

Supporting Information

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Supporting Information

Organocatalytic *anti*-Mannich Reactions with Dihydroxyacetone and Acyclic Dihydroxyacetone Derivatives: A Facile Route to Amino Sugars

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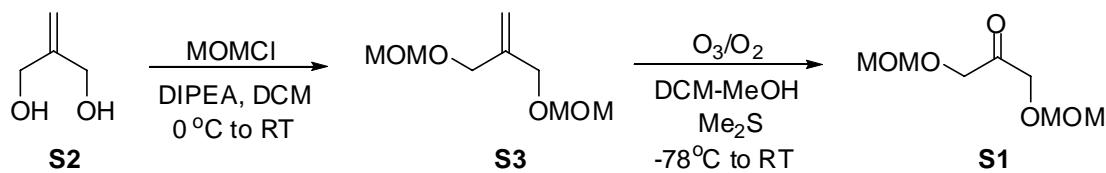
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General: For thin layer chromatography (TLC), silica gel plates VWR GL60 F254 were used and compounds were visualized by irradiation with UV light and/or by treatment with a solution of phosphomolybdic acid (25 g), Ce(SO₄)₂•H₂O (10 g), and conc. H₂SO₄ (60 mL) in H₂O (940 mL) followed by heating or by treatment with a solution of *p*-anisaldehyde (23 mL), conc. H₂SO₄ (35 mL), and acetic acid (10 mL) in ethanol (900 mL) followed by heating. Flash column chromatography was performed using Bodman silica gel 32-63, 60Å. ¹H NMR and ¹³C NMR spectra were recorded on Bruker DRX-500 and Varian INOVA-400. Proton chemical shifts are given in relative to tetramethylsilane (0.00 ppm) in CDCl₃ or to the residual proton signals of the deuterated solvent in CD₃OD (3.31 ppm). Carbon chemical shifts were internally referenced to the deuterated solvent signals in CDCl₃ (77.00 ppm) or CD₃OD (49.00 ppm). High-resolution mass spectra were recorded on an Agilent ESI-TOF mass spectrometer. Enantiomeric excesses were determined by chiral-phase HPLC using a Hitachi instrument.

Preparation of protected dihydroxyacetones:

1,3-Dihydroxyacetone dimer was purchased from Aldrich and was used without further purification. Bn-protected dihydroxyacetone was prepared according to the literature methods.^{1,2} TBS-protected dihydroxyacetone was prepared according to the literature method.³ MOM-protected dihydroxyacetone **S1** was prepared as following procedure (**Scheme S1**).

Scheme 1: Synthesis of MOM-protected dihydroxyacetone **S1.**



6-Methylene-2,4,8,10-tetraoxaundecane (S3**)**

To an ice-cold solution of 2-methylene-1,3-propanediol **S2** (3.5 g, 39.8 mmol) in methylene chloride (25 mL) was added diisopropylethyl amine (17.3 mL, 99.4 mmol). After stirring for 10 min at the same temperature, chloromethyl methyl ether (6.65 mL, 87.5 mmol) was added drop wise and the reaction mixture was stirred for 10 h at room temperature. It was then poured into dilute HCl. The mixture was extracted with methylene chloride and the organic layers were combined and washed with brine, dried (Na_2SO_4), concentrated, and purified by flash column chromatography (hexanes/ethyl acetate) to afford the desired product **S3**.

2,4,8,10-Tetraoxaundecan-6-one (S1**):**

Ozone was passed through a methylene chloride-methanol (25 mL, 5:1) solution of **S3** (5.2 g, 29.5 mmol) at -78 °C, till the reaction mixture turned pale blue in color. Dimethyl sulfide (8.6 mL, 118 mmol) was then added to the reaction mixture and stirred for 12 h at room temperature. Solvents were removed under reduced pressure and the reaction mixture was purified by flash column chromatography (hexanes/ethyl acetate) to afford **S1** in 72% yield for two steps.

General procedure for Two-component Mannich-type reactions of DHA (Table 1 and Table 2):

Reactions were performed in a closed system (a vial with a cap). An inert atmosphere of nitrogen or argon was not necessary for the reactions. To a solution of *N*-PMP-protected preformed imine (0.5 mmol, 1 equiv) in anhydrous 1-methyl-2-pyrrolidinone (NMP, 0.5 mL) was added 1,3-dihydroxyacetone dimer (0.5 mmol, 1 equiv; 2 equiv as monomer) followed by O-*t*Bu-L-Thr (0.1 mmol, 0.2 equiv) and 5-methyl-1*H*-tetrazole (0.05 mmol, 0.1 equiv) at room temperature. The reaction was stirred at room temperature for the time as indicated in Table 1 and Table 2. The reaction mixture

was worked up by addition of saturated ammonium chloride, and extracted with AcOEt. The organic layers were washed with water and brine, dried over Na₂SO₄, filtered, concentrated in vacuo, and purified by flash column chromatography (AcOEt/hexane) to afford the corresponding Mannich adducts as an *anti/syn* mixture. Racemic standards were prepared using (\pm)-tryptophan and phenylphosphinic acid.

General procedure for Three-component Mannich reactions of DHA (Table 2, entries 5-7):

A mixture of NMP (0.5 mL), *p*-anisidine (0.5 mmol, 1 equiv) and aldehyde (0.5 mmol, 1 equiv), was stirred at 25°C for 0.5 h. Then, to the mixture, added 1,3-dihydroxyacetone dimer (0.5 mmol, 1 equiv; 2 equiv as monomer), O-*t*Bu-L-Thr (0.1 mmol, 0.2 equiv) and additive 5-methyl-1*H*-tetrazole (0.05 mmol, 0.1 equiv). The reaction was stirred at room temperature for the time as indicated in Table 2. The reaction mixture was worked up by addition of saturated ammonium chloride, and extracted with AcOEt. The combined organic layers were washed with water and brine, dried over Na₂SO₄, filtered, concentrated in vacuo, and purified by flash column chromatography (AcOEt/hexane) to afford the corresponding Mannich adducts as an *anti/syn* mixture. Racemic standards were prepared using (\pm)-tryptophan and phenylphosphinic acid.

