

Advanced
**Synthesis &
Catalysis**

Supporting Information

© Copyright Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, 2007

Supporting information

for

Substituted Benzocarbocycles by Palladium-Catalyzed Cascade Reactions Featuring a C(sp³)-H Activation Step

Julien Hitce, Olivier Baudoin*

ICBMS, Université Lyon 1
bât. CPE, 43 boulevard du 11 novembre 1918, Villeurbanne, F-69622, France

olivier.baudoin@univ-lyon1.fr

Table of Contents

I. Synthesis of the Dihalophenylacetonitrile Substrates

Characterization data for compounds 1b-d.

II. C(sp³)-H Activation / Heck Cyclization / Heck Arylation Sequence

Characterization data for compounds 5a-b, 5d-h.

III. C(sp³)-H Activation / Heck Reactions / Hydrogenation Sequences

Characterization data for compounds 6b-d.

Transmission electronic microscopy (TEM).

IV. Inhibition of Microtubule Assembly

V. Copies of NMR Spectra for target compounds 3a, 4a, 5a-h, 6a-d including NOESY spectra of compounds 5b, 5g, 6a, 6d.

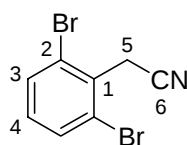
I. Preparation of the Dihalophenylacetonitrile Substrates^[1] (Scheme 2, Tables 1-3)

Representative procedure (Preparation of compound **1b**)

N-bromosuccinimide (4.04 g, 22.7 mmol) and benzoyl peroxide (191 mg, 0.79 mmol) were added to a stirred solution of 2,6-dibromotoluene (5.15 g, 20.6 mmol) in carbon tetrachloride (50 mL). The mixture was refluxed for 2 h. After cooling to r.t. it was filtered through celite and the solvent was removed under vacuum. The resulting white powder was dissolved in acetonitrile (100 mL). Trimethylsilyl cyanide (4.1 mL, 30.9 mmol) and tetrabutylammonium fluoride (31 mL of a 1 M solution in THF, 30.9 mmol) were carefully added at r.t. and the mixture was stirred for 2 h. The solvent was then removed under reduced pressure and the residue was purified by flash chromatography (silica gel, heptanes/diethyl ether 9:1) to afford 2,6-dibromophenylacetonitrile as a white solid (4.60 g, 81 % over 2 steps).

Lithium bis(trimethylsilyl)amide (67 mL of a 1 M solution in THF, 67.0 mmol) was added dropwise to a solution of 2,6-dibromophenylacetonitrile (4.60 g, 16.7 mmol) and DMPU^[2] (8.1 mL, 66.9 mmol) in THF (180 mL) under argon. After the reaction mixture had been stirred for 45 min at r.t., iodoethane (5.4 mL, 66.9 mmol) was added with a syringe and the reaction mixture was stirred for 2 h. The reaction was quenched with a saturated NH₄Cl solution (100 mL) and the aqueous layer was extracted with diethyl ether (150 mL × 3). The combined organic layers were washed with a 1 M HCl solution (150 mL) and with a saturated solution of NaHCO₃ (150 mL). The extracts were then dried over magnesium sulfate and evaporated under reduced pressure. The residue was purified by flash chromatography (silica gel, heptanes/ethyl acetate 95:5) to afford **1b** as a colourless oil (4.45 g, 80 %).

Products characterisation

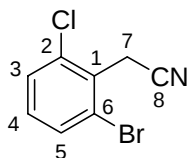


2,6-dibromophenylacetonitrile

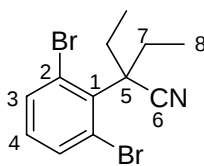
¹H NMR (300 MHz, CDCl₃) δ = 4.06 (s, 2H, H₅), 7.10 (t, *J* = 8.0 Hz, 1H, H₄), 7.60 (d, *J* = 8.0 Hz, 2H, H₃) ppm; ¹³C NMR (75.5 MHz, CDCl₃) δ = 25.9 (C₅), 115.4 (C₆), 125.3 (C₂), 130.2 (C₁), 131.1 (C₄), 132.6 (C₃) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 → 250 °C at 8 °C/min): 9.7 min (*m/z*: 274 [M⁺•]); IR (film) ν = 2257, 2974 cm⁻¹.

[1] a) Bromination: M. Tashiro, K. Nakayama, *OPPI Briefs* **1984**, 16, 379-381; b) Cyanation: E. D. Soli, A. S. Manoso, M. C. Patterson, P. DeShong, D. A. Favor, R. Hirschmann, A. B. Smith, III, *J. Org. Chem.* **1999**, 64, 3171-3177.

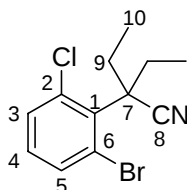
[2] DMPU: 1,3-dimethyl-3,4,5,6-tetrahydro-2(1*H*)-pyrimidinone.

**2-bromo-6-chlorophenylacetonitrile**

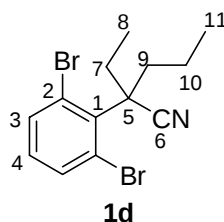
^1H NMR (500 MHz, CDCl_3) δ = 4.06 (s, 2H, H_7), 7.18 (dd, J = 8.2, 8.2 Hz, H_4), 7.42 (dd, J = 8.1, 1.2 Hz, H_3), 7.55 (dd, J = 7.9, 1.3 Hz, H_5) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 22.8 (C_7), 115.4 (C_8), 125.5 (C_6), 128.3 (C_1), 129.2 (C_3), 130.7 (C_4), 131.9 (C_5), 135.5 (C_2) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 8.1 min (m/z : 229 [$\text{M}^{+\bullet}$]); IR (film) ν = 2257, 2952 cm^{-1} .

**1b**

^1H NMR (300 MHz, CDCl_3) δ = 1.08 (dd, J = 7.4, 7.4 Hz, 6H, H_8), 2.05 (dq, J = 14.7, 7.4 Hz, 2H, H_{7a}), 2.95 (dq, J = 14.6, 7.3 Hz, 2H, H_{7b}), 6.93 (t, J = 7.5 Hz, 1H, H_4), 7.68 (d, J = 7.5 Hz, 2H, H_3) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.0 (C_8), 32.0 (C_7), 52.5 (C_5), 123.0 (C_6), 124.0 (C_2), 129.5 (C_4), 134.0 (C_1), 136.0 (C_3) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 13.0 min (m/z : 329 [$\text{M}^{+\bullet}$]); IR (film) ν = 1458, 1576, 2225, 2970 cm^{-1} .

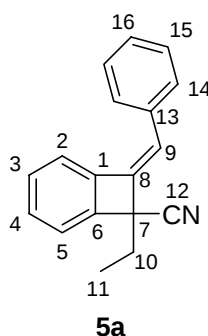
**1c**

^1H NMR (300 MHz, CDCl_3) δ = 1.06 (dd, J = 7.7, 7.7 Hz, 6H, H_{10}), 2.09 (dq, J = 14.7, 7.4 Hz, 2H, H_{9a}), 2.88 (dq, J = 14.6, 7.3 Hz, 2H, H_{9b}), 7.03 (dd, J = 8.0, 8.0 Hz, 1H, H_4), 7.37 (dd, J = 8.0, 1.5 Hz, 1H, H_3), 7.58 (dd, J = 8.0, 1.5 Hz, 1H, H_5) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.0 (C_{10}), 32.3 (C_9), 51.5 (C_7), 123.2 (C_8), 123.7 (C_6), 129.2 (C_4), 132.0 (C_3), 132.9 (C_1), 135.4 (C_2 , C_5) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 12.0 min (m/z : 285 [$\text{M}^{+\bullet}$]); IR (film) ν = 1458, 1573, 2225, 2970 cm^{-1} .

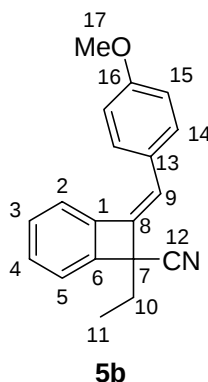


^1H NMR (500 MHz, CDCl_3) δ = 0.97 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 1.08 (dd, J = 7.4, 7.4 Hz, 3H, H_8), 1.30-1.40 (m, 1H, H_{10a}), 1.57-1.68 (m, 1H, H_{10b}), 1.95-2.04 (m, 1H, H_{9a}), 2.10 (dq, J = 14.6, 7.5 Hz, 1H, H_{7a}), 2.85-2.98 (m, 2H, H_{7b} , H_{9b}), 6.93 (t, J = 7.9 Hz, 1H, H_4), 7.63 (d, J = 8.2 Hz, 2H, H_3) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.0 (C_8), 14.0 (C_{11}), 18.9 (C_{10}), 32.2 (C_7), 40.7 (C_9), 51.7 (C_5), 123.2 (C_6), 123.8 (C_2), 129.4 (C_4), 134.2 (C_1), 135.9 (C_3) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 13.8 min (m/z : 343 [$\text{M}^{+\bullet}$]); IR (film) ν = 1464, 2224, 2961 cm^{-1} .

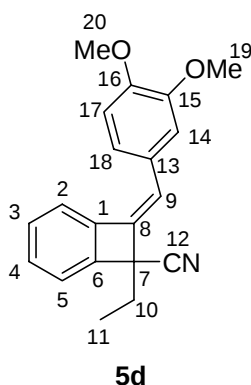
II. $\text{C}(\text{sp}^3)\text{-H}$ Activation / Heck Cyclization / Heck Arylation Process (Table 2)



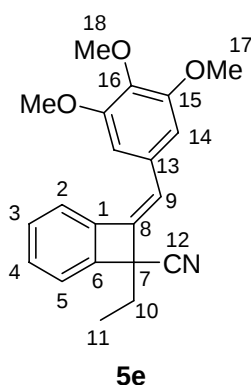
^1H NMR (500 MHz, CDCl_3) δ = 1.23 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.04 (dq, J = 14.5, 7.4, 1H, H_{10a}), 2.26 (dq, J = 14.5, 7.3 Hz, 1H, H_{10b}), 6.54 (s, 1H, H_9), 7.29-7.46 (m, 5H, H_3 , H_4 , H_{15} , H_{16} and H_2 or H_5), 7.51 (d, J = 7.4 Hz, 1H, H_2 or H_5), 7.56 (d, J = 7.9 Hz, 2H, H_{14}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.6 (C_{11}), 30.0 (C_{10}), 51.1 (C_7), 120.8 (C_{12}), 121.8, 121.9 (C_{16} , CH_{arom}), 122.2 (C_9), 128.0 (C_{14}), 128.1 (CH_{arom}), 128.7 (C_{15}), 130.0, 130.1 (CH_{arom}), 135.7 (C_1), 138.8 (C_8), 142.6 (C_{13}), 145.2 (C_6) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 15.3 min (m/z : 245 [$\text{M}^{+\bullet}$]); IR (film) ν = 1459, 1599, 2229, 2968 cm^{-1} .



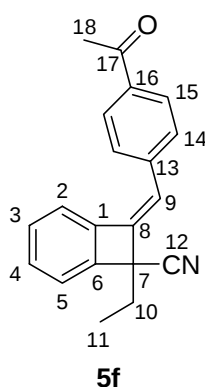
^1H NMR (500 MHz, CDCl_3) δ = 1.23 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.03 (dq, J = 14.8, 7.4 Hz, 1H, H_{10a}), 2.25 (dq, J = 14.7, 7.4 Hz, 1H, H_{10b}), 3.86 (s, 3H, H_{17}), 6.48 (s, 1H, H_9), 6.97 (d, J = 8.7 Hz, 2H, H_{15}), 7.32-7.41 (m, 3H, H_{arom}), 7.49-7.52 (m, 3H, H_{14} , H_{arom}) ppm; ^{13}C NMR (125.8 MHz, CDCl_3) δ = 10.5 (C_{11}), 30.1 (C_{10}), 51.0 (C_7), 55.4 (C_{17}), 114.2 (C_{15}), 121.0 (C_{12}), 121.6, 121.8 (CH_{arom}), 121.9 (C_9), 128.4 (C_1), 129.4 (C_{14}), 129.7, 129.9 (CH_{arom}), 136.9 (C_8), 142.8 (C_{13}), 145.1 (C_6), 159.6 (C_{16}) ppm; HRMS (ESI, MeOH + CH_2Cl_2) calcd for $\text{C}_{19}\text{H}_{17}\text{NONa}$ [$(\text{M} + \text{Na})^+$]: 298.1208; found: 298.1194; IR (film) ν = 1247, 1509, 1605, 2228, 2967 cm^{-1} .



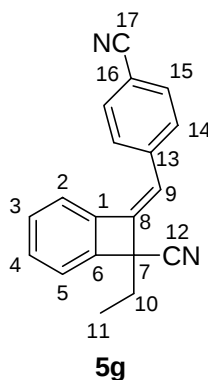
^1H NMR (300 MHz, CDCl_3) δ = 1.24 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.03 (dq, J = 14.3, 7.3 Hz, 1H, H_{10a}), 2.26 (dq, J = 14.3, 7.4 Hz, 1H, H_{10b}), 3.93 (s, 3H, H_{19} or H_{20}), 3.96 (s, 3H, H_{19} or H_{20}), 6.48 (s, 1H, H_9), 6.94 (d, J = 8.3 Hz, 1H, H_{17}), 7.10 (d, J = 2.0 Hz, 1H, H_{14}), 7.14 (dd, J = 8.3 Hz, 2.0 Hz, 1H, H_{18}), 7.32-7.42 (m, 3H, H_{arom}), 7.49-7.53 (m, 1H, H_{arom}); ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.4 (C_{11}), 30.0 (C_{10}), 51.0 (C_7), 56.0, 56.1 (C_{19} , C_{20}), 110.9 (C_{14}), 111.2 (C_{17}), 120.9 (C_{12}), 121.2 (C_{18}), 121.4, 121.9 (CH_{arom}), 122.1 (C_9), 128.6 (C_1), 129.8, 129.9 (CH_{arom}), 137.0 (C_8), 142.7 (C_{13}), 145.1 (C_6), 149.0, 149.1 (C_{15} , C_{16}) ppm; HRMS (ESI, MeOH + CH_2Cl_2) calcd for $\text{C}_{20}\text{H}_{19}\text{NO}_2\text{Na}$ [$(\text{M} + \text{Na})^+$]: 328.1313; found: 328.1294; IR (film) ν = 1236, 1258, 1512, 1599, 2227, 2966 cm^{-1} .



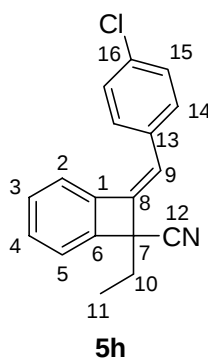
^1H NMR (500 MHz, CDCl_3) δ = 1.24 (dd, J = 7.6, 7.6 Hz, 3H, H_{11}), 2.04 (dq, J = 14.6, 7.4 Hz, 1H, H_{10a}), 2.26 (dq, J = 14.3, 7.3 Hz, 1H, H_{10b}), 3.90 (s, 3H, H_{18}), 3.93 (s, 6H, H_{17}), 6.48 (s, 1H, H_9), 6.80 (s, 1H, H_{14}), 7.34-7.43 (m, 3H, H_{3-5}), 7.52 (d, J = 7.4 Hz, 1H, H_2) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.5 (C_{11}), 30.1 (C_{10}), 51.1 (C_7), 56.3 (C_{17}), 61.0 (C_{18}), 105.4 (C_{14}), 120.7 (C_{12}), 121.5 (C_2), 122.0 (CH_{arom}), 122.2 (C_9), 130.0, 130.1 (CH_{arom}), 131.3 (C_{13}), 138.1 (C_8), 138.3 (C_{16}), 142.5 (C_1), 145.3 (C_6), 153.5 (C_{15}) ppm; HRMS (ESI, MeOH + CH_2Cl_2) calcd for $\text{C}_{21}\text{H}_{21}\text{NO}_3\text{Na}$ $[(\text{M} + \text{Na})^+]$: 358.1419; found: 358.1382; IR (film) ν = 1520, 1604, 2228, 2967 cm^{-1} .



^1H NMR (300 MHz, CDCl_3) δ = 1.24 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.06 (dq, J = 14.5, 7.3 Hz, 1H, H_{10a}), 2.28 (dq, J = 14.5, 7.2 Hz, 1H, H_{10b}), 2.63 (s, 3H, H_{18}), 6.57 (s, 1H, H_9), 7.37-7.46 (m, 3H, H_{arom}), 7.49-7.55 (m, 1H, H_{arom}), 7.65 (d, J = 8.2 Hz, 2H, H_{14}), 8.03 (d, J = 8.4 Hz, 2H, H_{15}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.4 (C_{11}), 26.6 (C_{18}), 30.0 (C_{10}), 51.3 (C_7), 120.5 (C_{12}), 121.0 (C_9), 122.1 ($2 \times \text{CH}_{\text{arom}}$), 128.0 (C_{14}), 128.8 (C_{15}), 130.2, 130.9 (CH_{arom}), 136.3 (C_{16}), 140.5 (C_{13}), 141.1 (C_8), 142.0 (C_1), 145.5 (C_6), 197.4 (C_{17}) ppm; HRMS (ESI, MeOH + CH_2Cl_2) calcd for $\text{C}_{20}\text{H}_{17}\text{NONa}$ $[(\text{M} + \text{Na})^+]$: 310.1208; found: 310.1178; IR (film) ν = 1600, 1678, 2230, 2968 cm^{-1} .



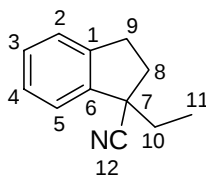
^1H NMR (500 MHz, CDCl_3) δ = 1.24 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.05 (dq, J = 14.7, 7.4 Hz, 1H, H_{10a}), 2.28 (dq, J = 14.7, 7.3 Hz, 1H, H_{10b}), 6.54 (s, 1H, H_9), 7.38-7.51 (m, 4H, H_{2-5}), 7.65 (d, J = 8.5 Hz, 2H, H_{14}), 7.72 (d, J = 8.3 Hz, 2H, H_{15}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.3 (C_{11}), 30.0 (C_{10}), 51.4 (C_7), 111.3 (C_{16}), 118.7 (C_{17}), 120.1 (C_9), 120.2 (C_{12}), 122.1, 122.2 (CH_{arom}), 128.4 (C_{14}), 130.3, 131.3 (CH_{arom}), 132.5 (C_{15}), 140.3 (C_{13}), 141.6 (C_1), 142.1 (C_8), 145.6 (C_6) ppm; GC/MS (CI (NH_3), DB-5ms column, 1 min at 150 $^\circ\text{C}$ then 150 $\rightarrow$ 280 $^\circ\text{C}$ at 8 $^\circ\text{C}/\text{min}$): 15.6 min (m/z : 288 [$(\text{M} + \text{NH}_4)^+$]); IR (film) ν = 1603, 2226, 2969 cm^{-1} .



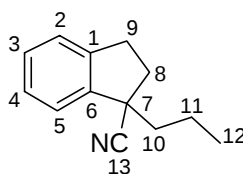
^1H NMR (300 MHz, CDCl_3) δ = 1.22 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.02 (dq, J = 15.0, 7.4 Hz, 1H, H_{10a}), 2.25 (dq, J = 14.8 Hz, 7.4 Hz, 1H, H_{10b}), 6.48 (s, 1H, H_9), 7.35-7.50 (m, 8H, H_{arom}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 10.4 (C_{11}), 30.0 (C_{10}), 51.2 (C_7), 120.6 (C_{12}), 120.9 (C_9), 121.8, 122.0 (CH_{arom}), 128.9 (C_{14}), 129.2 (C_{15}), 130.1, 130.4 (CH_{arom}), 133.8, 134.2 (C_1 , C_{16}), 139.4 (C_8), 142.2 (C_{13}), 145.3 (C_6) ppm; GC/MS (EI, DB-5ms column, 1 min at 150 $^\circ\text{C}$ then 150 $\rightarrow$ 280 $^\circ\text{C}$ at 8 $^\circ\text{C}/\text{min}$): 14.0 min (m/z : 279 [$\text{M}^{+\bullet}$]); IR (film) ν = 1092, 1592, 2230, 2968 cm^{-1} .

III. $\text{C}(\text{sp}^3)\text{-H}$ Activation / Heck Reactions / Hydrogenation One-Pot Sequences (Table 3)

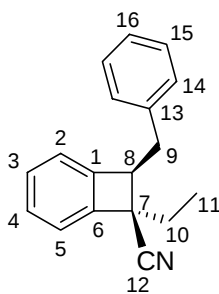
Products characterization

**6b**

^1H NMR (300 MHz, CDCl_3) δ = 1.15 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 1.79 (dq, J = 14.7, 7.4 Hz, 1H, H_{10a}), 2.01 (dq, J = 14.9, 7.4 Hz, 1H, H_{10b}), 2.26 (ddd, J = 13.0, 7.6, 5.5 Hz, 1H, H_{8a}), 2.61 (ddd, J = 13.0, 8.6, 7.2 Hz, 1H, H_{8b}), 2.94-3.14 (m, 2H, H_9), 7.25-7.30 (m, 3H, H_{arom}), 7.36-7.42 (m, 1H, H_{arom}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 9.8 (C_{11}), 30.2 (C_9), 31.7 (C_{10}), 36.9 (C_8), 47.9 (C_7), 123.2 (C_{12}), 123.8, 125.1, 127.1, 128.6 (C_{2-5}), 142.3, 142.5 (C_1 , C_6) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 6.6 min (m/z : 171 [$\text{M}^{+\bullet}$]); IR (film) ν = 1458, 2232, 2969 cm^{-1} .

