

Advanced
**Synthesis &
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Supporting Information

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**Enantioselective Michael Addition of Dicyanoolefins to α , β -Unsaturated Aldehydes
in Aqueous Medium**

Jun Lu, Feng Liu, Teck-Peng Loh *

Division of Chemistry and Biological Chemistry, School of Physical and Mathematical Sciences,
Nanyang Technological University. 1 Nanyang Link, Singapore 637371

Fax: (+65) 6791 1961 E-mail: teckpeng@ntu.edu.sg

Supporting Information

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General Methods:

Analytical thin layer chromatography (TLC) was performed using Merck 60 F254 precoated silica gel plate (0.2 mm thickness). Subsequent to elution, plates were visualized using UV radiation (254 nm) on Spectroline Model ENF-24061/F 254 nm. Further visualization was possible by staining with acidic solution of ceric molybdate or ethanol solution of ninhydrin.

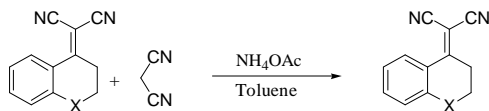
Flash chromatography was performed using Merck silica gel 60 with freshly distilled solvents. Columns were typically packed as slurry and equilibrated with the appropriate solvent system prior to use.

Infrared spectra were recorded on a Bio-Rad FTS 165 FTIR spectrometer. Liquid samples were examined as film between NaCl salt plates.

Proton nuclear magnetic resonance spectra (^1H NMR) were recorded on a Bruker Avance DPX 300 and Bruker AMX 400 spectrophotometer (CDCl_3 as solvent). Chemical shifts for ^1H NMR spectra are reported as δ in units of parts per million (ppm) downfield from SiMe_4 (δ 0.0) and relative to the signal of chloroform-*d* (δ 7.2600, singlet). Multiplicities were given as: s (singlet); d (doublet); t (triplet); q (quartet); or m (multiplets). The number of protons (n) for a given resonance is indicated by $n\text{H}$. Coupling constants are reported as a J value in Hz. Carbon nuclear magnetic resonance spectra (^{13}C NMR) are reported as δ in units of parts per million (ppm) downfield from SiMe_4 (δ 0.0) and relative to the signal of chloroform-*d* (δ 77.03, triplet). The proportion of diastereomers was determined from the integration of ^1H NMR and ^{13}C NMR spectra.

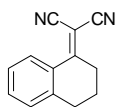
Mass spectral analyses were carried out on a VG 7035 micromass mass spectrophotometer at a source temperature of 200°C and at an ion current of 70 eV. Mass spectral data were reported in units of mass to charge (m/z) and % intensity.

General procedures for the preparation of olefins:



Malononitrile (25 mmol) and the ketone (27.5 mmol) were dissolved in 30 mL of toluene containing ammonium acetate (500 mg) and glacial acetic acid (2 mL) in a 50 mL flask. By refluxing vigorously, the water formed in the reaction was removed by a Dean and Stark trap placed under the reflux condenser. After the completion of the reaction, the reaction mixture was diluted with EtOAc and washed with brine. The organic phase was dried over Na₂SO₄ and the solvent was evaporated under reduced pressure. The residue was recrystallized from alcohol to give pure products.

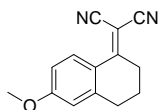
2-(2,3-Dihydronaphthalen-4(1H)-ylidene) malononitrile



¹H NMR (400 MHz, CDCl₃): *d* 8.20 (d, *J* = 10.5 Hz, 1H), 7.52-7.46 (m, 1H), 7.36-7.26 (m, 2H), 3.02 (t, *J* = 8.7 Hz, 2H), 2.89 (t, *J* = 8.3 Hz, 2H), 2.02 (t, *J* = 8.6 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): *d* 172.6, 142.0, 133.7, 130.0, 129.5, 127.9, 126.9, 114.0, 113.4, 79.8, 33.1, 29.7, 22.2.

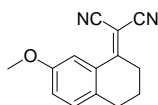
2-(6-Methoxy-2,3-dihydronaphthalen-4(1H)-ylidene) malononitrile



¹H NMR (400 MHz, CDCl₃): *d* 8.30 (d, *J* = 8.9 Hz, 1H), 6.86 (d, *J* = 8.9 Hz, 1H), 6.76 (s, 1H), 3.88 (s, 3H), 2.99 (t, *J* = 6.4 Hz, 2H), 2.85 (t, 6.2 Hz, 2H), 1.99-1.93 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): *d* 171.1, 163.5, 146.9, 130.4, 129.6, 126.3, 114.2, 113.0, 112.9, 55.4, 38.9, 30.2, 23.4.

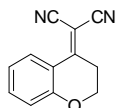
2-(7-Methoxy-2,3-dihydronaphthalen-4(1H)-ylidene) malononitrile



¹H NMR (400 MHz, CDCl₃): *d* 7.73 (d, *J* = 2.4 Hz, 1H), 7.18 (d, *J* = 8.5 Hz, 1H), 7.06 (dd, *J* = 8.5, 2.5 Hz, 1H), 3.84 (s, 3H), 3.00 (t, *J* = 6.5 Hz, 2H), 2.83 (t, *J* = 6.2 Hz, 2H), 1.99-1.96 (m, 2H).

¹³C NMR (100 MHz, CDCl₃): *d* 172.6, 157.9, 134.5, 130.5, 130.4, 121.9, 114.2, 113.5, 110.9, 79.6, 55.7, 33.1, 28.9, 22.5.

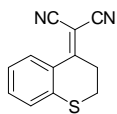
2-(2,3-Dihydrochromen-4-ylidene) malononitrile



¹H NMR (400 MHz, CDCl₃): *d* 8.36 (dd, *J* = 8.2, 1.4 Hz, 1H), 7.53-7.48 (m, 1H), 7.07-7.03 (m, 1H), 6.98 (dd, *J* = 8.4, 0.9 Hz, 1H), 4.39 (t, *J* = 6.3 Hz, 2H), 3.15 (t, *J* = 6.3 Hz, 2H).

¹³C NMR (100 MHz, CDCl₃): *d* 163.4, 157.4, 136.8, 127.3, 121.7, 118.7, 117.2, 113.8, 113.2, 64.7, 30.9.

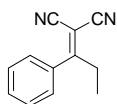
2-(2,3-Dihydrothiochromen-4-ylidene) malononitrile



^1H NMR (400 MHz, CDCl_3): d 7.96 (d, $J = 8.0$ Hz, 1H), 7.42-7.40 (m, 1H), 7.38-7.27 (m, 1H), 7.22-7.18 (m, 1H), 3.31 (t, $J = 6.5$ Hz, 2H), 3.16 (t, $J = 6.4$ Hz, 2H).

^{13}C NMR (100 MHz, CDCl_3): d 169.8, 138.9, 133.5, 129.4, 128.3, 128.0, 125.0, 113.4, 112.8, 81.9, 32.7, 25.7.

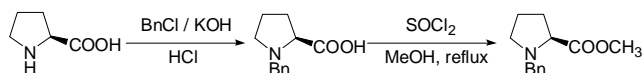
2-(1-Phenylpropylidene)malononitrile



^1H NMR (400 MHz, CDCl_3): d 7.56-7.46 (m, 5H), 2.98 (q, $J = 7.5$ Hz, 2H), 1.11 (t, $J = 7.5$ Hz, 3H).

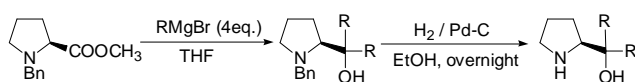
^{13}C NMR (100 MHz, CDCl_3): d 181.7, 134.6, 132.0, 129.2, 127.5, 112.8, 112.5, 84.3, 31.1, 12.8.

Preparation of dialkyl-(S)-pyrrolidin-2-yl-methanol



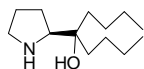
A solution of 17.3 g (0.15 mol) of L-proline and 25.4 g (0.45 mol) of KOH in isopropanol (100 mL) was prepared with stirring at 40°C. As soon as the solution became transparent, slow addition of freshly distilled BnCl (18.9 mL, 0.16 mol) was commenced under stirring at the same temperature. The dropwise addition took 3 h to complete, and the stirring was continued for another 6 h at the same temperature. The reaction mixture was neutralized with concentrated aqueous HCl until pH 5–6, then was added to the reaction mixture CHCl₃ (40 ml) with stirring. The mixture was left overnight, then filtered and the precipitate was washed with CHCl₃. The CHCl₃ solutions were combined and evaporated, the residue was treated with acetone and the precipitate of crude BP filtered and additionally washed with acetone. The solid was dried over P₂O₅ to yield the unpurified benzylated acid (29.2g, unpurified yield), which was used further without any purification.

SOCl₂ (4 mL, 55 mmol) was added during 10 min at 0 °C to a mixture of N-benzyl-L-proline (12 g, 50 mmol) in 40 mL MeOH and afterwards stirred at reflux for 1h. MeOH and excess of SOCl₂ were evaporated under reduced pressure; EA (150 mL) was added and washed with saturated sodium bicarbonate solution (50 mL×4). The organic phase was dried over anhydrous NaSO₄ and evaporated under reduce pressure, remaining a brown oil. The product was used in the following step with further purification. Yield: 9.9 g (90 %). ¹HNMR (400 MHz, CDCl₃): δ 7.38-7.23 (m, 5H), 3.83 (d, J = 29.7 Hz, 1H), 3.59 (s, 3H), 3.46 (d, J = 35 Hz, 1H), 3.26-3.23 (m, 1H), 3.08-3.04 (m, 1H), 2.42-2.36 (m, 1H), 2.18-2.08 (m, 1H), 2.00-1.86 (m, 2H), 1.81-1.74 (m, 1H).



The corresponding alkylmagnesium bromide (80 mmol) in THF was cooled at 0°C and (S)-methyl 1-benzylpyrrolidine-2-carboxylate (20 mmol in THF) was added dropwise over 20 min. Then the reaction mixture was stirred overnight at rt. The reaction was quenched with saturated NH₄Cl. The supernatant liquid was collected leaving behind a white precipitate which was extracted with dichloromethane. The combined organic extracts were washed with brine and dried over anhydrous Na₂SO₄. The solvent was evaporated under reduced pressure and the product obtained was used in the following step without further purification.

The crude dialkyl-(1-benzylpyrrolidin-2-yl)methanol (10 mmol) was dissolved in 10 mL of ethanol and palladium, (10 wt. % on activated carbon, 20% mol % weight) was added. The reaction mixture was stirred under hydrogen atmosphere (1 atm) at room temperature overnight. The solution was filtered over celite in a Büchner funnel and the solvent evaporated under reduced pressure. The crude obtained was purified by column chromatography.