General procedure for the O-*t*Bu-L-Thr-catalyzed Mannich reaction of Bn-protected DHA and N-PMP-protected preformed imine (Table 3).

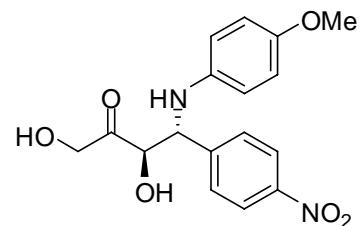
To a solution of Bn-protected-dihydroxyacetone (0.45 mmol, 1.5 equiv) in anhydrous NMP (0.3 mL), imine (0.3 mmol, 1 equiv) and O-*t*Bu-L-Thr (0.06 mmol, 0.2 equiv) were added followed by 5-methyl-1*H*-tetrazole (0.03 mmol, 0.1 equiv) at room temperature. The reaction was stirred at room temperature until the imine was consumed as monitored by TLC. The reaction mixture was worked up by addition of saturated ammonium chloride, and extracted with AcOEt. The organic layers were washed with water and brine, dried over Na₂SO₄, filtered, concentrated in vacuo, and purified by flash column chromatography (AcOEt/hexane) to afford the corresponding Mannich adducts as an *anti/syn* mixture. Racemic standards were prepared using (\pm)-tryptophan and phenylphosphinic acid.

General procedure for the O-*t*Bu-L-Thr-catalyzed Mannich reaction of MOM-protected DHA and N-PMP-protected preformed imine (Table 4).

To a solution of MOM-protected-dihydroxyacetone (0.5 mmol, 2 equiv) in anhydrous NMP (0.25 mL), imine (0.25 mmol, 1 equiv) and O-*t*Bu-L-Thr (0.05 mmol, 0.2 equiv) were added followed by 5-methyl-1*H*-tetrazole (0.025 mmol, 0.1 equiv) at room temperature. The reaction was stirred at room temperature until the imine was consumed as monitored by TLC. The reaction mixture was worked up by addition of saturated ammonium chloride, and extracted with AcOEt. The organic layers were washed with water and brine, dried over Na₂SO₄, filtered, concentrated in vacuo, and purified by flash column chromatography (AcOEt/hexane) to afford the corresponding Mannich adducts as an *anti/syn* mixture. Racemic standards were prepared using (\pm)-tryptophan and phenylphosphinic acid.

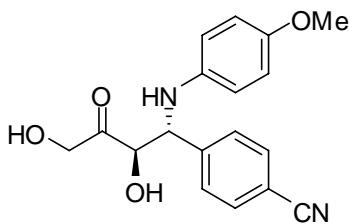
Characterizations of Mannich products 8 and 9 were previously reported.

(3*R*,4*R*)-1,3-Dihydroxy-4-(*p*-methoxyphenylamino)-4-(*p*-nitrophenyl)butan-2-one (1)



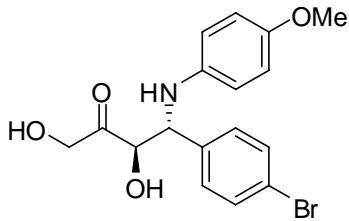
¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.69 (s, 3H), 4.10 (d, 1H, *J* = 20.0 Hz), 4.41 (d, 1H, *J* = 20.0 Hz), 4.77 (d, 1H, *J* = 4.5 Hz), 4.87 (d, 1H, *J* = 4.5 Hz), 6.55 (d, 2H, *J* = 9.0 Hz), 6.70 (d, 2H, *J* = 9.0 Hz), 7.46 (d, 2H, *J* = 8.5 Hz), 8.12 (d, 2H, *J* = 8.5 Hz). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 55.6, 60.6, 67.1, 77.1, 115.0, 116.0, 123.8, 128.7, 139.0, 147.6, 153.3, 210.2. HRMS: calcd for C₁₇H₁₉N₂O₆ (MH⁺) 347.1238, found 347.1236. HPLC (Daicel Chiraldpak OJ-H, hexane/*i*-PrOH = 75:25, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 78.4 min, t_R (*anti* minor enantiomer) = 52.2 min, t_R (*syn* major enantiomer) = 28.8 min, t_R (*syn* minor enantiomer) = 34.8 min.

(3*R*,4*R*)-4-(*p*-Cyanophenyl)-1,3-dihydroxy-4-(*p*-methoxyphenylamino)butan-2-one (2)



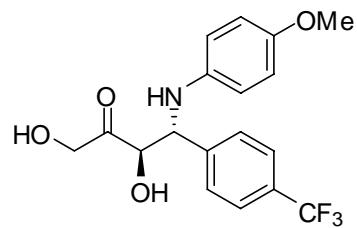
¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.69 (s, 3H), 4.08 (d, 1H, *J* = 20.0 Hz), 4.39 (d, 1H, *J* = 20.0 Hz), 4.74 (d, 1H, *J* = 4.0 Hz), 4.82 (d, 1H, *J* = 4.0 Hz), 6.54 (d, 2H, *J* = 9.0 Hz), 6.70 (d, 2H, *J* = 9.0 Hz), 7.41 (d, 2H, *J* = 8.5 Hz), 7.56 (d, 2H, *J* = 8.5 Hz). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 55.6, 60.8, 67.1, 77.1, 111.9, 114.9, 115.9, 118.4, 128.5, 132.4, 139.1, 143.2, 153.2, 210.3. HRMS: calcd for C₁₈H₁₉N₂O₄ (MH⁺) 327.1339, found 327.1340. HPLC (Daicel Chiraldpak OJ-H, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 82.4 min, t_R (*anti* minor enantiomer) = 76.5 min, t_R (*syn* major enantiomer) = 39.8 min, t_R (*syn* minor enantiomer) = 46.2 min.