**6c**

^1H NMR (500 MHz, CDCl_3) δ = 0.97 (dd, J = 7.2, 7.2 Hz, 3H, H_{12}), 1.50-1.71 (m, 3H, H_{10a} , H_{11}), 1.91 (ddd, J = 18.0, 12.5, 4.3 Hz, 1H, H_{10b}), 2.24 (ddd, J = 13.0, 7.9, 5.4 Hz, 1H, H_{8a}), 2.59 (ddd, J = 12.9, 8.6, 7.2 Hz, 1H, H_{8b}), 2.95-3.09 (m, 2H, H_9), 7.23-7.30 (m, 3H, H_{arom}), 7.34-7.40 (m, 1H, H_{arom}) ppm; ^{13}C NMR (75.5 MHz, CDCl_3) δ = 14.0 (C_{12}), 18.9 (C_{11}), 30.2 (C_9), 37.5 (C_8), 40.8 (C_{10}), 47.2 (C_7), 123.4 (C_{13}), 123.8, 125.1, 127.1, 128.5 (C_{2-5}), 142.4, 142.5 (C_1 , C_6) ppm; GC/MS (EI, DB-5ms column, 1 min at 120 °C then 120 $\rightarrow$ 250 °C at 8 °C/min): 7.7 min (m/z : 185 [$\text{M}^{+\bullet}$]); IR (film) ν = 1458, 2232, 2960 cm^{-1} .

**6d**

^1H NMR (500 MHz, CDCl_3) δ = 1.17 (dd, J = 7.4, 7.4 Hz, 3H, H_{11}), 2.00 (q, J = 7.4 Hz, 2H, H_{10}), 3.09 (dd, J = 13.7, 10.1 Hz, 1H, H_{9a}), 3.35 (dd, J = 13.8, 6.6 Hz, 1H, H_{9b}), 3.65 (dd, J = 10.1, 6.6 Hz, 1H, H_8), 6.79 (d, J = 6.3 Hz, 1H, H_2), 7.21-7.39 (m, 8H, H_{arom}) ppm; ^{13}C NMR (125.8 MHz, CDCl_3) δ = 10.7 (C_{11}), 30.8 (C_{10}), 38.1 (C_9), 50.0 (C_7), 54.7 (C_8), 120.5 (C_{12}), 121.9 (CH_{arom}), 123.5

(C₂), 126.7, 128.5 (CH_{arom}), 128.6 (C₁₅), 129.1 (C₁₄), 129.2 (CH_{arom}), 138.9 (C₁₃), 142.3 (C₆), 145.7 (C₁) ppm; GC/MS (CI (NH₃), DB-5ms column, 1 min at 120 °C then 120 → 250 °C at 8 °C/min): 13.7 min (m/z: 265 [(M + NH₄)⁺]); IR (film) ν = 1455, 1496, 2230, 2966 cm⁻¹.

Transmission Electronic Microscopy

Electron microscopy observations were performed with a scanning transmission electron microscope STEM VG HB501. Particle size distribution was measured at 100% substrate conversion (for the reaction **1a** → **1b**, see Scheme 1) from 8 different images, each corresponding to a different sample zone, using the ImageJ 1.33 analysis program from the National Institutes of Health, USA (<http://rsb.info.nih.gov/ij>). The average particle diameter was 3.1 ± 0.2 nm.

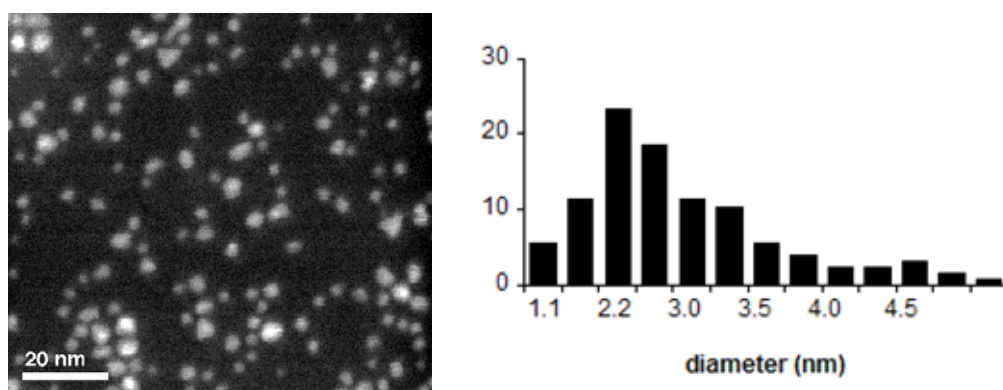
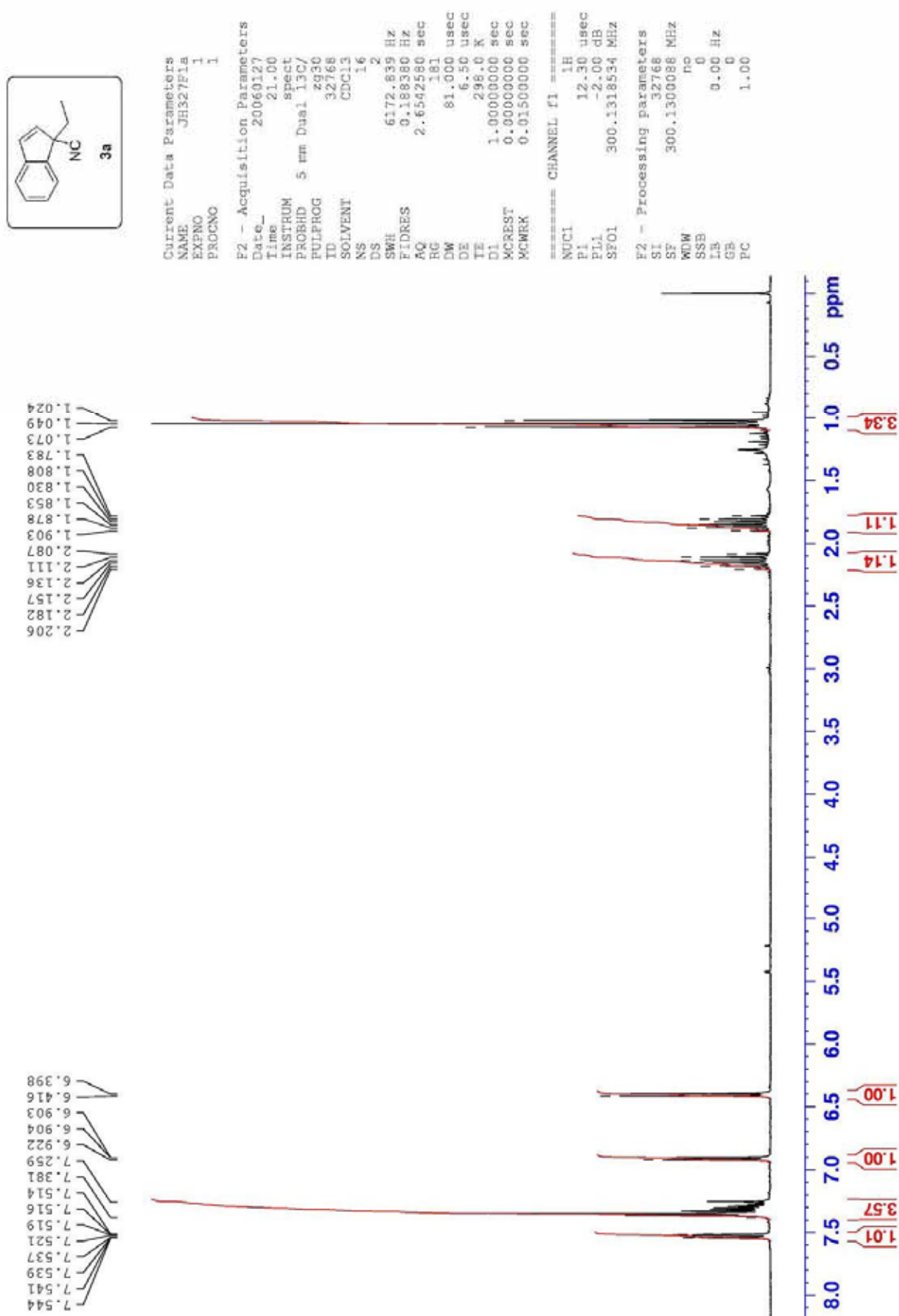
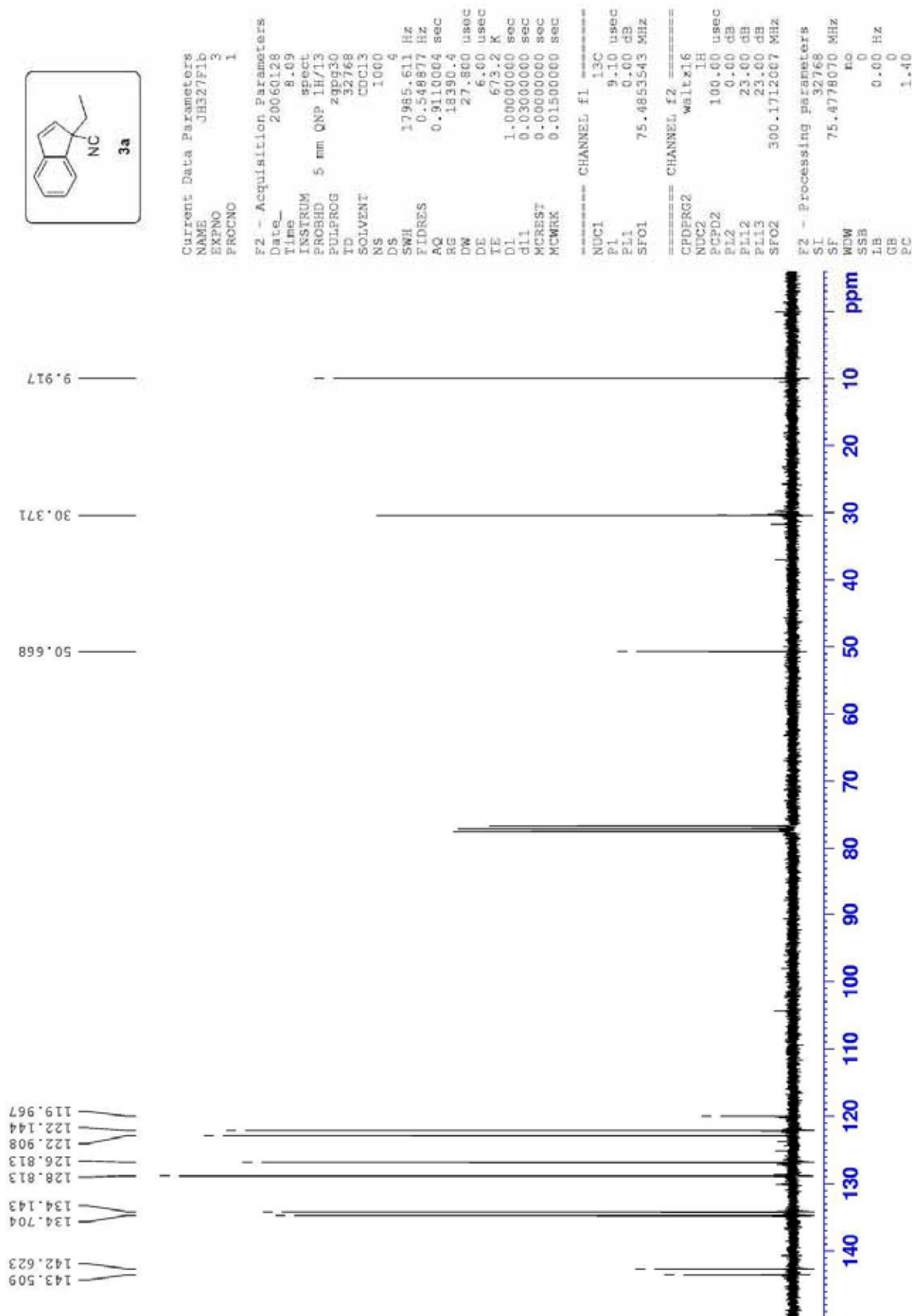


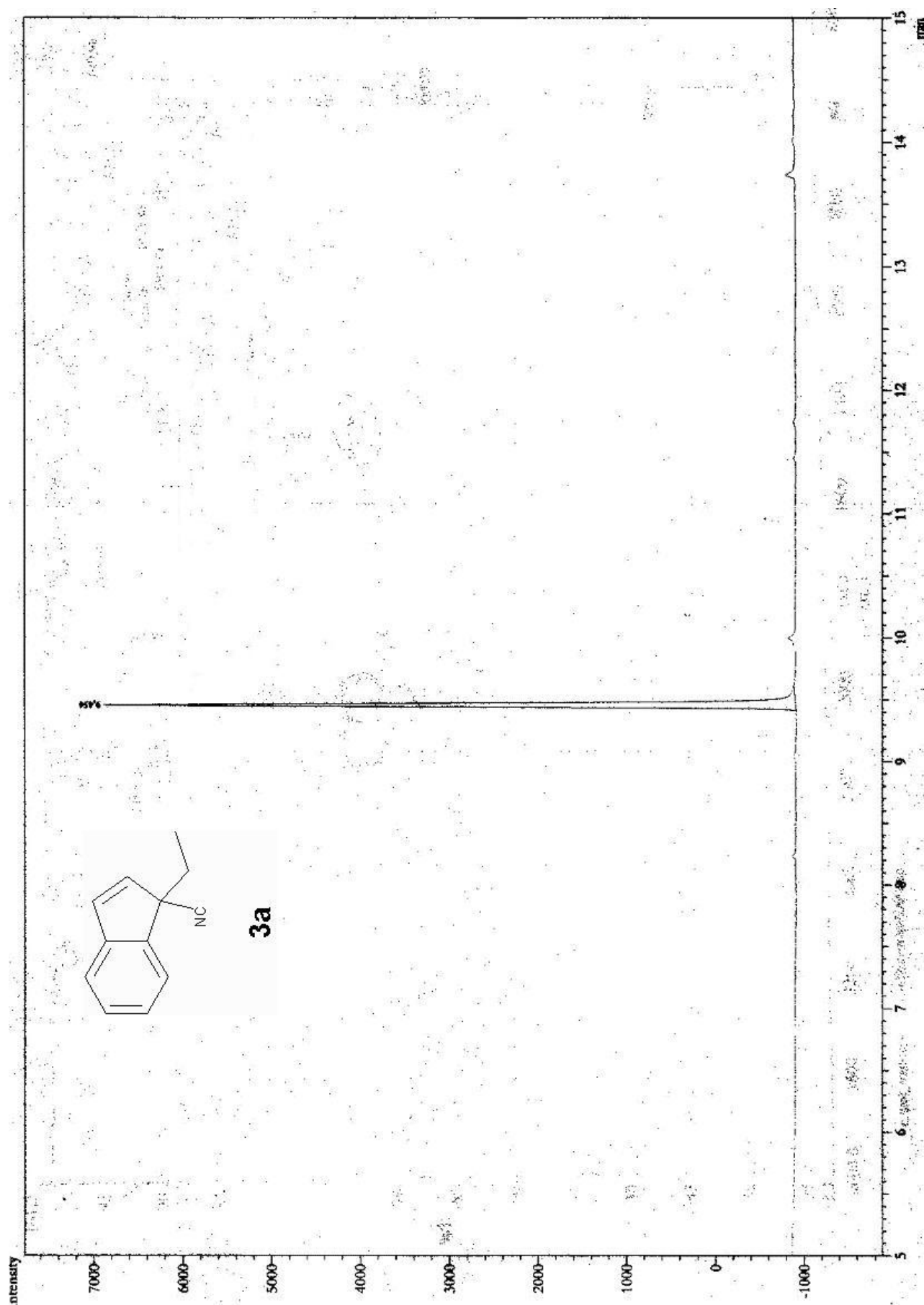
Figure S1. TEM Image and Size Distribution of Palladium Nanoparticles Formed During the Reaction **1a** → **1b** (100 % conversion)

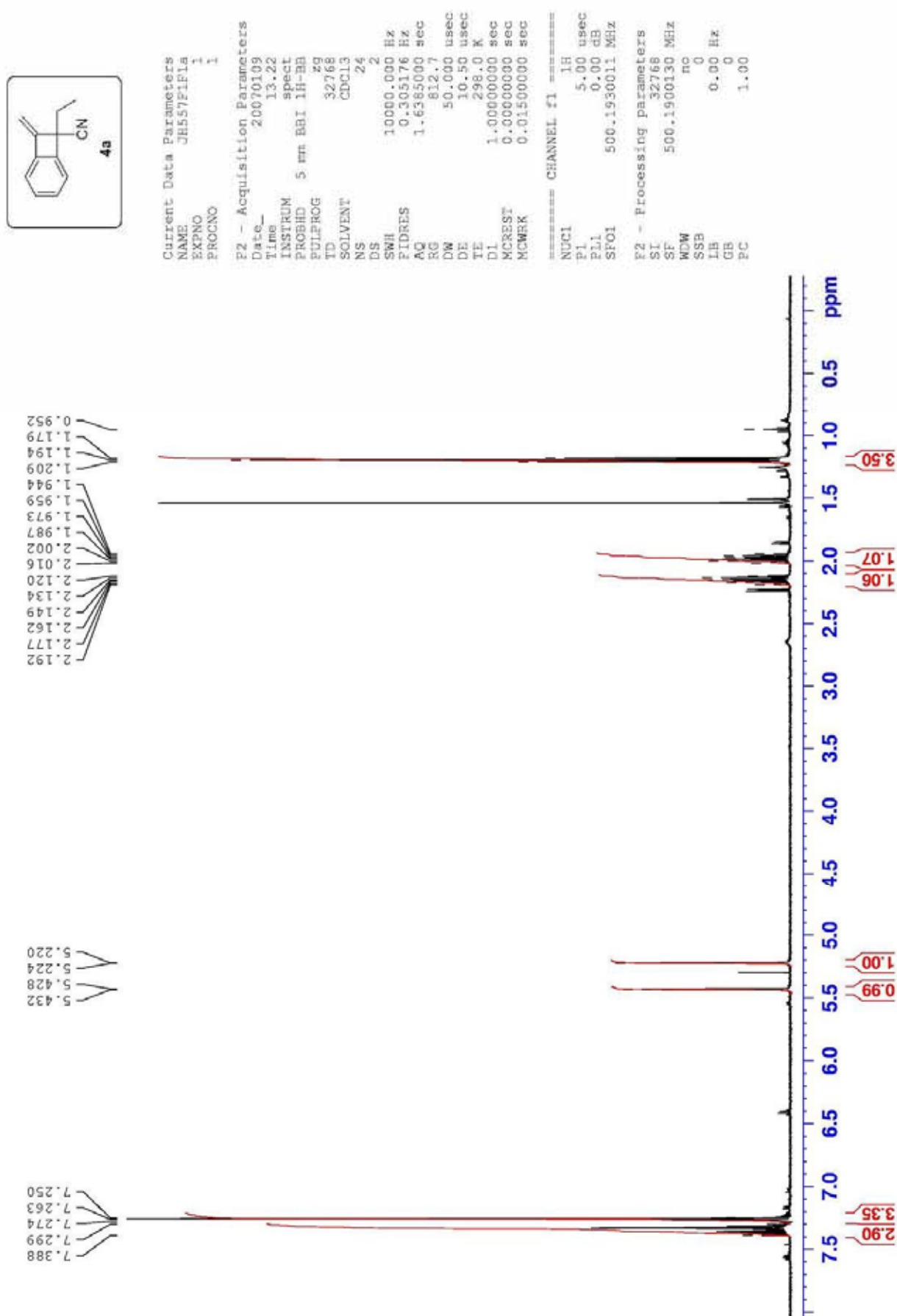
IV. Inhibition of Microtubule Assembly (Figure 1)

