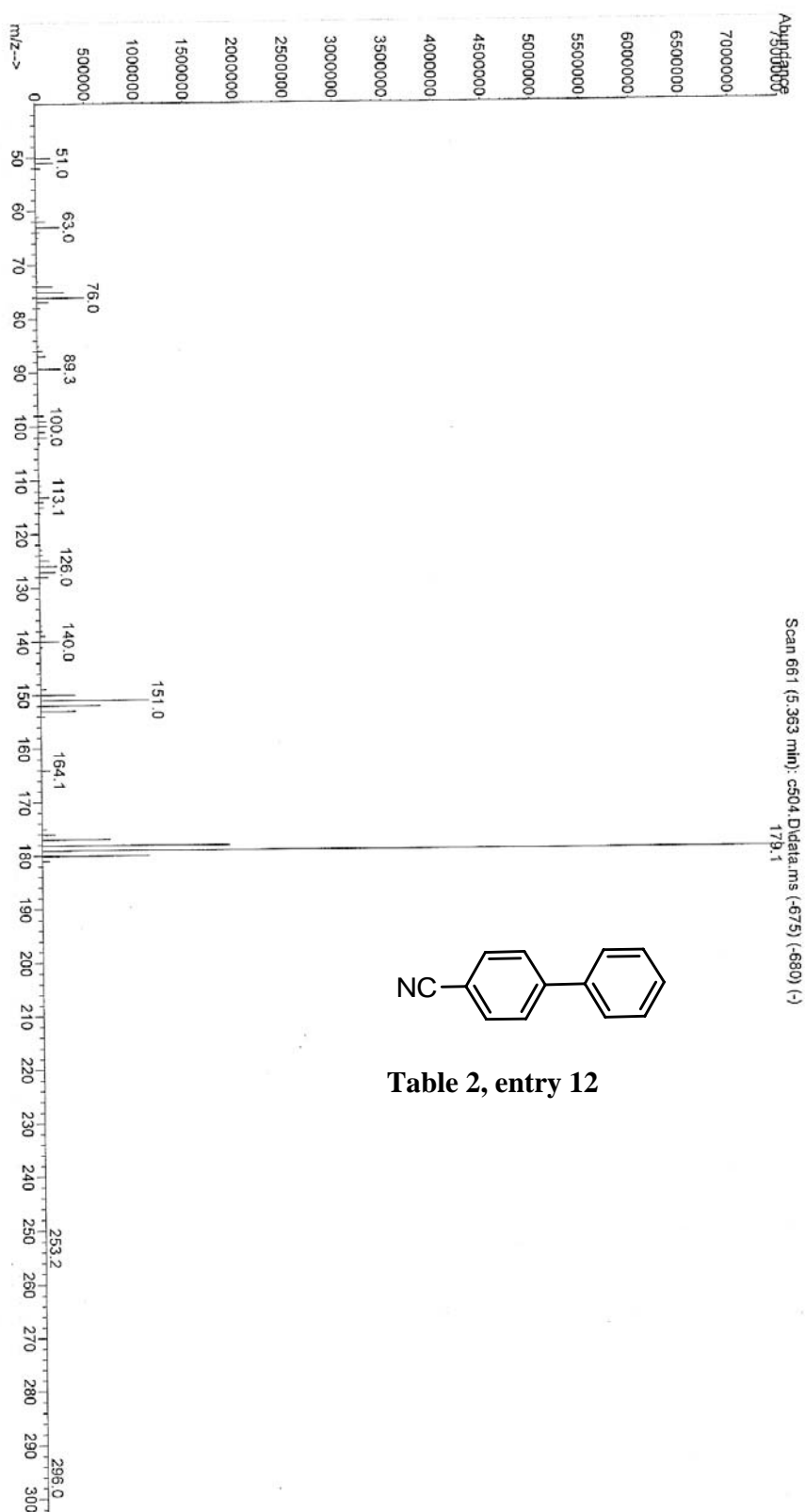


Table 2, entry 12

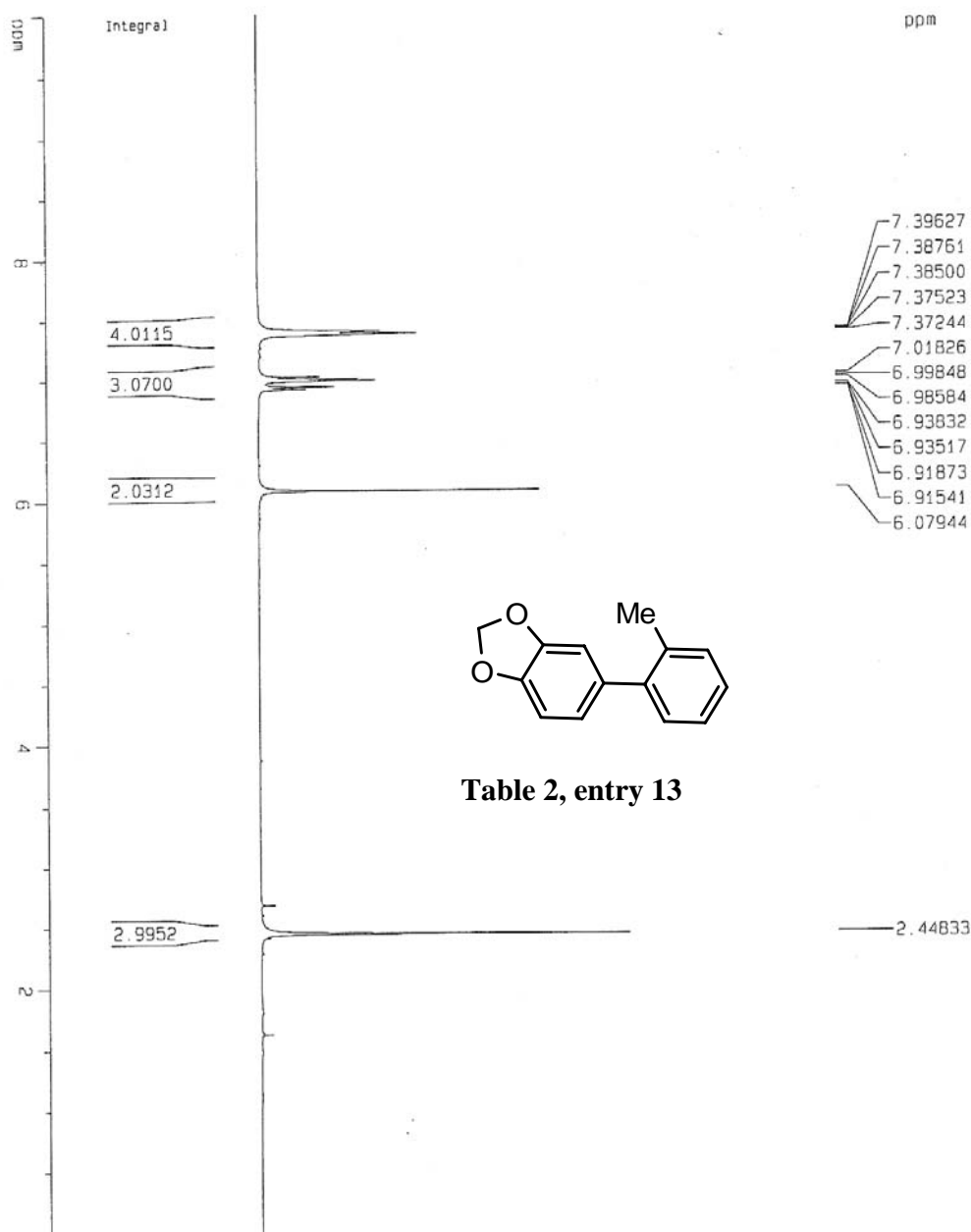


Table 2, entry 13

Current Data Parameters
NAME Coupling
EXPNO 126
PROCNO 1

F2 - Acquisition Parameters
Date_ 20071102
Time 14.43
INSTRUM dp400
PROBHD 5 mm QNP 1H
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 0
SWH 4401.409 HZ
FIDRES 0.134320 HZ
AQ 3.7224948 sec
RG 20.2
DM 113.600 usec
DE 4.50 usec
TE 300.0 K
D1 1.00000000 sec

===== CHANNEL f1 =====
NUC1 1H
P1 9.50 usec
PL1 -6.00 dB
SFO1 400.132006 MHz

F2 - Processing parameters
SI 16384
SF 400.130085 MHz
WDW EM
SSB 0
LB 0.40 HZ
GB 0
PC 1.00

1D NMR plot parameters
CX 19.63 cm
F1P 10.000 ppm
F1 4001.30 HZ
F2P -0.000 ppm
F2 -0.00 HZ
PPMCM 0.50930 ppm/cm
HZCM 203.78769 HZ/cm

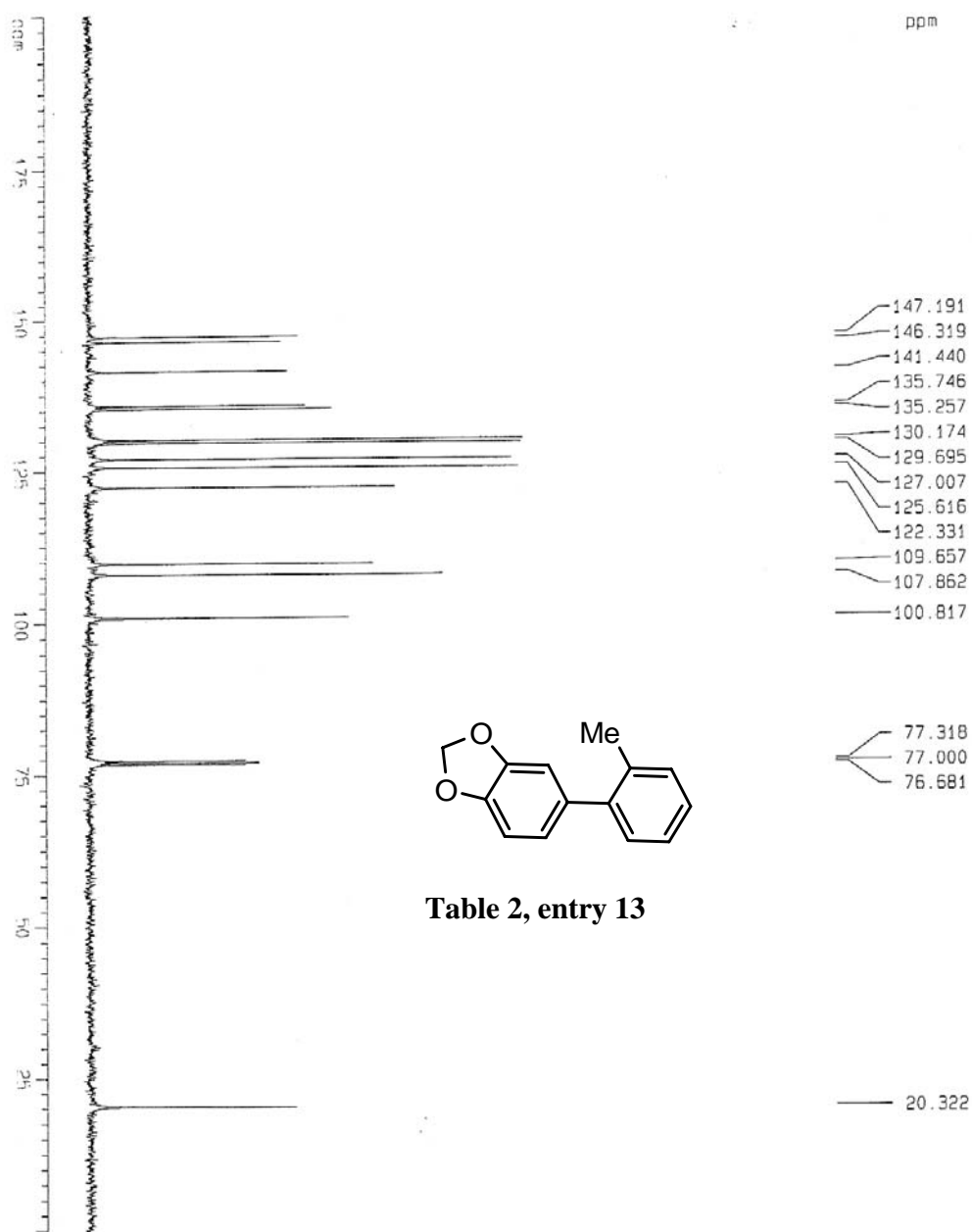


Table 2, entry 13

Current Data Parameters

NAME	COUPLING
EXPNO	B1
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071102	14.46

INSTRUM 60x400
PROBHD 5 mm GNP 1H
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 32
DS 0
SWH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.6083827 sec
RG 8192
DM 19.900 usec
DE 4.50 usec
TE 300.0 K
d11 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128032 MHz
WDW	EM
SSB	0
LB	4.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm/cm
HZCM	1031.92615 Hz/cm

File : C:\msdchem\1\DATA\Kenson\c473.D
Operator :
Acquired : 5 Nov 2007 22:56 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 5

Scan 680 (5.472 min): c473.D\data.ms (-690) (-)