(3*R*,4*R*)-4-(*p*-Bromophenyl)-1,3-dihydroxy-4-(*p*-methoxyphenylamino)butan-2-one (3)



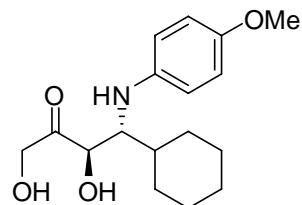
¹H NMR (500 MHz, CDCl₃): *anti*- δ 3.69 (s, 3H), 3.99 (d, 1H, *J* = 20.0 Hz), 4.31 (d, 1H, *J* = 20.0 Hz), 4.67 (d, 1H, *J* = 3.5 Hz), 4.71 (d, 1H, *J* = 3.5 Hz), 6.55 (d, 2H, *J* = 8.5 Hz), 6.70 (d, 2H, *J* = 8.5 Hz), 7.14 (d, 2H, *J* = 8.5 Hz), 7.41 (d, 2H, *J* = 8.5 Hz). *Syn*- δ 3.67 (s, 3H), 4.57 (m, 3H), 4.79 (s, 1H), 6.46 (d, 2H, *J* = 8.5 Hz), 6.66 (d, 2H, *J* = 8.5 Hz), 7.19 (d, 2H, *J* = 8.5 Hz), 7.43 (d, 2H, *J* = 8.5 Hz). ¹³C NMR (125 MHz, CDCl₃): mixture of diastereomers (*anti:syn* = 2:1) δ 55.6, 59.3, 60.6, 66.5, 67.1, 77.1, 78.8, 114.9, 115.5, 116.0, 1216, 122.2, 128.8, 129.2, 131.8, 136.3, 138.0, 139.4, 152.7, 153.0, 209.8, 210.6. HRMS: calcd for C₁₇H₁₉BrNO₄ (MH⁺) 380.0492, found 380.0493. HPLC (Daicel Chiraldpak AD, hexane/*i*-PrOH = 95:5, flow rate 0.8 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 93.9 min, t_R (*anti* minor enantiomer) = 79.7 min, t_R (*syn* major enantiomer) = 85.9 min, t_R (*syn* minor enantiomer) = 72.5 min.

(3*R*,4*R*)-1,3-Dihydroxy-4-(*p*-methoxyphenylamino)-4-(*p*-trifluoromethylphenyl)butan-2-one (4)



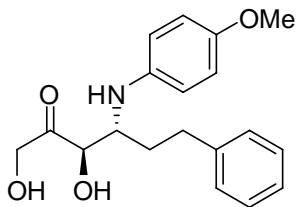
¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.70 (s, 3H), 4.07 (d, 1H, *J* = 20.0 Hz), 4.36 (d, 1H, *J* = 20.0 Hz), 4.74 (d, 1H, *J* = 4.0 Hz), 4.82 (d, 1H, *J* = 4.0 Hz), 6.57 (d, 2H, *J* = 9.0 Hz), 6.72 (d, 2H, *J* = 9.0 Hz), 7.41 (d, 2H, *J* = 8.5 Hz), 7.56 (d, 2H, *J* = 8.5 Hz). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 55.6, 60.9, 67.1, 77.1, 114.9, 116.0, 123.8 (q, *J* = 270.6 Hz), 125.7 (q, *J* = 3.8 Hz), 128.0, 130.4 (q, *J* = 32.5 Hz), 139.3, 141.4, 153.2, 210.2. HRMS: calcd for C₁₈H₁₉F₃NO₄ (MH⁺) 370.1261, found 370.1261. HPLC (Daicel Chiraldpak OJ-H, hexane/*i*-PrOH = 90:10, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 54.7 min, t_R (*anti* minor enantiomer) = 46.6 min, t_R (*syn* major enantiomer) = 28.6 min, t_R (*syn* minor enantiomer) = 33.3 min.

(3*R*,4*R*)-4-Cyclohexyl-1,3-dihydroxy-4-(4-methoxyphenylamino)butan-2-one (5)



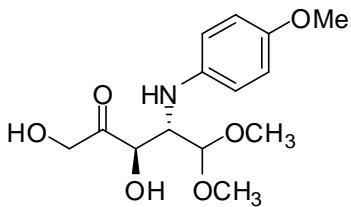
¹H NMR (400 MHz, CDCl₃): major diastereomer δ 0.99-1.30 (m, 5H), 1.50-1.90 (m, 6H), 3.20-3.50 (br s, 2H), 3.41 (t, 1H, *J* = 7.6 Hz), 3.50 (dd, 1H, *J* = 10.0 Hz, 2.8 Hz), 3.72 (s, 3H), 4.36 (d, 2H, *J* = 22.8 Hz), 4.44 (d, 1H, *J* = 4.0 Hz), 4.47 (d, 1H, *J* = 2.8 Hz), 6.50 (d, 2H, *J* = 12.0 Hz), 6.71 (2H, *J* = 11.6 Hz). ¹³C NMR (100 MHz, CDCl₃): major diastereomer δ 26.13, 26.17, 26.18, 26.23, 30.3, 41.2, 55.6, 61.1, 66.0, 75.2, 114.7, 114.9, 141.5, 152.2, 211.9. HRMS: calcd for C₁₇H₂₆NO₄ (MH⁺) 308.1856, found 308.1852. HPLC (Daicel Chiraldpak OJ-H, hexane/*i*-PrOH = 90:10, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 67.3 min, t_R (*anti* minor enantiomer) = 47.3 min, t_R (*syn* major enantiomer) = 31.4 min, t_R (*syn* minor enantiomer) = 28.0 min.