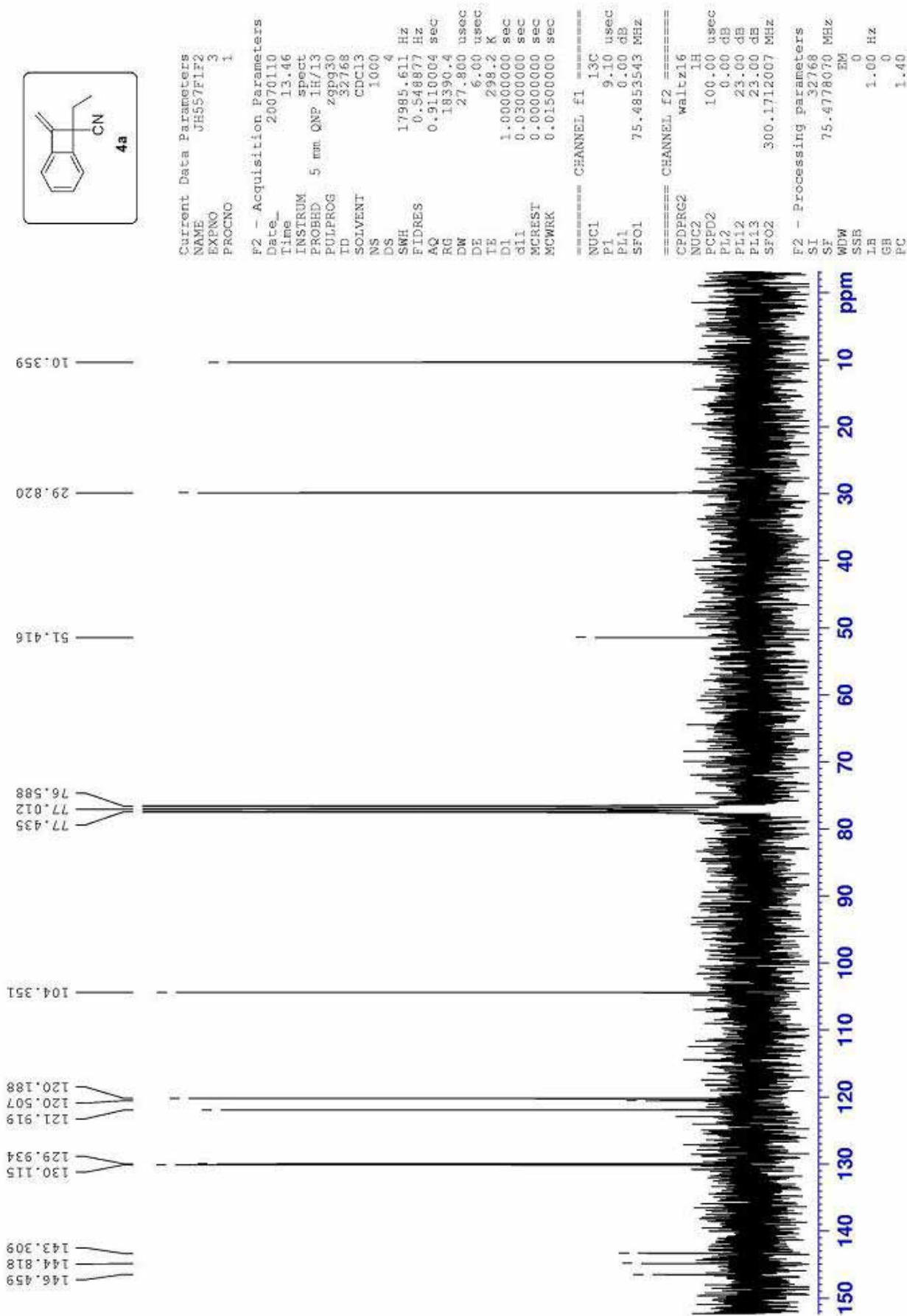
The drug, dissolved in DMSO at different concentrations, was pre-incubated with a solution of tubulin at 37 °C for 10 min, then the solution was cooled to 0 °C for 5 min in order to achieve complete tubulin depolymerization. The solution was then placed in a temperature-controlled cell at 37 °C (microtubule assembly) and the increase of the optical density was monitored in a UV spectrophotometer at 350 nm for 1 min. The maximum rate of assembly was recorded and compared to a sample without drug. The IC₅₀ is the concentration of compound required to inhibit 50 % of the rate of microtubule assembly. It was calculated from the effect of several concentrations and compared to the IC₅₀ of colchicine obtained within the same day with the same tubulin preparation. Reported values are averages of three experiments.

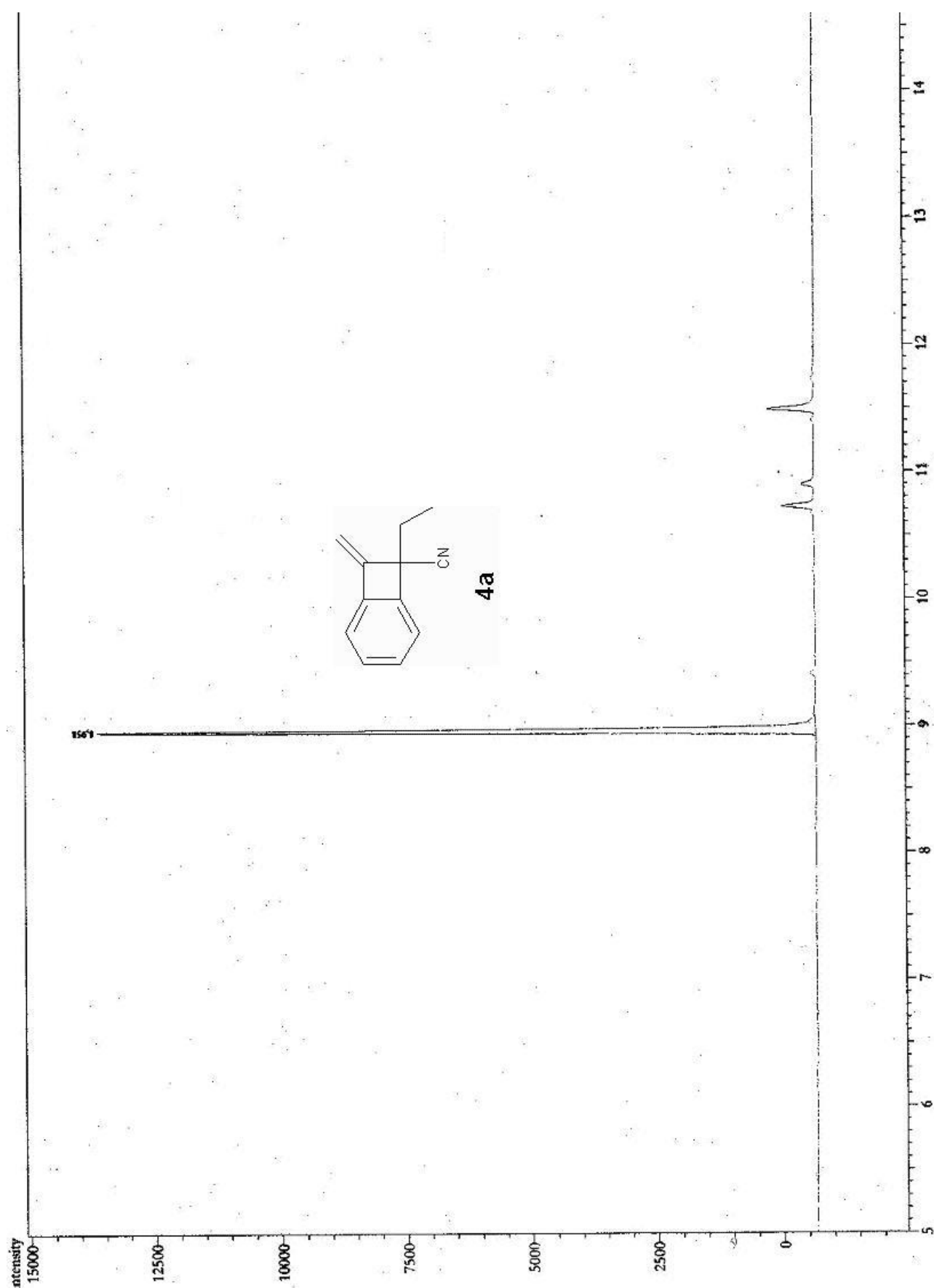


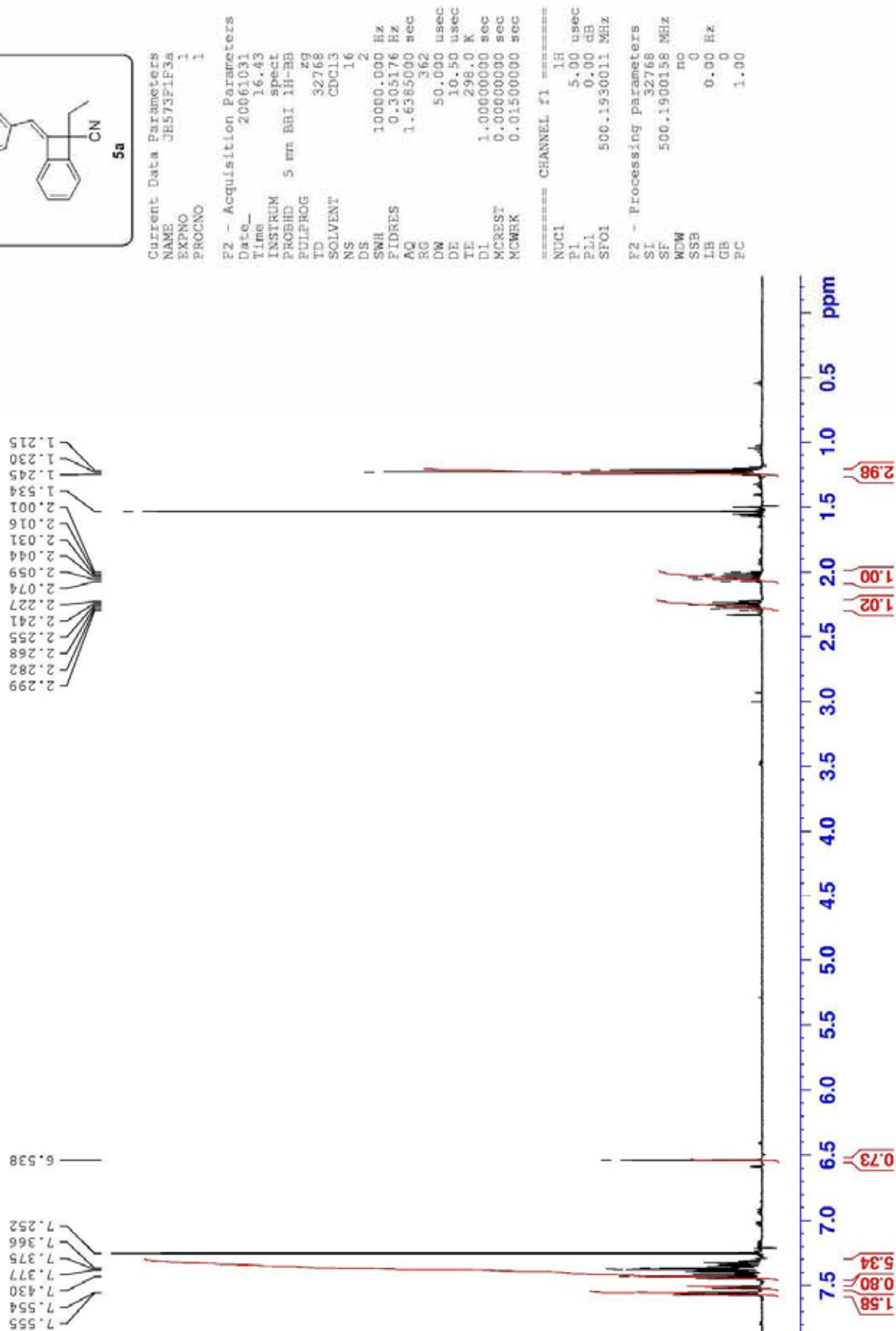
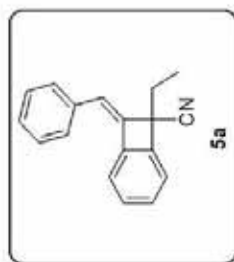


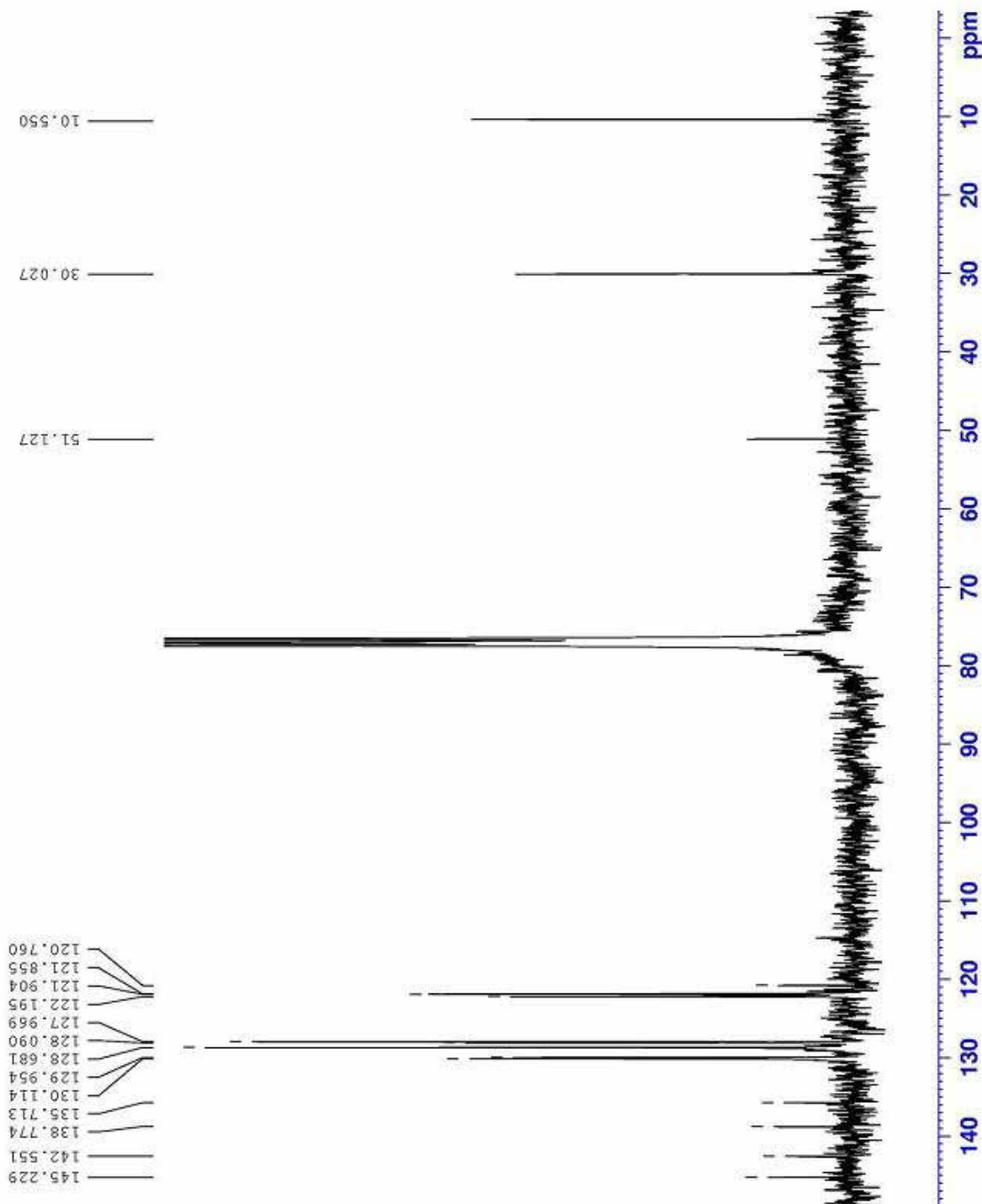
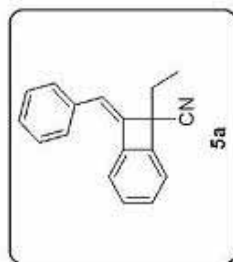












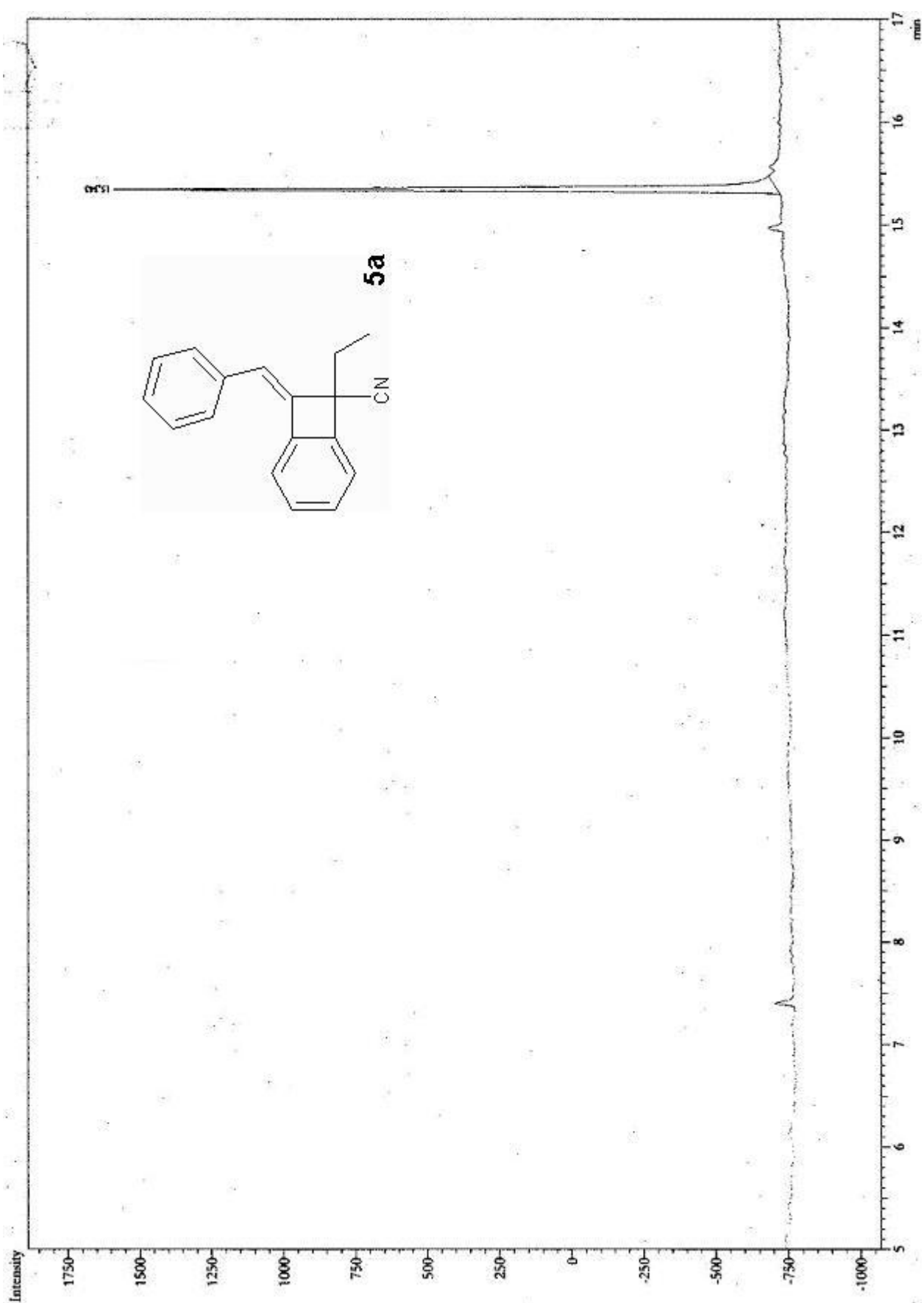
Current Data Parameters
 NAME JH573F1F3D
 EXPNO 2
 PROCNO 1

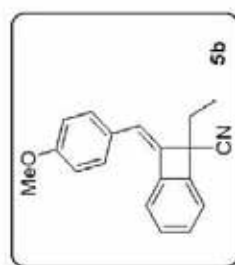
F2 - Acquisition Parameters
 Date_ 20061102
 Time 6.14
 INSTRUM spect
 PROSHD 5 mm Dual 13C/
 PULPROG zgpg30
 TD 16384
 SOLVENT CDCl3
 NS 25000
 DS 4
 SWH 17985.611 Hz
 FIDRES 1.097755 Hz
 AQ 0.4555252 sec
 RG 812.7
 DW 27.800 usec
 DE 6.50 usec
 TE 298.0 K
 D1 0.69999999 sec
 d11 0.03000000 sec
 DELTA 0.59999996 sec
 MCREST 0.00000000 sec
 MCWRX 0.01500000 sec

CHANNEL f1 13C
 P1 9.70 usec
 PL1 -1.00 dB
 SFO1 75.4752953 MHz

CHANNEL f2
 waltz16
 CPDPRG2
 NUC2 1H
 P1 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 SFO2 300.1318530 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677490 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.40