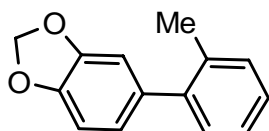
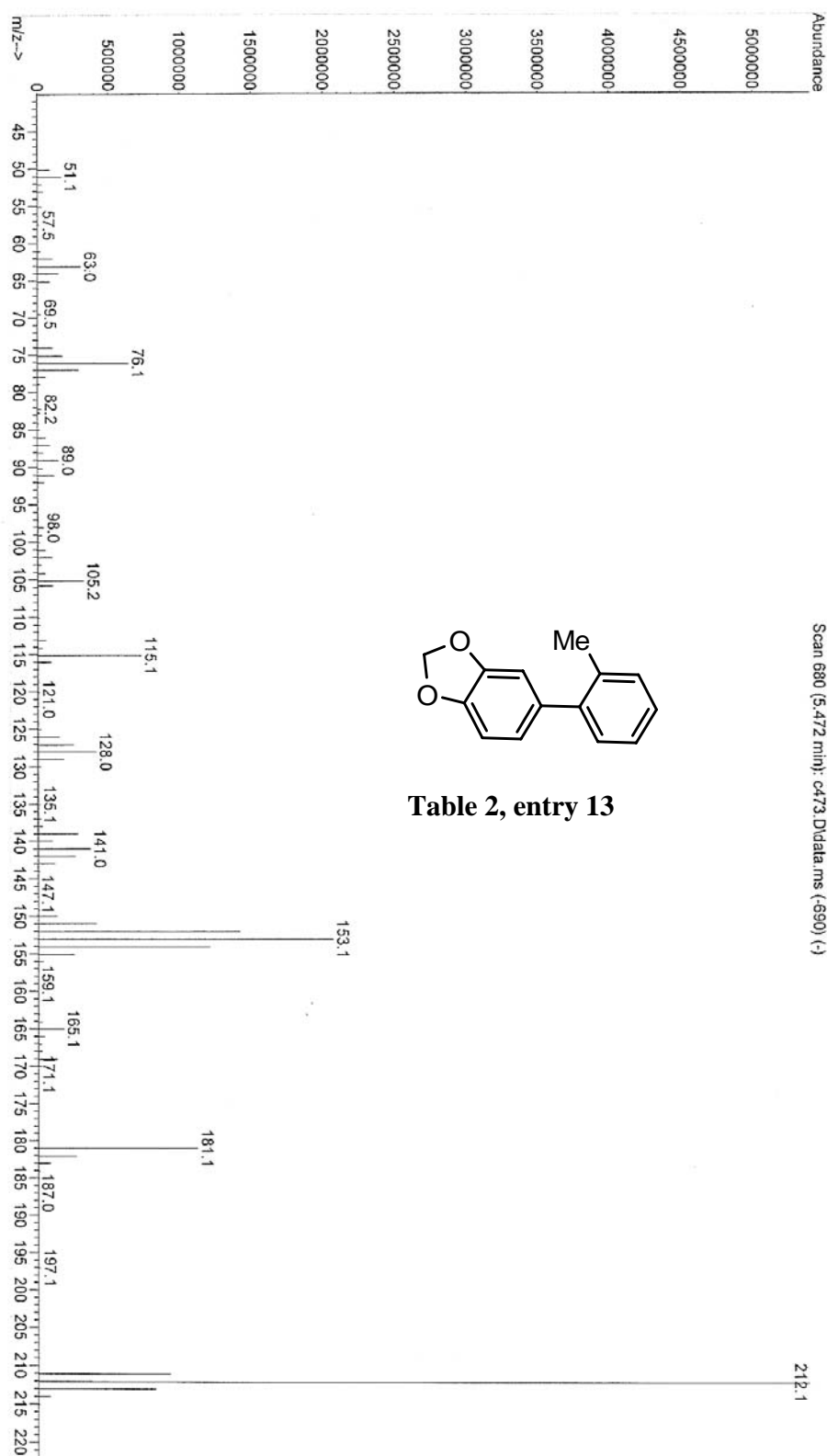
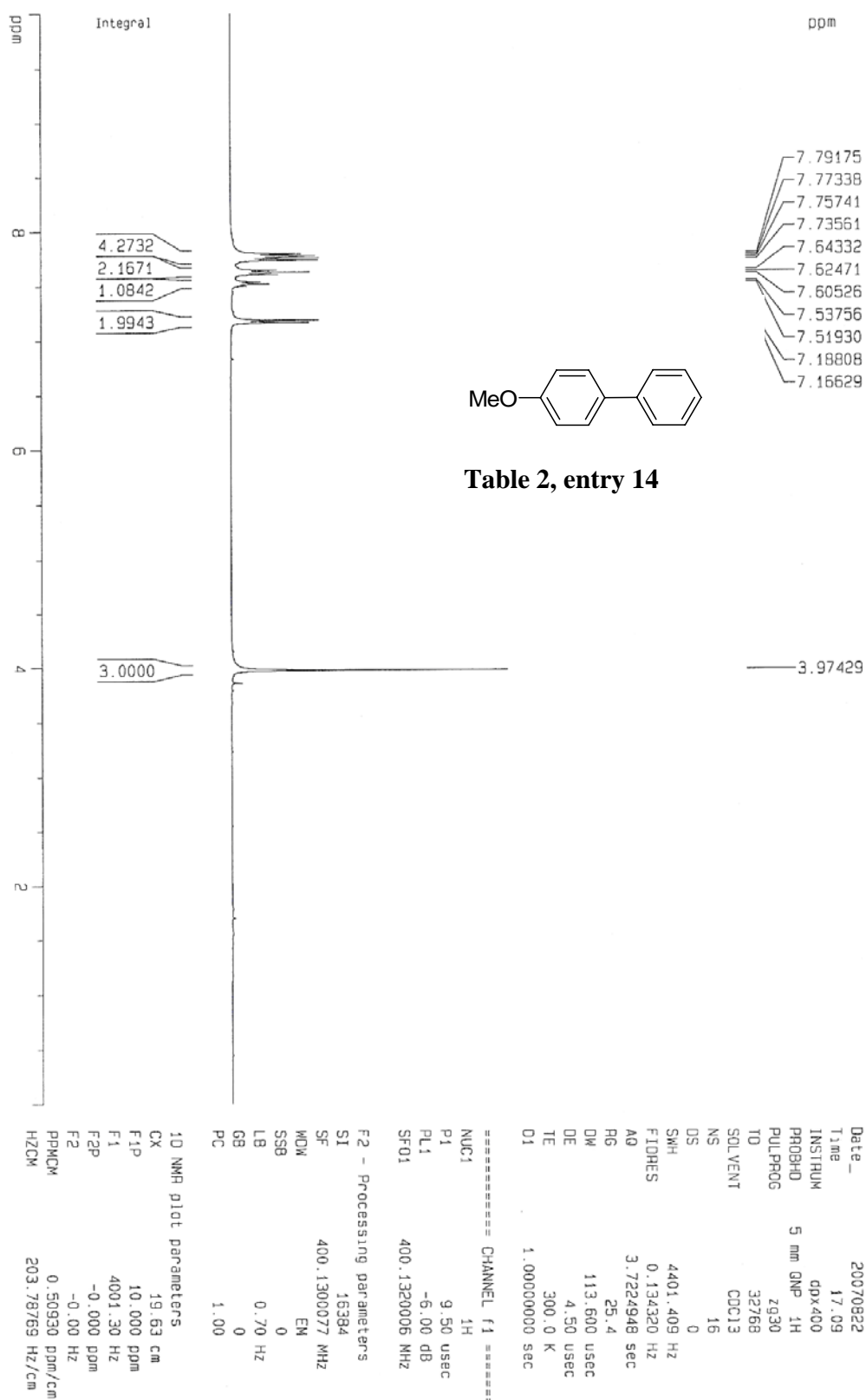


Table 2, entry 13





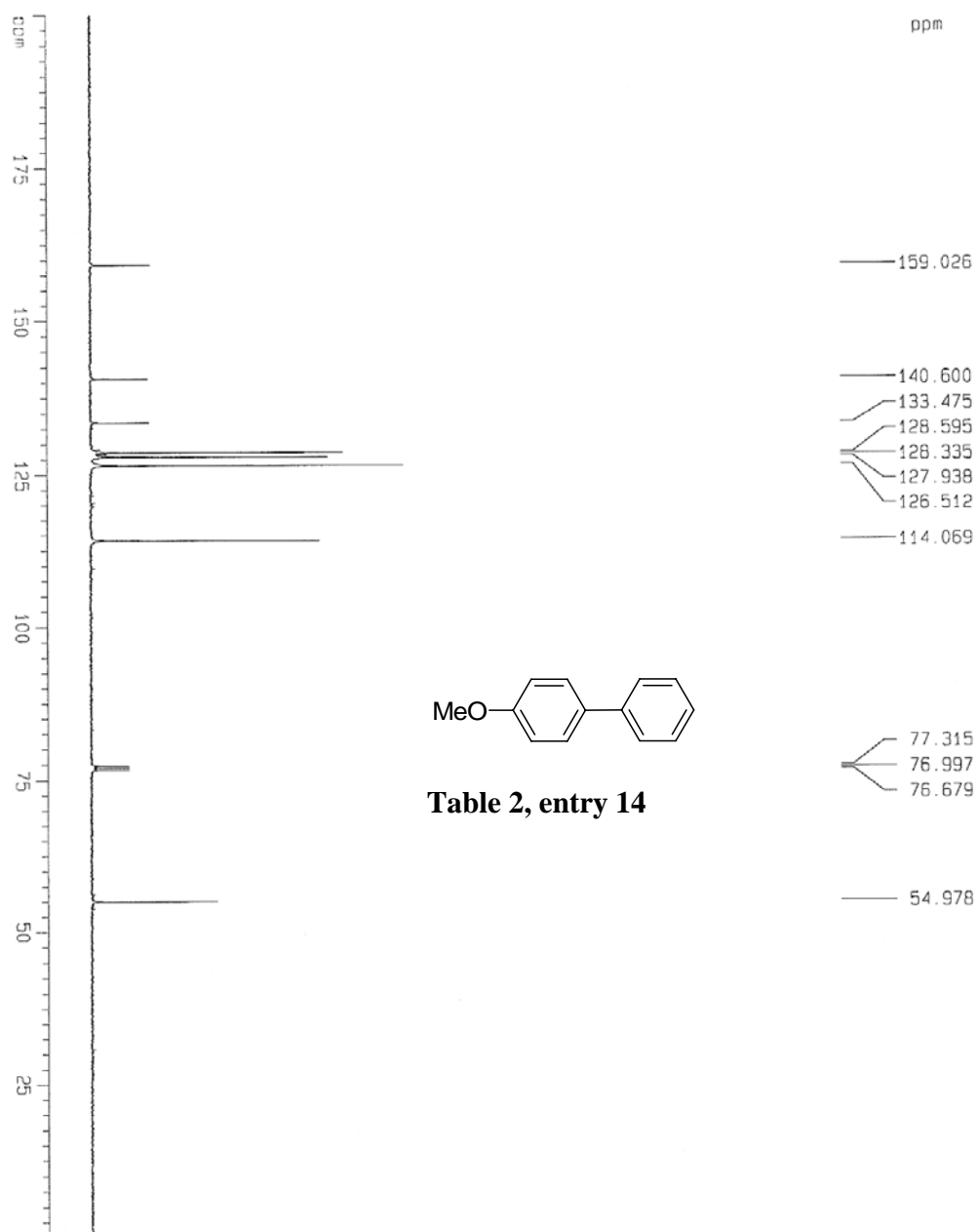


Table 2, entry 14

Current Data Parameters

NAME	Coupling
EXPNO	56
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070822	17.11

INSTRUM gpcx400
PROBHD 5 mm QNP 1H
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 32
DS 0
SWH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.6083827 sec
RG 8192
DW 19.900 usec
DE 4.50 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231253 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128128 MHz
KDN	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm/cm
HZCM	1031.92639 Hz/cm

File : C:\msdchem\1\DATA\snow\c379.D
Operator :
Acquired : 21 Aug 2007 14:34 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 7

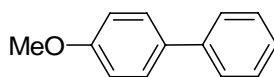
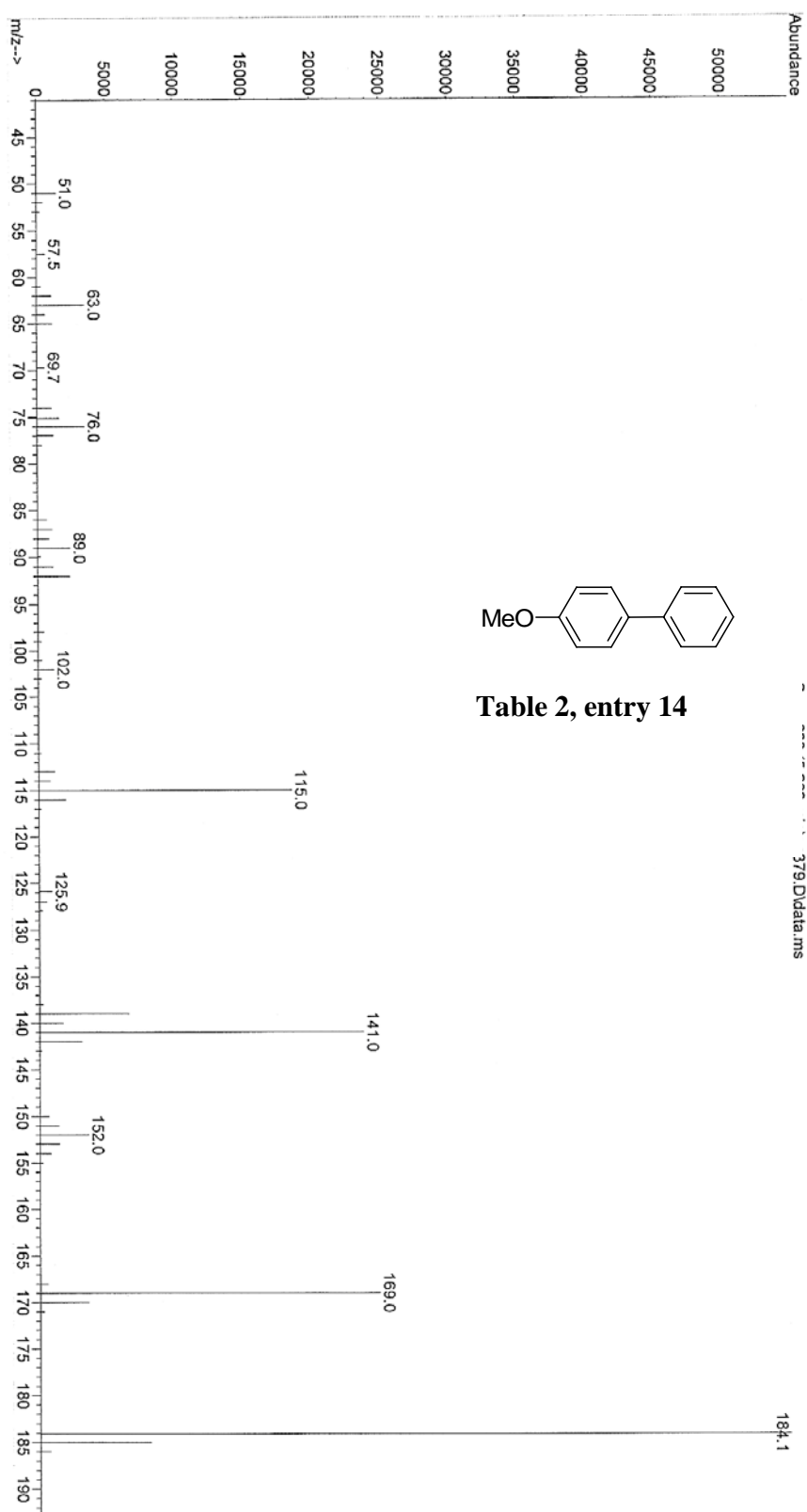
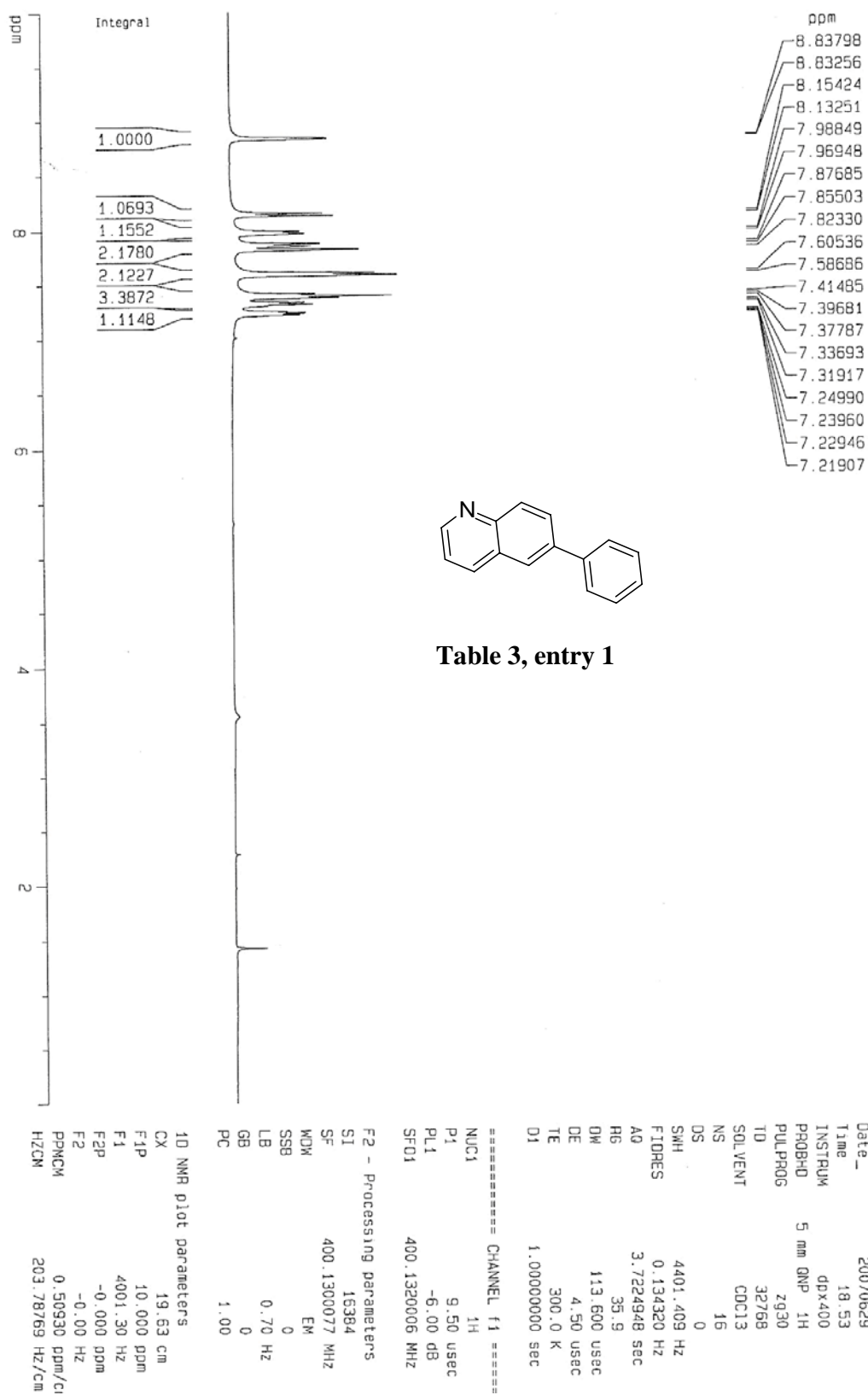