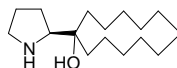


[α]_D²⁵ = -31.4 (c = 2.5, DCM)

¹HNMR (400 MHz, CDCl₃) δ: 3.49-3.36 (m, 1H), 3.08-2.92 (m, 1H), 2.90-2.76 (m, 2H), 1.75-1.58 (m, 4H), 1.40-1.30 (m, 3H), 1.35-1.06 (m, 8H), 0.79 (t, J = 6.5 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 73.33, 64.32, 46.42, 37.36, 34.74, 25.82, 25.79, 25.57, 25.19, 23.36, 23.34, 14.03, 13.95.

MS: C₁₃H₂₇NO 214.20 [M+H]⁺.



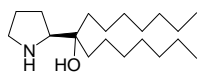
[α]_D²⁵ = -15.4 (c 1.6, DCM).

¹HNMR (400 MHz, CDCl₃) δ: 3.14-3.08 (m, 1H), 2.99-2.91 (m, 2H), 1.80-1.53 (m, 4H), 1.49-1.18 (m, 20H), 0.89 (t, J = 6.8 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) δ: 73.48, 64.28, 46.49, 37.80, 35.05, 31.85, 31.76, 30.06, 30.04, 25.93, 25.25, 23.63,

23.39, 22.65, 22.62, 14.05.

MS: C₁₇H₃₅NO 270.25 [M+H]⁺.

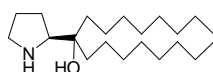


[a]_D²⁵ = -13.8 (c = 2.3, DCM)

¹H-NMR (400 MHz, CDCl₃) d: 3.15-3.12 (m, 1H), 3.04-2.93 (m, 2H), 1.82-1.70 (m, 3H), 1.57-1.43 (m, 3H), 1.40-1.06 (m, 22H), 0.88 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) d: 73.52, 64.39, 46.46, 37.78, 35.07, 31.90, 31.86, 29.71, 29.66, 29.34, 29.24, 29.11, 25.90, 25.29, 23.70, 23.44, 22.69, 14.11.

MS: C₁₉H₃₉NO 298.26 [M+1]⁺.



[a]_D²⁵ = -10.4 (c = 2.2, DCM)

¹H-NMR (400 MHz, CDCl₃) d: 3.29-3.23 (m, 1H), 3.08-2.98 (m, 2H), 1.90-1.74 (m, 3H), 1.58-1.44 (m, 3H), 1.38-1.12 (m, 26H), 0.86 (t, J = 6.7 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) d: 73.55, 65.08, 46.18, 37.48, 35.15, 31.92, 31.88, 30.33, 30.30, 29.70, 29.63, 29.36, 29.28, 25.45, 25.32, 23.63, 23.39, 22.69, 22.67, 14.12.

MS: C₂₁H₄₃NO 326.35 [M+H]⁺.



[a]_D²⁵ = -6.4 (c = 1.8, DCM)

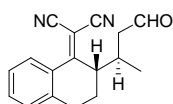
¹H-NMR (400 MHz, CDCl₃) d: 3.18-3.04 (m, 1H), 2.96-2.86 (m, 2H), 1.72-1.61 (m, 3H), 1.40-1.22 (m, 37H), 0.87 (t, J = 7.2 Hz, 6H).

¹³C-NMR (100 MHz, CDCl₃) d: 73.50, 64.04, 46.61, 37.95, 35.02, 32.86, 31.96, 30.47, 30.44, 29.73, 29.69, 29.61, 29.48, 29.39, 26.13, 25.80, 25.26, 23.73, 23.47, 22.71, 14.09.

MS: C₂₅H₅₁NO 382.33 [M+H]⁺.

Direct Vinylogous Michael Addition Reaction:

Typical Procedure (Table 2, entry 1): A mixture of **6a** (19.4 mg, 0.1 mmol), **7a** (32 μ L, 0.4 mmol), cat. (0.02 mmol) and PNBA (3.4 mg, 0.02 mmol) in brine (0.5 mL) was stirred for 36 h at 0°C. Then the reaction was quenched by adding 0.5 mL 1M HCl. The mixture was extracted with EtOAc, dried with anhydrous sodium sulfate. The crude product was purified by column chromatography on silica gel to give the desired product **8aa** in 82% yield with 90% ee, determined by HPLC (Chiralpak AS, 0% 2-propanol/hexane, 1 mL/min): $t_{\text{minor}} = 18.756$ min, $t_{\text{major}} = 26.674$ min.



8aa $[\alpha]_D^{22} = -368$ (c 1.02, DCM), 90% ee;

$R_f = 0.32$ (Hex:EA = 4 : 1);

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.64 (s, 1H), 7.93 (d, $J = 7.9$ Hz, 1H), 7.50 (t, $J = 7.5$ Hz, 1H), 7.34 (t, $J = 7.6$ Hz, 1H), 7.28-7.26 (m, 1H), 3.04-3.02 (m, 1H), 3.00-2.96 (m, 1H), 2.91-2.84 (m, 1H), 2.45-2.41 (m, 2H), 2.38-2.19 (m, 2H), 2.12-2.04 (m, 1H), 1.05 (d, $J = 6.6$ Hz, 3H);

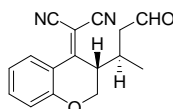
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.4, 177.4, 140.2, 133.8, 129.5, 129.2, 128.3, 127.0, 113.5, 113.3,

81.1, 48.7, 47.1, 28.8, 24.9, 24.5, 17.3;

IR (KBr): 2226, 1762 cm^{-1} ;

MS: $\text{C}_{17}\text{H}_{16}\text{N}_2\text{O}$ 264.12 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 18.756$ min, $t_{\text{major}} = 26.674$ min.



8ba $[\alpha]_D^{22} = -475$ (c 0.98, DCM), 92% ee;

$R_f = 0.24$ (Hex:EA = 4 : 1);

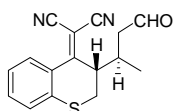
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.63 (s, 1H), 8.13 (d, $J = 8.2$ Hz, 1H), 7.53 (t, $J = 7.8$ Hz, 1H), 7.06 (t, $J = 7.6$ Hz, 1H), 6.96 (d, $J = 8.2$ Hz, 1H), 4.68 (dd, $J = 1.4, 11.2$ Hz, 1H), 4.26 (dd, $J = 2.1, 12.6$ Hz, 1H), 2.85 (d, $J = 10.1$ Hz, 1H), 2.53-2.33 (m, 3H), 1.19 (d, $J = 6.5$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 199.9, 167.9, 156.2, 136.9, 127.9, 121.9, 118.2, 115.7, 113.2, 78.5, 66.3, 48.0, 44.5, 27.8, 17.9;

IR (KBr): 2225, 1720 cm^{-1} ;

MS: $\text{C}_{16}\text{H}_{14}\text{N}_2\text{O}_2$ 266.09 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 16.176$ min, $t_{\text{major}} = 21.493$ min.



8ca $[\alpha]_D^{22} = -514$ (c 0.89, DCM), 91% ee;

$R_f = 0.16$ (Hex:EA = 4 : 1);

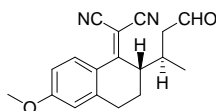
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.58 (s, 1H), 7.80 (d, $J = 7.9$ Hz, 1H), 7.39 (t, $J = 7.6$ Hz, 1H), 7.23-7.15 (m, 2H), 3.48-3.43 (m, 1H), 3.27-3.22 (m, 2H), 2.47-2.23 (m, 3H), 1.15 (d, $J = 6.3$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 199.8, 173.4, 138.2, 133.8, 130.3, 127.2, 125.6, 124.9, 113.0, 112.6, 82.6, 48.1, 43.9, 28.5, 27.9, 17.5;

IR (KBr): 2229, 1721 cm^{-1} ;

MS: $\text{C}_{16}\text{H}_{14}\text{N}_2\text{OS}$ 282.09 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 21.464$ min, $t_{\text{major}} = 39.825$ min.



8da $[\alpha]_D^{22} = -171$ (c 0.85, DCM), 88% ee;

$R_f = 0.18$ (Hex:EA = 4 : 1);

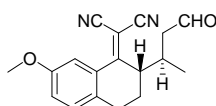
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.62 (s, 1H), 7.78 (d, $J = 8.8$ Hz, 1H), 6.84 (d, $J = 8.8$ Hz, 1H), 6.74 (s, 1H), 3.86 (s, 3H), 3.06-2.82 (m, 3H), 2.45-2.38 (m, 2H), 2.36-2.21 (m, 2H), 2.09-1.99 (m, 1H), 1.04 (d, $J = 6.6$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.6, 176.4, 163.9, 142.8, 130.6, 121.9, 114.4, 114.2, 113.9, 113.1, 77.8, 55.6, 48.6, 47.0, 28.6, 25.1, 24.5, 17.5;

IR (KBr): 2224, 1725 cm^{-1} ;

MS: $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}_2$ 294.14 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 20.088$ min, $t_{\text{major}} = 24.691$ min.



8ea $[\alpha]_D^{22} = -48$ (c = 0.32, DCM), 87% ee;

$R_f = 0.17$ (Hex:EA = 4 : 1);

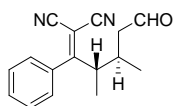
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.64 (s, 1H), 7.42 (d, $J = 2.5$ Hz, 1H), 7.16 (d, $J = 8.5$ Hz, 1H), 7.08-7.05 (m, 1H), 3.84 (s, 3H), 3.11-3.06 (m, 1H), 2.98-2.89 (m, 1H), 2.83-2.77 (m, 1H), 2.45-2.35 (m, 1H), 2.31-2.19 (m, 2H), 2.10-2.01 (m, 1H), 1.06 (d, $J = 6.5$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.5, 177.5, 157.9, 132.3, 130.6, 129.6, 121.7, 113.6, 113.3, 111.6, 80.7, 55.7, 48.7, 47.1, 28.8, 24.9, 24.1, 17.4;

IR (KBr): 2221, 1724 cm^{-1} ;

MS: $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}_2$ 294.4 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 12.136$ min, $t_{\text{major}} = 22.946$ min.



8fa [a]²⁴_D = -19 (c = 0.26, DCM), 75% ee;

R_f = 0.29 (Hex:EA = 4 : 1);

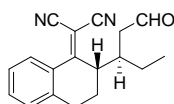
¹H NMR (400 MHz, CDCl₃): δ 9.75 (s, 1H), 7.53-7.48 (m, 3H), 7.30-7.28 (m, 2H), 3.16-3.09 (m, 1H), 2.63-2.58 (m, 1H), 2.39-2.21 (m, 2H), 1.22 (d, J = 6.9 Hz, 3H), 1.07 (d, J = 6.5 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃): δ 200.6, 183.9, 134.1, 131.0, 129.1, 127.1, 111.9, 88.4, 48.0, 47.3, 31.2, 19.3, 17.3;

IR (KBr): 2228, 1726 cm⁻¹;

MS: C₁₆H₁₆N₂O 252.14 [M]⁺.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min) after converted to the alcohol, *t*_{minor} = 10.742 min, *t*_{major} = 13.389 min.