Current Data Parameters
 NAME JH560F2E1a
 EXPNO 1
 PROCNO 1

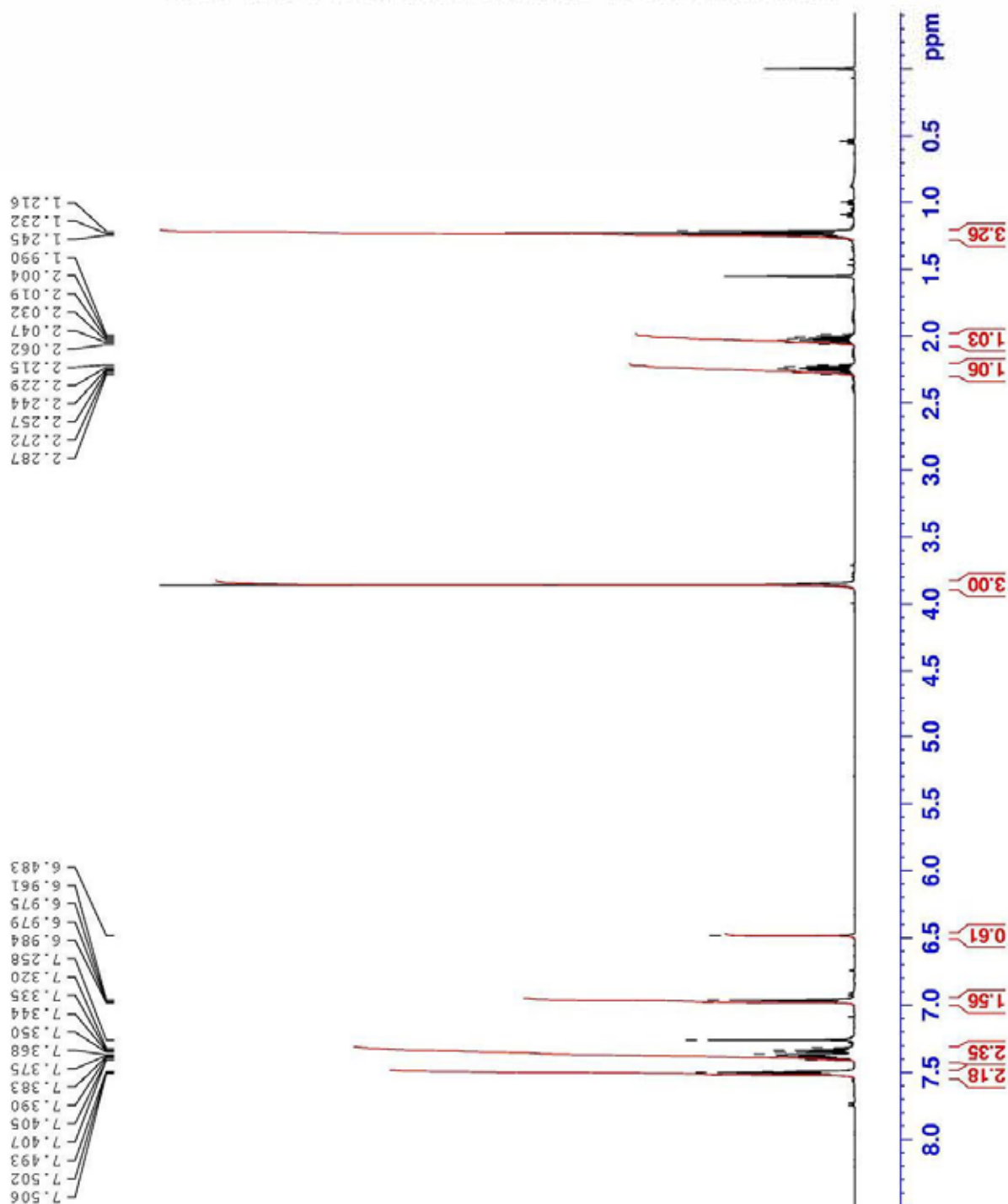
F2 - Acquisition Parameters

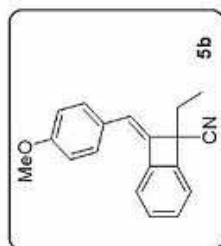
Date_ 20061018
 Time 13.32
 INSTRUM spect
 PROBHD 5 mm BBI 1H-BB
 PULPROG zg
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.305176 Hz
 AQ 1.6385000 sec
 RG 71.8
 DW 50.000 usec
 DE 10.50 usec
 TE 298.0 K
 DI 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 5.00 usec
 PL1 0.00 dB
 SFO1 500.1930011 MHz

F2 - Processing parameters

SI 32768
 SF 500.1900121 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00





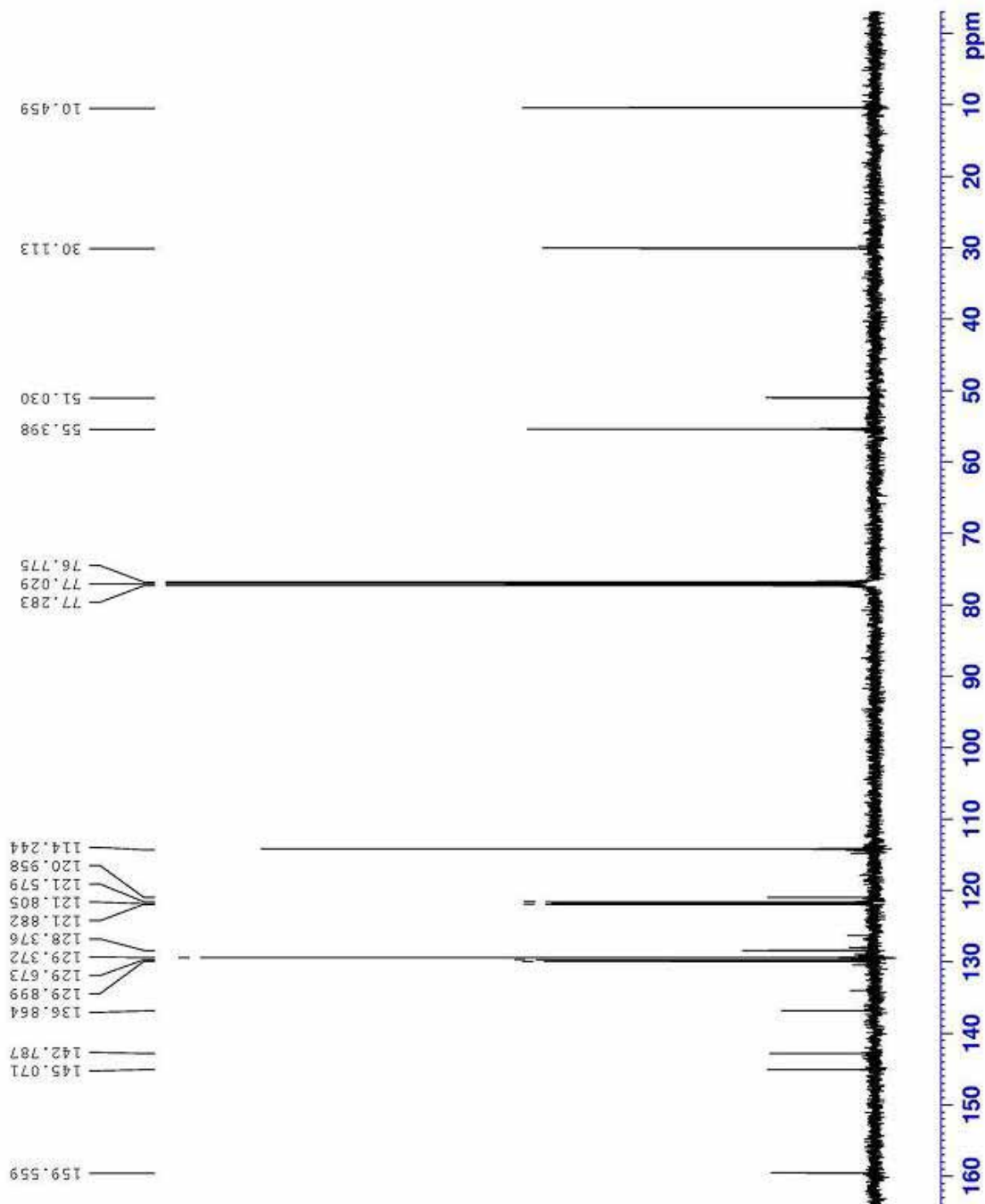
Current Data Parameters
 NAME JH440F4D
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060619
 Time 13.17
 INSTRUM spect
 PROSHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 16384
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 30030.029 Hz
 FIDRES 1.832888 Hz
 AQ 0.2728603 sec
 RG 3160.6
 DE 16.650 usec
 TE 300.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec
 DELTA 0.89999998 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

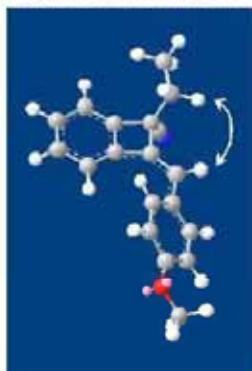
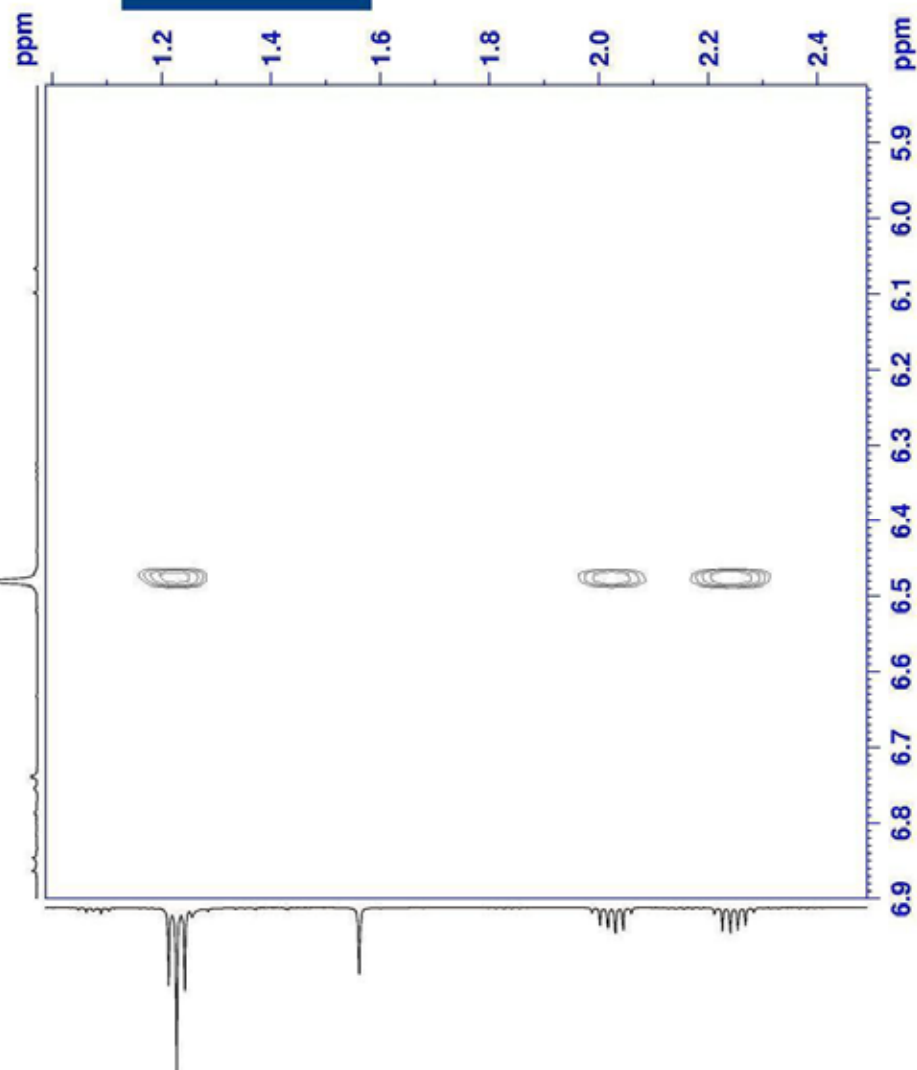
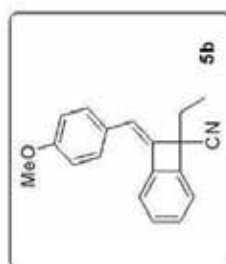
CHANNEL f1 13C
 NUC1 13C
 P1 9.00 usec
 PL1 0.00 dB
 SFO1 125.7703643 MHz

CHANNEL f2 waltz16
 NUC2 1H
 P2 80.00 usec
 PCPD2 0.00 dB
 PL2 17.00 dB
 PL12 17.00 dB
 PL13 17.00 dB
 SFO2 500.130880 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577890 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



NOESY spectrum for compound 5b



Current Data Parameters
NAME JH44074b
EXPNO 8
PROCNO 1

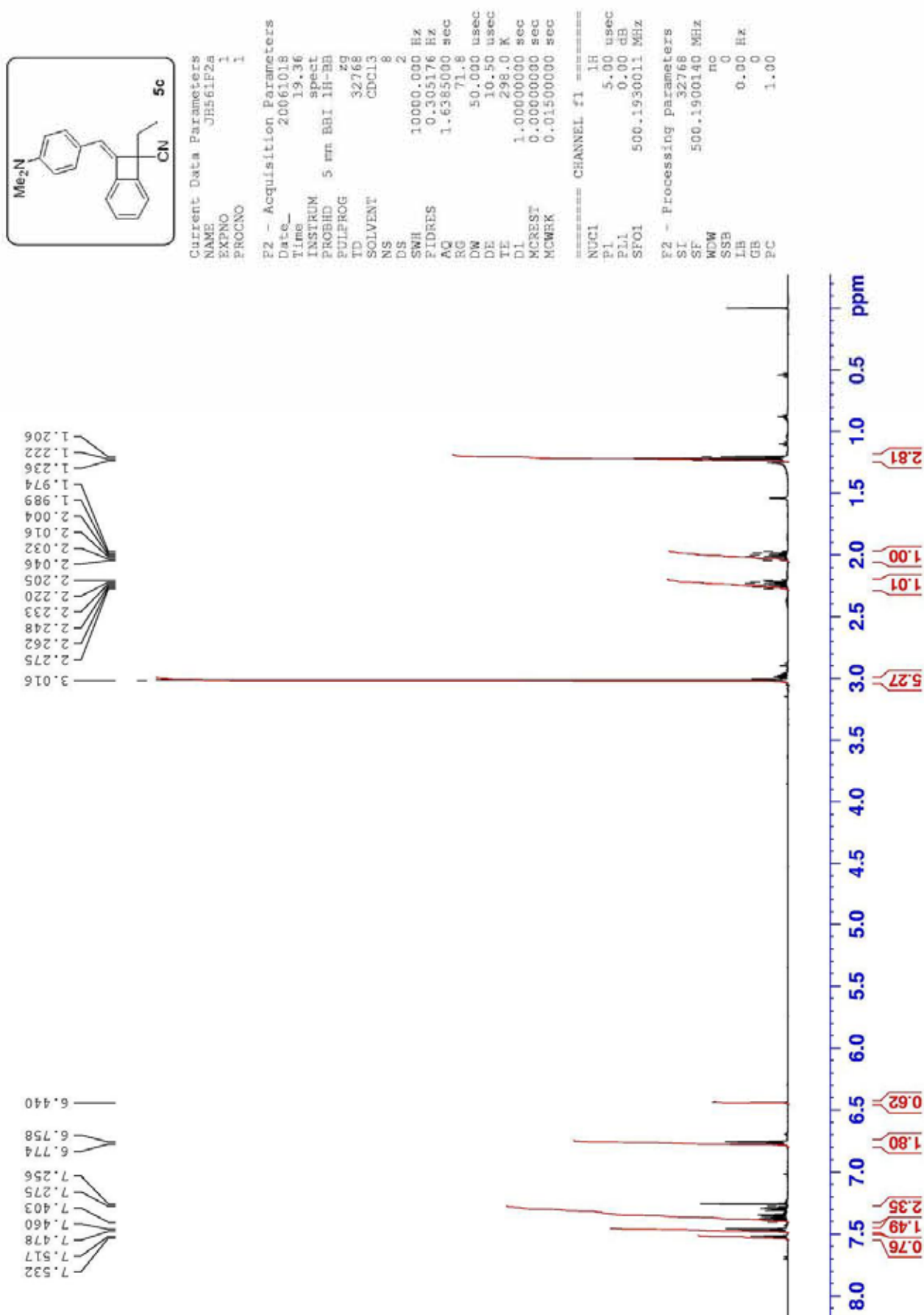
F2 - Acquisition Parameters
Date_ 20060619
Time 14.02
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG noesyph
TD 2048
SOLVENT CDCl3
NS 16
DS 4
SWH 4528.985 Hz
FIDRES 2.211519 Hz
AQ 0.2262596 sec
RG 114
DW 110.400 usec
DE 6.00 usec
TE 300.0 K
d0 0.0009576 sec
D1 1.27501297 sec
D8 0.50000000 sec
IN0 0.0002080 sec
MCHES1
MCWRR
STICN1 128

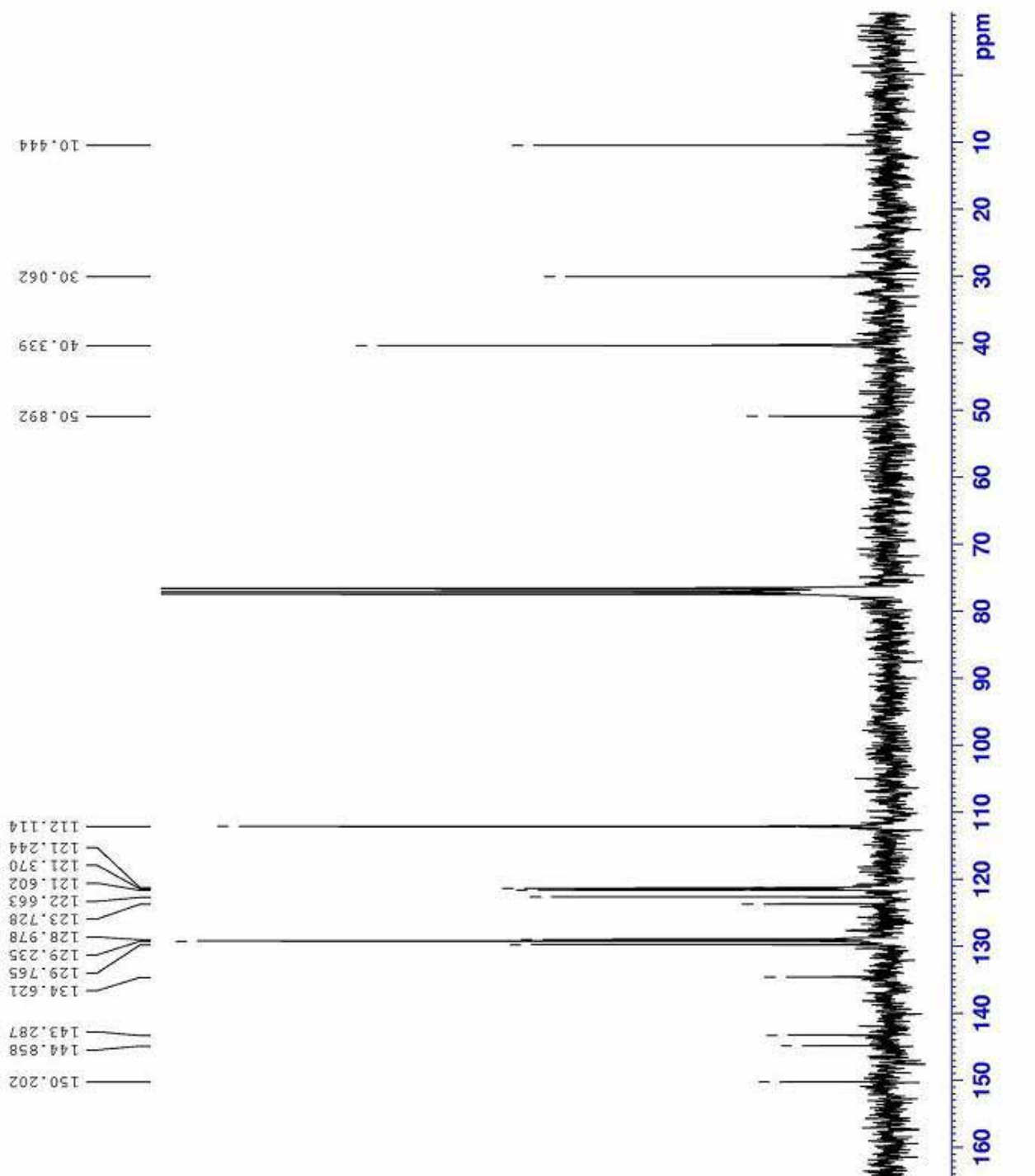
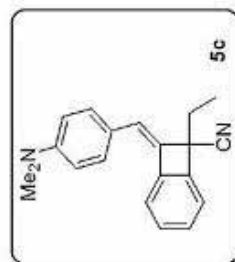
CHANNEL F1
NUC1 ^1H
P1 11.50 usec
PL1 0.00 dB
SF01 500.1319625 MHz