Table 2, entry 14





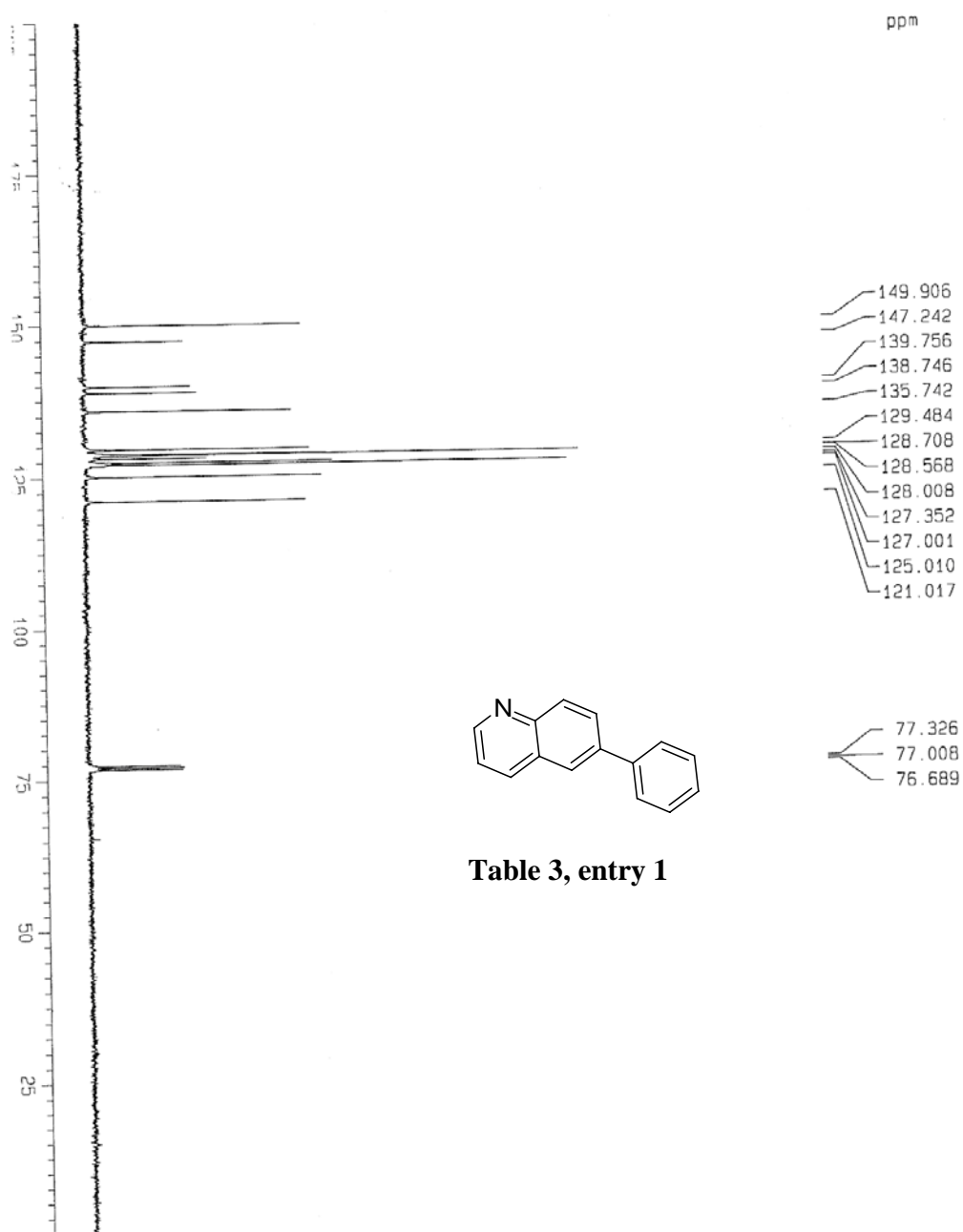


Table 3, entry 1

Current Data Parameters

NAME	Coupling
EXPNO	51
PROCNO	1

F2 - Acquisition Parameters

Date_	20070629
Time	18.55
INSTRUM	gpcx400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	32
DS	0
SWH	25125.629 Hz
FIDRES	0.191593 Hz
AQ	2.6093827 sec
RG	8192
DM	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	¹³ C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CHOPRG2	waltz16
NUC2	¹ H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128089 MHz
WDW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.55 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm
HZCM	1031.92627 Hz

File : C:\msdchem\1\DATA\cmso\CT\c360-78hr.D
Operator : fbm
Acquired : 26 Jun 2007 18:29 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 1

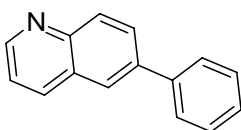
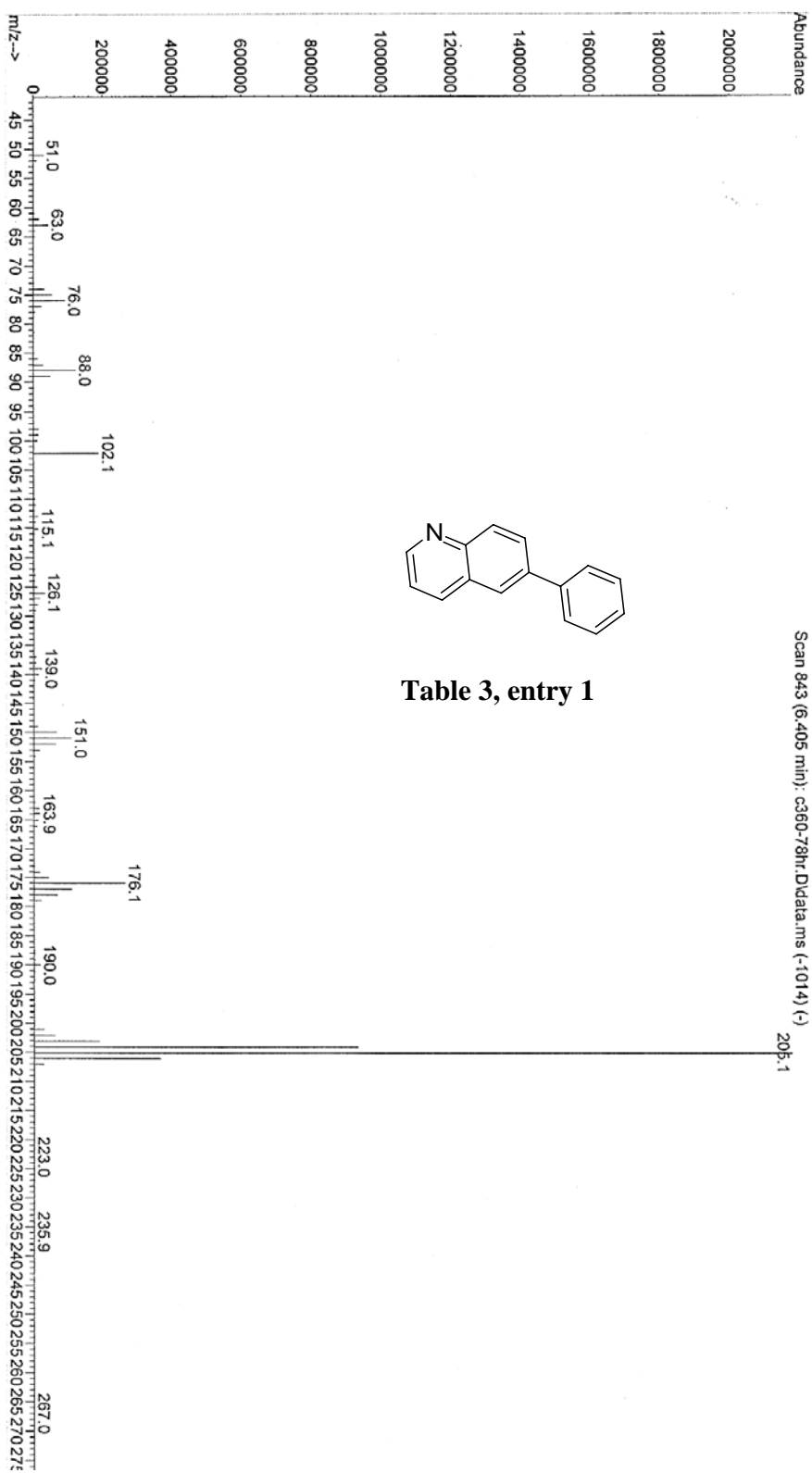
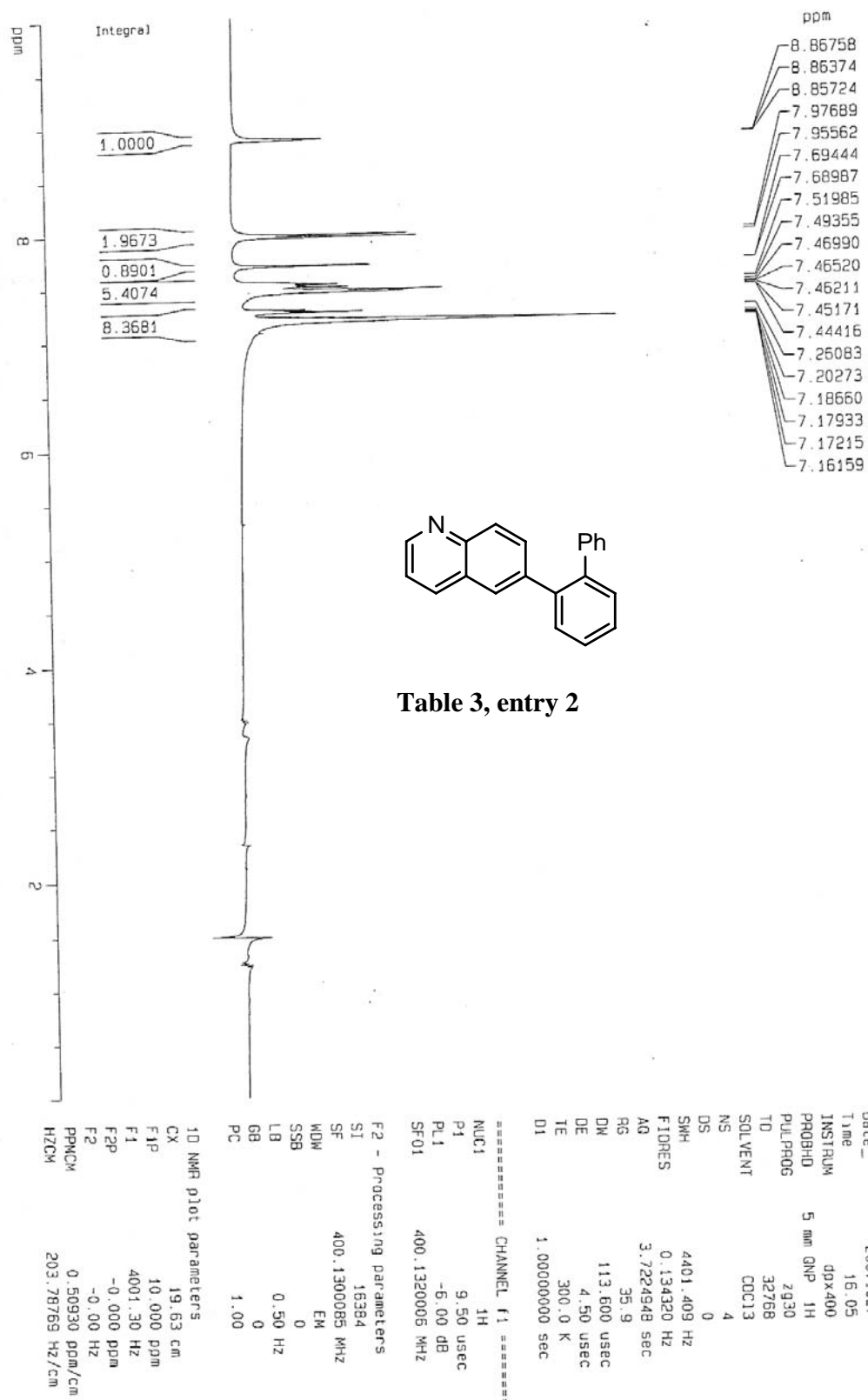


Table 3, entry 1





4. US FM 10/6/11/V1

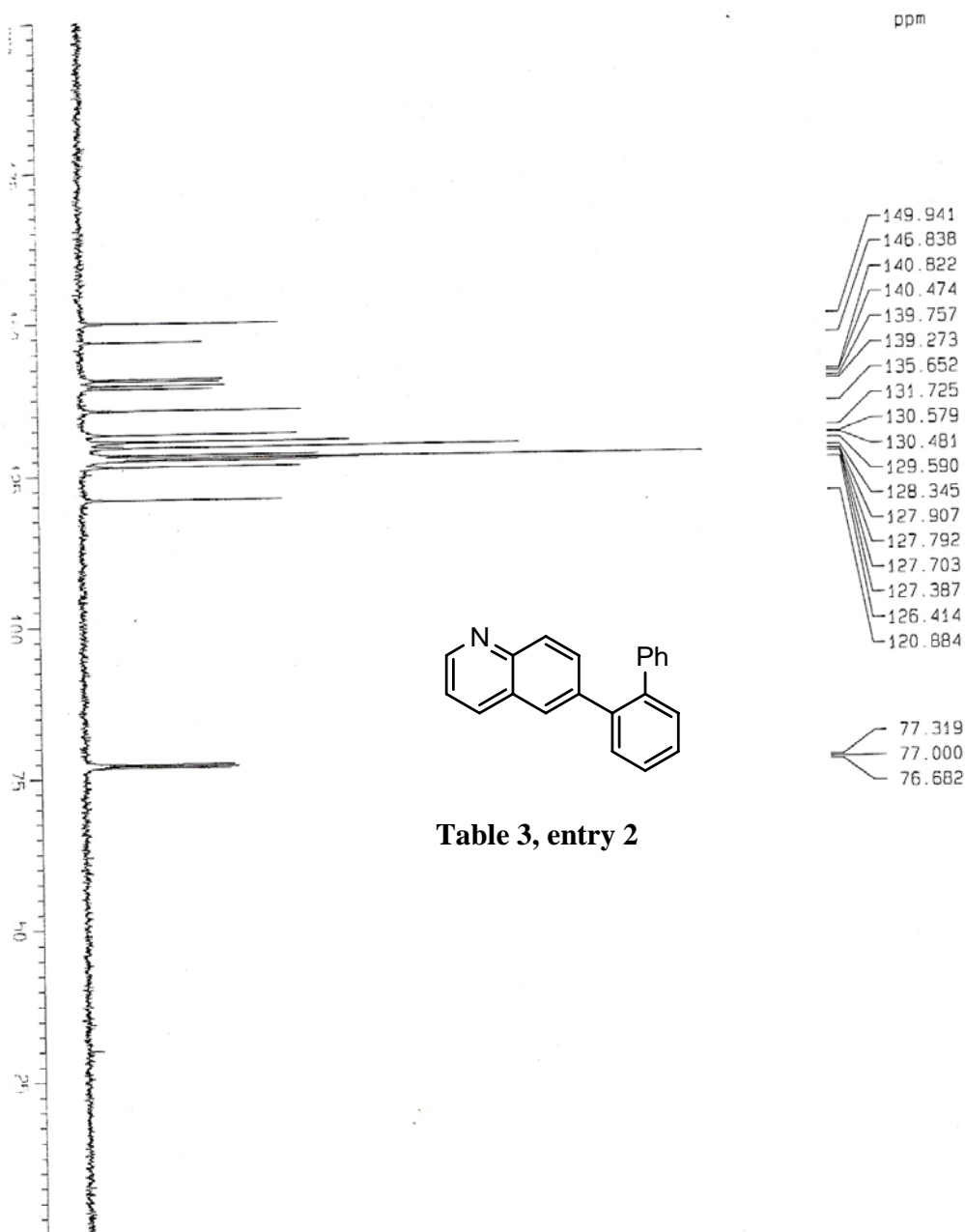


Table 3, entry 2

Current Data Parameters

NAME	Value	Unit
EXPNO	76	
PROCNO	1	

F2 - Acquisition Parameters

Date_	Time	INSTRUM	PROBHD	PULPROG	TD	SOLVENT	NS	DS	SWH	FIDRES	AQ	RG	OW	DE	TE	D1
20071027	16.06	dp400	5 mm QNP 1H	zgpg30	131072	CDCl3	32	0	25125.629 Hz	0.191693 Hz	2.6083627 sec	8192	19.900 usec	4.50 usec	300.0 K	3.00000000 sec
																0.03000000 sec

===== CHANNEL f1 =====

NUC1	P1	PL1	SFO1
13C	5.80 usec	-6.00 dB	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	NUC2	PCPD2	PL2	SFO2
waltz16	1H	71.00 usec	120.00 dB	400.1326008 MHz

F2 - Processing parameters

SF	WDW	SSB	LB	GB	PC
65536	EM	0	4.00 Hz	0	1.00

1D NMR plot parameters

CX	F1P	F1	F2P	F2	PPMCM	HZCM
19.50 cm	200.000 ppm	20122.56 Hz	-0.000 ppm	-0.00 Hz	10.25641 ppm/	1031.92627 Hz/c

File : C:\msdchem\1\DATA\cmso\CT\c462sd.D
Operator :
Acquired : 10 Nov 2007 16:01 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 2