8ab [a]²³_D = -269 (c 0.58, DCM), 84% ee;

R_f = 0.39 (Hex:EA = 4 : 1);

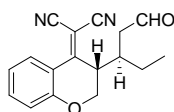
¹H NMR (400 MHz, CDCl₃): δ 9.61 (s, 1H), 7.91 (d, J = 7.8 Hz, 1H), 7.50 (t, J = 7.1 Hz, 1H), 7.35-7.26 (m, 2H), 3.27-3.23 (m, 1H), 2.98-2.88 (m, 2H), 2.56-2.49 (m, 1H), 2.22-2.16 (m, 2H), 2.11-2.04 (m, 1H), 1.61-1.54 (m, 1H), 1.49-1.42 (m, 1H), 1.36-1.28 (m, 1H), 0.89 (t, J = 7.5 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃): δ 200.6, 177.5, 140.2, 133.7, 129.4, 129.3, 128.5, 126.9, 113.5, 113.3, 80.9, 45.2, 44.7, 34.0, 25.1, 24.6, 23.1, 9.6;

IR (KBr): 2219, 1743 cm⁻¹;

MS: C₁₈H₁₈N₂O 301.11 [M+Na]⁺.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), *t*_{minor} = 14.866 min, *t*_{major} = 18.309 min.



8bb [a]²⁴_D = -291 (c = 2.4, DCM), 88% ee;

R_f = 0.45 (Hex:EA = 4 : 1);

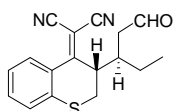
¹H NMR (400 MHz, CDCl₃): δ 9.58 (s, 1H), 8.11 (d, J = 8.2 Hz, 1H), 7.51 (t, J = 7.8 Hz, 1H), 7.04 (t, J = 8.2 Hz, 1H), 6.98 (d, J = 7.8 Hz, 1H), 4.64 (dd, J = 1.7, 12.6 Hz, 1H), 4.24 (dd, J = 2.2, 12.6 Hz, 1H), 2.98 (d, J = 8.3 Hz, 1H), 2.60-2.53 (m, 1H), 2.38-2.23 (m, 2H), 1.75-1.68 (m, 2H), 0.93 (t, J = 7.5 Hz, 3H);

¹³C NMR (100 MHz, CDCl₃): δ 199.9, 168.1, 156.2, 136.9, 127.9, 121.8, 118.1, 115.8, 113.2, 113.1, 78.5, 66.3, 44.6, 42.1, 32.6, 23.5, 9.46;

IR (KBr): 2226, 1719 cm⁻¹;

MS: C₁₇H₁₆N₂O₂ 280.12 [M]⁺.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), *t*_{minor} = 14.009 min, *t*_{major} = 19.113 min.



8cb $[\alpha]_D^{24} = -492$ ($c = 1.3$, DCM), 90% ee;

$R_f = 0.27$ (Hex:EA = 4 : 1);

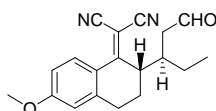
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.54 (s, 1H), 7.78 (d, $J = 8.0$ Hz, 1H), 7.40 (t, $J = 7.6$ Hz, 1H), 7.23-7.16 (m, 2H), 3.40 (dd, $J = 1.8, 12.5$ Hz, 1H), 3.21 (dd, $J = 2.3, 12.6$ Hz, 1H), 2.48-2.36 (m, 2H), 2.23-2.18 (m, 1H), 1.73-1.54 (m, 3H), 0.96 (t, $J = 7.5$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 199.9, 173.3, 138.2, 133.8, 130.4, 127.1, 125.8, 124.9, 113.0, 112.6, 82.7, 44.5, 41.4, 32.7, 28.5, 23.1, 9.4;

IR (KBr): 2223, 1725 cm^{-1} ;

MS: $\text{C}_{17}\text{H}_{16}\text{N}_2\text{OS}$ 296.12 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 19.394$ min, $t_{\text{major}} = 31.557$ min.



8db $[\alpha]_D^{24} = -198$ ($c = 2.1$, DCM), 84% ee;

$R_f = 0.22$ (Hex:EA = 4 : 1);

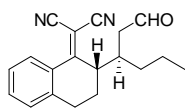
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.60 (s, 1H), 7.94 (d, $J = 8.9$ Hz, 1H), 6.84 (dd, $J = 2.6, 8.9$ Hz, 1H), 6.74 (s, 1H), 3.86 (s, 3H), 3.21-3.16 (m, 1H), 2.98-2.94 (m, 1H), 2.88-2.82 (m, 1H), 2.55-2.49 (m, 1H), 2.28-2.18 (m, 3H), 2.06-2.01 (m, 1H), 1.61-1.55 (m, 1H), 1.50-1.42 (m, 1H), 0.89 (t, $J = 7.5$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.7, 176.5, 163.9, 142.7, 130.7, 122.1, 114.3, 114.2, 113.9, 113.0, 77.9, 55.6, 45.2, 44.5, 33.7, 25.3, 24.5, 23.1, 9.4;

IR (KBr): 2225, 1726 cm^{-1} ;

MS: $\text{C}_{19}\text{H}_{20}\text{N}_2\text{O}_2$ 308.13 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 19.145$ min, $t_{\text{major}} = 21.763$ min.



8ac $[\alpha]_D^{23} = -269$ ($c = 0.58$, DCM), 85% ee;

$R_f = 0.59$ (Hex:EA = 4 : 1);

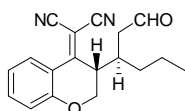
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.61 (s, 1H), 7.89 (d, $J = 7.9$ Hz, 1H), 7.50 (t, $J = 7.2$ Hz, 1H), 7.33 (t, $J = 7.6$ Hz, 1H), 7.26 (d, $J = 8.5$ Hz, 1H), 3.28-3.23 (m, 1H), 3.01-2.94 (m, 1H), 2.89-2.82 (m, 1H), 2.53-2.45 (m, 1H), 2.30-2.23 (m, 2H), 2.19-2.13 (m, 1H), 2.11-2.04 (m, 1H), 1.46-1.28 (m, 4H), 1.27-1.19 (m, 1H), 0.89 (t, $J = 7.0$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.6, 177.5, 140.2, 133.7, 129.5, 129.4, 128.4, 126.9, 113.5, 113.3, 81.0, 46.0, 45.3, 33.2, 32.9, 25.2, 24.5, 18.8, 14.3;

IR (KBr): 2225, 1730 cm^{-1} ;

MS: $\text{C}_{19}\text{H}_{20}\text{N}_2\text{O}$ 292.10 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 12.974$ min, $t_{\text{major}} = 15.776$ min.



8bc $[\alpha]_{\text{D}}^{23} = -529$ (c 0.63, DCM), 87% ee;

$R_f = 0.35$ (Hex:EA = 4 : 1);

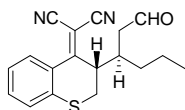
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.57 (s, 1H), 8.08 (d, $J = 7.1$ Hz, 1H), 7.51 (t, $J = 8.4$ Hz, 1H), 7.05 (t, $J = 7.5$ Hz, 1H), 6.95 (d, $J = 8.4$ Hz, 1H), 4.64 (dd, $J = 1.4, 12.3$ Hz, 1H), 4.24 (dd, $J = 2.2, 12.6$ Hz, 1H), 2.98 (d, $J = 9.9$ Hz, 1H), 2.57-2.49 (m, 1H), 2.39-2.32 (m, 2H), 1.65-1.42 (m, 2H), 1.40-1.24 (m, 2H), 0.93 (t, $J = 7.3$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.0, 168.2, 156.2, 136.9, 127.9, 121.8, 118.1, 115.8, 113.2, 78.6, 66.4, 45.3, 42.7, 33.4, 31.8, 18.8, 14.2;

IR (KBr): 2228, 1744 cm^{-1} ;

MS: $\text{C}_{18}\text{H}_{18}\text{N}_2\text{O}_2$ 294.14 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 12.723$ min, $t_{\text{major}} = 16.203$ min.



8cc $[\alpha]_{\text{D}}^{23} = -278$ (c 2.8, DCM), 84% ee;

$R_f = 0.28$ (Hex:EA = 4 : 1);

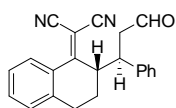
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.54 (s, 1H), 7.77 (d, $J = 8.0$ Hz, 1H), 7.40 (t, $J = 7.6$ Hz, 1H), 7.24-7.16 (m, 2H), 3.45-3.42 (m, 2H), 3.24-3.20 (m, 1H), 2.45-2.42 (m, 2H), 2.27-2.19 (m, 1H), 1.38-1.14 (m, 4H), 0.92 (t, $J = 7.1$ Hz, 3H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 200.7, 174.1, 137.8, 133.6, 130.2, 127.2, 126.1, 125.1, 113.1, 112.5, 82.9, 44.5, 42.7, 34.2, 32.7, 28.8, 19.6, 13.9;

IR (KBr): 2221, 1724 cm^{-1} ;

MS: $\text{C}_{18}\text{H}_{18}\text{N}_2\text{OS}$ 310.12 $[\text{M}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 17.132$ min, $t_{\text{major}} = 24.571$ min.



8ad $[\alpha]_{\text{D}}^{23} = -176$ (c 1.8, DCM), 85% ee;

$R_f = 0.31$ (Hex:EA = 4 : 1);

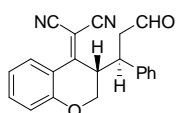
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.40 (s, 1H), 8.03 (d, $J = 7.9$ Hz, 1H), 7.55 (t, $J = 7.2$ Hz, 1H), 7.39-7.25 (m, 7H), 3.59-3.54 (m, 1H), 3.20-3.13 (m, 1H), 3.08-2.92 (m, 2H), 2.87-2.81 (m, 1H), 2.56-2.51 (m, 1H), 1.98-1.88 (m, 1H), 1.79-1.74 (m, 1H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 199.6, 176.5, 140.0, 139.9, 134.2, 129.9, 129.2, 128.7, 128.4, 127.8, 127.1, 113.5, 113.4, 80.8, 48.1, 47.3, 43.5, 40.4, 25.3, 24.3;

IR (KBr): 2213, 1748 cm^{-1} ;

MS: $\text{C}_{22}\text{H}_{18}\text{N}_2\text{O}$ 327.13 $[\text{M}+\text{H}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{major}} = 20.732$ min, $t_{\text{minor}} = 30.280$ min.