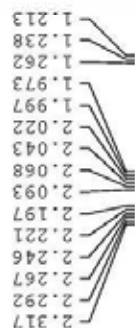
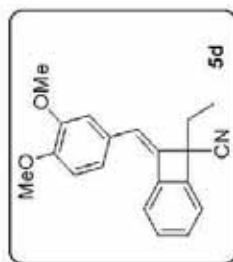
F1 - Acquisition Parameters
ND0 1
TD 256
SF01 500.132 MHz
FIDRES 17.691349 Hz
SW 9.056 pps
FMODE States-TPI

F2 - Processing Parameters
SI 1024
SF 500.1300153 MHz
WDW QSI
SSB 2
LB 0.00 Hz
GB 0
PC 1.00

F1 - Processing Parameters
SI 1024
MC2 States-TPI
SF 500.1300153 MHz
WDW QSI
SSB 2
LB 0.00 Hz
GB 0





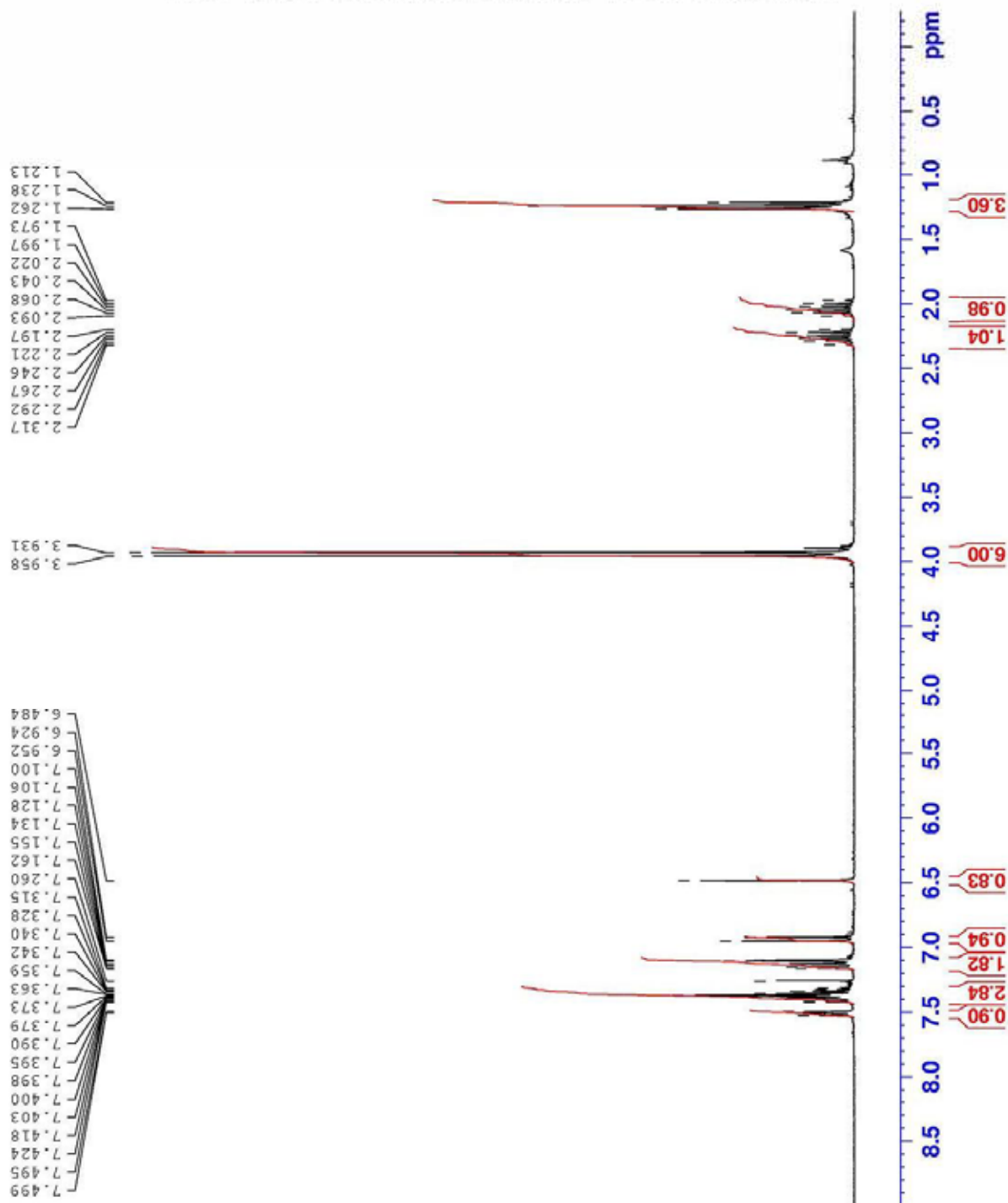


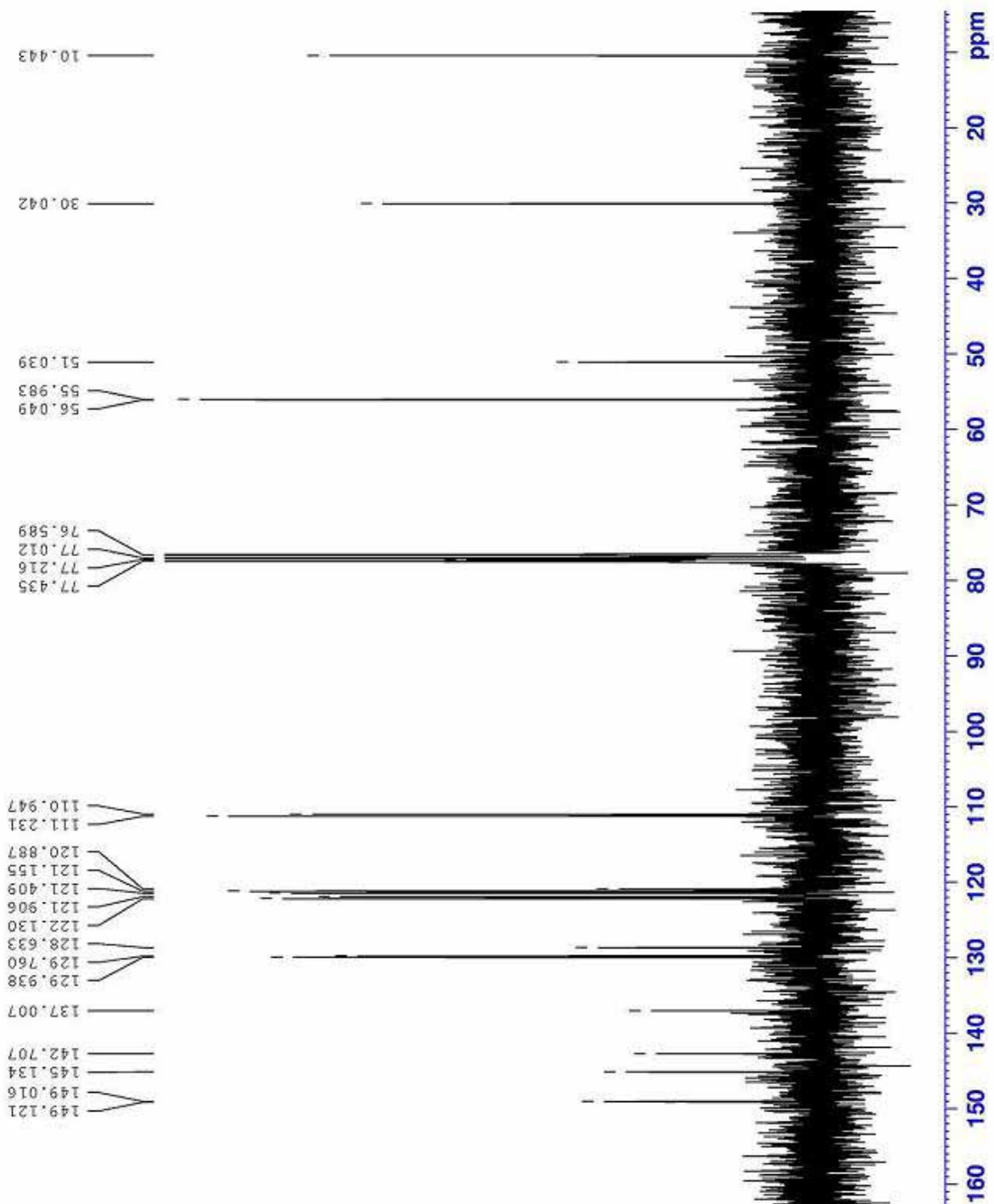
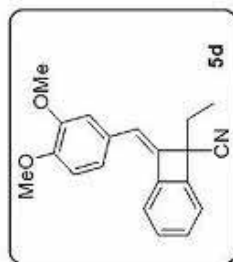
Current Data Parameters
 NAME JH566bisEla
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20061025
 Time 12.50
 INSTRUM spect
 PROBHD 5 mm Dual 13C/
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.188380 Hz
 AQ 2.6542580 sec
 RG 181
 DW 81.000 usec
 DE 6.50 usec
 TE 298.0 K
 DI 1.00000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.30 usec
 PL1 -2.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300061 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00





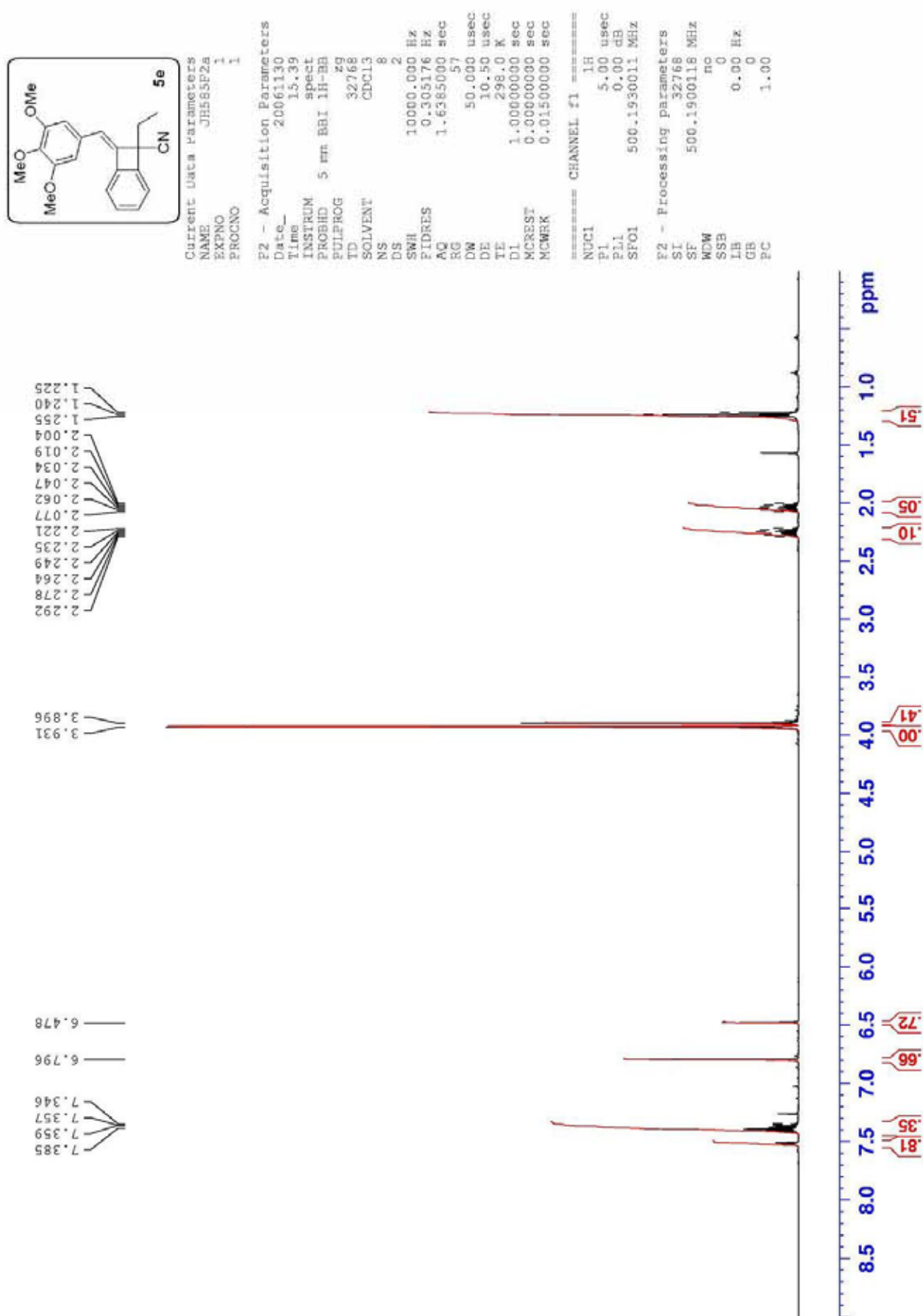
Current Data Parameters
 NAME JH524F2F3b
 EXPNO 3
 PROCNO 1

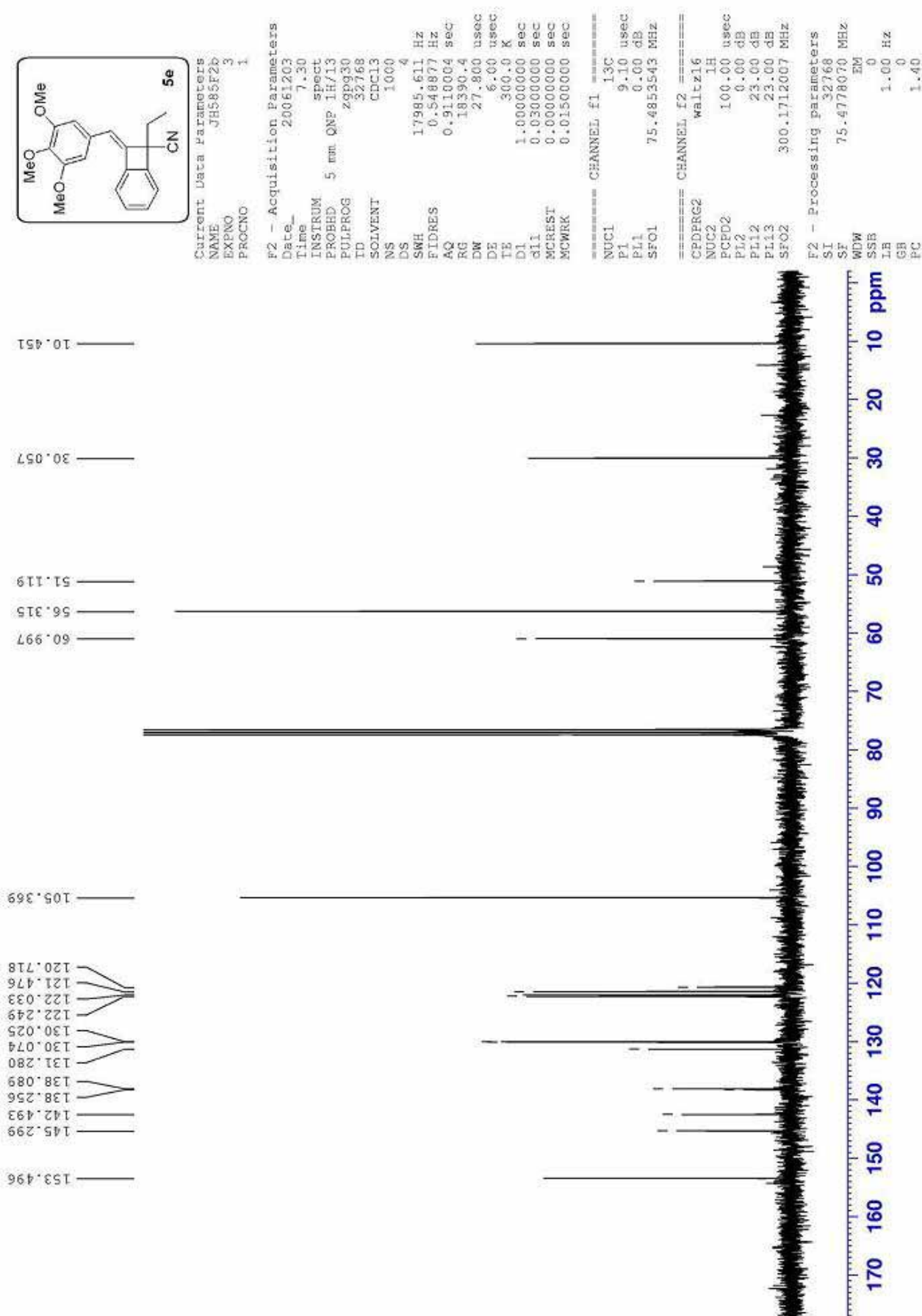
F2 - Acquisition Parameters
 Date_ 20060917
 Time 5.26
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 ID 32768
 SOLVENT CDCl₃
 NS 1000
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.548877 Hz
 AQ 0.9110004 sec
 RG 18390.4
 DW 27.800 usec
 DE 6.00 usec
 TE 299.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

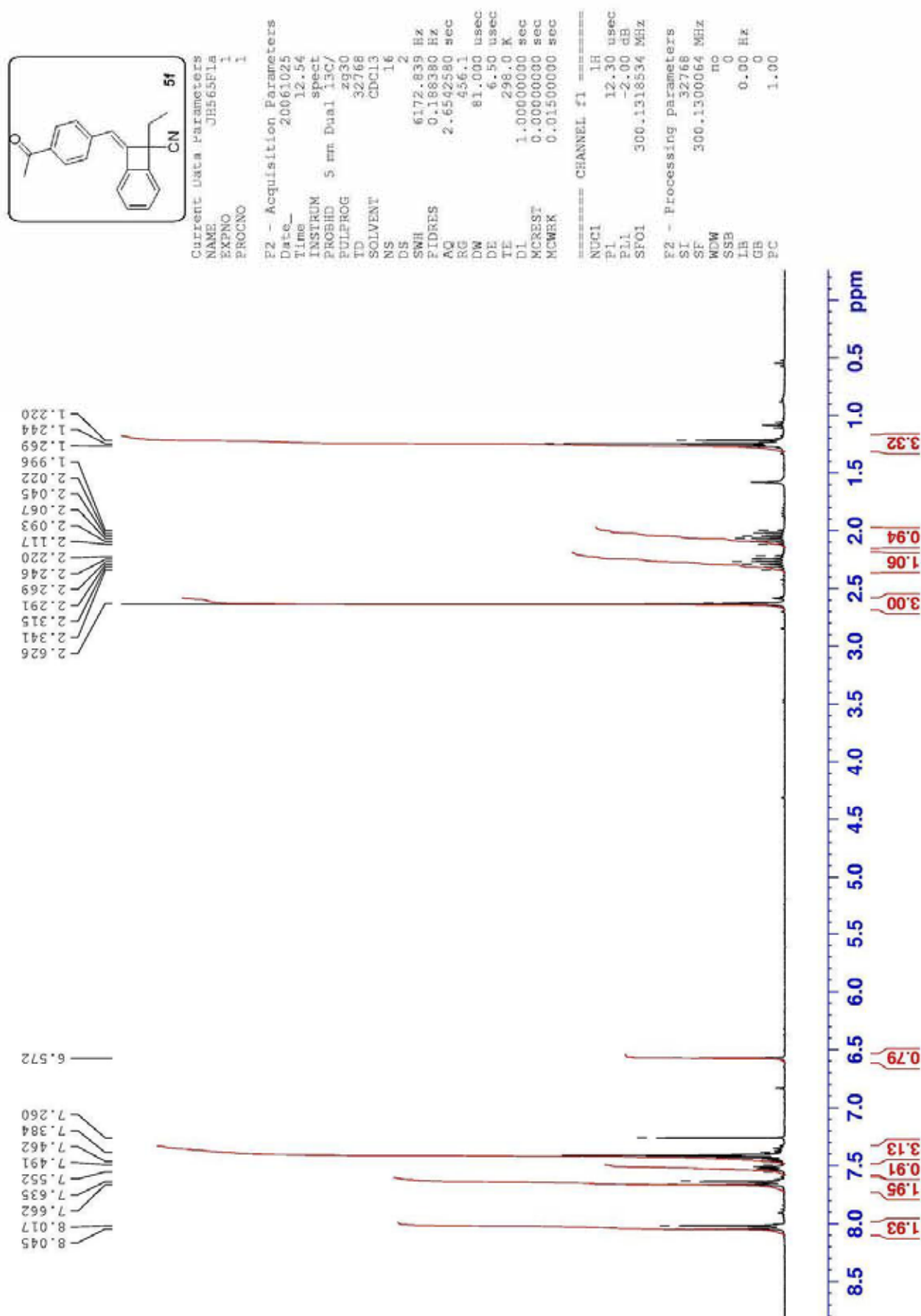
CHANNEL f1 13C
 NUC1 13C
 P1 9.10 usec
 PL1 0.00 dB
 SFO1 75.4853543 MHz