(3*R*,4*R*)-1,3-Dihydroxy-4-(4-methoxyphenylamino)-6-phenylhexan-2-one (6)



¹H NMR (400 MHz, CDCl₃): major diastereomer δ 1.46-1.60 (m, 1H), 1.78-2.10 (m, 1H), 2.56-2.90 (m, 2H), 3.06 (brs, 1H), 3.56 (td, 1H, J = 13.6 Hz, 3.3 Hz), 3.75 (s, 3H), 3.81 (d, 1H, J = 14.8 Hz), 4.44 (d, 2H, J = 12.4 Hz), 4.43 (dd, 1H, J = 17.6 Hz, 4.0 Hz), 6.61 (d, 2H, J = 12.0 Hz), 6.80 (d, 2H, J = 11.6 Hz), 7.10-7.32 (m, 5H). ¹³C NMR (100 MHz, CDCl₃): major diastereomer δ 31.4, 32.0, 55.7, 56.8, 66.6, 75.6, 115.1, 116.1, 126.21, 126.25, 128.3, 128.48, 128.54, 128.56, 211.0. HRMS: calcd for C₁₉H₂₄NO₄ (MH⁺) 330.1700, found 308.1709. HPLC (Daicel Chiralpak AD, hexane/i-PrOH = 93:7, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 30.1 min, t_R (*anti* minor enantiomer) = 47.1 min, t_R (*syn* major enantiomer) = 39.5 min, t_R (*syn* minor enantiomer) = 33.4 min.

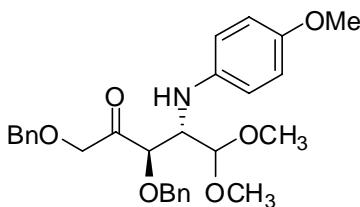
(3*R*,4*S*)-1,3-Dihydroxy-5,5-dimethoxy-4-(*p*-methoxyphenylamino)pentan-2-one (7)



¹H NMR (500 MHz, CDCl₃): mixture of diastereomers (*anti:syn* = 1:1) δ 3.32 (s, 3H x 1/2), 3.41 (s, 3H x 1/2), 3.43 (3H x 1/2), 3.45 (s, 3H x 1/2), 3.73 (s, 3H x 1/2), 3.74 (s, 3H x 1/2), 3.89 (t, 1H x 1/2, J = 3.0 Hz), 4.03 (dd, 1H x 1/2, J = 3.0 Hz, 5.0 Hz), 4.36-4.66 (m, 4H), 6.61 (d, 2H x 1/2, J = 9.0 Hz), 6.69 (d, 2H x 1/2, J = 9.0 Hz), 6.75 (d, 2H x 1/2, J = 9.0 Hz), 6.79 (d, 2H x 1/2, J = 9.0 Hz). ¹³C NMR (125 MHz, CDCl₃): mixture of diastereomers (*anti:syn* = 1:1) δ 55.0, 55.6, 56.4, 57.0, 57.3, 58.3, 58.4, 66.6, 67.2, 74.8, 75.4, 105.8, 105.9, 114.9, 115.0, 115.7, 115.8, 139.6, 140.1, 153.0, 153.2, 211.5, 212.1. HRMS: calcd for C₁₄H₂₂NO₆ (MH⁺) 300.1442, found 300.1442. HPLC (Daicel Chiralpak OJ-H, hexane/i-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 21.8 min, t_R (*anti* minor enantiomer) = 19.8 min, t_R (*syn* major enantiomer) = 27.9 min, t_R (*syn* minor enantiomer) =

16.7 min.

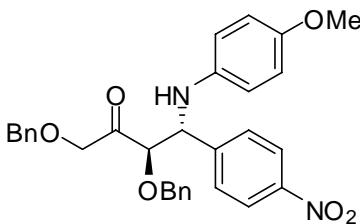
(3*R*,4*S*)-1,3-Bis(benzyloxy)-5,5-dimethoxy-4-(*p*-methoxyphenylamino)pentan-2-one(10)



¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.26 (s, 3H), 3.28 (s, 3H), 3.71 (s, 1H), 4.08 (dd, 1H, *J* = 3.5, 7.0 Hz, 4.18 (d, 1H, *J* = 17.0 Hz), 4.33 (d, 1H, *J* = 3.5 Hz), 4.35 (d, 1H, *J* = 4.5 Hz), 4.38(d, 1H, *J* = 10.0 Hz), 4.53-4.55 (m, 3H), 4.63 (d, 2H, *J* = 11.5 Hz), 6.55 (d, 2H, *J* = 9.0 Hz), 6.66 (d, 2H, *J* = 9.0 Hz), 7.21-7.35 (m, 10 Hz). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 52.2, 55.4, 55.7, 58.2, 73.4, 74.0, 74.4, 81.6, 102.7, 114.9, 115.1, 127.9, 128.0, 128.1, 128.4, 137.3, 137.4, 140.4, 152.6, 206.7. HRMS: calcd for C₂₈H₃₄NO₆ (MH⁺) 480.2381, found 480.2374. HPLC (Daicel Chiralcel AD, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm): t_R (*anti* major enantiomer) = 18.0 min; t_R (*anti* minor enantiomer) = 20.2 min, t_R (*syn* major enantiomer) = 11.8 min, t_R (*syn* minor enantiomer) = 20.2 min. Based on the dr determined by ¹H NMR and the ee of *syn* enantiomer which obtained from OD-H column (see the following), area of t_R 20.2 min was corrected to give ee value.

HPLC (Daicel Chiralcel OD-H, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm): t_R (*anti* enantiomer) = 16.9 min, t_R (*syn* major enantiomer) = 11.3 min, t_R (*syn* minor enantiomer) = 21.2 min.

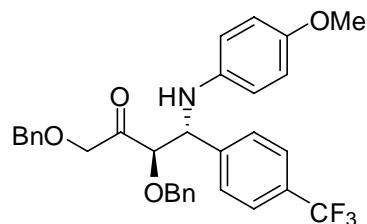
(3*R*,4*R*)-1,3-Bis(benzyloxy)-4-(*p*-methoxyphenylamino)-4-(*p*-nitrophenyl)butan-2-one (11)



¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.68 (s, 3H), 3.89 (d, 1H, *J* = 18 Hz), 4.11 (d, 1H, *J* = 18 Hz), 4.24 (brs, 1H), 4.33 (d, 1H, *J* = 11.5 Hz), 4.38 (d, 1H, *J* = 5.5 Hz), 4.46 (s, 2H), 4.57 (d, 1H, *J* = 11.5 Hz), 4.78 (d, 1H, *J* = 5.5 Hz), 6.37 (d, 2H, *J* = 9.0 Hz), 6.65 (d, 2H, *J* = 9.0 Hz), 7.16-7.34 (m,