Scan 1156 (8.196 min): c462sd.D\data.ms

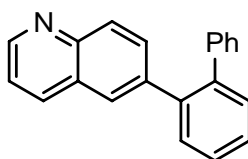
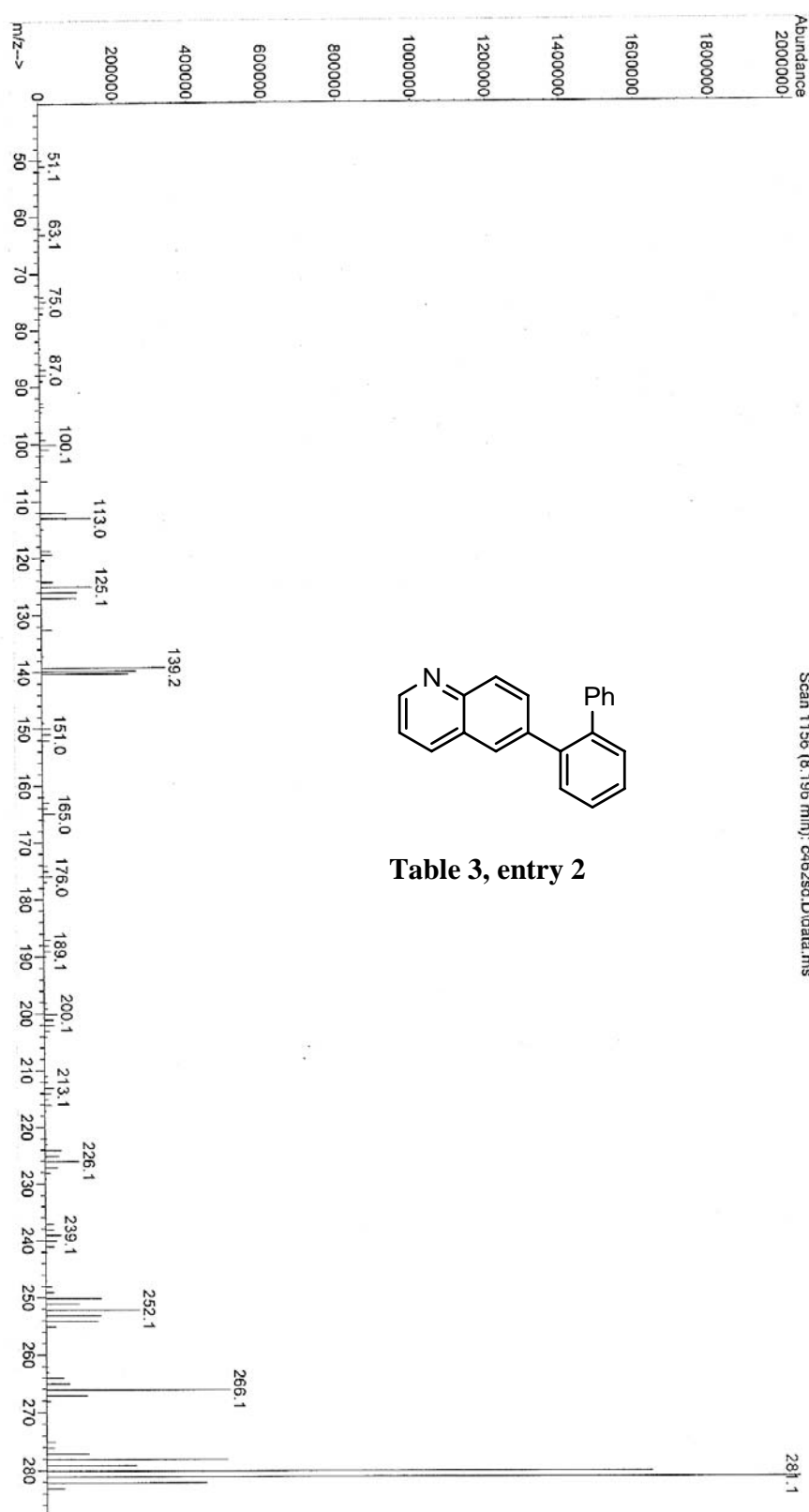
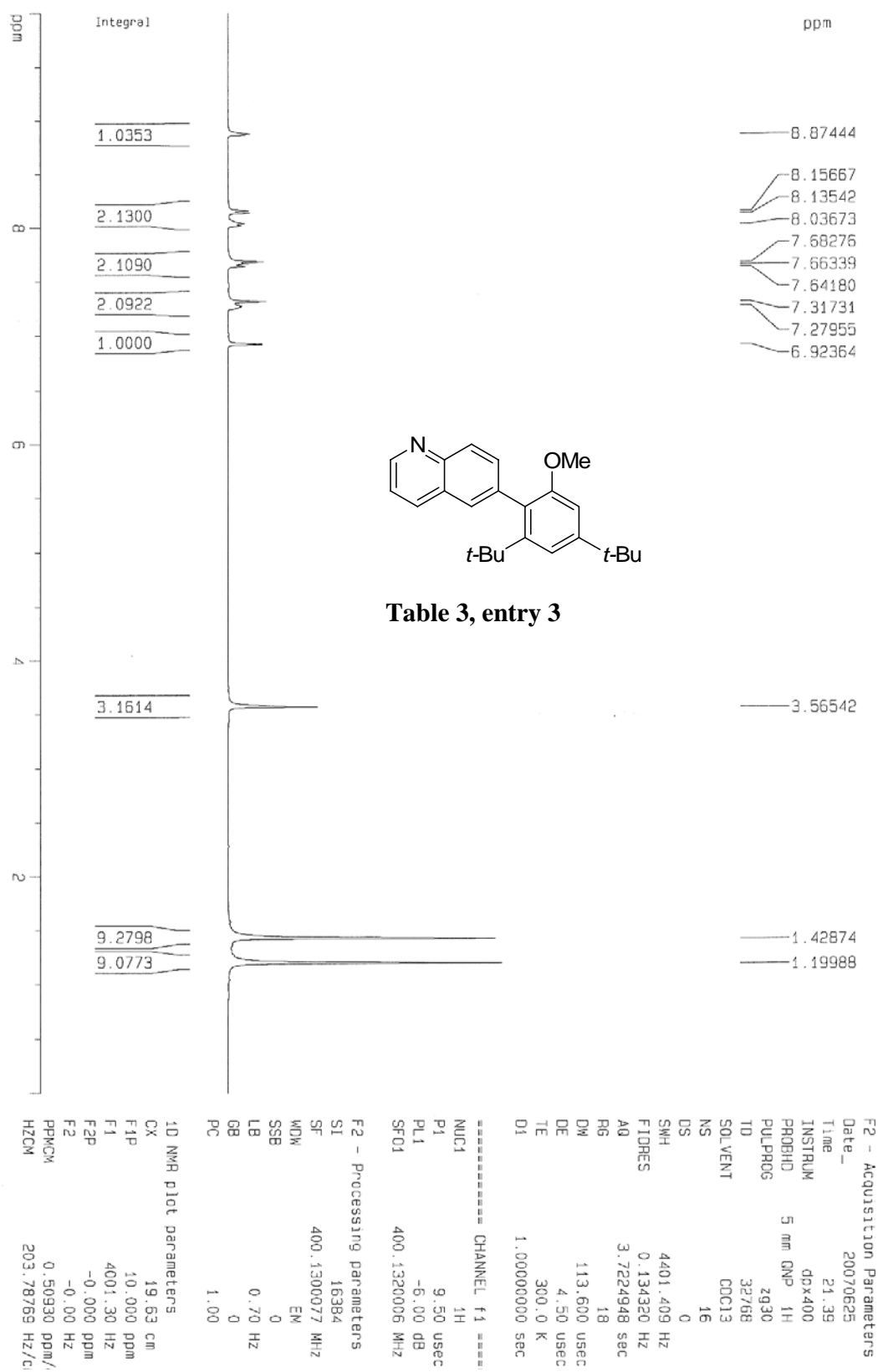


Table 3, entry 2





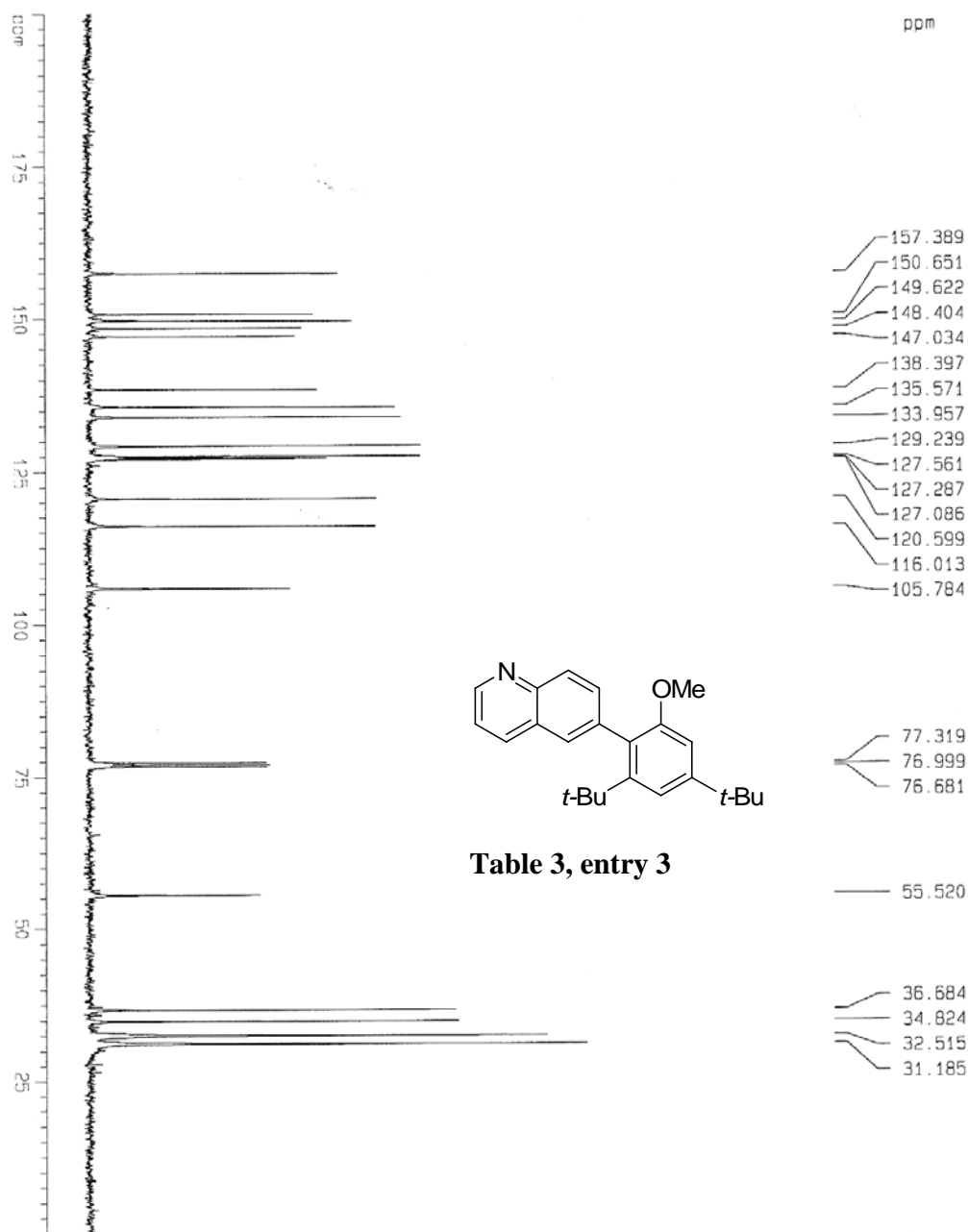


Table 3, entry 3

Current Data Parameters

NAME	COUPLING
EXPNO	45
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070625	21.41
INSTRUM	gpcx400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	64
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DW	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128082 MHz
MDW	EM
SSB	0
LB	4.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm/
HZCM	1031.92627 Hz/c

File : C:\msdchem\1\DATA\cms0\CT\c363.D
Operator :
Acquired : 25 Jun 2007 14:58 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 2

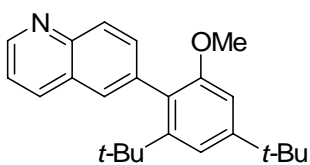
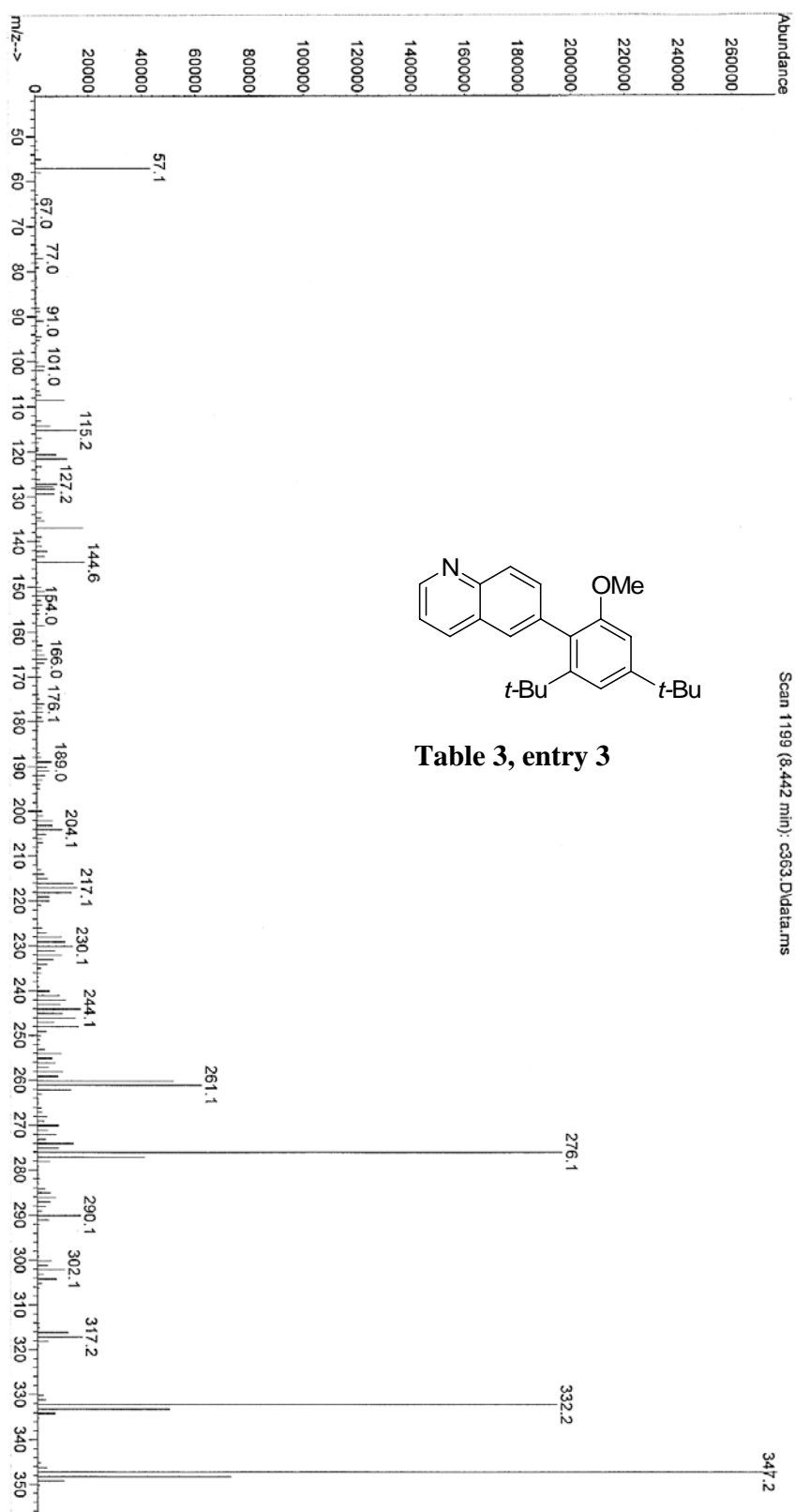