8bd $[\alpha]_D^{22} = -218$ (c 1.08, DCM), 87% ee;

$R_f = 0.26$ (Hex:EA = 4 : 1);

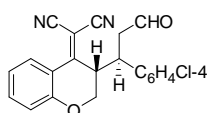
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.44 (s, 1H), 8.20 (d, $J = 8.1$ Hz, 1H), 7.56 (t, $J = 7.8$ Hz, 1H), 7.41-7.37 (m, 2H), 7.33-7.29 (m, 3H), 7.10 (t, $J = 7.7$ Hz, 1H), 7.02 (d, $J = 8.4$ Hz, 1H), 4.12-4.02 (m, 2H), 3.45-3.38 (m, 1H), 3.28-3.18 (m, 1H), 3.12-3.05 (m, 1H), 2.65-2.59 (m, 1H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 199.2, 167.1, 156.3, 139.2, 137.3, 129.4, 128.1, 128.0, 122.0, 118.4, 115.3, 113.2, 78.7, 66.5, 46.8, 45.2, 39.5;

IR (KBr): 2224, 1719 cm^{-1} ;

MS: $\text{C}_{21}\text{H}_{16}\text{N}_2\text{O}_2$ 327.15 $[\text{M}-\text{H}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 19.582$ min, $t_{\text{major}} = 22.589$ min.



8be $[\alpha]_D^{24} = -153$ (c 0.67, DCM), 88% ee.

$R_f = 0.38$ (Hex:EA = 4 : 1);

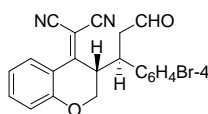
$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.44 (s, 1H), 8.19 (d, $J = 8.1$ Hz, 1H), 7.56 (t, $J = 8.2$ Hz, 1H), 7.53 (d, $J = 8.2$ Hz, 2H), 7.22 (d, $J = 8.2$ Hz, 2H), 7.10 (t, $J = 8.4$ Hz, 1H), 7.03 (d, $J = 8.4$ Hz, 1H), 4.13-4.01 (m, 2H), 3.45-3.35 (m, 1H), 3.19-3.02 (m, 2H), 2.68-2.63 (m, 1H);

$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 198.6, 166.7, 156.2, 137.8, 137.4, 132.5, 129.9, 127.9, 122.2, 118.4, 116.2, 115.2, 113.4, 113.0, 78.8, 66.4, 46.7, 44.9, 38.6;

IR (KBr): 2216, 1720 cm^{-1} ;

MS: $\text{C}_{21}\text{H}_{15}\text{ClN}_2\text{O}_2$ 363.11 $[\text{M}+\text{H}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 20.517$ min, $t_{\text{major}} = 23.797$ min.



8bf $[\alpha]_D^{24} = -174$ (c 0.45, DCM), 85% ee;

$R_f = 0.33$ (Hex:EA = 4 : 1);

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.46 (s, 1H), 8.20 (d, $J = 8.1$ Hz, 1H), 7.57 (t, $J = 7.8$ Hz, 1H), 7.53 (d, $J = 8.2$ Hz, 2H), 7.20 (d, $J = 8.2$ Hz, 2H), 7.11 (t, $J = 8.3$ Hz, 1H), 7.03 (d, $J = 8.4$ Hz, 1H), 4.12-4.01 (m, 2H), 3.44-3.37 (m, 1H), 3.19-3.04 (m, 2H), 2.66-2.61 (m, 1H);

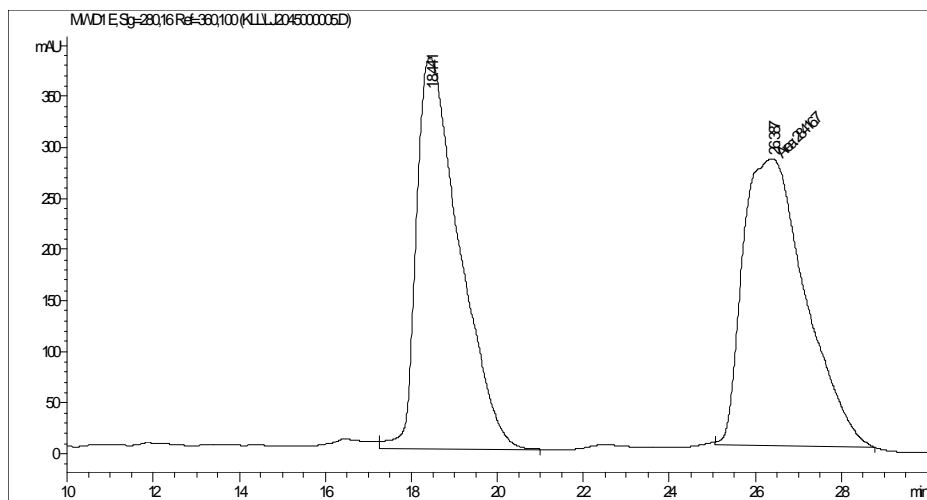
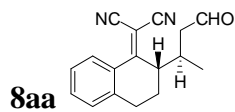
$^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 198.6, 166.7, 156.2, 138.4, 137.4, 132.5, 129.9, 127.9, 122.2, 118.4, 116.2, 115.2, 113.4, 113.1, 78.8, 66.4, 46.7, 44.9, 38.8;

IR (KBr): 2209, 1719 cm^{-1} ;

MS: $\text{C}_{21}\text{H}_{15}\text{BrN}_2\text{O}_2$ 429.03 $[\text{M}+\text{Na}]^+$.

The enantiomeric ratio was determined by HPLC on Chiralpak AS column (20% 2-propanol/hexane, 1 mL/min), $t_{\text{minor}} = 22.685$ min, $t_{\text{major}} = 25.682$ min.

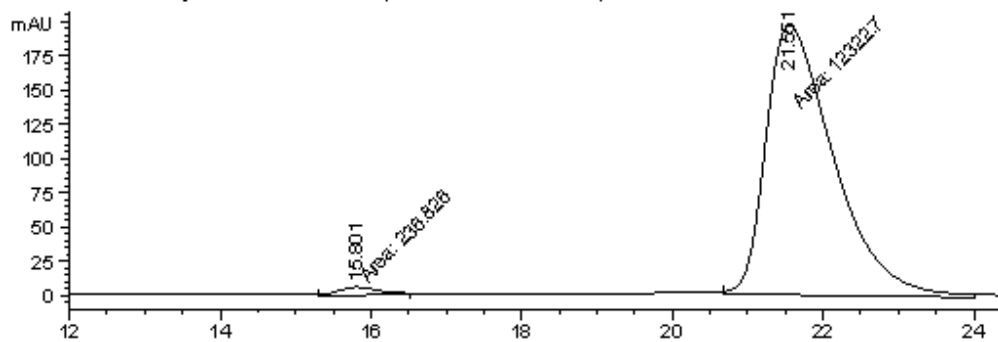
HPLC spectra of products:



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.441	VB	1.0312	2.65250e4	383.35468	47.5160
2	26.387	MM	1.7159	2.92984e4	284.57108	52.4840

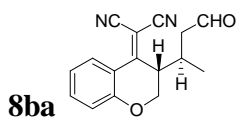
Totals : 5.58234e4 667.92575

MWD1 D, Sig=320,16 Ref=360,100 (LUJUNLJ204600001.D)

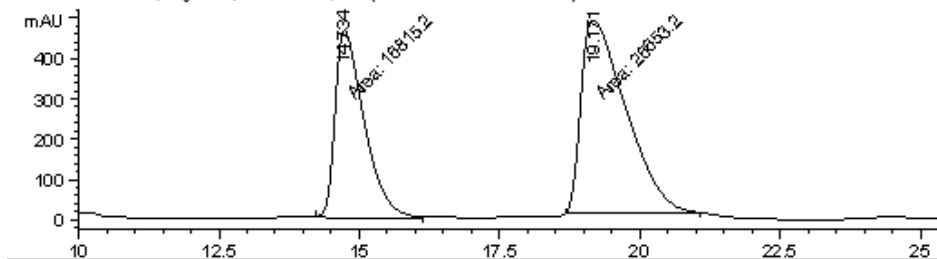


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.801	MM	0.6492	236.82565	6.08029	1.8856
2	21.551	MM	1.0425	1.23227e4	197.00284	98.1144

Totals : 1.25595e4 203.08313



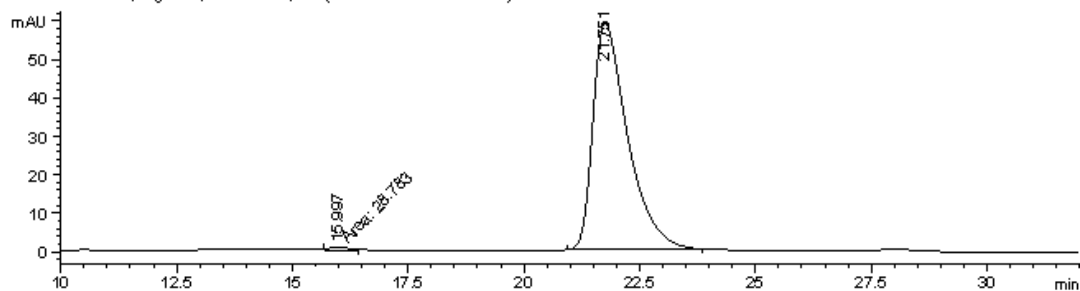
MWVD1.D, Sig=320,16 Ref=360,100 (LUJU NVLJ2134000003.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.734	MM	0.6034	1.68152e4	464.48425	38.6837
2	19.171	MM	0.9279	2.66532e4	478.71790	61.3163

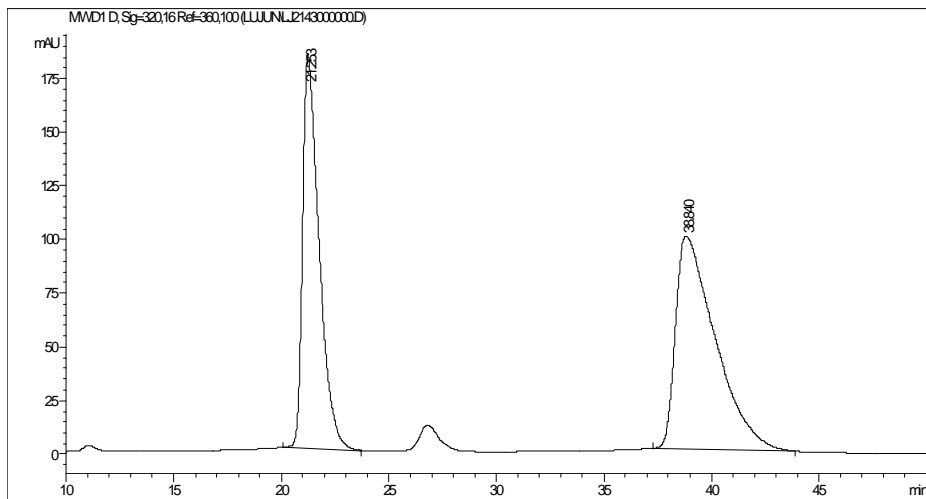
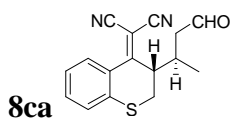
Totals : 4.34683e4 943.20215