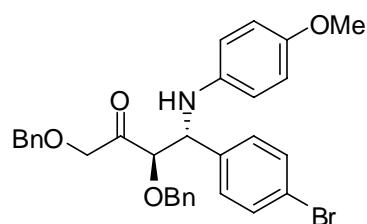
10H), 7.42 (d, 2H, J = 9.0 Hz), 8.08 (d, 2H, J = 9.0 Hz). ^{13}C NMR (125 MHz, CDCl_3): major diastereomer δ 55.6, 59.7, 73.3, 73.6, 74.0, 84.4, 114.8, 115.4, 123.6, 128.0, 128.2, 128.3, 128.6, 128.72, 128.74, 136.2, 136.5, 139.2, 146.4, 147.5, 152.9, 207.4. HRMS: calcd for $\text{C}_{31}\text{H}_{31}\text{N}_2\text{O}_6$ (MH^+) 527.2177, found 527.2176. HPLC (Daicel Chiralpak AD, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 44.8 min, t_R (*anti* minor enantiomer) = 37.3 min, t_R (*syn* major enantiomer) = 15.8 min, t_R (*syn* minor enantiomer) = 18.8 min.

(3*R*,4*R*)-1,3-Bis(benzyloxy)-4-(*p*-methoxyphenylamino)-4-(*p*-trifluoromethylphenyl)butan-2-one (12)



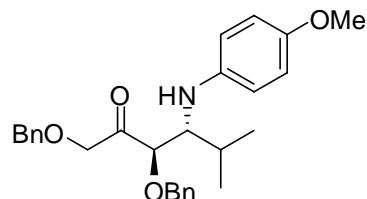
^1H NMR (500 MHz, CDCl_3): major diastereomer δ 3.67 (s, 3H), 3.88 (d, 1H, J = 18 Hz), 4.11 (d, 1H, J = 18 Hz), 4.29-4.37 (m, 4H), 4.44 (d, 1H, J = 12.0 Hz), 4.54 (d, 1H, J = 12.0 Hz), 4.74 (d, 1H, J = 6.0 Hz), 6.40 (d, 2H, J = 9.0 Hz), 6.65 (d, 2H, J = 9.0 Hz), 7.11-7.36 (m, 10H), 7.39 (d, 2H, J = 8.5 Hz), 7.51 (d, 2H, J = 8.5 Hz) ^{13}C NMR (125 MHz, CDCl_3): major diastereomer δ 55.5, 59.8, 73.3, 73.5, 73.9, 84.6, 114.8, 115.4, 124.0 (q, J = 270.4 Hz), 125.3 (q, J = 3.8 Hz), 127.9, 128.1, 128.2, 128.4, 128.5, 128.6, 129.9 (q, J = 32.0 Hz), 136.4, 136.7, 139.5, 142.9, 152.7, 207.7. HRMS: calcd for $\text{C}_{32}\text{H}_{31}\text{F}_3\text{NO}_4$ (MH^+) 550.2200, found 550.2200. HPLC (Daicel Chiralpak OD-H, hexane/*i*-PrOH = 50:50, flow rate 0.5 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 36.9 min, t_R (*anti* minor enantiomer) = 24.3 min, t_R (*syn* major enantiomer) = 18.6 min, t_R (*syn* minor enantiomer) = 28.8 min.

(3*R*,4*R*)-1,3-Bis(benzyloxy)-4-(*p*-bromophenyl)-4-(*p*-methoxyphenylamino)butan-2-one (13)



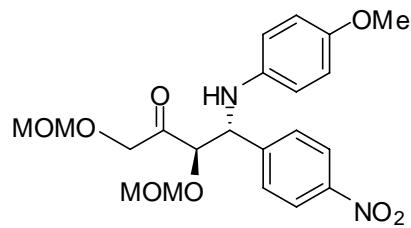
¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.62 (s, 3H), 3.79 (d, 1H, *J* = 18.5 Hz), 4.04 (d, 1H, *J* = 18.5 Hz), 4.26-4.31 (m, 3H), 4.38 (d, 1H, *J* = 11.5 Hz), 4.48 (d, 1H, *J* = 11.5 Hz), 4.60 (d, 1H, *J* = 5.0 Hz), 6.36 (d, 2H, *J* = 9.0 Hz), 6.60 (d, 2H, *J* = 9.0 Hz), 7.09-7.33 (m, 14H). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 55.6, 59.7, 73.2, 73.6, 73.9, 84.7, 114.8, 115.4, 121.8, 127.95, 128.04, 128.1, 128.4, 128.5, 128.6, 129.5, 131.6, 136.5, 136.8, 137.7, 139.6, 152.7, 207.9. HRMS: calcd for C₃₁H₃₁BrNO₄(MH⁺) 560.1431, found 560.1431. HPLC (Daicel Chiralpak AD, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 23.7 min, t_R (*anti* minor enantiomer) = 17.7 min, t_R (*syn* major enantiomer) = 10.4 min, t_R (*syn* minor enantiomer) = 13.7 min.

(3*R*,4*R*)-1,3-Bis(benzyloxy)-4-(*p*-methoxyphenylamino)-5-methylhexan-2-one (14)