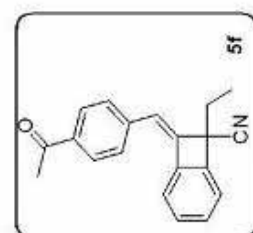
CHANNEL f2 1H
 NUC2 1H
 P2 100.00 usec
 PL2 0.00 dB
 PL12 23.00 dB
 PL13 23.00 dB
 SFO2 300.1712007 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4778070 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40









Current Data Parameters
 NAME JH530F3c
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters

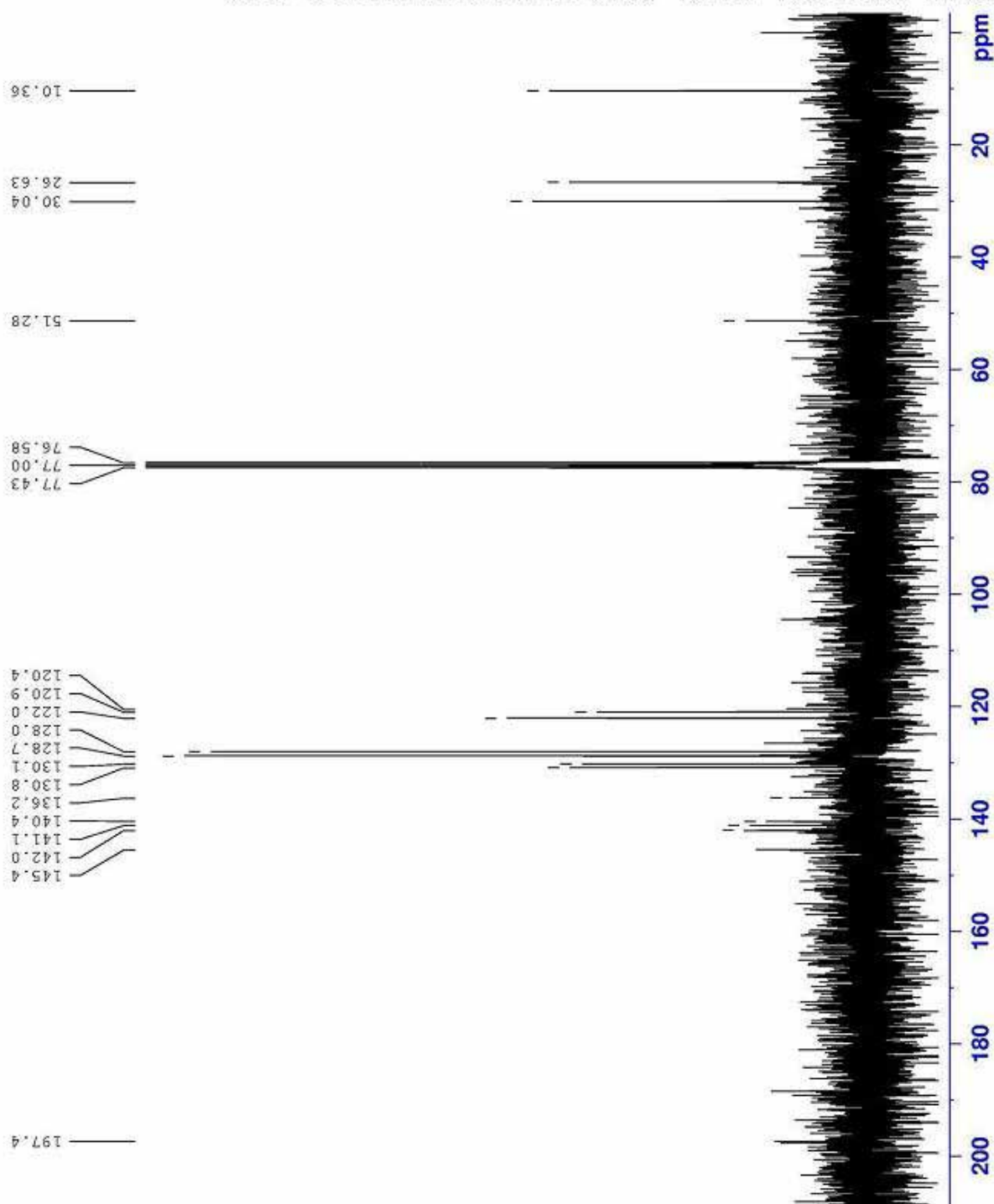
Date_ 20060918
 Time 11.43
 INSTRUM spect
 PROBD 5 mm QNP 1H/13
 PULPROG zgpg30
 ID 32768
 SOLVENT CDCl3
 NS 1000
 DS 4
 SWH 17985.611 Hz
 FIDRES 0.548877 Hz
 AQ 0.9110004 sec
 RG 18390.4
 DW 27.800 usec
 DE 6.00 usec
 TE 299.2 K
 D1 1.00000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

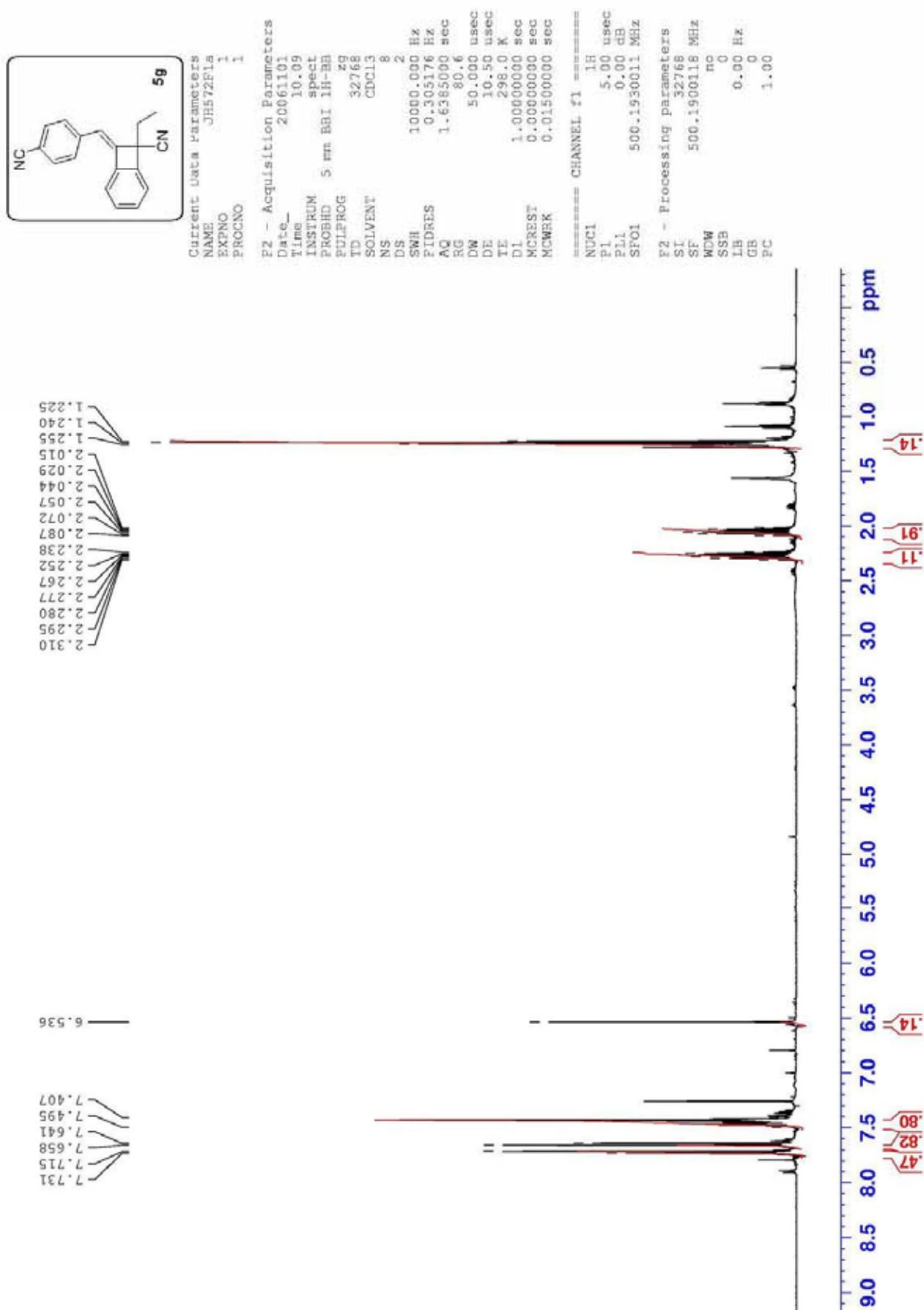
CHANNEL f1
 NUC1 13C
 P1 9.10 usec
 PL1 0.00 dB
 SFO1 75.4853543 MHz

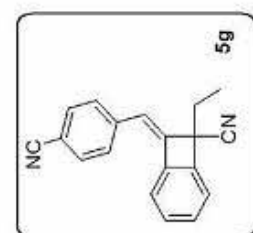
CHANNEL f2
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 23.00 dB
 PL13 23.00 dB
 SFO2 300.1712007 MHz

F2 - Processing parameters

SI 32768
 SF 75.4778070 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40







Current data Parameters
 NAME JHS23F0e
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

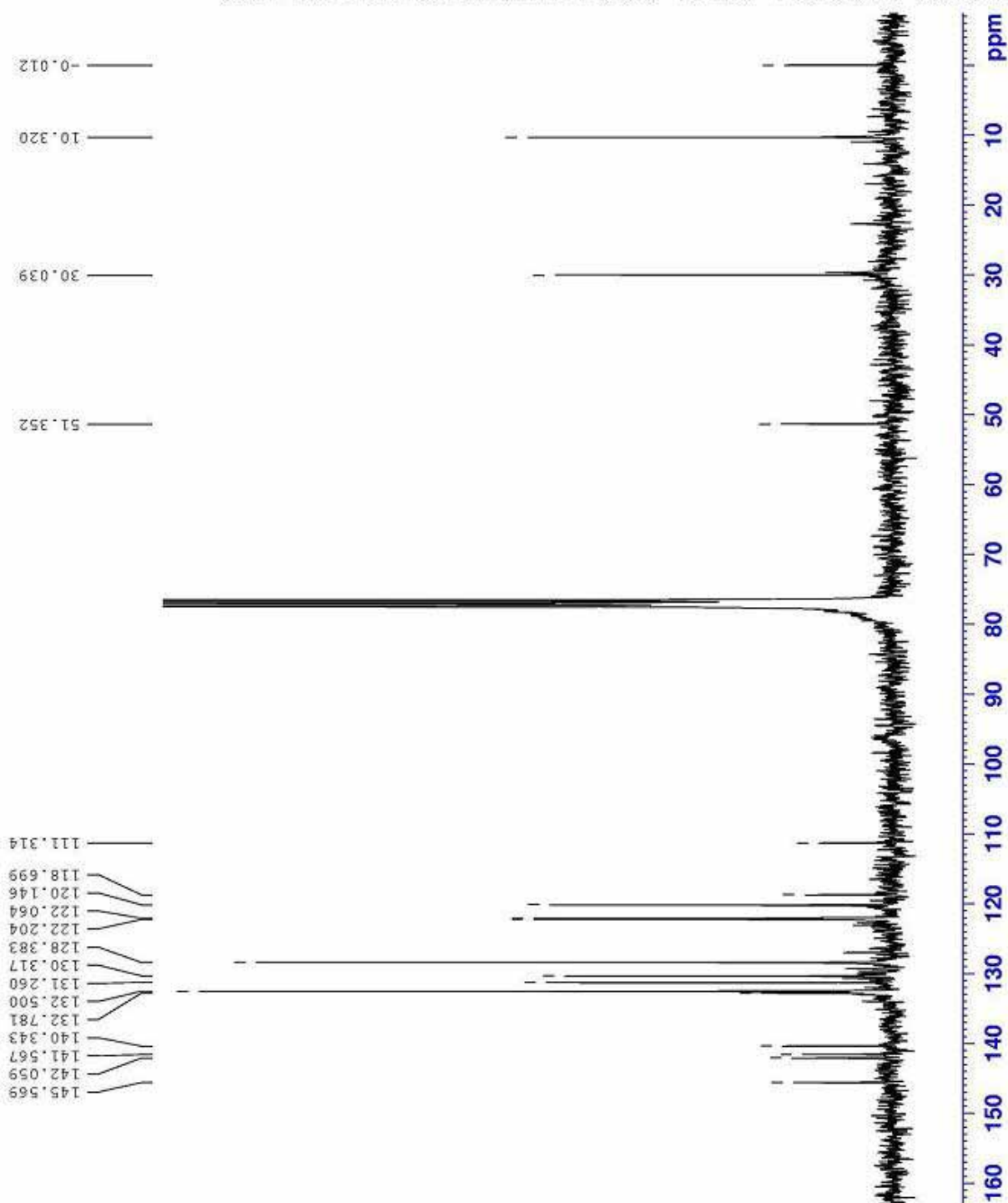
Date_ 20060916
 Time 4.57
 INSTRUM spect
 PROSHD 5 mm Dual 13C/
 PULPROG zgpg30
 TD 16384
 SOLVENT CDCl3
 NS 14000
 DS 4
 SWH 17985.611 Hz
 FIDRES 1.097755 Hz
 AQ 0.4555252 sec
 RG 812.7
 DW 27.800 usec
 DE 6.50 usec
 TE 298.0 K
 D1 0.69999999 sec
 d11 0.03000000 sec
 DELTA 0.59999996 sec
 MCREST 0.00000000 sec
 MCWRR 0.01500000 sec

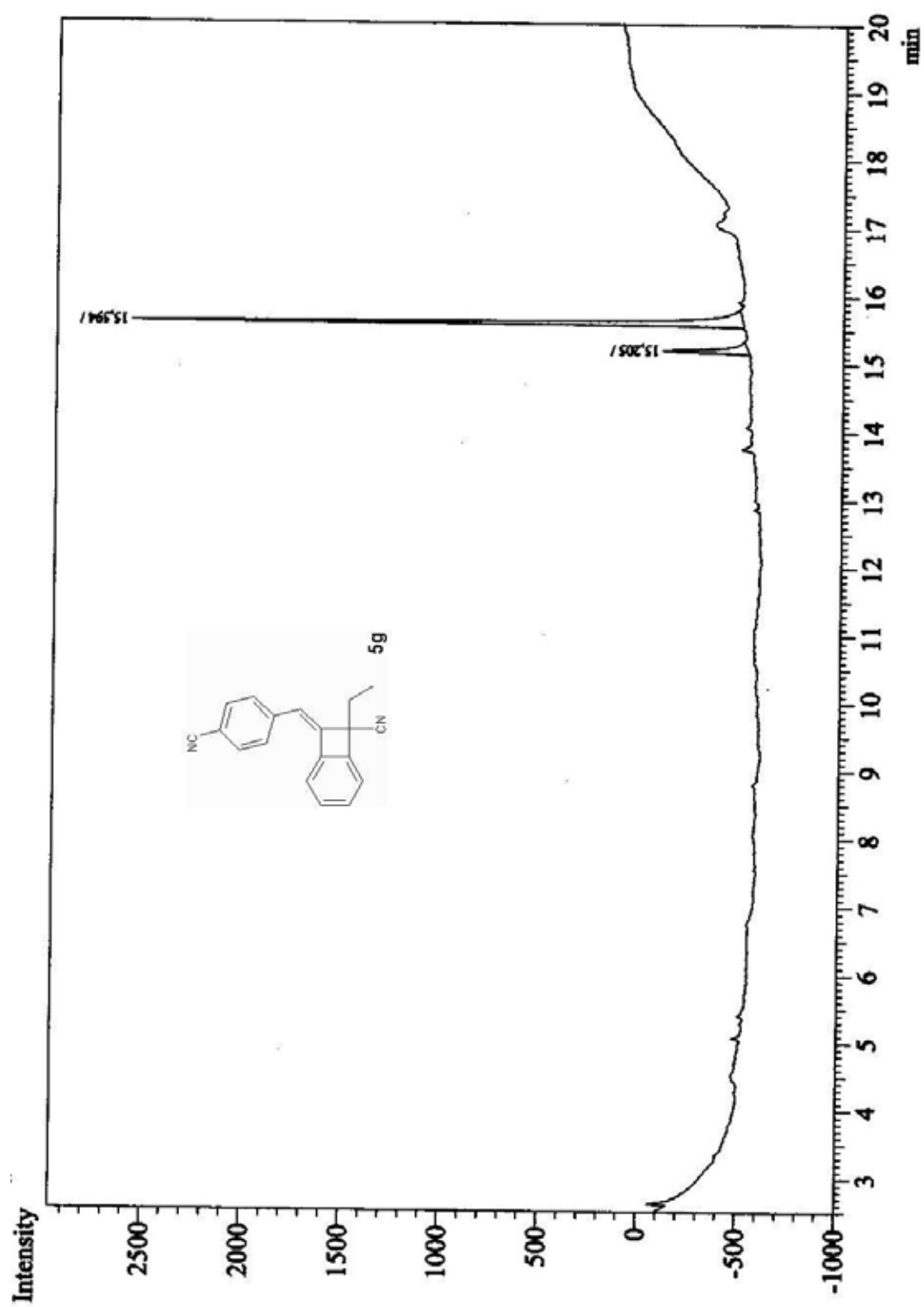
CHANNEL f1 13C
 NUC1 13C
 P1 9.70 usec
 PL1 -1.00 dB
 SFO1 75.4752953 MHz

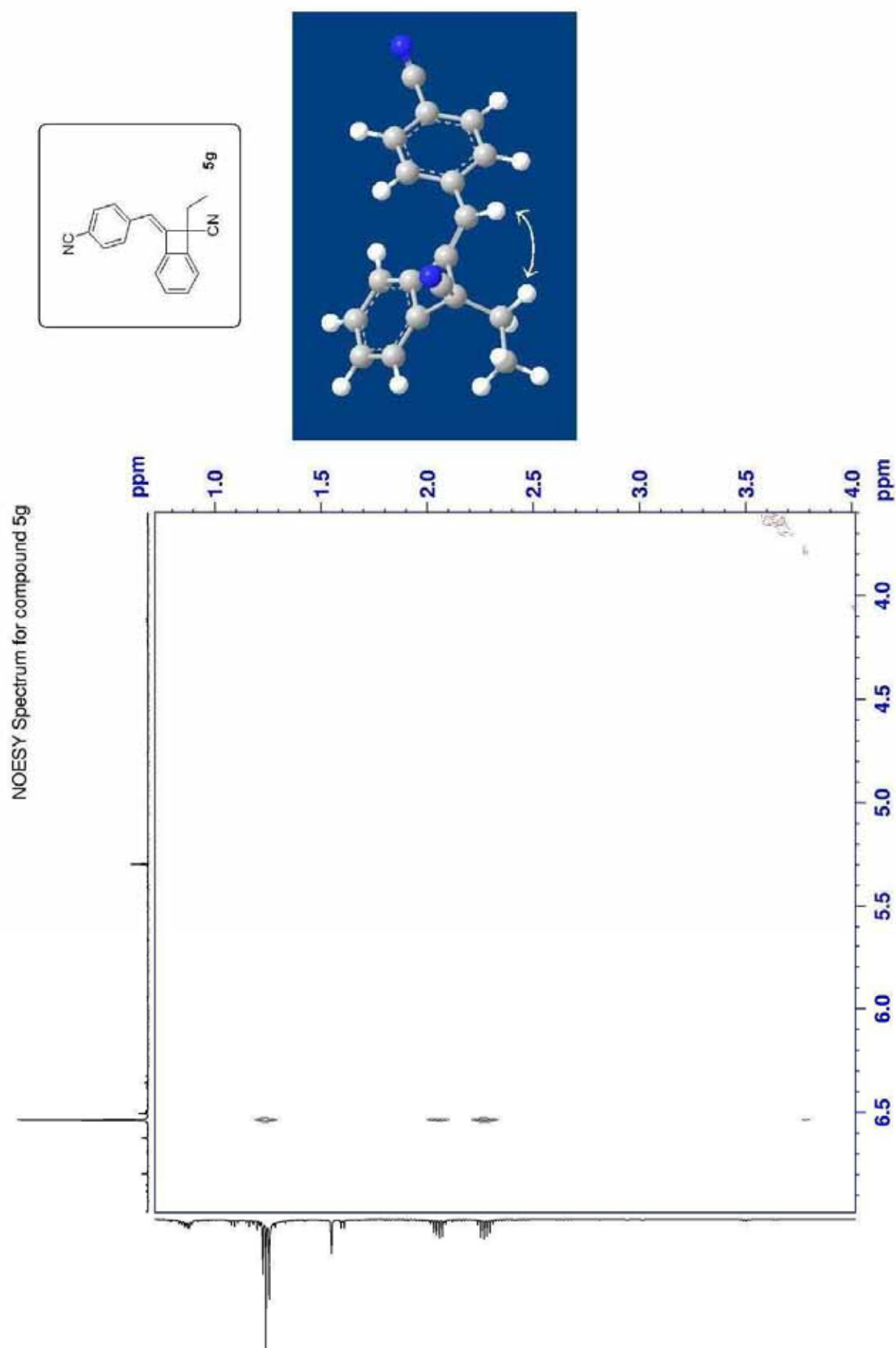
CHANNEL f2 waltz16
 CPDPRG2 1H
 NUC2 1H
 P2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 SFO2 300.1318530 MHz