Table 3, entry 3



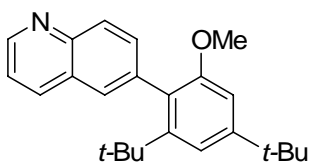
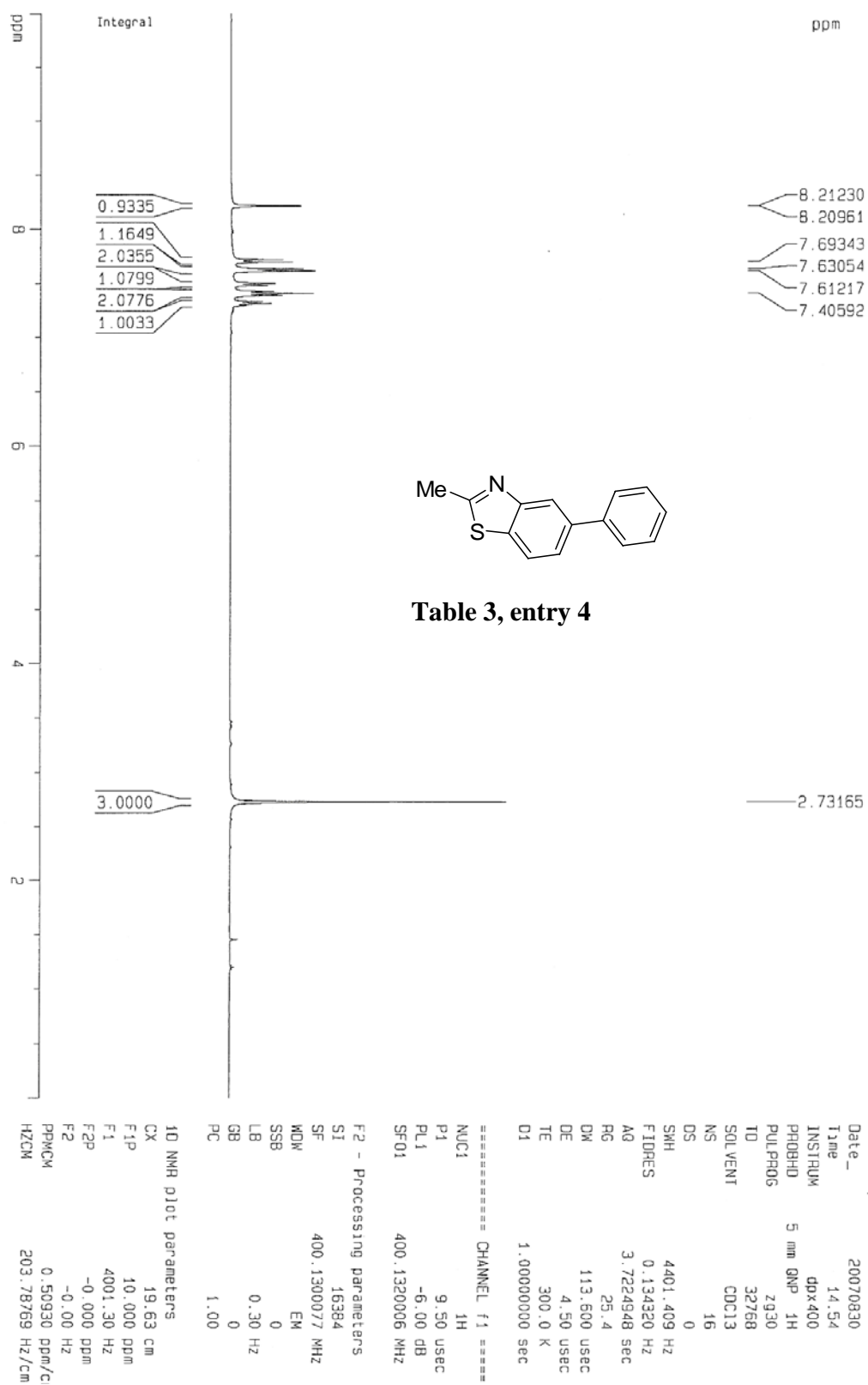


Table 3, entry 3

Molecular formula:
C₂₄H₂₉N₁O₁
[M]⁺ (theoretical)
= 347.2244



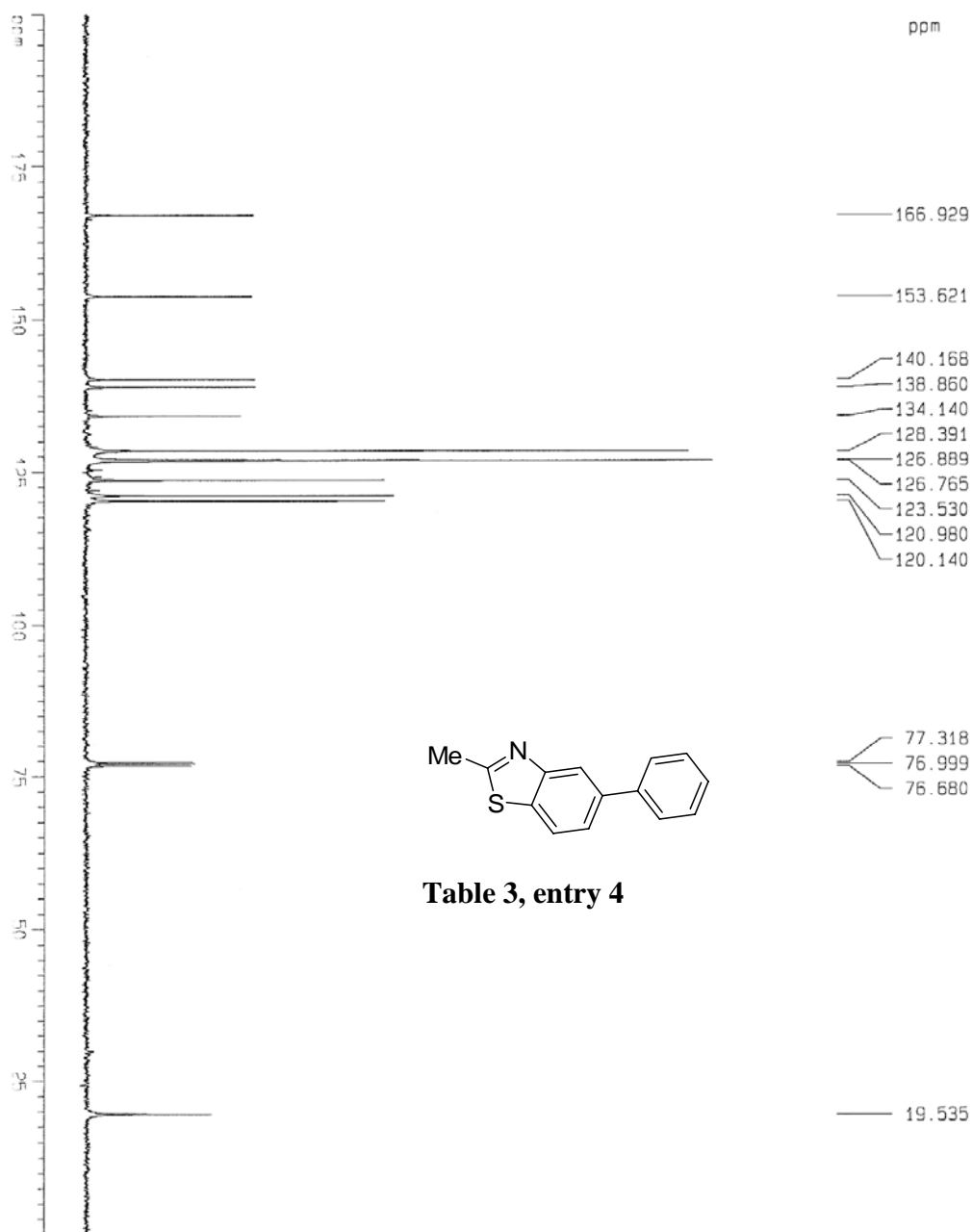


Table 3, entry 4

Current Data Parameters

NAME	COUPLING
EXPNO	59
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20070630	14.56
INSTRUM	dp400
PROBHD	5 mm GNP 4H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	32
DS	0
SWH	25125.629 Hz
FIDRES	0.191593 Hz
AQ	2.6083827 sec
RG	8192
DM	19.900 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-5.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

F2 - Processing parameters

SI	65536
SF	100.6128262 MHz
WDW	EM
SSB	0
LB	4.00 Hz
GB	0
PC	1.00

10 NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	10.25641 ppm/ci
HZCM	1031.92639 Hz/cm

File : C:\msdchem\1\DATA\cmso\CT\c385.D
Operator :
Acquired : 30 Aug 2007 13:12 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 6

Scan 876 (6.594 min): c385.D\data.ms (-1114) (-)

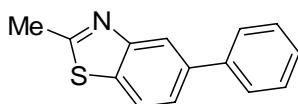
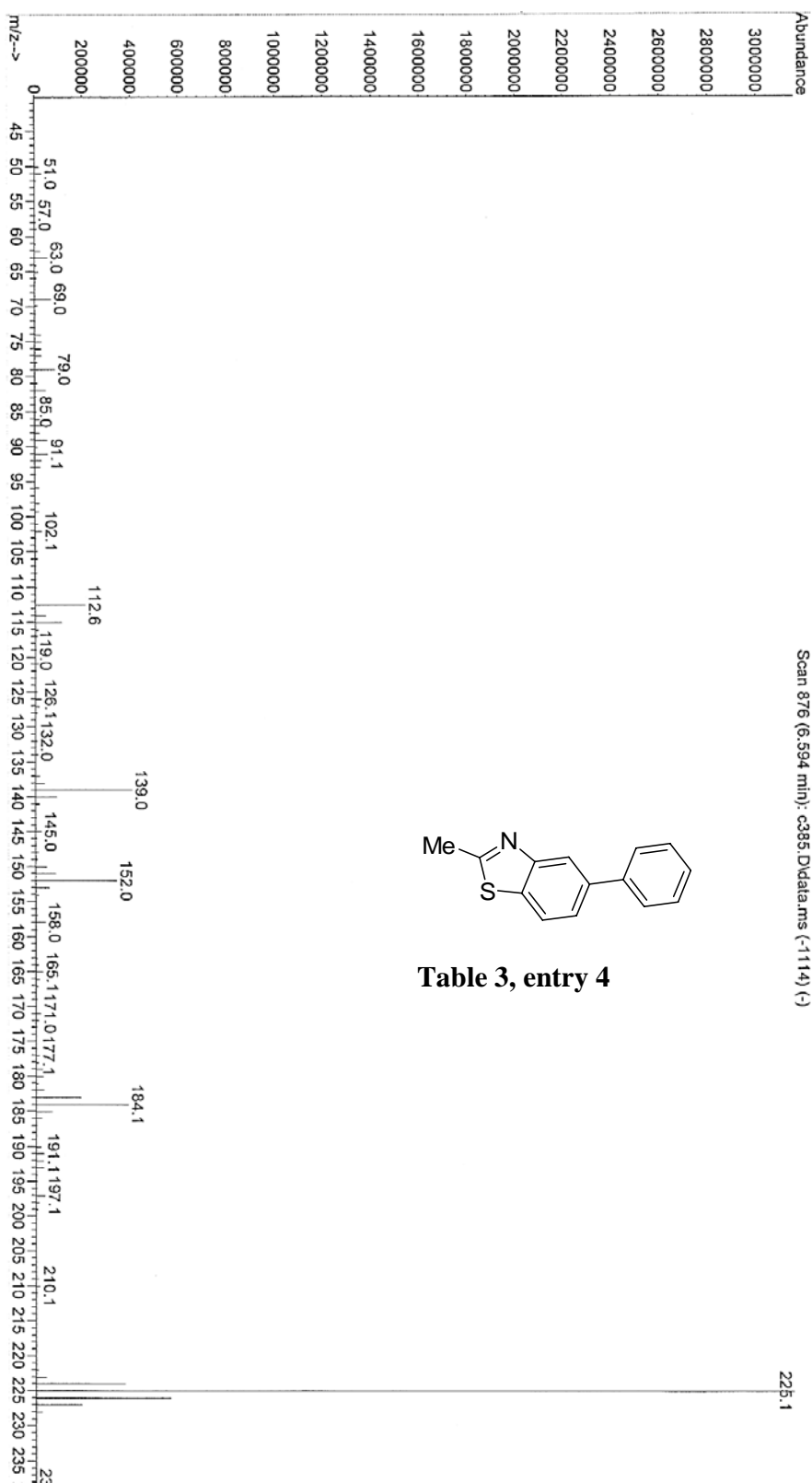
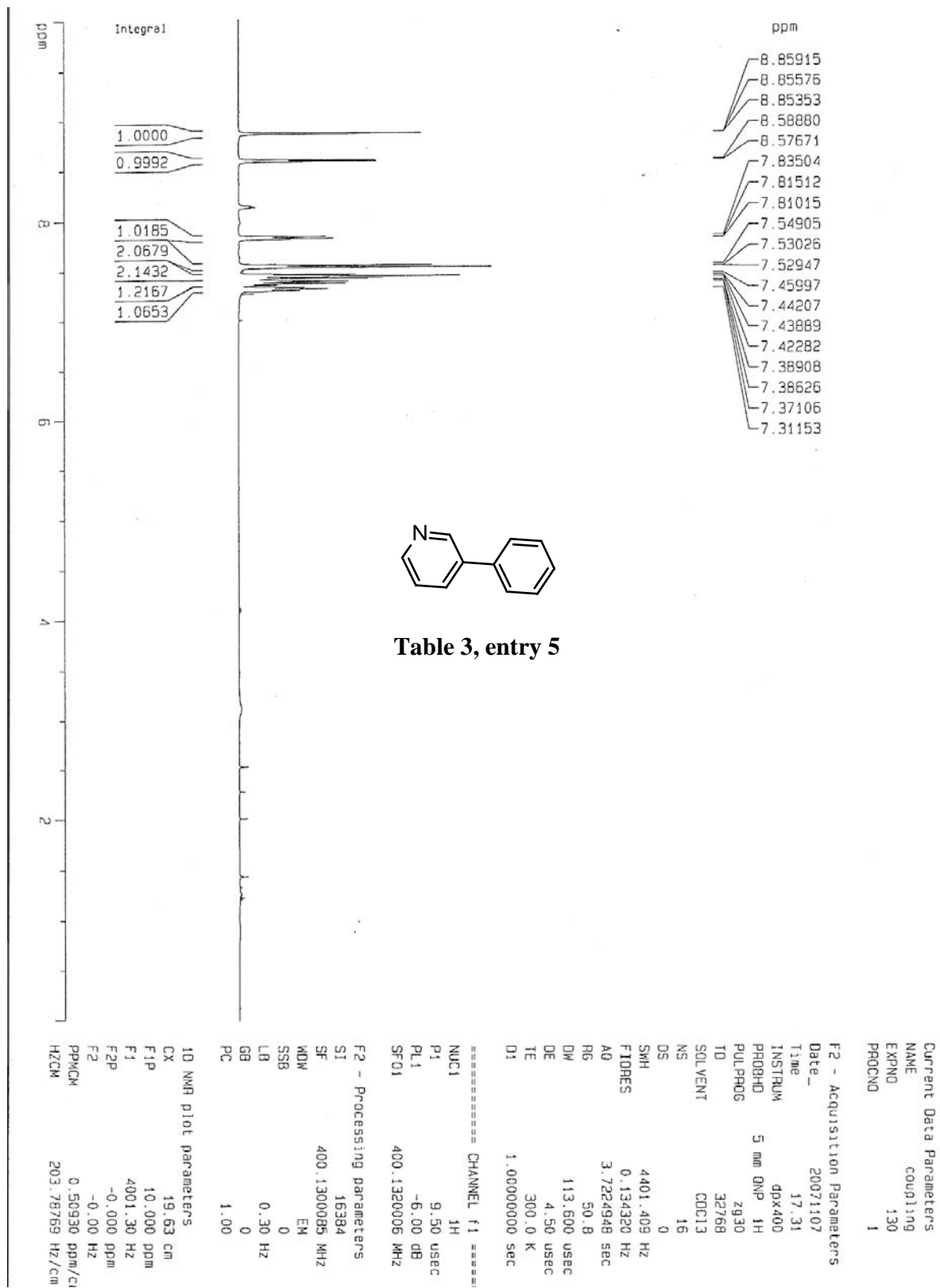