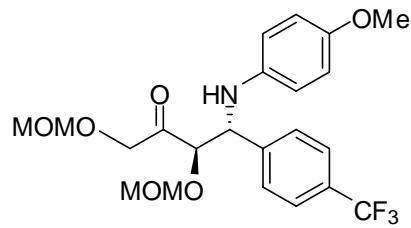
¹H NMR (500 MHz, CDCl₃): mixture of diastereomers (*anti:syn* = 2:1), * denotes *syn* diastereomer δ 0.88 (d, 3H* x 1/3, *J* = 6.8 Hz), 0.90 (d, 3H x 2/3, *J* = 6.8 Hz), 0.92 (d, 3H x 2/3, *J* = 6.8 Hz), 0.93 (d, 3H* x 1/3, *J* = 6.8 Hz), 1.95 (m, 1H* x 1/3), 2.00 (m, 1H x 2/3), 3.48 (d, 1H* x 1/3, *J* = 7.5 Hz), 3.63 (t, 1H x 2/3, *J* = 4.5 Hz), 3.70 (s, 1H* x 1/3), 3.71 (s, 1H x 2/3), 4.12-4.64 (m, 7H), 6.43 (d, 2H* x 1/3, *J* = 9.0 Hz), 6.49 (d, 2H x 2/3, *J* = 9.0 Hz), 6.67 (d, 2H* x 1/3, *J* = 9.0 Hz), 6.69 (d, 2H x 2/3, *J* = 9.0 Hz), 7.20-7.37 (m, 10 Hz). ¹³C NMR (125 MHz, CDCl₃): mixture of diastereomers (*anti:syn* = 2:1) δ 17.10, 19.83, 20.06, 20.71, 29.88, 31.77, 55.69, 61.21, 62.27, 73.13, 73.23, 73.37, 73.49, 73.57, 73.73, 74.33, 83.76, 83.53, 114.32, 114.87, 114.92, 127.83, 127.88, 127.90, 127.95, 128.05, 128.08, 128.21, 128.25, 128.39, 128.42, 128.49, 128.50, 128.51, 136.98, 137.04, 137.17, 137.20, 141.54, 142.14, 151.83, 152.28, 209.19, 209.91. HRMS: calcd for C₂₈H₃₄NO₄(MH⁺) 448.2482, found 448.2470. HPLC (Daicel Chiralpak AD, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 7.3 min, t_R (*anti* minor enantiomer) = 8.5 min, t_R (*syn* major enantiomer) = 10.8 min, t_R (*syn* minor enantiomer) = 6.2 min.

(3*R*,4*R*)-1,3-Bis(methoxymethyl)-4-(*p*-methoxyphenylamino)-4-(*p*-nitrophenyl)butan-2-one (15)



¹H NMR (500 MHz, CDCl₃): major diastereomer δ 3.30 (s, 3H), 3.34 (s, 3H), 3.68 (s, 3H), 4.02 (d, 1H, *J* = 18.3 Hz), 4.27 (d, 1H, *J* = 18.3 Hz), 4.50 (d, 1H, *J* = 5.8 Hz), 4.62 (m, 5H), 4.88 (s, 1H), 6.48 (d, 2H, *J* = 9.0 Hz), 6.68 (d, 2H, *J* = 9.0 Hz), 7.52 (d, 2H, *J* = 9.0 Hz), 8.16 (d, 2H, *J* = 9.0 Hz). ¹³C NMR (125 MHz, CDCl₃): major diastereomer δ 55.6, 55.7, 56.5, 59.7, 71.2, 83.7, 96.4, 97.9, 114.9, 115.3, 123.6, 128.7, 139.3, 146.4, 147.5, 152.9, 206.3. HRMS: calcd for C₂₁H₂₇N₂O₈ (MH⁺) 435.1762, found 435.1759. HPLC (Daicel Chiralcel AD, hexane/*i*-PrOH = 80:20, flow rate 1.0 mL/min, λ = 254 nm): t_R (*anti* major enantiomer) = 57.7 min, t_R (*anti* minor enantiomer) = 27.4 min, (*syn* major enantiomer) = 17.7 min, t_R (*syn* minor enantiomer) = 15.0 min.

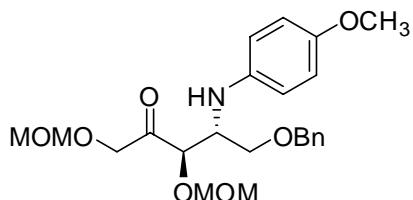
(3*R*,4*R*)-1,3-Bis(methoxymethyl)-4-(*p*-methoxyphenylamino)-4-(*p*-trifluoromethylphenyl)butan-2-one (16)



¹H NMR (400 MHz, CDCl₃): major diastereomer δ 3.27 (s, 3H), 3.33 (s, 3H), 3.70 (s, 3H), 3.76 (d, 1H, *J* = 14.8 Hz), 3.94 (d, 1H, *J* = 24.4 Hz), 4.21 (d, 1H, *J* = 24.4 Hz), 4.47-4.70 (m, 5H), 6.50 (d, 2H, *J* = 12.0 Hz), 6.70 (d, 2H, *J* = 12.0 Hz), 7.47 (d, 2H, *J* = 11.6 Hz), 7.57 (d, 2H, *J* = 11.6 Hz). ¹³C NMR (100 MHz, CDCl₃): major diastereomer δ 55.5, 55.6, 56.4, 59.8, 71.2, 83.8, 96.3, 97.8, 114.8, 115.0, 115.2, 125.4, 125.5, 127.5, 128.1, 139.6, 152.6, 206.5. HRMS: calcd for C₂₂H₂₇F₃NO₆ (MH⁺) 458.1785, found 458.1789. HPLC (Daicel Chiralpak AD, hexane/*i*-PrOH = 90:10, flow rate 1.0 mL/min, λ = 254

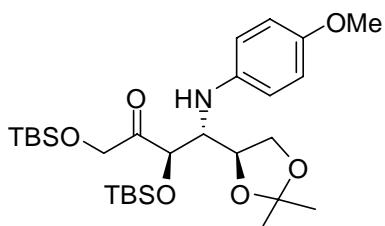
nm), t_R (*anti* major enantiomer) = 63.6 min, t_R (*anti* minor enantiomer) = 29.1 min, t_R (*syn* major enantiomer) = 15.8 min, t_R (*syn* minor enantiomer) = 14.0 min.