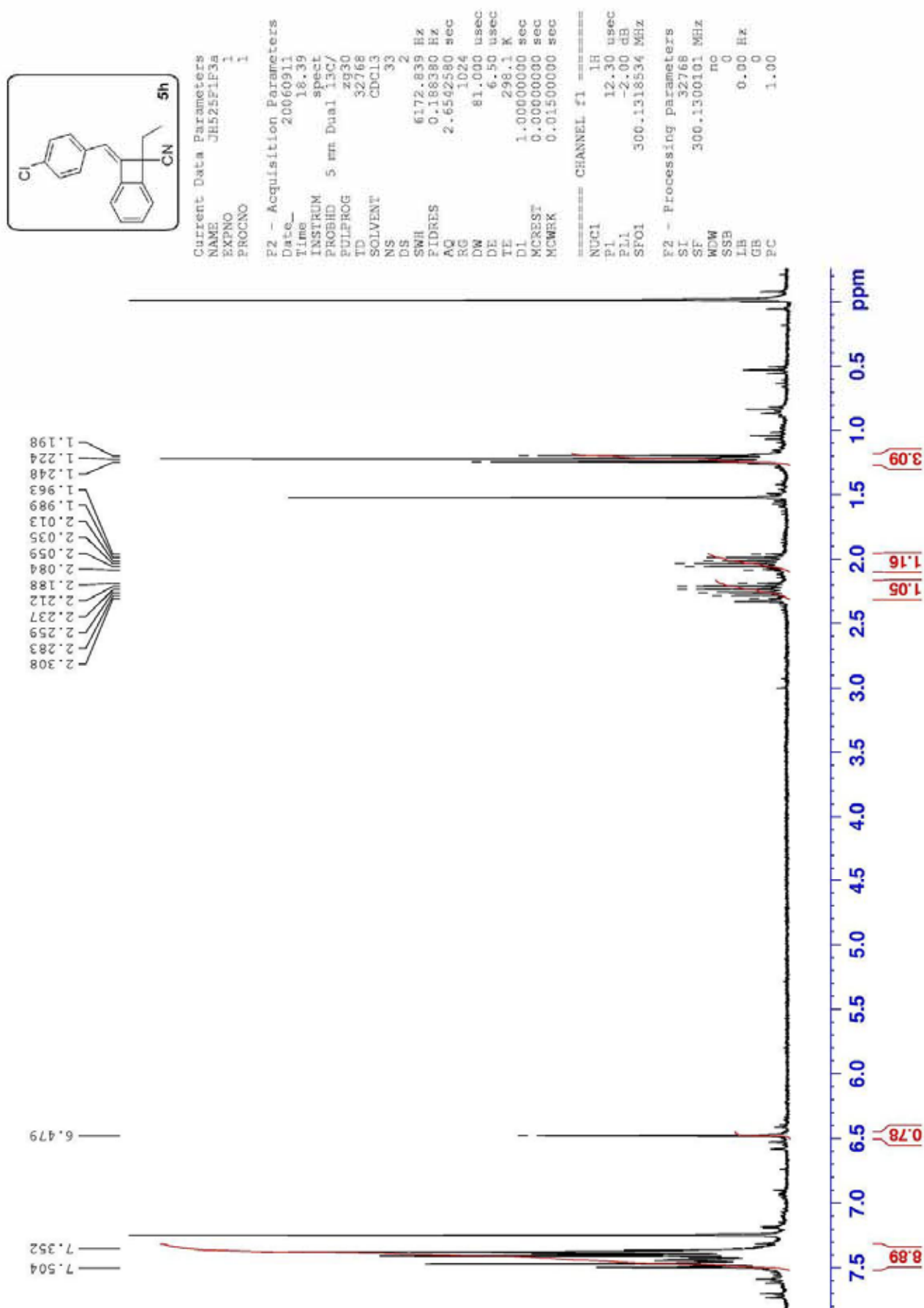
F2 - Processing parameters

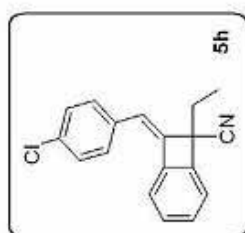
SI 32768
 SF 75.4677490 MHz
 EM
 WDW 0
 SSB 3.00 Hz
 LB 0
 GB 0
 PC 1.40











Current data parameters
 NAME JH570F2f1a
 EXPNO 2
 PROCNO 1

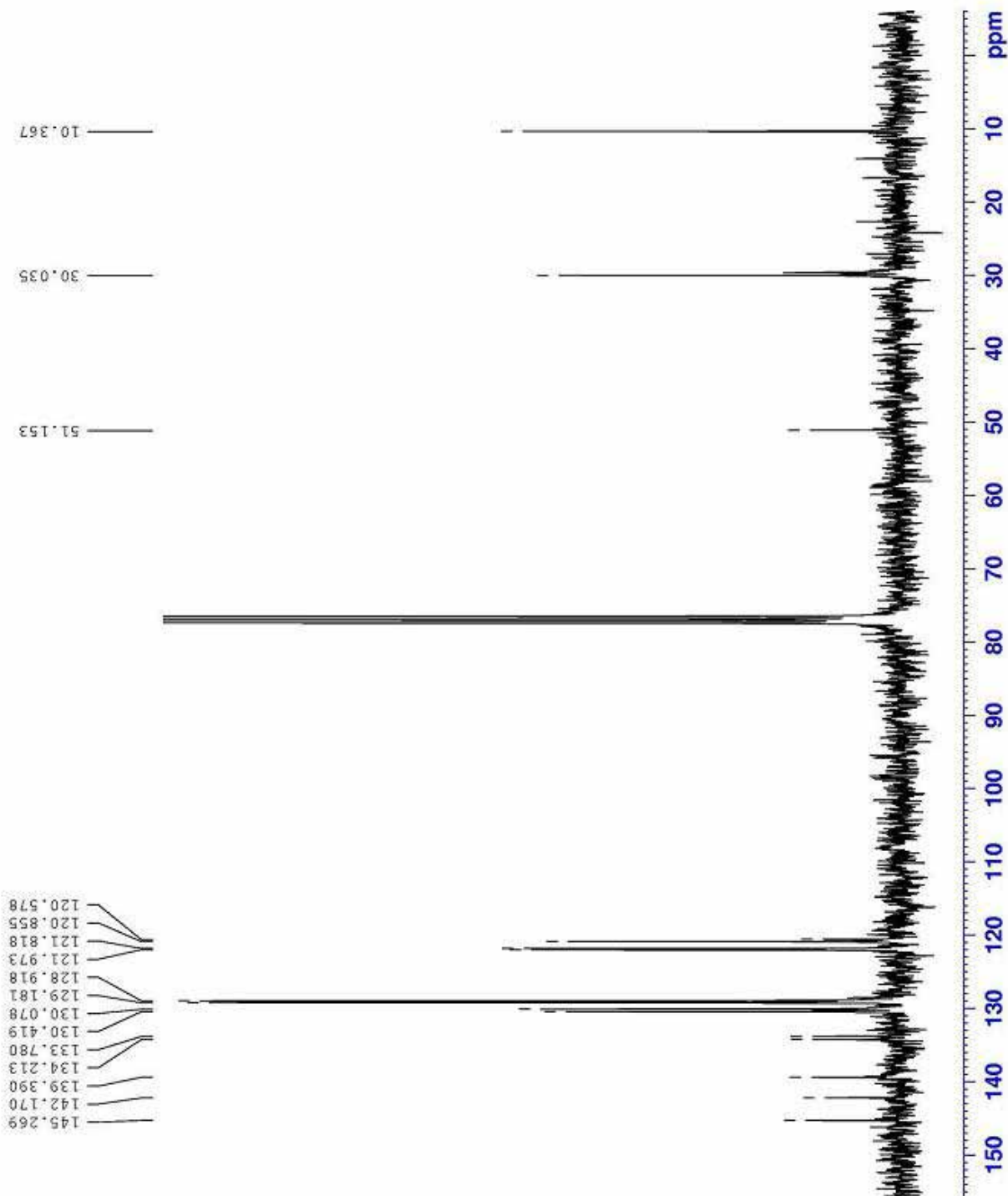
F2 - Acquisition Parameters

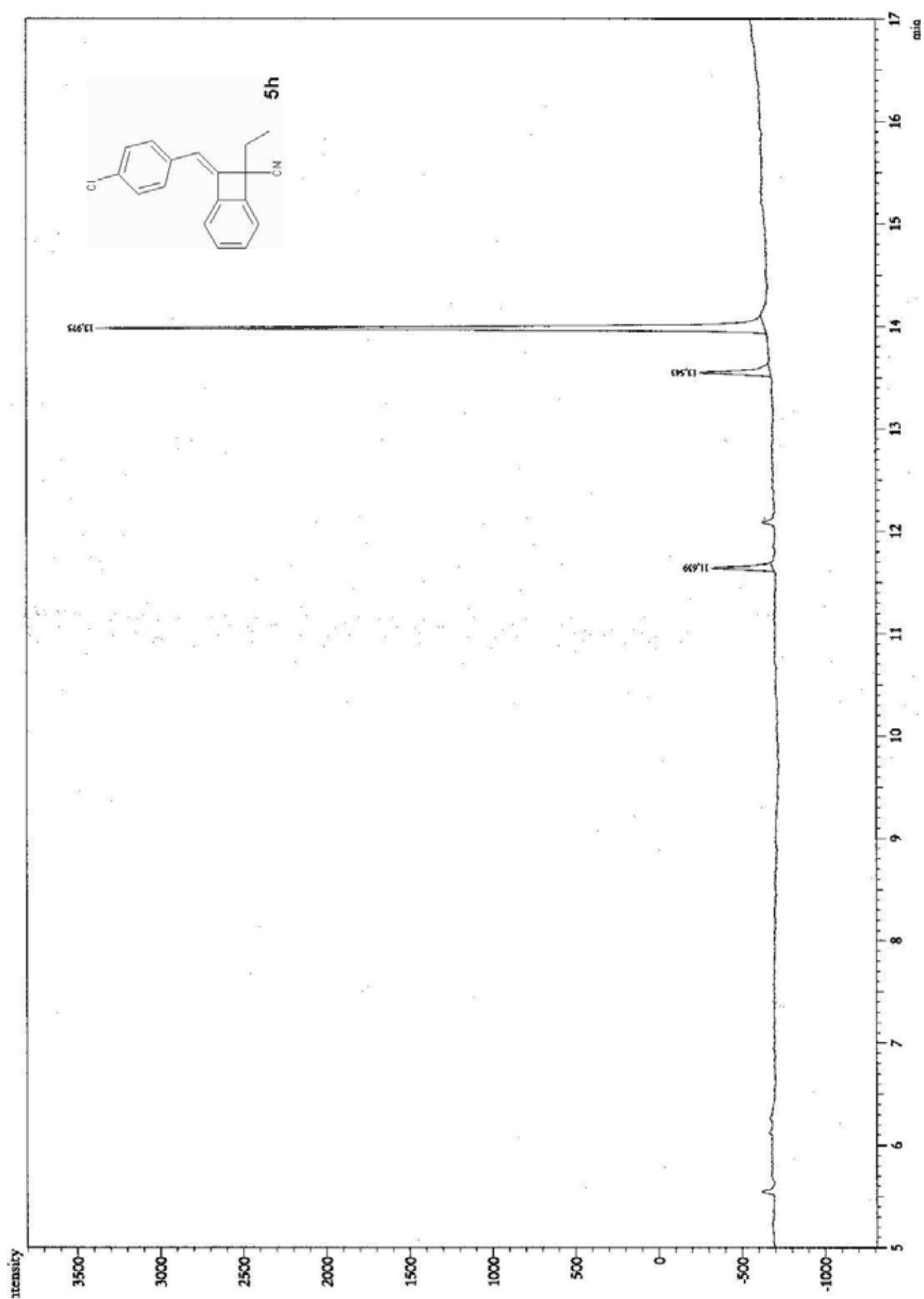
Date_ 20061030
 Time 23.10
 INSTRUM spect
 PROSHD 5 mm Dual 13C/
 PULPROG zgpg30
 TD 16384
 SOLVENT CDCl3
 NS 620
 DS 4
 SWH 17985.611 Hz
 FIDRES 1.097755 Hz
 AQ 0.4555252 sec
 RG 812.7
 DW 27.800 usec
 DE 6.50 usec
 TE 298.0 K
 d1 0.69999999 sec
 d11 0.03000000 sec
 DELTA 0.59999996 sec
 MCREST 0.00000000 sec
 MCWRX 0.01500000 sec

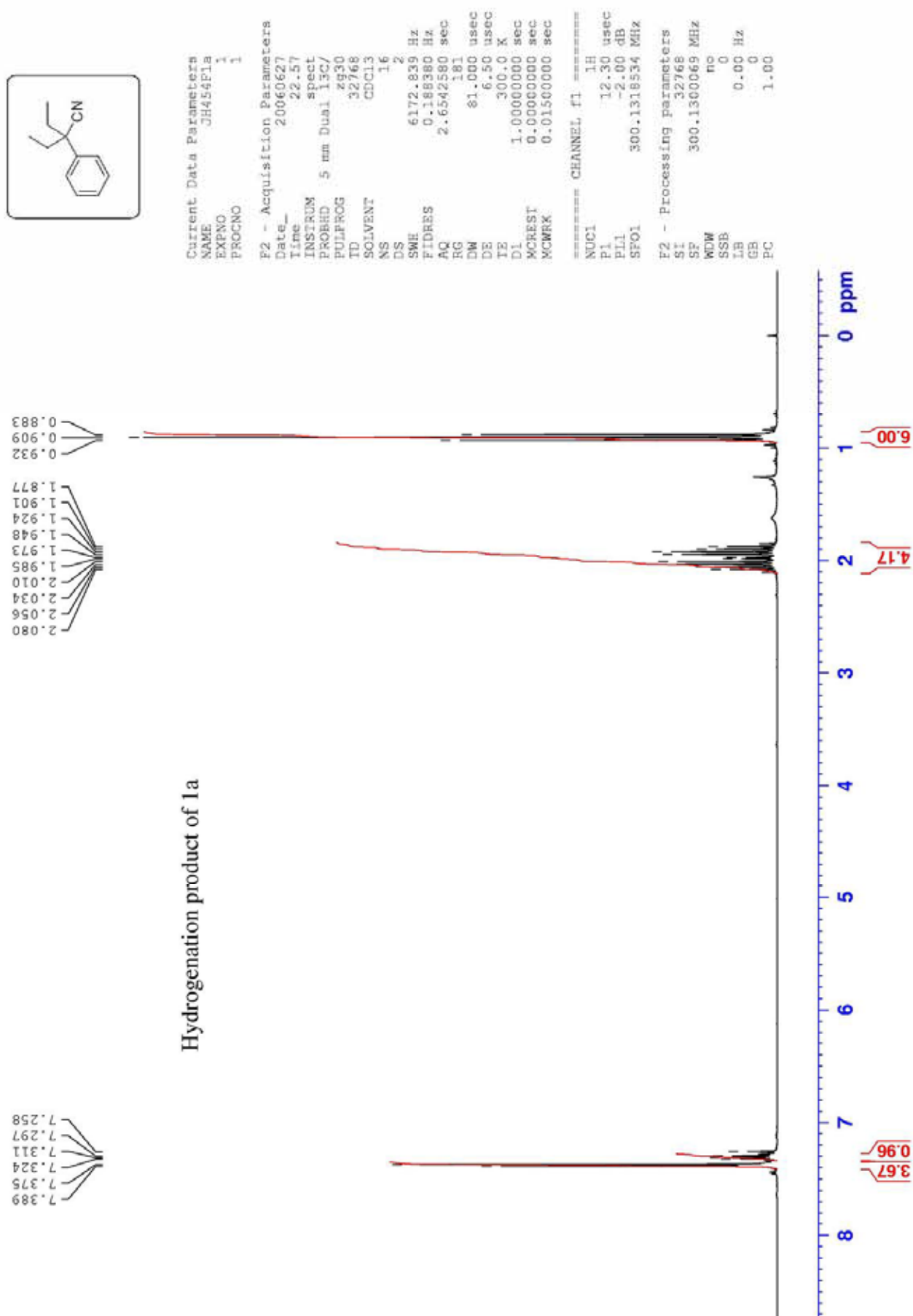
CHANNEL f1 13C
 NUC1 13C
 P1 9.70 usec
 PL1 -1.00 dB
 SFO1 75.4752953 MHz

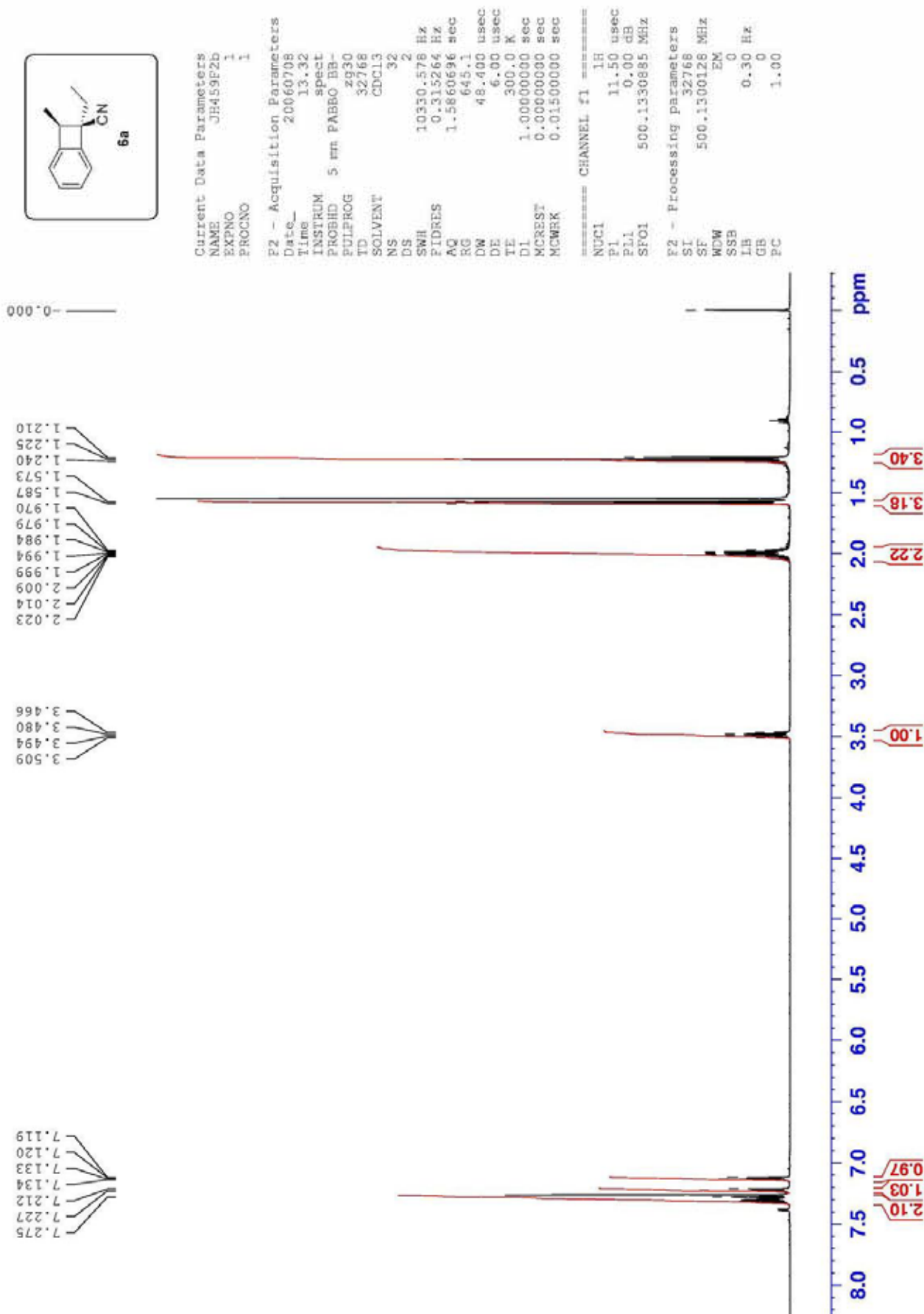
CHANNEL f2 waltz16
 CPDPRG2 1H
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 14.50 dB
 PL13 14.50 dB
 SFO2 300.1318530 MHz