Table 3, entry 4





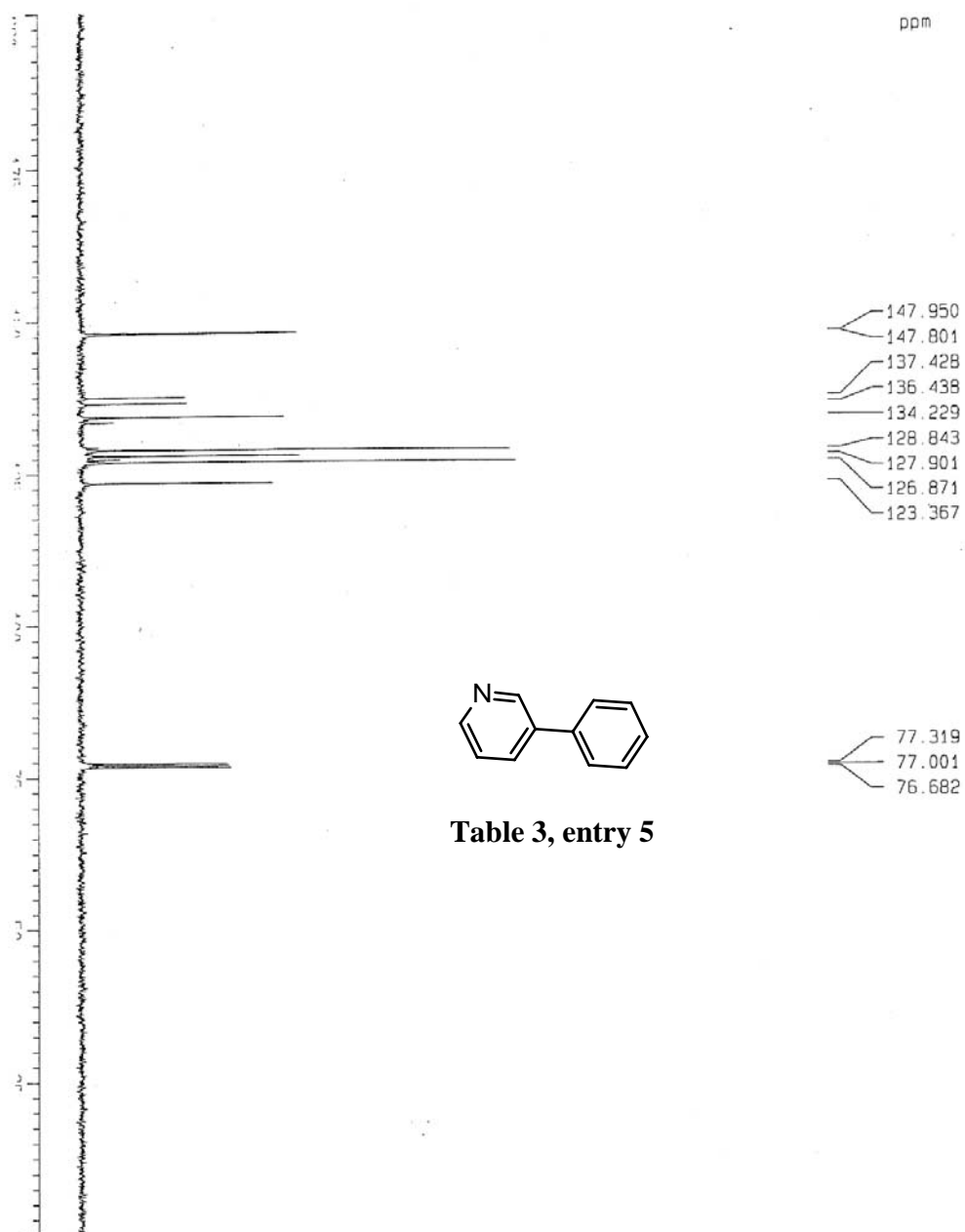


Table 3, entry 5

```

Current Data Parameters
NAME          Coupling
EXPNO         84
PROCNO        1

F2 - Acquisition Parameters
Date_         20071107
Time          17.33
INSTRUM       qnp400
PROBHD        5 mm QNP 1H
PULPROG       zgpg30
TD            131072
SOLVENT       CDCl3
NS            32
DS            0
SWH           25125.629 HZ
FIDRES        0.191593 HZ
AQ            2.6083827 sec
RG            8192
DM            19.500 usec
DE            4.50 usec
TE            300.0 K
D1            3.00000000 sec
d11           0.03000000 sec

===== CHANNEL f1 =====
NUC1          13C
P1            5.80 usec
PL1           -6.00 dB
SFO1          100.6231253 MHz

===== CHANNEL f2 =====
COPYPAG2      wait215
NUC2          1H
PCPD2         71.00 usec
PL2           120.00 dB
PL12          17.00 dB
SFO2          400.1326008 MHz

F2 - Processing parameters
SI            65536
SF            100.6127959 MHz
WDW           EM
SSB           0
LB            4.00 HZ
GB            0
PC            1.00

1D NMR plot parameters
CX            19.50 cm
F1P           200.000 ppm
F1            20122.56 HZ
F2P           -0.000 ppm
F2            -0.00 HZ
PPMCM         10.25641 ppm/cm
HZCM          1031.92515 Hz/cm

```

File : C:\msdchem\1\DATA\cms0\CT\c496.D
Operator :
Acquired : 7 Nov 2007 14:21 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 2

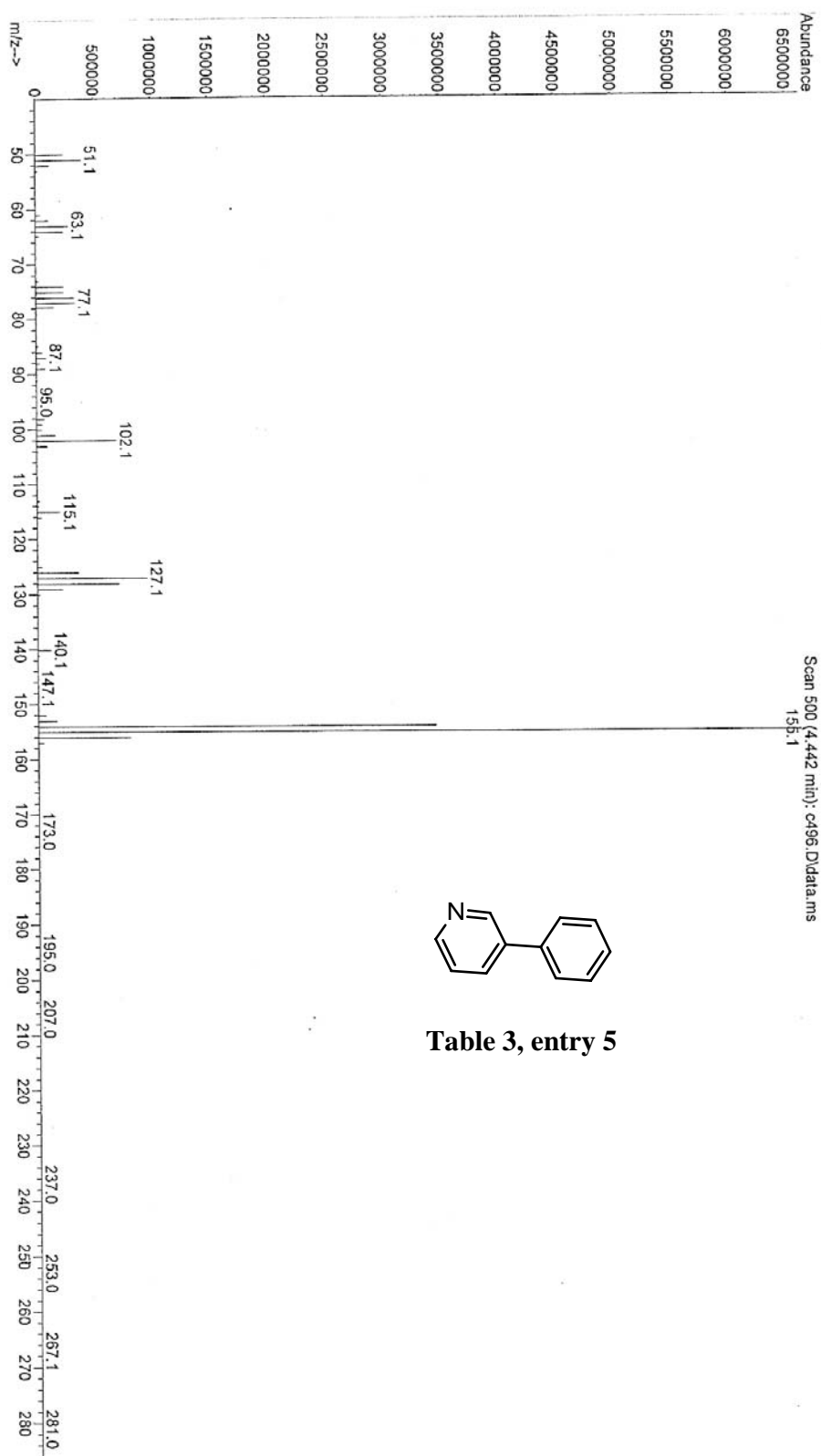
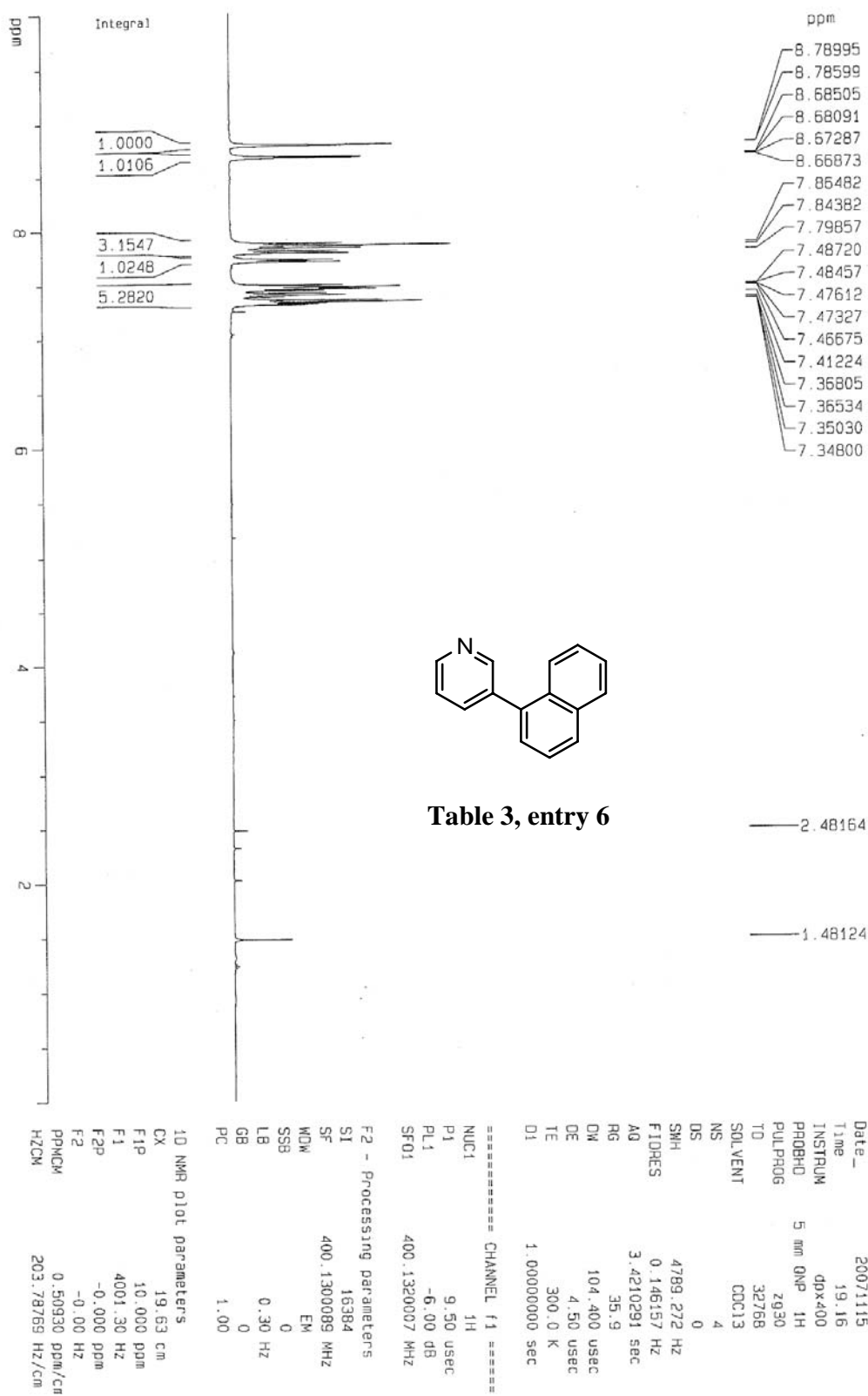


Table 3, entry 5



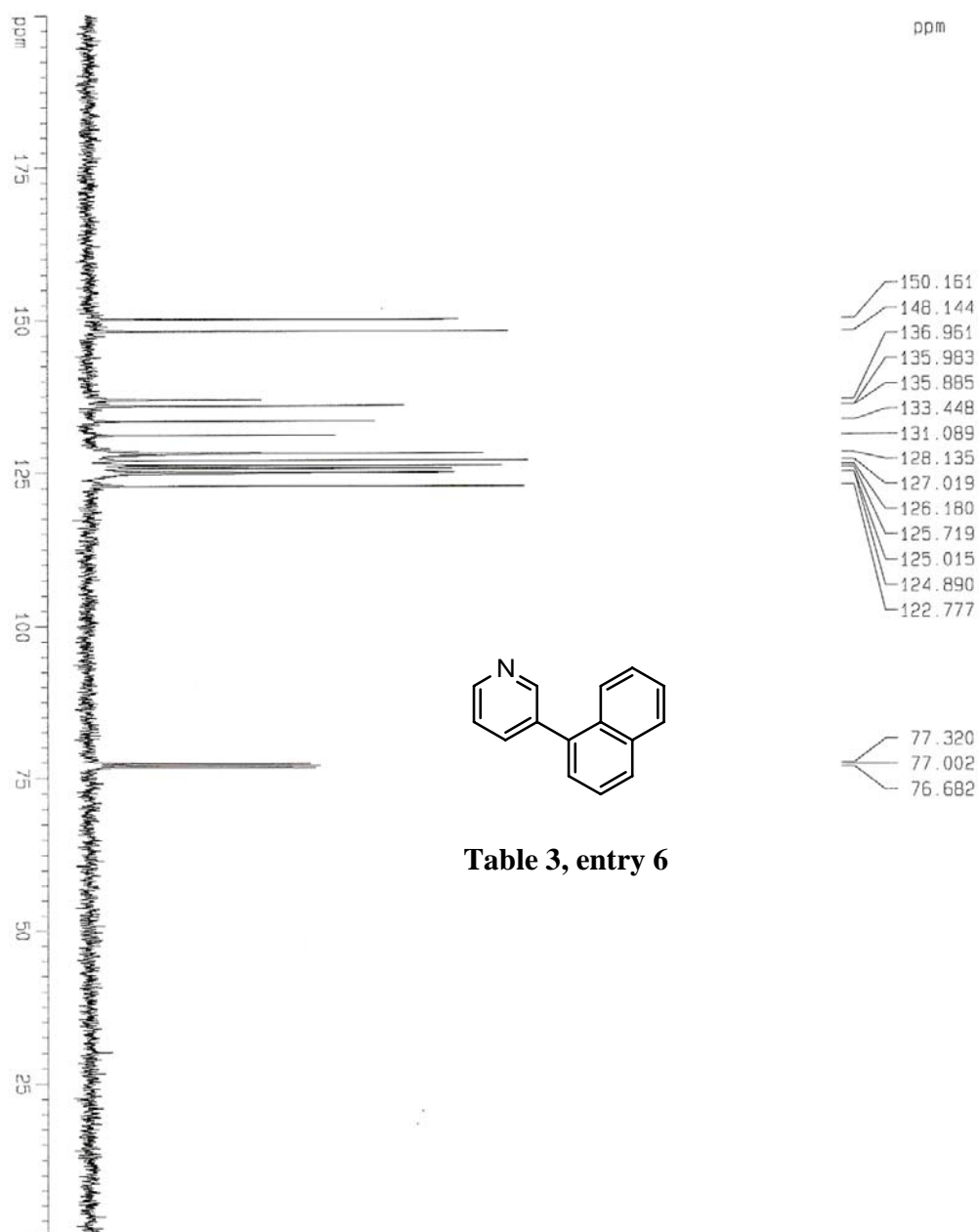


Table 3, entry 6

Current Data Parameters

NAME	Coupling
EXPNO	101
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071117	14.40
INSTRUM	dp400
PROBHD	5 mm QNP 1H
PULPROG	zgpg30
TD	131072
SOLVENT	CDCl3
NS	16
DS	0
SWH	25125.629 Hz
FIDRES	0.191693 Hz
AQ	2.6083827 sec
RG	8192
DM	19.908 usec
DE	4.50 usec
TE	300.0 K
D1	3.00000000 sec
d11	0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-6.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

CPDPRG2	waltz16
NUC2 <td>1H</td>	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1332010 MHz