(3*R*,4*R*)-5-Benzyl-1,3-bis(methoxymethyl)-4-(*p*-methoxyphenylamino)pentan-2-one(17)



¹H NMR (400 MHz, CDCl₃): major diastereomer δ 3.30 (s, 3H), 3.35 (s, 3H), 3.51-3.63 (m, 1H), 3.73 (s, 3H), 4.00-4.13 (m, 1H), 4.30-4.70 (m, 10 H), 6.61 (d, 2H, J = 8.8 Hz), 6.75 (d, 2H, J = 8.8 Hz), 7.20-7.40 (m, 5H). ¹³C NMR (100 MHz, CDCl₃): major diastereomer δ 55.5, 55.7, 56.3, 56.6, 67.7, 71.6, 73.2, 80.4, 96.3, 97.8, 114.6, 114.9, 115.8, 127.7, 127.8, 137.7, 140.2, 152.8, 206.6. HRMS: calcd for C₂₃H₃₂NO₇ (MH⁺) 434.2173, found 434.2174. HPLC (Daicel Chiraldak AS-H, hexane/*i*-PrOH = 92:8, flow rate 1.0 mL/min, λ = 254 nm), t_R (*anti* major enantiomer) = 42.9 min, t_R (*anti* minor enantiomer) = 35.4 min, t_R (*syn* major enantiomer) = 30.0 min, t_R (*syn* minor enantiomer) = 19.2 min.

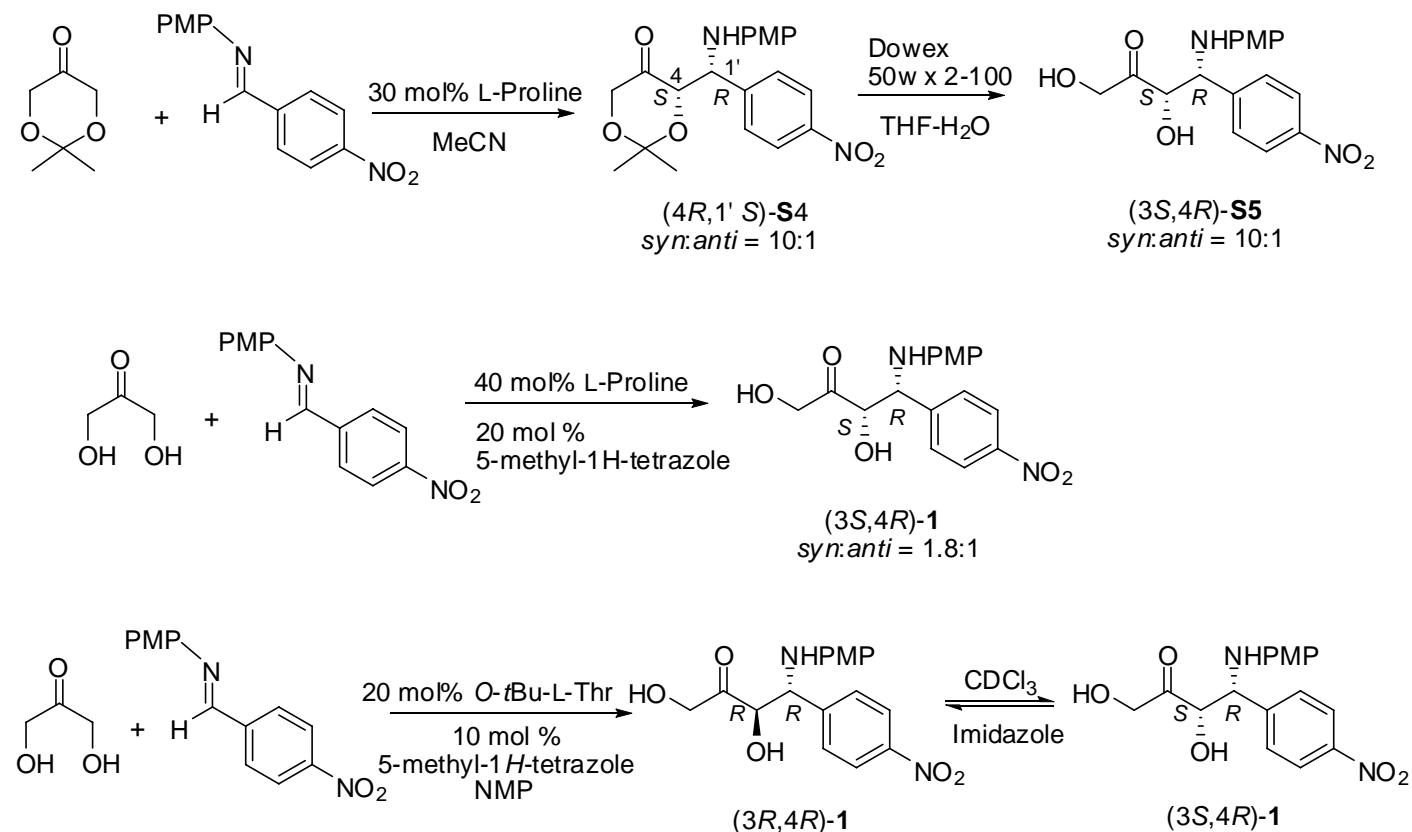
**(3*R*,4*R*)-1,3-Bis(*tert*-butyldimethylsiloxy)-4-[*(R*)-2,2-dimethyl-1,3-dioxolan-4-yl]-4-(*p*-methoxyphe
nylaminoo)-2-one (18)**



¹H NMR (500 MHz, CDCl₃): δ -0.05 (s, 3H), 0.02 (s, 3H), 0.08 (s, 3H), 0.09 (s, 3H), 0.91 (s, 9H), 0.96 (s, 9H), 1.30 (s, 3H), 1.35 (s, 3H), 3.41 (d, 1H, J = 10.0 Hz), 3.74 (s, 3H), 3.85 (dd, 1H, J = 9.0 Hz, 5.5 Hz), 3.89 (t, 1H, J = 9.0 Hz), 4.08 (dd, 1H, J = 8.5 Hz, 6.0 Hz), 4.14-4.19 (m, 1H), 4.37 (d, 1H, J = 17.3 Hz), 4.51 (d, 1H, J = 6.0 Hz), 4.75 (d, 1H, J = 17.3 Hz), 6.64 (d, 2H, J = 10.0 Hz), 6.77 (d, 2H, J = 10.0 Hz), ¹³C NMR (125 MHz, CDCl₃): δ -5.4, -5.3, -4.9, -4.7, 18.2, 18.5, 25.4, 25.8, 25.9, 26.2, 55.7, 60.9, 67.9, 68.4, 74.2, 76.7, 109.8, 114.9, 115.1, 139.8, 152.6, 209.3. HRMS: calcd for C₂₈H₅₂NO₆Si₂ (MH⁺) 554.3328, found 554.3337.