MWVD1.D, Sig=320,16 Ref=360,100 (LUJU NG127-1000001.D)



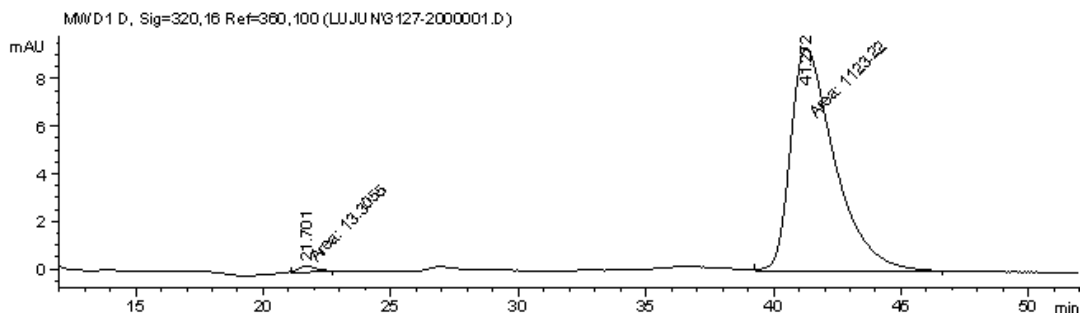
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.997	MM	0.5243	28.78300	9.14992e-1	0.9249
2	21.751	BB	0.7792	3083.30664	59.05229	99.0751

Totals : 3112.08964 59.96728



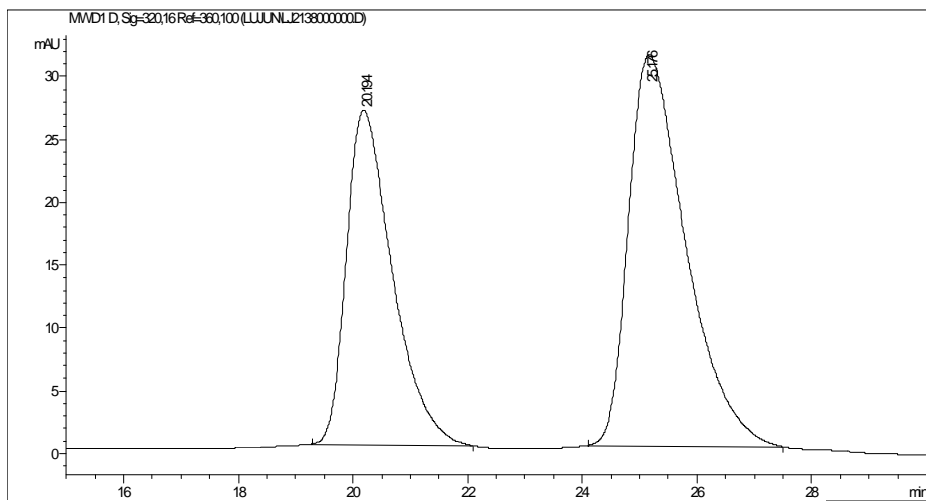
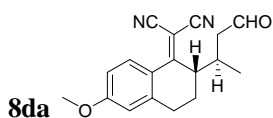
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.253	BB	0.8013	9826.14746	183.93390	43.5414
2	38.840	BB	1.7987	1.27412e4	99.31683	56.4586

Totals : 2.25673e4 283.25073



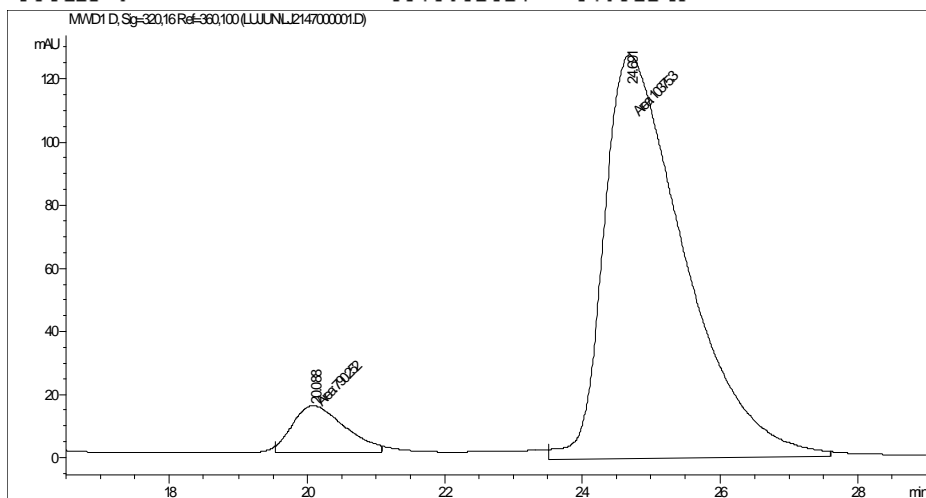
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.701	MM	0.8366	13.30549	2.65072e-1	1.1707
2	41.272	MM	1.9940	1123.22412	9.38844	98.8293

Totals : 1136.52962 9.65352



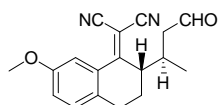
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.194	BB	0.8240	1495.50134	26.67845	40.7103
2	25.176	BB	1.0082	2178.02393	31.13301	59.2897

Totals : 3673.52527 57.81146

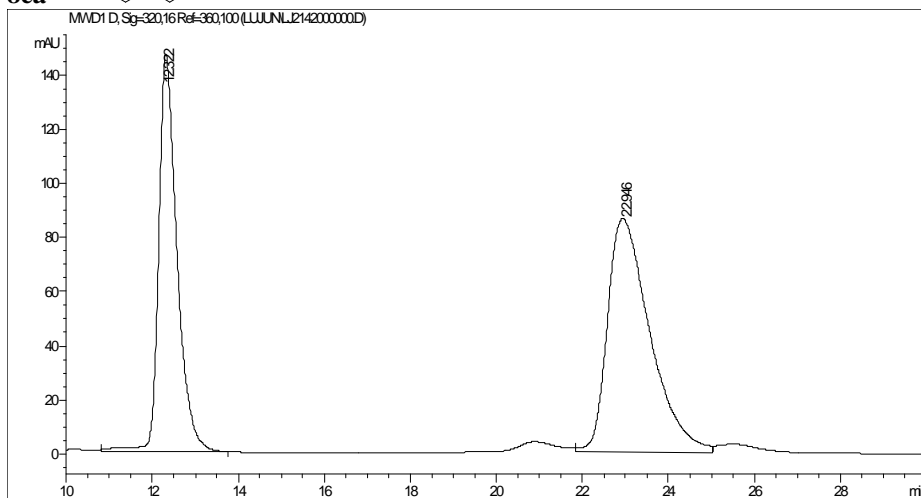


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.088	MM	0.8115	670.05554	13.76126	6.1565
2	24.691	MM	1.3409	1.02137e4	126.95211	93.8435

Totals : 1.08837e4 140.71337

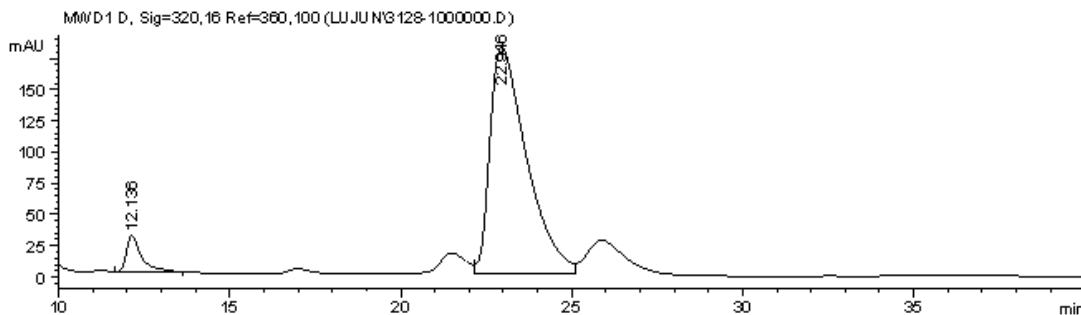


8ea



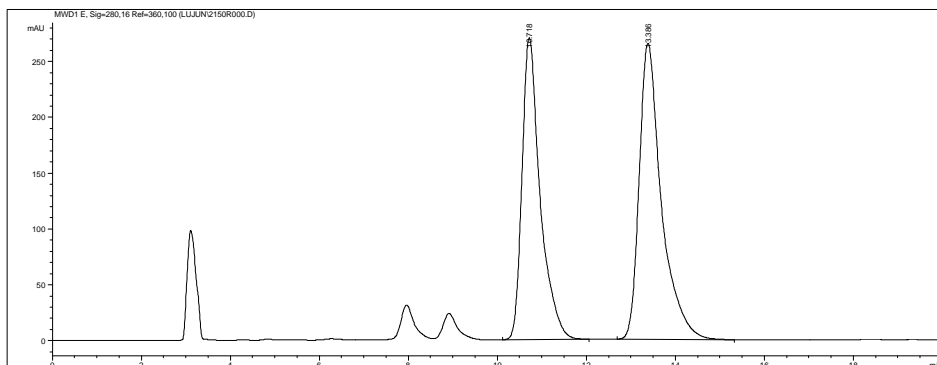
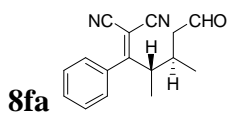
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.322	BB	0.4412	4350.37939	147.04796	42.3114
2	22.946	VB	1.0330	5931.44287	86.60807	57.6886

Totals : 1.02818e4 233.65603



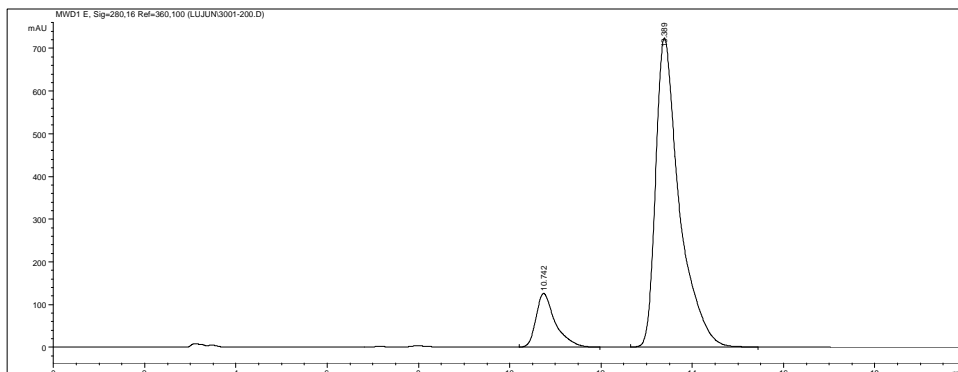
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.136	VB	0.4647	938.41187	29.37506	6.4351
2	22.946	VV	1.1204	1.36443e4	182.04903	93.5649