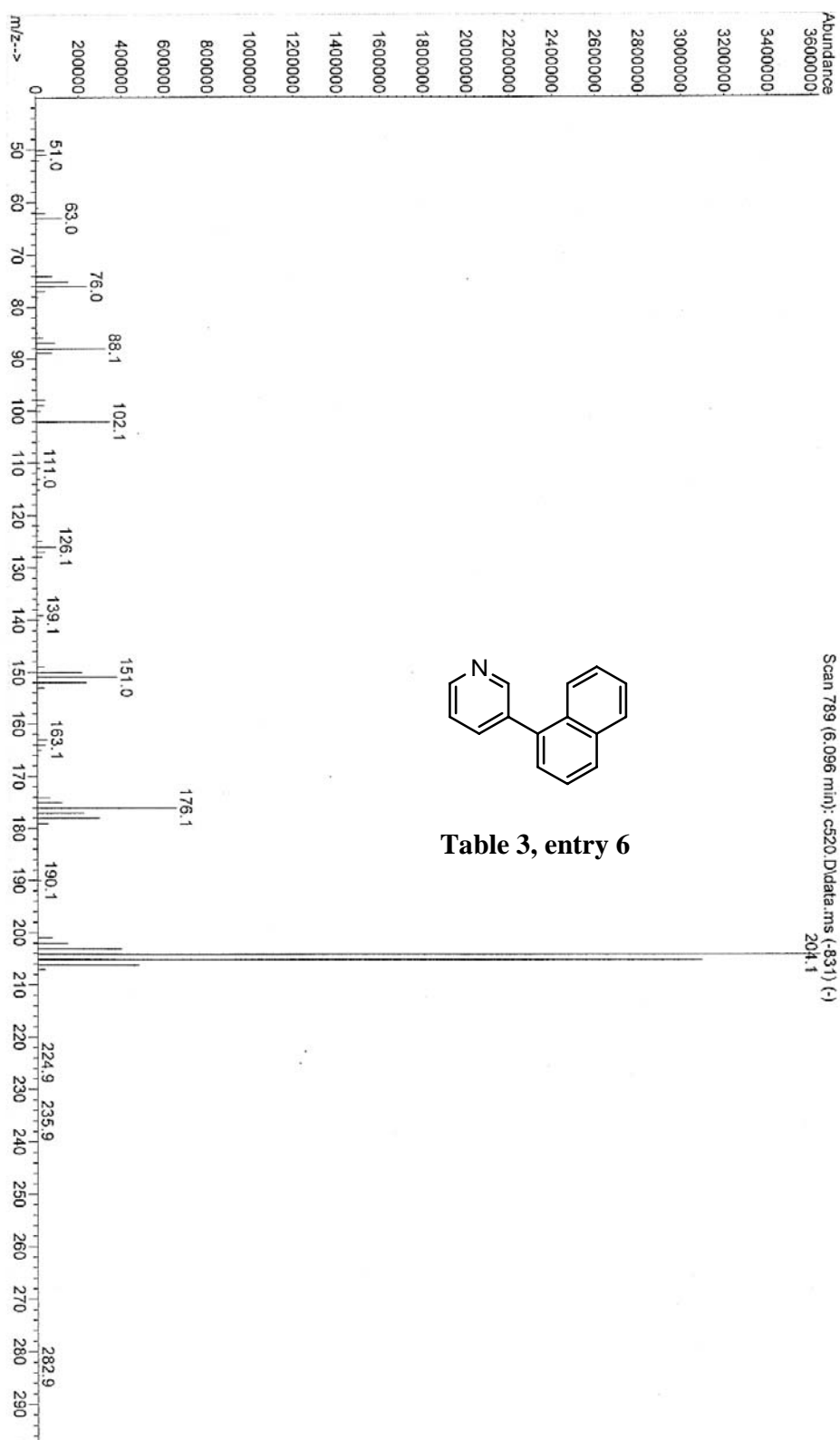
F2 - Processing parameters

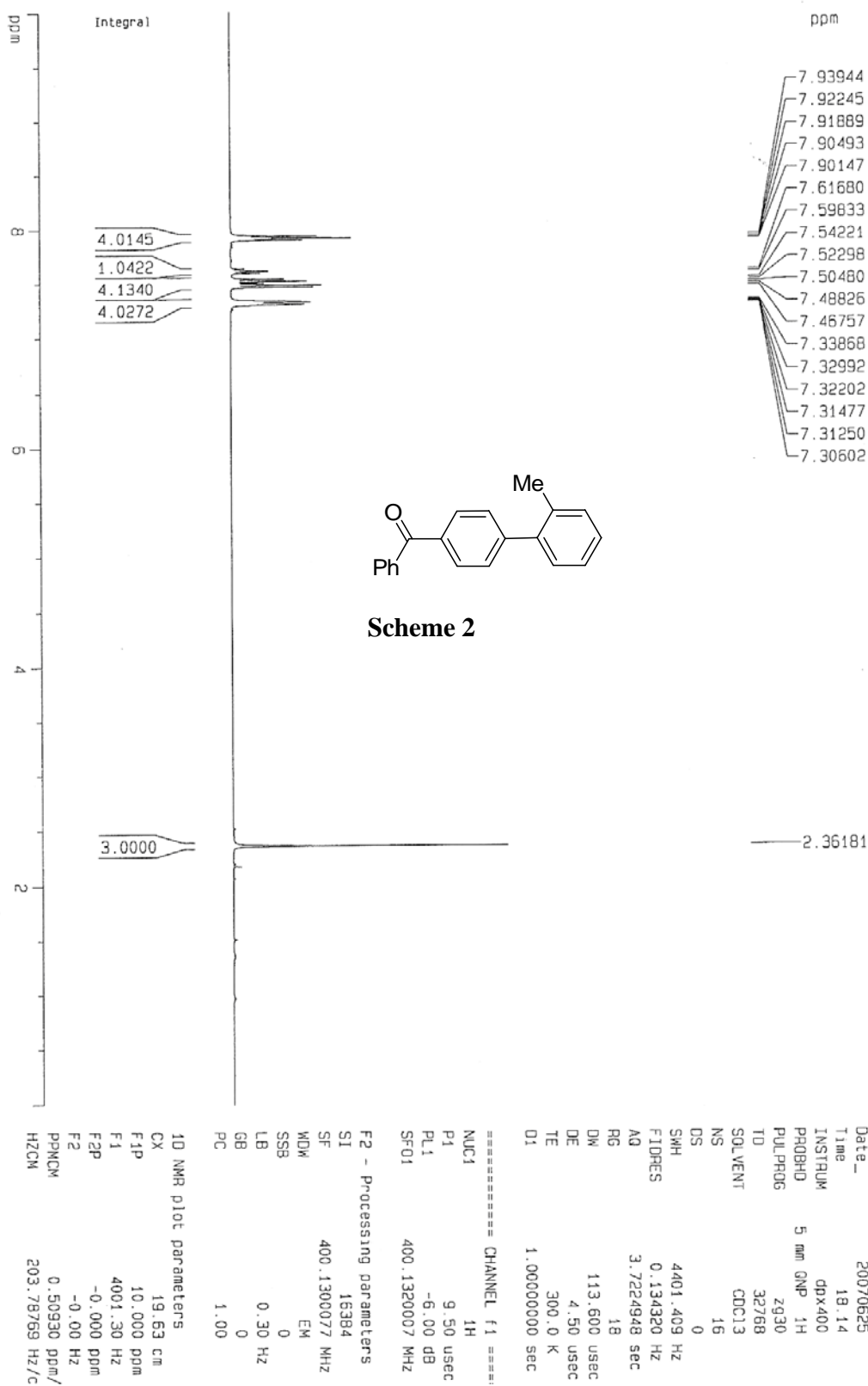
SI	65536
SF	100.6128116 MHz
WDW	EM
SSB	0
LB	3.50 Hz
GB	0
PC	1.00

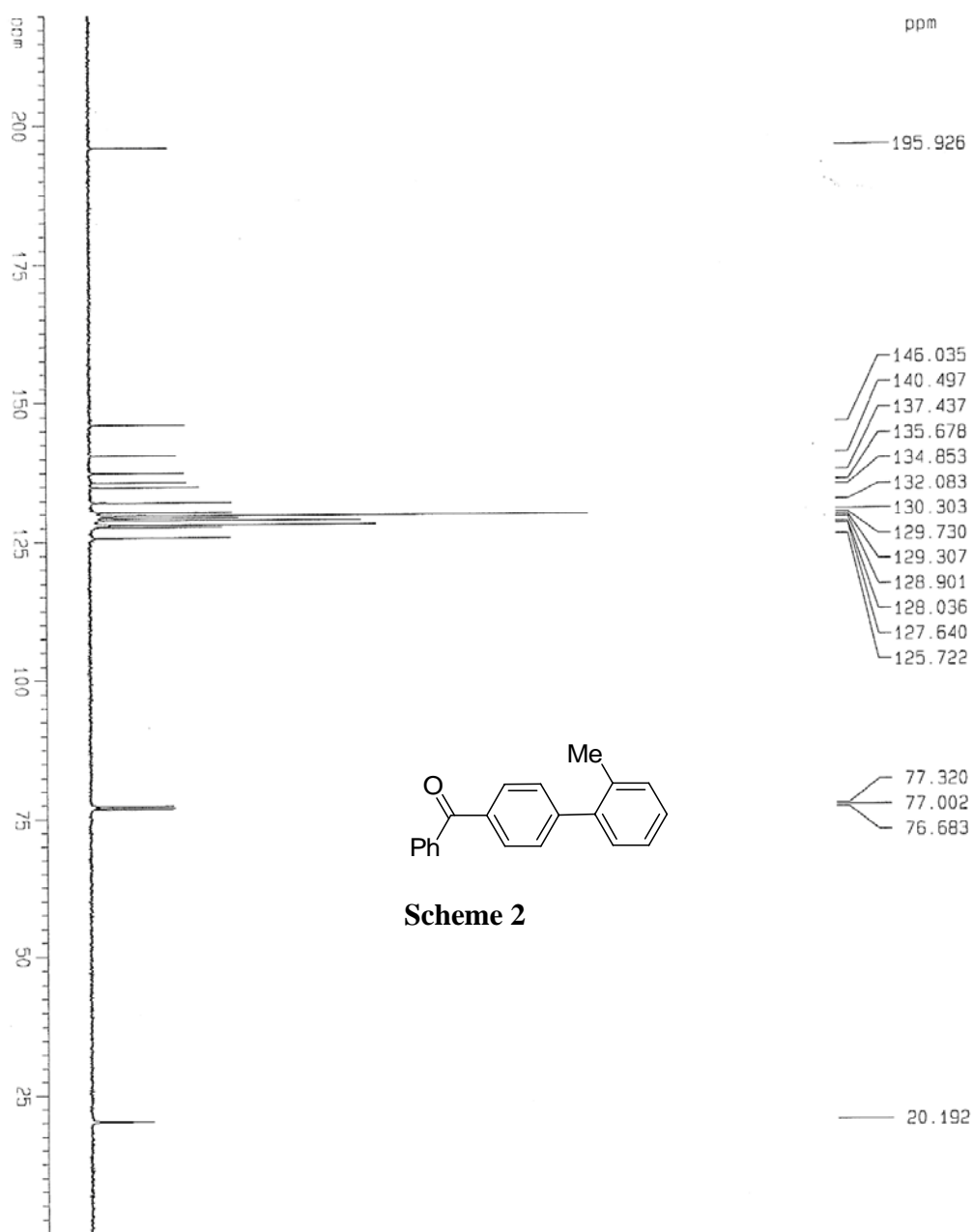
1D NMR plot parameters

CX	19.50 cm
F1P	200.000 ppm
F1	20122.56 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPACK	10.25641 ppm/cm
HZCM	1031.92527 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c520.D
Operator :
Acquired : 15 Nov 2007 13:14 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name:
Misc Info :
Vial Number: 1







Scheme 2

Current Data Parameters

NAME	Value
NAME	NAME
EXPO	44
PROCNO	1

F2 - Acquisition Parameters

Date_	Time	INSTRUM	PROBHD	PULPROG	TO	SOLVENT	NS	DS	SWH	FIDRES	RG	DW	DE	TE	D1	d11
20070525	18.16	CPX400	5 mm QNP 1H	zgpg30	131072	CDCl3	32	0	25125.629 Hz	0.191693 Hz	8192	19.900 usec	4.50 usec	300.0 K	3.00000000 sec	0.03000000 sec

===== CHANNEL f1 =====

NUC1	PL1	SFO1
13C	5.80 usec	100.6231263 MHz
PL1	-6.00 dB	

===== CHANNEL f2 =====

CPDPRG2	WALTZ16	NUC2	PL2	SFO2
waltz16		1H	71.00 usec	400.1326008 MHz
PL2			120.00 dB	
PL12			17.00 dB	