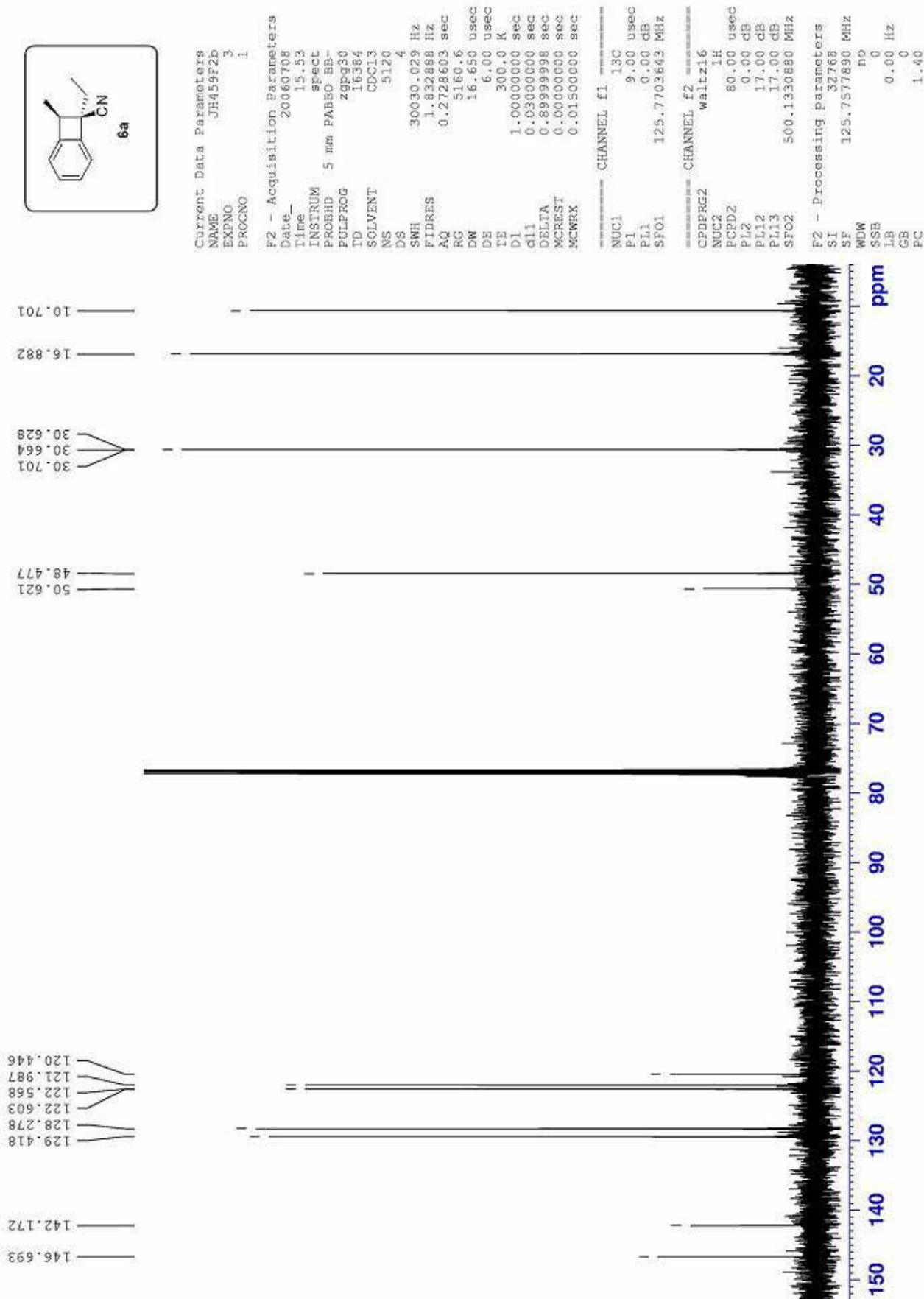
F2 - Processing parameters
 SI 32768
 SF 75.4677490 MHz
 WDW EM
 SSB 0
 LB 3.00 Hz
 GB 0
 PC 1.40

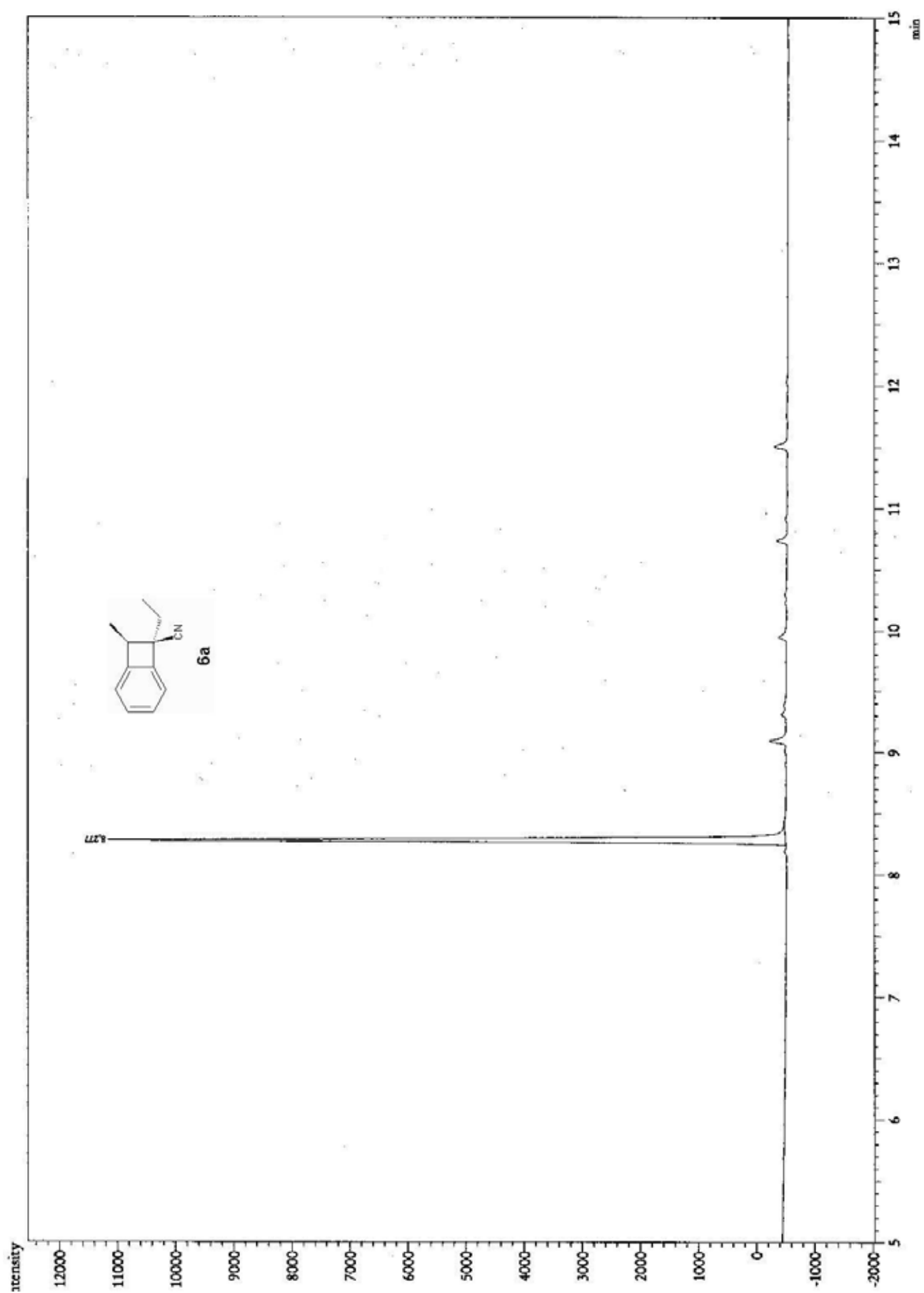




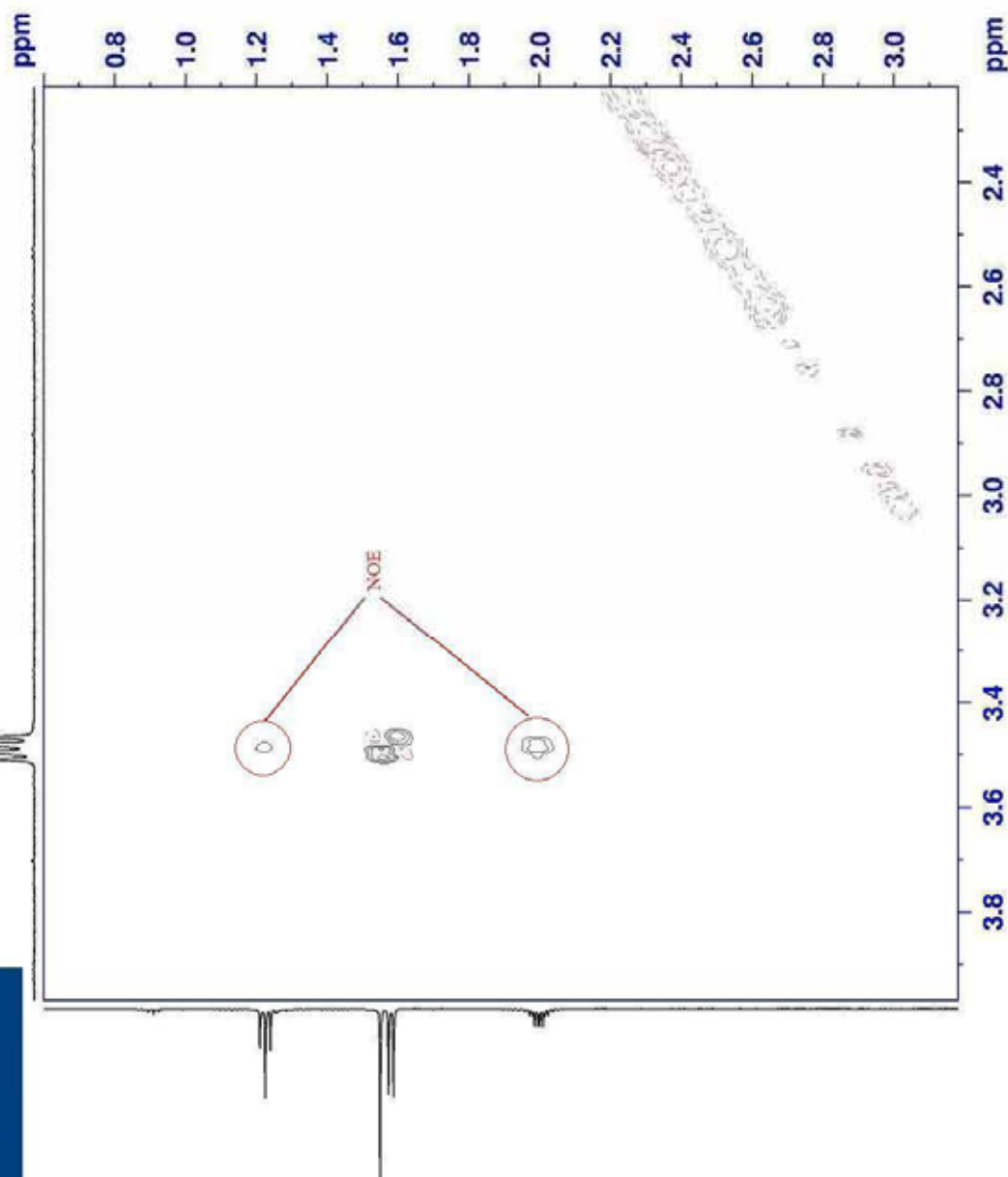
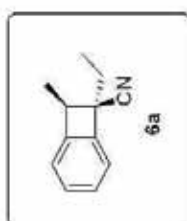








NOESY of compound 6a



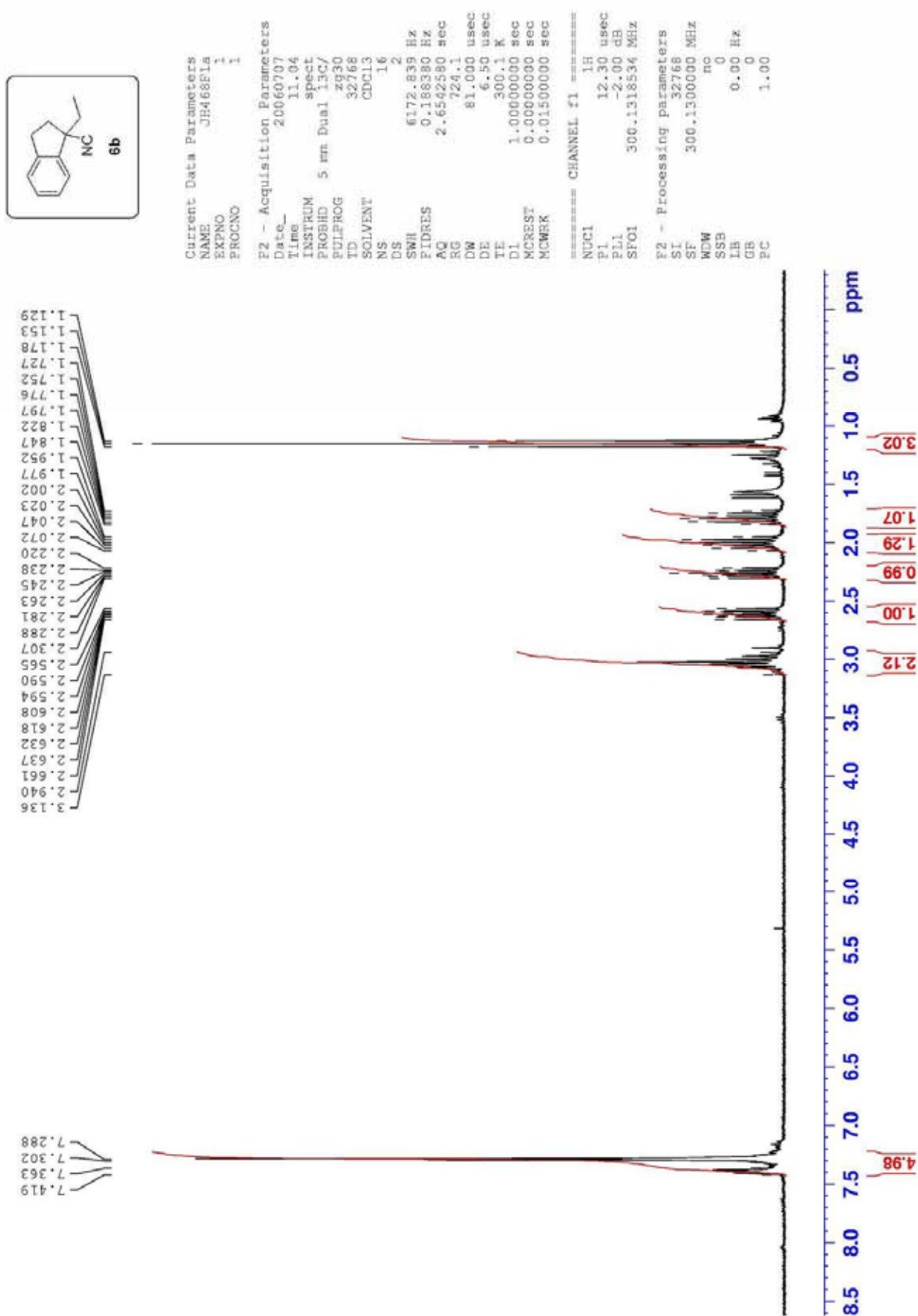
Current Data Parameters
 NAME JH459F2D
 EXPNO 8
 PROCNO 1

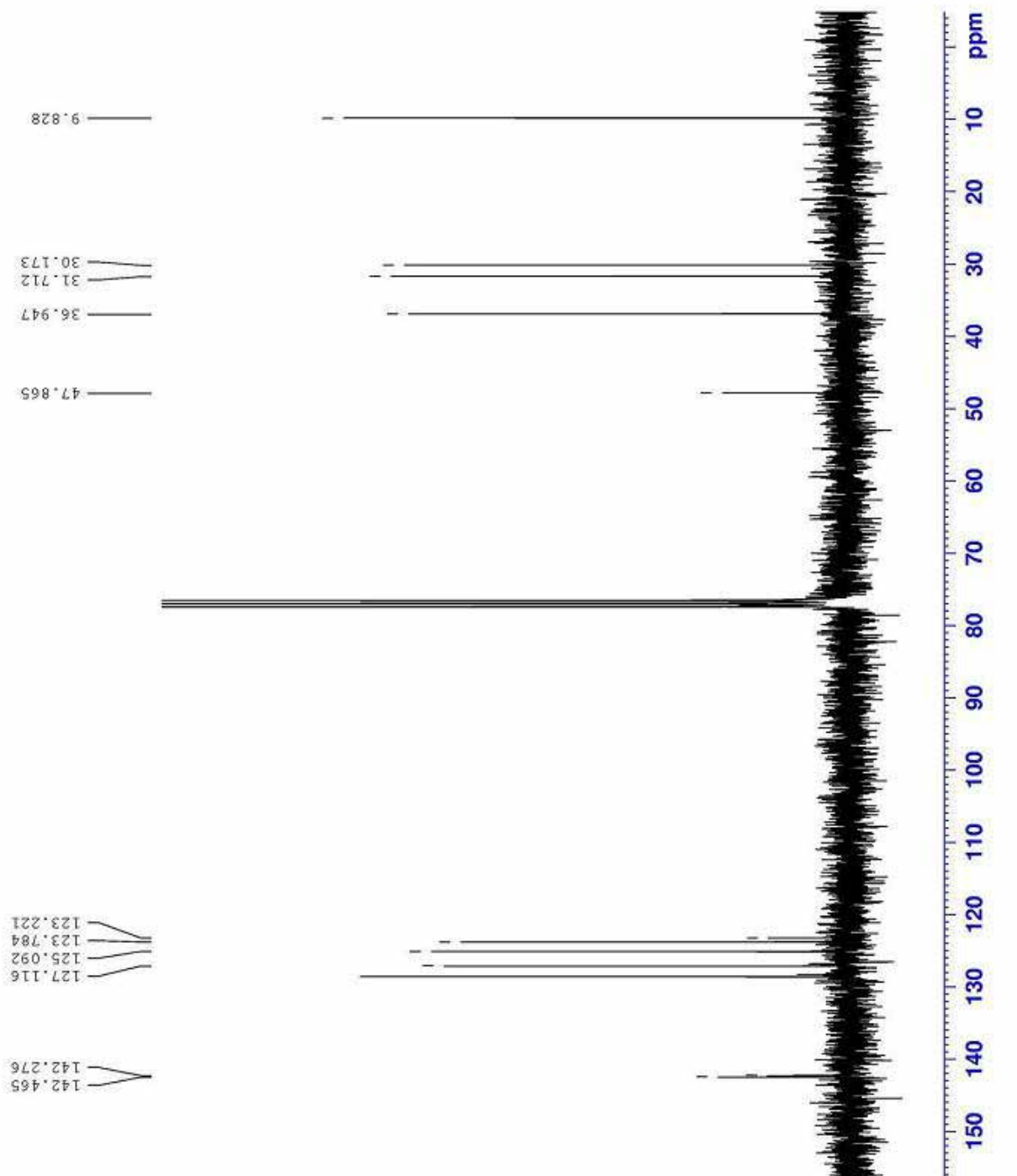
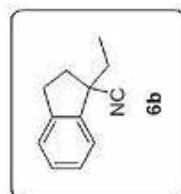
F2 - Acquisition Parameters
 Date_ 20060708
 Time 19.30
 INSTRUM spect
 PROBED 5 mm PABBO BB-
 FULPRG noesyph
 ID 2048
 SOLVENT CDCl3
 NS 16
 DS 4
 SWH 4240.822
 FIDRES 2.040245
 AQ 0.2393732
 RG 228.1
 DM 116.800
 DE 6.00
 TE 300.0
 D1 0.00010216
 D11 1.26067696
 D8 0.50000000
 INO 0.00823360
 MCKEST 0.00000000
 MCKRK 0.6303889
 STICNT 128

===== CHANNEL f1 =====
 NUC1 1H
 P1 11.50
 PL1 0.00
 SFO1 500.1318219

F1 - Acquisition Parameters
 NDO 1
 TD 256
 SFO1 500.1318
 FIDRES 16.721960
 SW 9.559
 FwMODE States-TPI
 F2 - Processing Parameters
 SI 1024
 SF 500.130131
 WDW QSINE
 SSB 2
 GB 0
 EC 1.00

F1 - Processing Parameters
 SI 1024
 MC2 States-TPI
 SF 500.130131
 WDW QSINE
 SSB 2
 GB 0





Current Data Parameters
 NAME JH468Flb
 EXFO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20060710
 Time 0.36
 INSTRUM spect
 PROHD 5 mm QNP 1H/13
 PULPROG zgpg30
 ID 32768
 SOLVENT CDCl3
 NS 1000
 DS 4
 SWE 17985.611 Hz
 FIDRES 0.548877 Hz
 AQ 0.9110004 sec
 RG 18390.4
 DW 27.800 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 d11 0.03000000 sec
 MCREST 0.00000000 sec
 MCWRK 0.01500000 sec

CHANNEL f1 13C
 NUC1
 P1 9.10 usec
 PL1 0.00 dB
 SF01 75.4853543 MHz

CHANNEL f2 1H
 CPDPRG2 waltz16
 NUC2
 PCPD2 100.00 usec
 PL2 0.00 dB
 PL12 23.00 dB
 PL13 23.00 dB
 SFO2 300.1712007 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4778070 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