Totals : 1.45827e4 211.42409



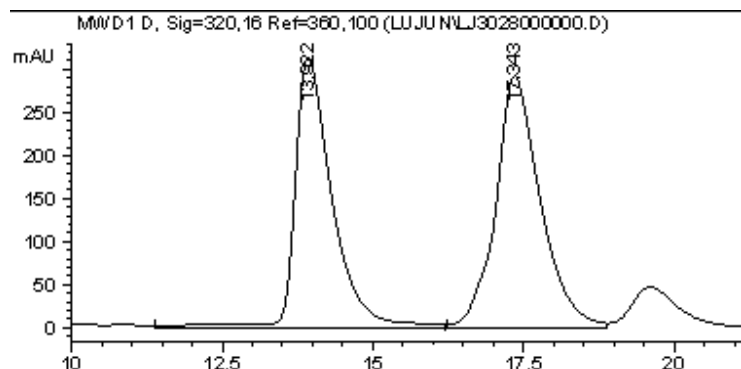
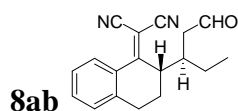
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.719	PB	0.4228	6710.71387	233.87077	45.4675
2	13.386	BB	0.5177	8048.66016	229.19559	54.5325

Totals : 1.47594e4 463.06636



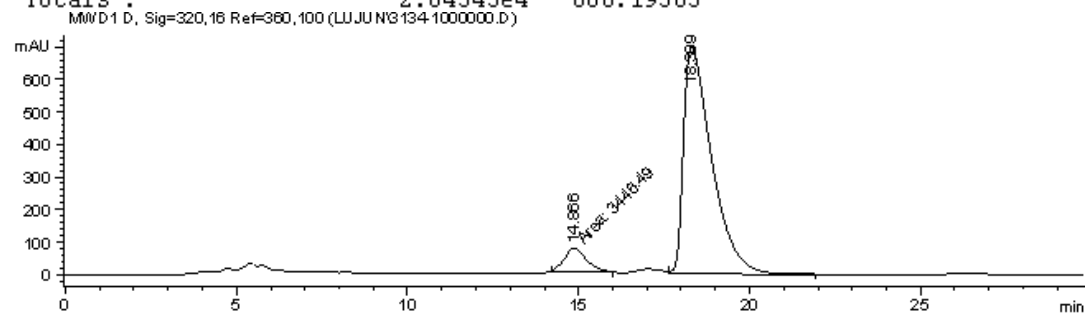
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.742	PB	0.4299	3653.79932	125.39944	12.2827
2	13.389	BB	0.5308	2.60936e4	723.51379	87.7173

Totals : 2.97474e4 848.91323



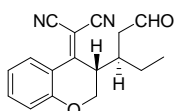
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.922	VB	0.6329	1.34304e4	314.09564	47.1998
2	17.343	BV	0.7601	1.50240e4	292.09741	52.8002

Totals : 2.84545e4 606.19305

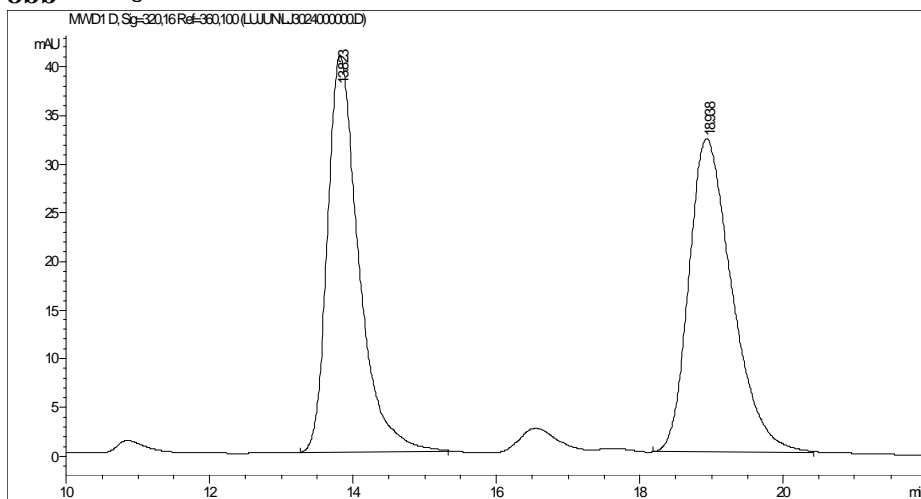


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.866	MM	0.7690	3446.48950	74.69913	7.9300
2	18.309	VB	0.8607	4.00152e4	698.09027	92.0700

Totals : 4.34617e4 772.78940

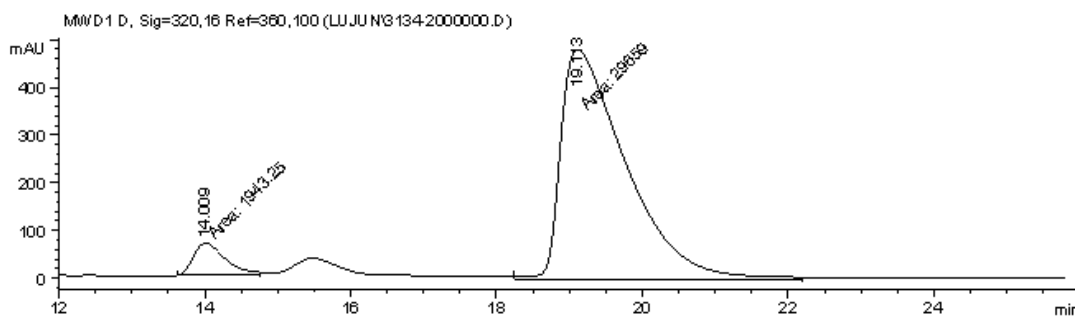


8bb



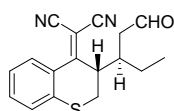
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.823	BB	0.4566	1240.91223	40.82467	47.9402
2	18.938	BB	0.6301	1347.54651	32.20778	52.0598

Totals : 2588.45874 73.03245

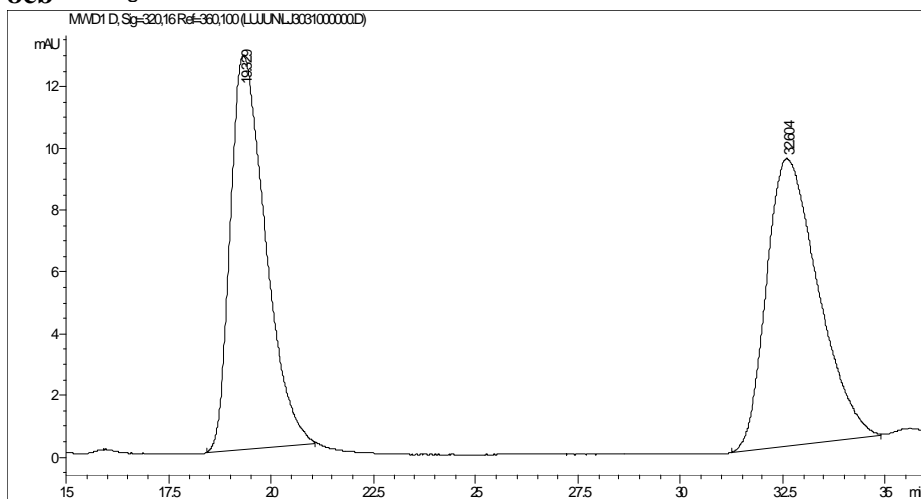


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.009	MM	0.4889	1943.24585	66.24547	6.1491
2	19.113	MM	1.0283	2.96590e4	480.73212	93.8509

Totals : 3.16022e4 546.97758

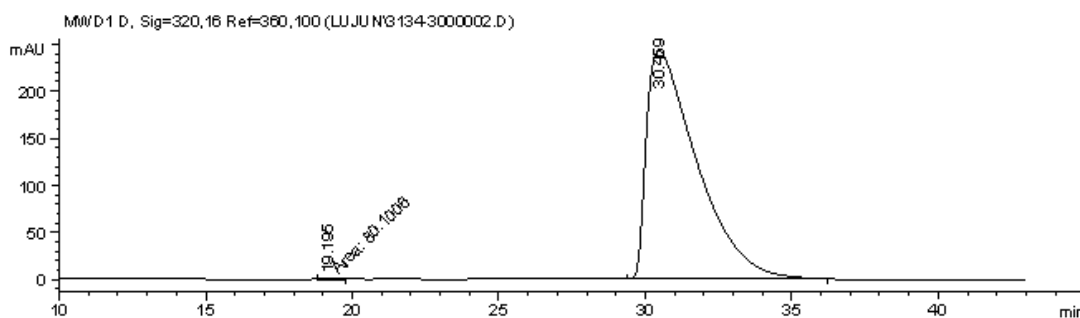


8cb



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.329	BB	0.8583	764.93243	12.77971	48.4356
2	32.604	BB	1.1621	814.34363	9.29515	51.5644

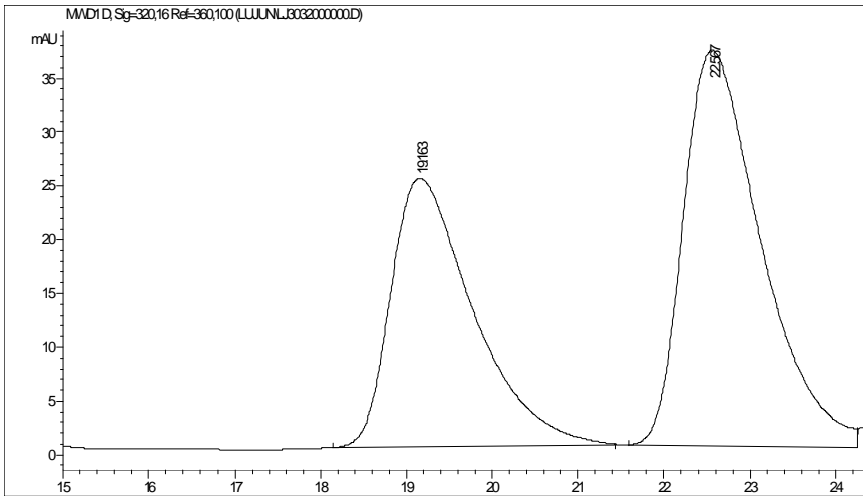
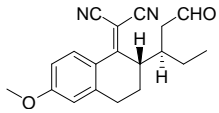
Totals : 1579.27606 22.07486



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.195	MM	0.5543	80.10063	1.73169	0.2750
2	30.459	BB	1.7438	2.90488e4	242.46143	99.7250