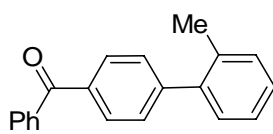
F2 - Processing parameters

SF	SI	WDW	SSB	LB	GB	PC
100.6126078 MHz	65536	EM	0	3.00 Hz	0	1.00

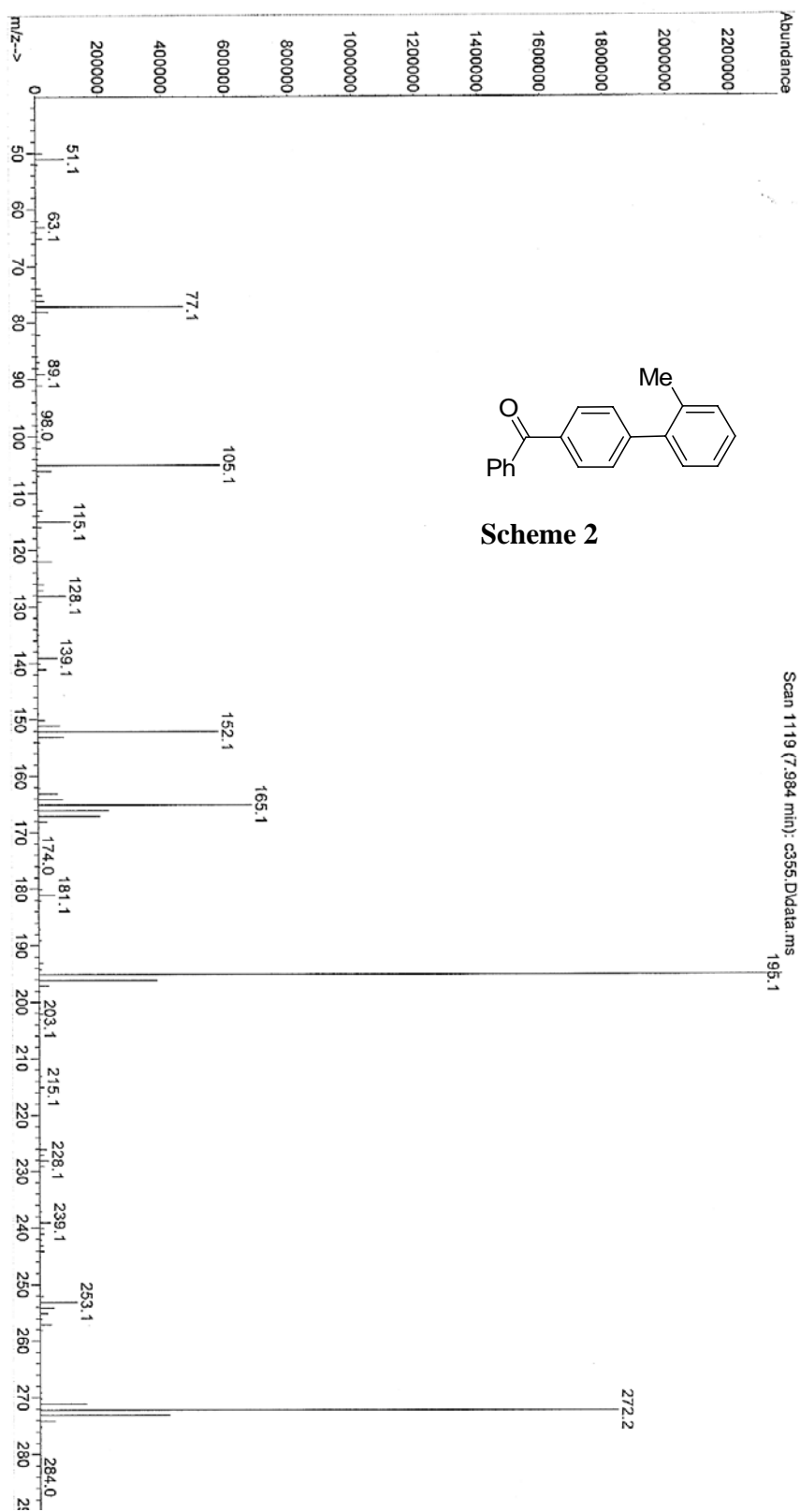
1D NMR plot parameters

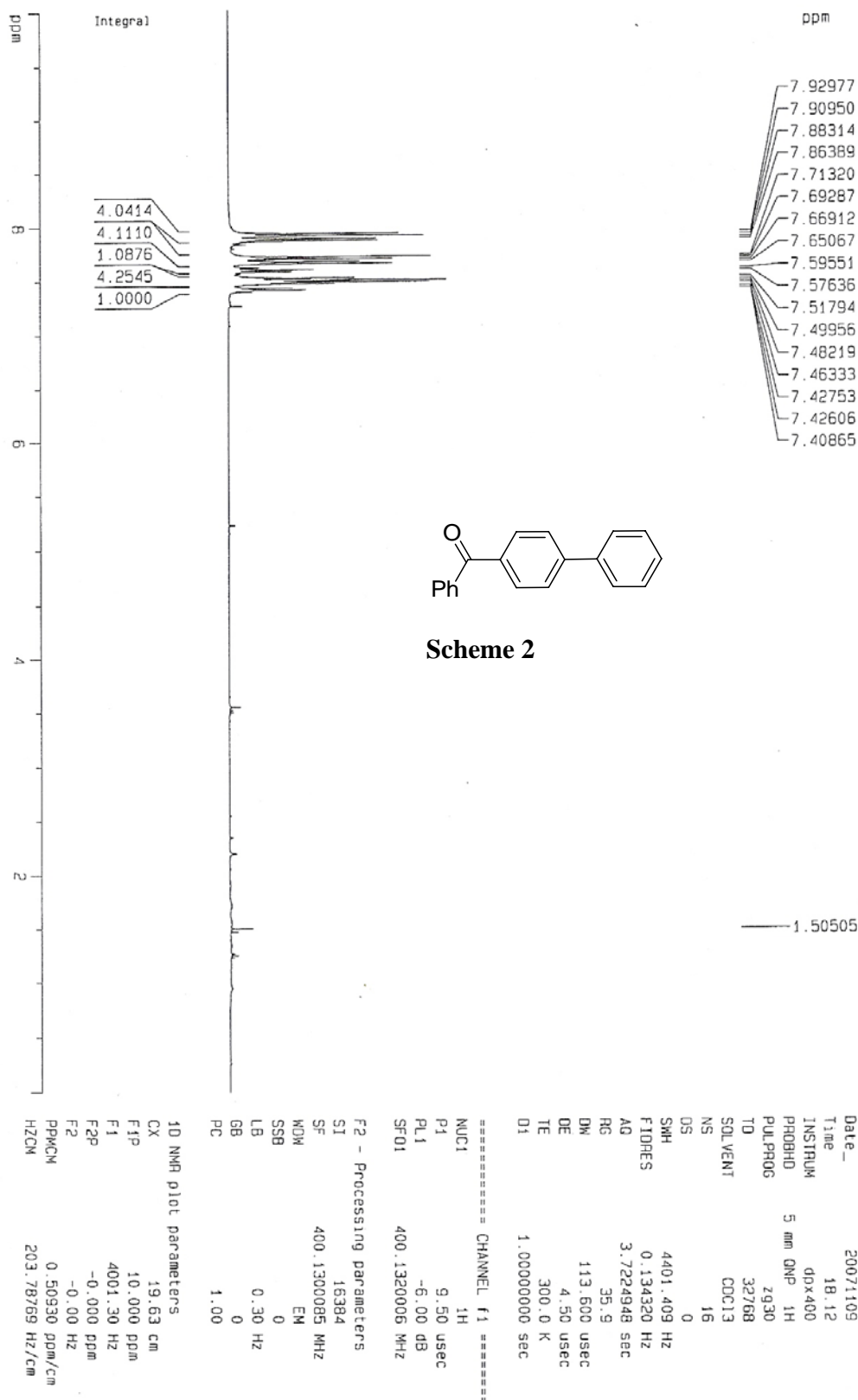
CK	F1P	F1	F2P	F2	PPMCH	HZCH
19.50 cm	220.000 ppm	22134.82 Hz	-0.000 ppm	-0.00 Hz	11.28205 ppm/	1135.11890 Hz/c

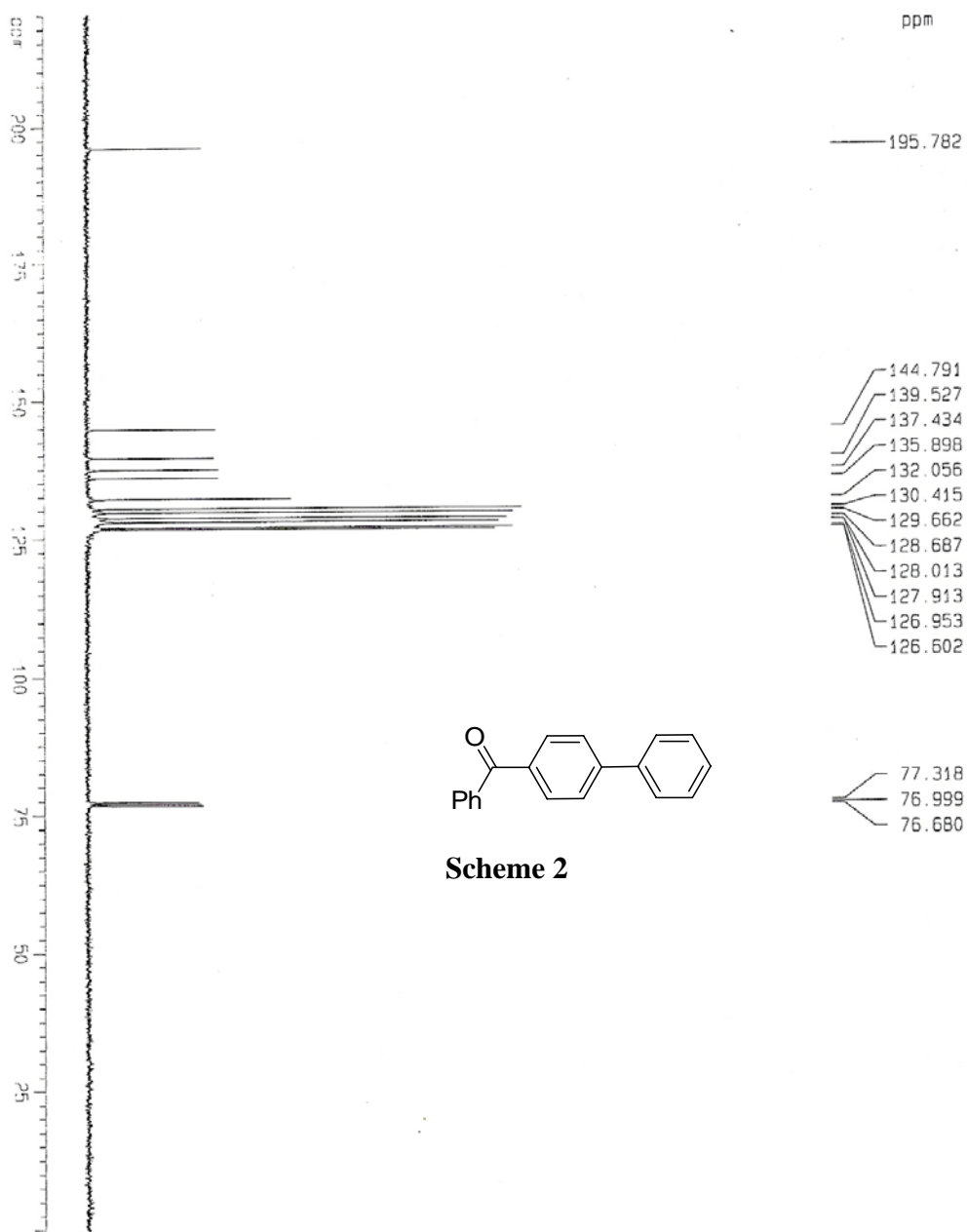
File : C:\msdchem\1\DATA\csmo\CT\c355.D
Operator :
Acquired : 22 Jun 2007 14:56 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name :
Misc Info :
Vial Number: 2



Scheme 2







Scheme 2

Current Data Parameters

NAME	Coupling
EXPNO	BB
PROCNO	1

F2 - Acquisition Parameters

Date_	Time
20071109	18.16

INSTRUM dpx400
PROBHD 5 mm QNP 1H
PULPROG zgpg30
TD 131072
SOLVENT CDCl3
NS 32
DS 0
SWH 25125.629 Hz
FIDRES 0.191693 Hz
AQ 2.6093827 sec
RG 8192
DM 19.900 usec
DE 4.50 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec

===== CHANNEL f1 =====

NUC1	13C
P1	5.80 usec
PL1	-5.00 dB
SFO1	100.6231263 MHz

===== CHANNEL f2 =====

OPPRG2	waltz16
NUC2	1H
PCPD2	71.00 usec
PL2	120.00 dB
PL12	17.00 dB
SFO2	400.1326008 MHz

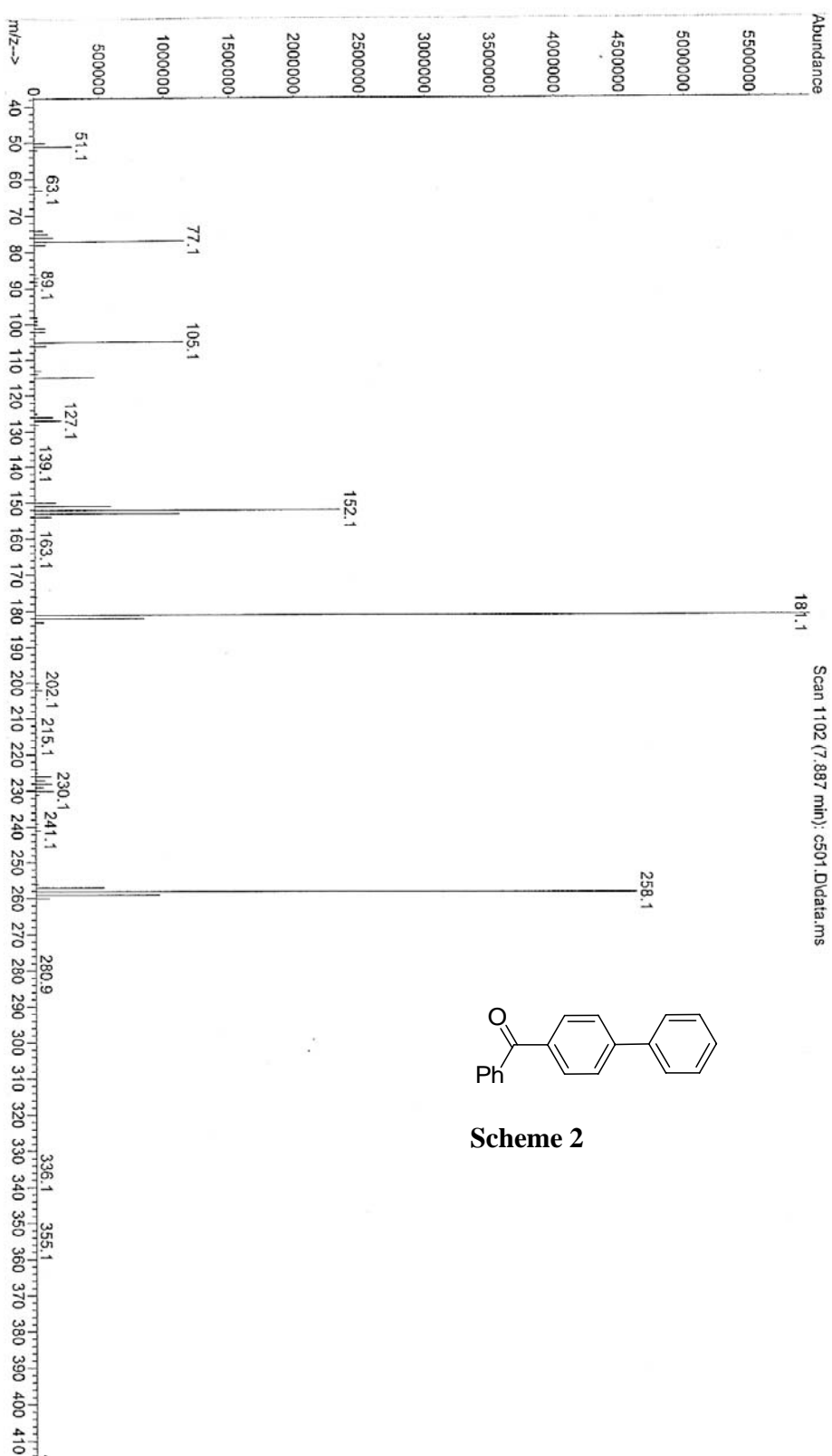
F2 - Processing Parameters

SF	65536
MW	EM
SSB	0
LB	3.00 Hz
GB	0
PC	1.00

1D NMR plot parameters

CK	19.50 cm
F1P	220.000 ppm
F1	22134.82 Hz
F2P	-0.000 ppm
F2	-0.00 Hz
PPMCM	11.28205 ppm/cm
HZCM	1135.11850 Hz/cm

File : C:\msdchem\1\DATA\cms0\CT\c501.D
Operator :
Acquired : 9 Nov 2007 16:00 using AcqMethod METHOD2.M
Instrument : 5973N
Sample Name: c501
Misc Info :
Vial Number: 1



Scheme 2

11. References.

- (1) Armarego, W. L. F.; Perrin, D. D. *Purification of Laboratory Chemicals.*, 4 Ed. ed., Butterworth-Heinemann: Oxford UK, **1996**.
- (2) Guo, M. J.; Varady, L.; Fokas, D.; Baldino, C.; Yu, L. B. *Tetrahedron Lett.* **2006**, 47, 3889.
- (3) Carnahan, J. C.; Closson, W. D.; Ganson, J. R.; Juckett, D. A.; Quaal, K. S. *J. Am. Chem. Soc.* **1976**, 98, 2526.
- (4) Percec, V.; Bae, J. Y.; Zhao, M. Y.; Hill, D. H. *J. Org. Chem.* **1995**, 60, 176.
- (5) Kaboudin, B. *Tetrahedron* **1999**, 55, 12865.
- (6) Bazile, Y.; Cointet, P. D.; Pigerol, C. *J. Het. Chem.* **1978**, 15, 859.
- (7) Cerfontain, H.; Ansink, H. R. W.; Coenjaarts, N. J.; Degraaf, E. J.; Koebergelder, A. *Recueil Des Travaux Chimiques Des Pays-Bas-Journal of the Royal Netherlands Chemical Society* **1989**, 108, 445.
- (8) Swain, N. A.; Brown, R. C. D.; Bruton, G. *J. Org. Chem.* **2004**, 69, 122.
- (9) Delgiudice, M. R.; Settimj, G.; Delfini, M. *Tetrahedron* **1984**, 40, 4067.
- (10) Tang, Z. Y.; Hu, Q. S. *J. Am. Chem. Soc.* **2004**, 126, 3058.
- (11) Moleele, S. S.; Michael, J. P.; de Koning, C. B. *Tetrahedron* **2006**, 62, 2831.
- (12) Zhang, H. C.; Kwong, F. Y.; Tian, Y.; Chan, K. S. *J. Org. Chem.* **1998**, 63, 6886.
- (13) Cho, C. H.; Yun, H. S.; Park, K. *J. Org. Chem.* **2003**, 68, 3017.
- (14) Wolfe, J. P.; Singer, R. A.; Yang, B. H.; Buchwald, S. L. *J. Am. Chem. Soc.* **1999**, 121, 9550.
- (15) Cooper, S. D.; Moseley, M. A.; Pellizzari, E. D. *Anal. Chem.* **1985**, 57, 2469.
- (16) Schultheiss, N.; Barnes, C. L.; Bosch, E. *Synth. Commun.* **2004**, 34, 1499.
- (17) Percec, V.; Golding, G. M.; Smidrkal, J.; Weichold, O. *J. Org. Chem.* **2004**, 69, 3447.
- (18) Miguez, J. M. A.; Adrio, L. A.; Sousa-Pedrares, A.; Vila, J. M.; Hii, K. K. *J. Org. Chem.* **2007**, 72, 7771.
- (19) Lacefield, W. B. M.; Winston, S., 7 pp. *Division of U.S. 3,928,604. CODEN: USXXAM US 3968251 19760706 Patent* **1976**.
- (20) Fedorova, O. A.; Andryukhina, E. N.; Gromov, S. P. *Synthesis* **2003**, 371.
- (21) Kataoka, N.; Shelby, Q.; Stambuli, J. P.; Hartwig, J. F., *J. Org. Chem.* **2002**, 67, 5553.

- (22) Wu, L.; Li, B. L.; Huang, Y. Y.; Zhou, H. F.; He, Y. M.; Fan, Q. H. *Org. Lett.* **2006**, 8, 3605.
- (23) Kataoka, N.; Shelby, Q.; Stambuli, J. P.; Hartwig, J. F., *J. Org. Chem.* **2002**, 67, 5553.
- (24) Nguyen, H. N.; Huang, X. H.; Buchwald, S. L. *J. Am. Chem. Soc.* **2003**, 125, 11818.
- (25) Shi, S. Y.; Zhang, Y. H. *J. Org. Chem.* **2007**, 72, 5927.
- (26) Sapountzis, I.; Lin, W. W.; Kofink, C. C.; Despotopoulou, C.; Knochel, P. *Angew. Chem. Int. Ed.* **2005**, 44, 1654.
- (27) Grabowsk, E.; Autrey, R. L. *Tetrahedron* **1969**, 25, 4315.
- (28) Leadbeater, N. E.; Smith, R. J. *Org. Lett.* **2006**, 8, 4588.
- (29) Cioffi, C. L.; Spencer, W. T.; Richards, J. J.; Herr, R. J. *J. Org. Chem.* **2004**, 69, 2210.