Determination of absolute stereochemistry of unprotected DHA *anti*-Mannich adduct 1:

Scheme S2

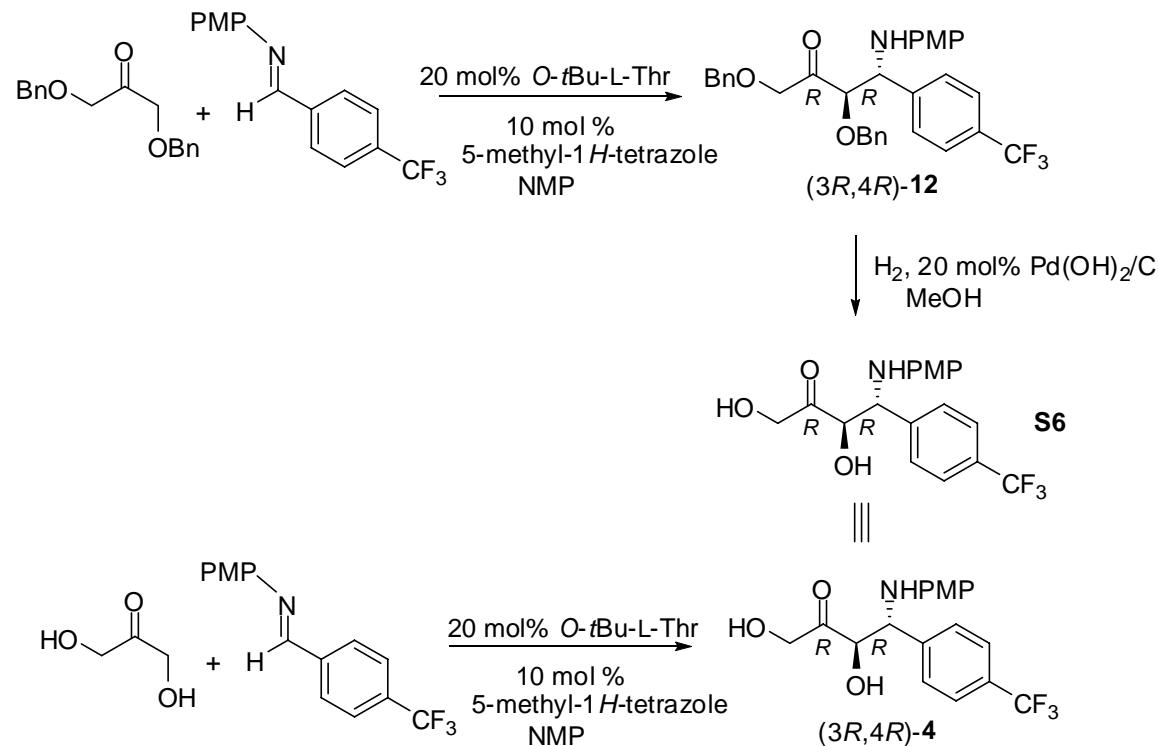


Syn-Mannich product (4*R*,1'*S*)-S4 (*syn:anti* = 10:1, *syn* 53% ee) was synthesized⁴ using (*S*)-proline, 2,2-dimethyl-1,3-dioxan-5-one, and *N*-PMP-protected preformed imine. This *syn*-Mannich product was transformed to (3*S*,4*R*)-S5 (*syn:anti* = 10:1, *syn* 54% ee) using reported⁵ procedures. Comparison of the HPLC traces confirmed the absolute configuration of Mannich product *syn*-1 obtained by the (*S*)-proline catalysis to be 3*R*,4*S*.

Imidazole isomerization⁶ of the *anti*-1 obtained from the O-*t*Bu-L-Thr-catalyzed reaction afforded the *syn*-product possessing a (3*R*,4*S*) configuration, which was the product of the (*S*)-proline-catalyzed reaction. This result confirmed that the major *anti*-product 1 generated from the O-*t*Bu-L-Thr-catalyzed reaction had a (3*R*,3*R*) configuration.

Determination of absolute stereochemistry of Bn-protected DHA *anti*-Mannich adduct 12:

Scheme S3



The Mannich Product **12** was hydrogenolyzed in methanol in the presence of 20% Pd(OH)₂/C under hydrogen to give the Mannich product **S6**. The major enantiomer of diol **S6** showed the same HPLC retention time as the major enantiomer of **4**, which was the O-*t*Bu-L-Thr catalyzed *anti*-Mannich adduct of dihydroxyacetone and *p*-trifluoromethylbenzaldehyde. Because the absolute configuration of **4** is (3*R*,4*R*) which had confirmed by the above method (See **Scheme 2**), major enantiomer of **12** must be (3*R*,4*R*).

References:

1. K. S. Kim, and S. D. Hong, *Tetrahedron Lett.*, 2000, **41**, 5909.
2. H. O. Kim, H. W. Baek, H. R. Moon, D.-K. Kim, M. W. Chun, and L. S. Jeong, *Org. Biomol. Chem.*, 2004, **2**, 1164.
3. M. Sodeoka, H. Yamada, and M. Shibasaki, *J. Am. Chem. Soc.*, 1990, **112**, 4906.
4. D. Enders, C. Grondal, M. Vrettou, and G. Raabe, *Angew. Chem. Int. Ed.*, 2005, **44**, 4079.
5. M. Kitamura, M. Isobe, Y. Ichikawa, and T. Goto, *J. Am. Chem. Soc.*, 1984, **106**, 3252.
6. D. E. Ward, M. Sales, and P. Sasikal, *J. Org. Chem.*, 2004, **69**, 4808.