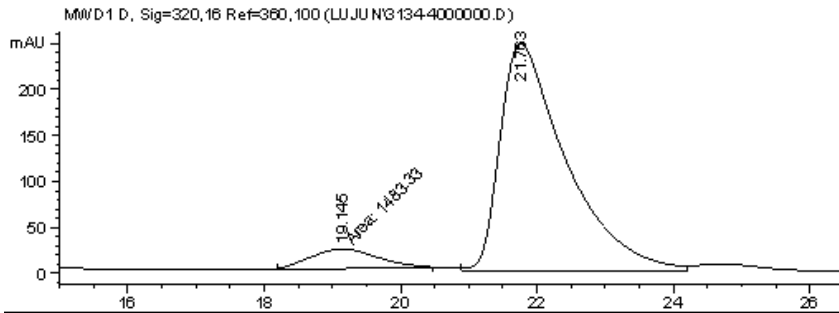
Totals : 2.91289e4 244.19312

8db



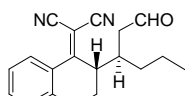
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.163	BB	0.9689	1662.22009	24.96114	41.7639
2	22.567	BB	0.9372	2317.82446	36.78011	58.2361

Totals : 3980.04456 61.74125

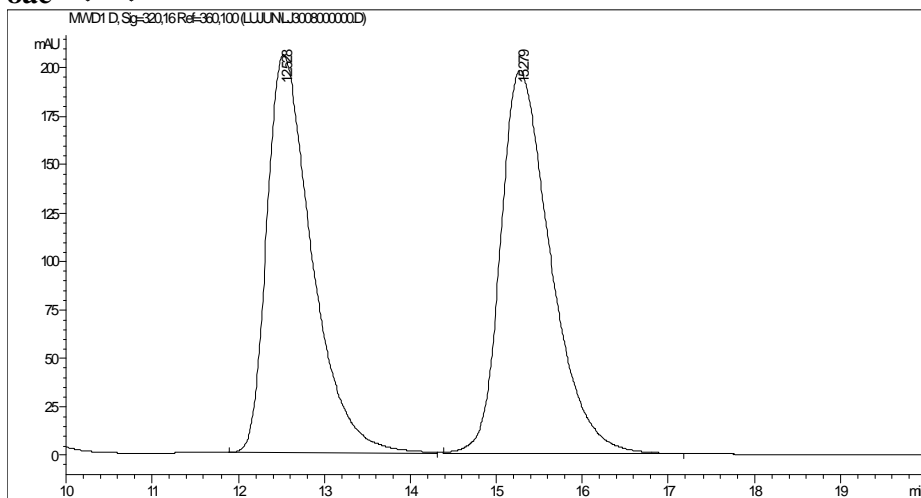


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.145	MM	1.1530	1483.32727	21.44216	8.1697
2	21.763	VB	0.9524	1.66731e4	247.35474	91.8303

Totals : 1.81564e4 268.79690

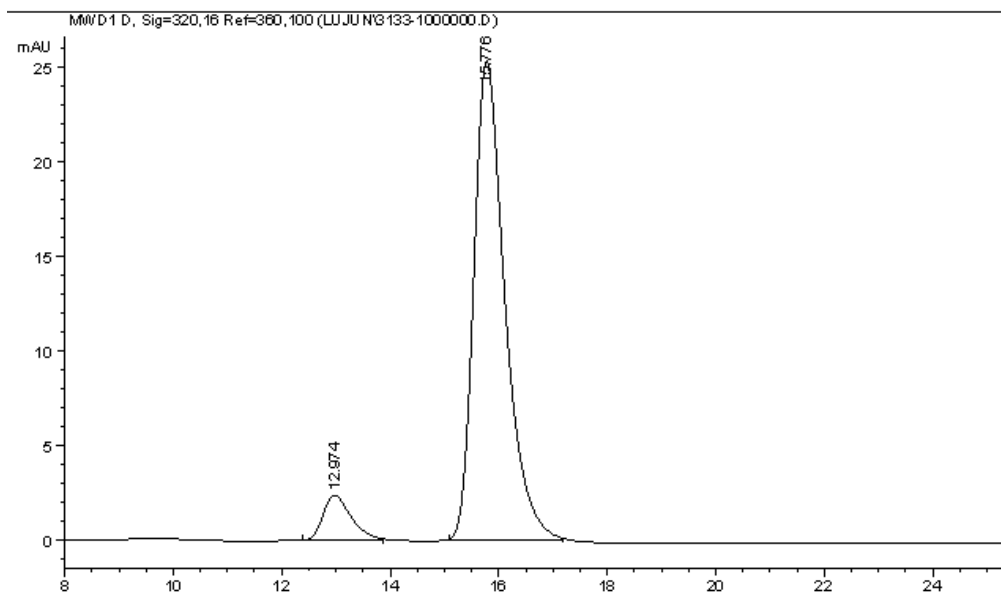


8ac



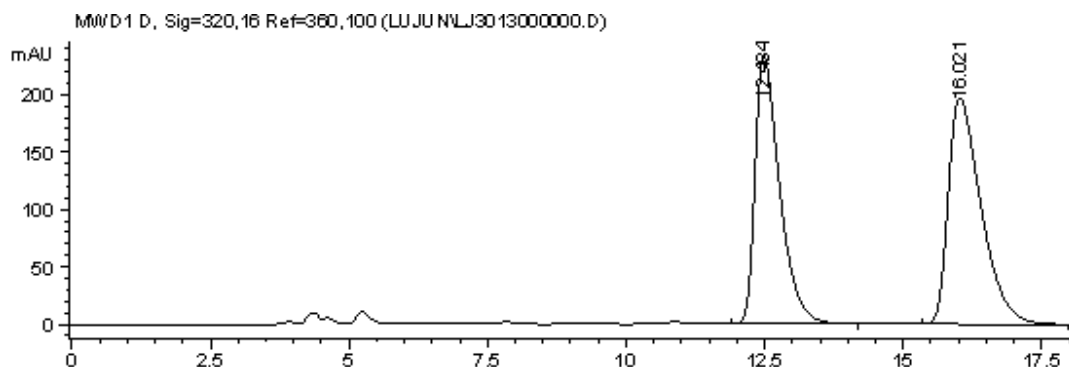
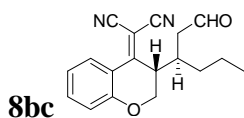
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.528	BB	0.5499	7456.85986	205.21529	48.3640
2	15.279	BB	0.6124	7961.35449	197.46574	51.6360

Totals : 1.54182e4 402.68103



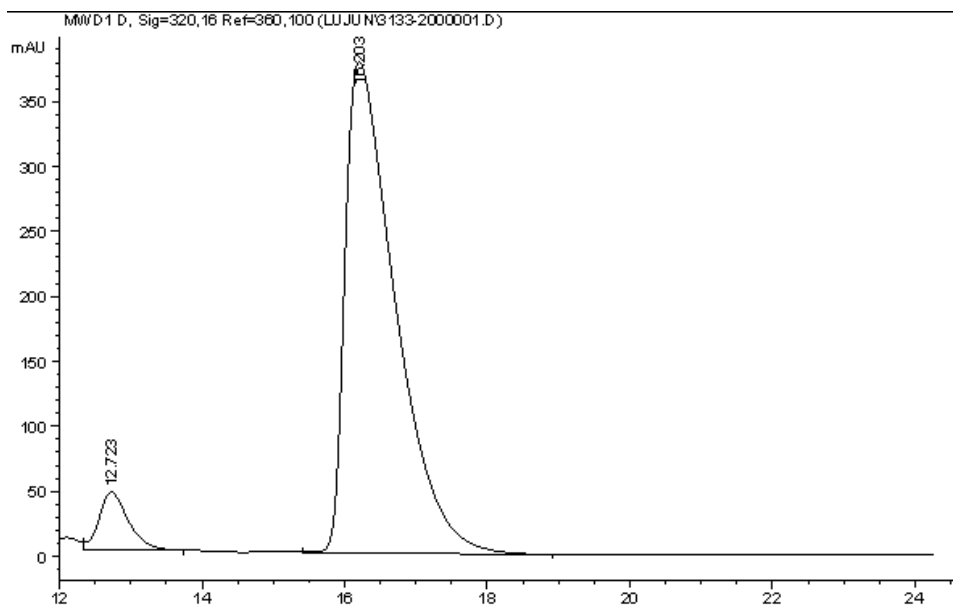
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.974	BB	0.5095	84.51665	2.37332	7.7437
2	15.776	BB	0.5996	1006.90967	25.33673	92.2563

Totals : 1091.42632 27.71005



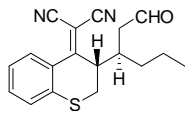
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.484	VB	0.4640	7220.45557	234.02757	47.1133
2	16.021	BB	0.6242	8105.28711	196.91902	52.8867

Totals : 1.53257e4 430.94659

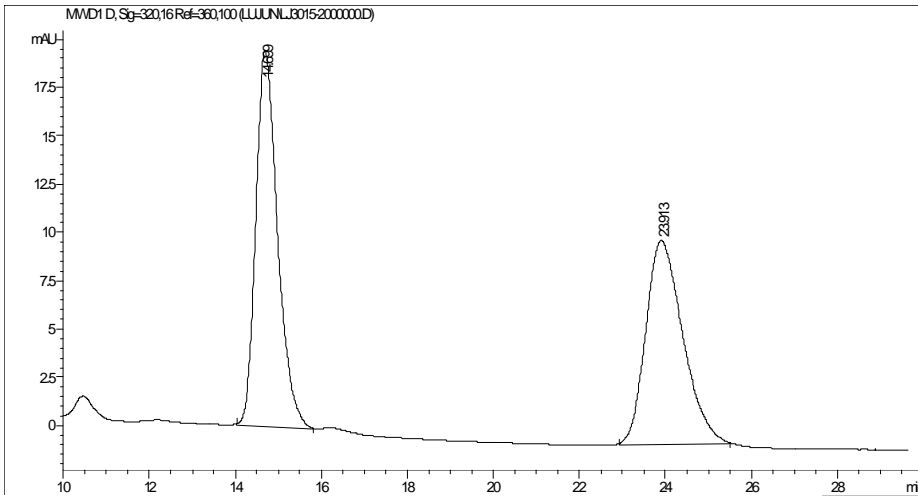


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	12.723	VB	0.4309	1287.24353	44.84673	6.5084
2	16.203	VB	0.7339	1.84908e4	378.53036	93.4916

Totals : 1.97781e4 423.37709

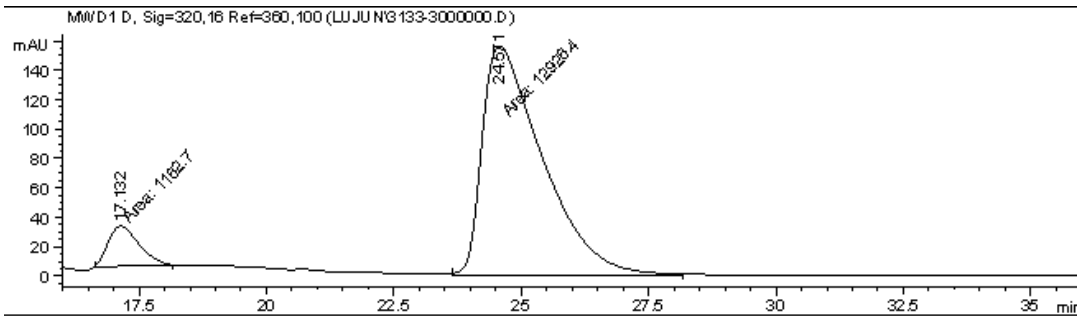


8cc



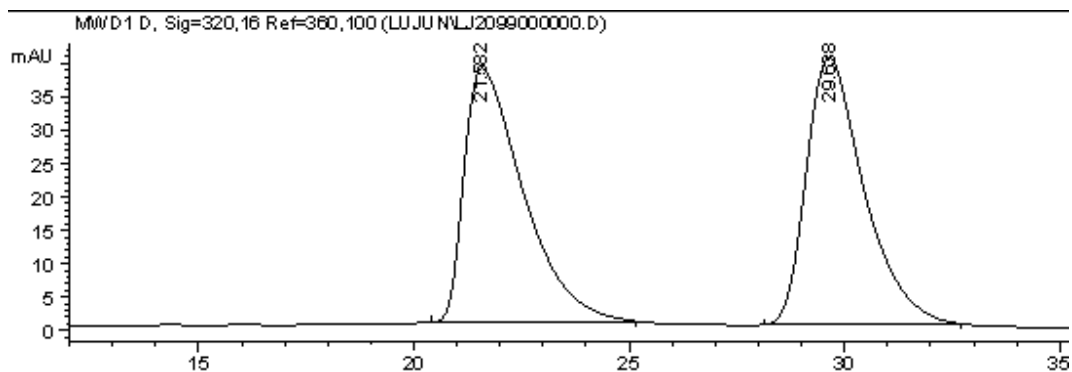
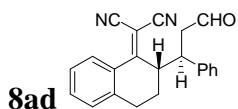
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.699	BB	0.5319	684.04742	19.47105	51.9720
2	23.913	BB	0.8702	632.13739	10.56251	48.0280

Totals : 1316.18481 30.03356



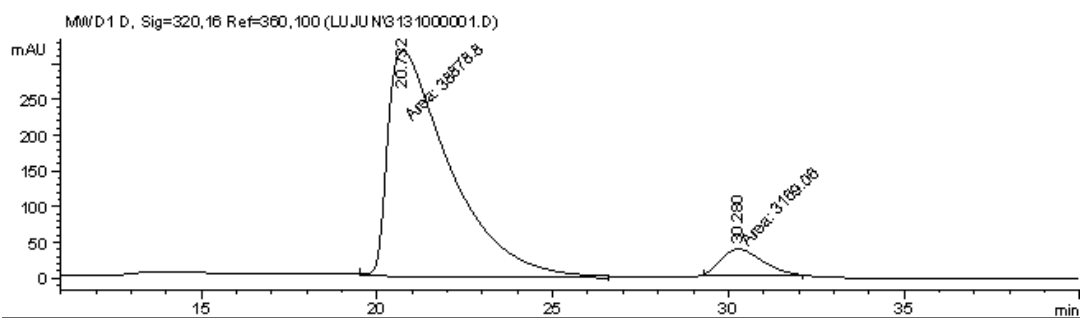
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.132	MM	0.7113	1162.69885	27.24472	8.2525
2	24.571	MM	1.3777	1.29264e4	156.37955	91.7475

Totals : 1.40891e4 183.62427



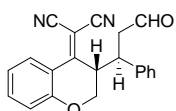
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.582	BB	1.4104	3643.73877	38.07954	49.3409
2	29.638	BB	1.3651	3741.08325	40.07685	50.6591

Totals : 7384.82202 78.15639

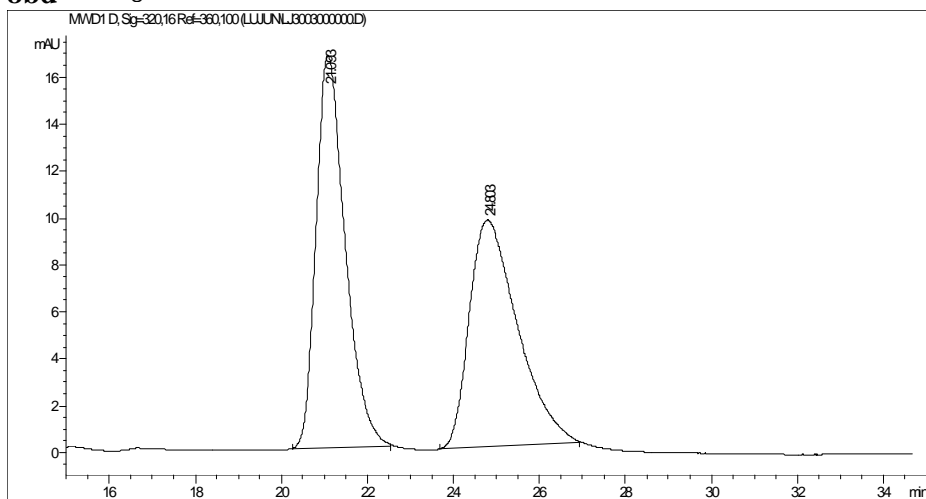


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.732	MM	2.0511	3.88788e4	315.92508	92.4632
2	30.280	MM	1.4028	3169.06030	37.65275	7.5368

Totals : 4.20478e4 353.57783

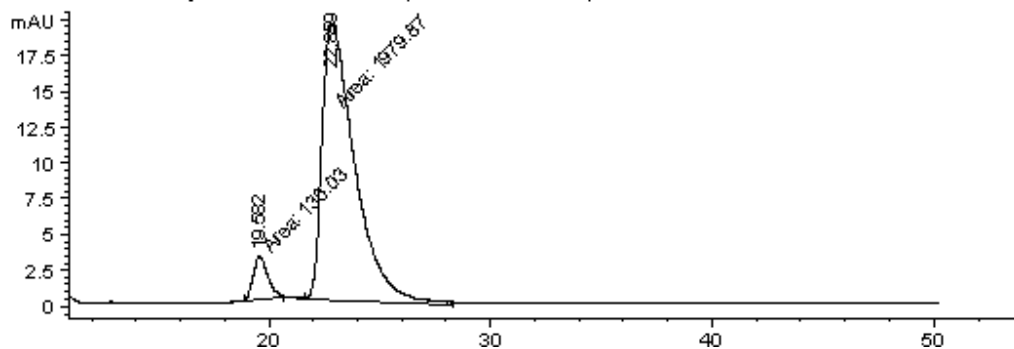


8bd



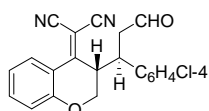
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.093	BB	0.7073	787.62866	16.72939	50.9622
2	24.803	BB	1.0954	757.88525	9.67999	49.0378

Totals : 1545.51392 26.40938
 MWVD1 D, Sig=320,16 Ref=360,100 (LUJUNLJ311722.D)



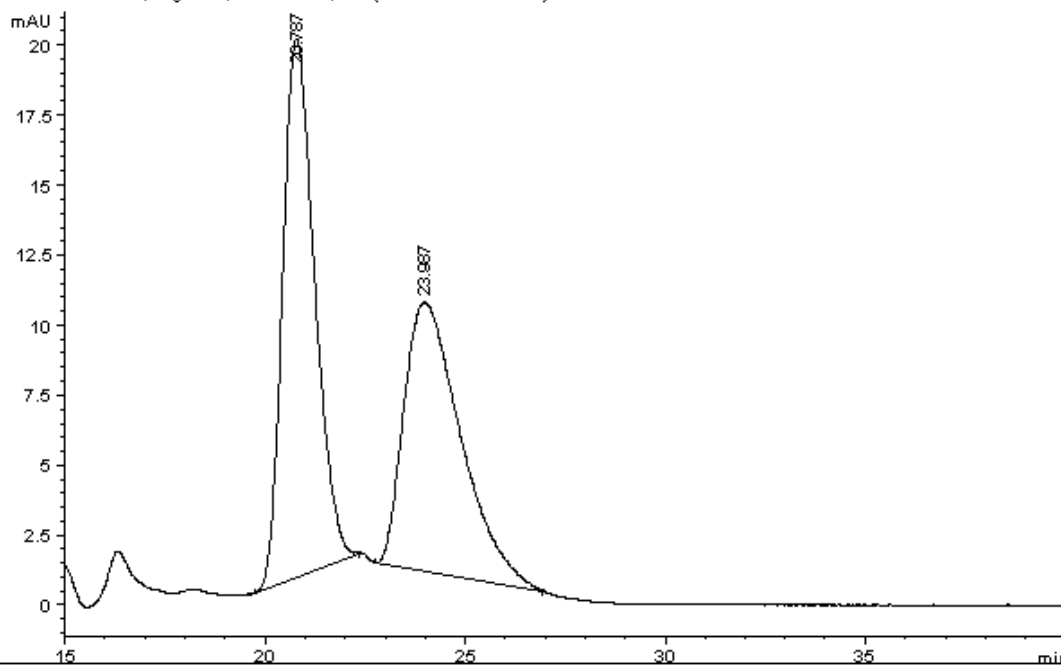
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.582	MM	0.7677	136.02953	2.95317	6.4289
2	22.859	MM	1.7166	1979.86804	19.22271	93.5711

Totals : 2115.89757 22.17588



8be

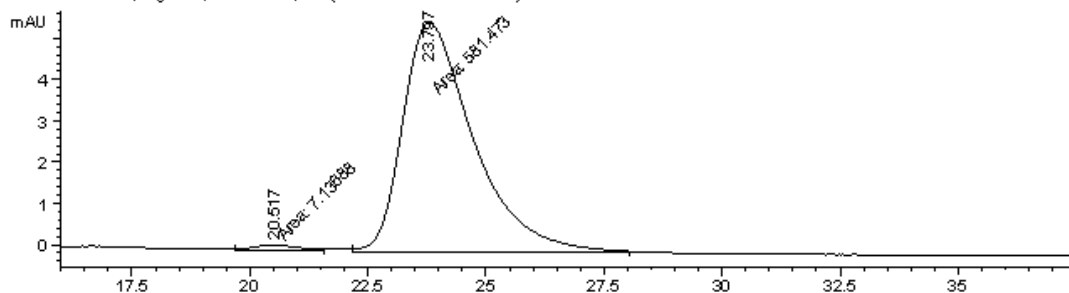
MWD1 D, Sig=320,16 Ref=360,100 (LUJUNLJ312520.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.787	PB	0.8056	1067.68127	19.23425	52.9542
2	23.987	PB	1.1760	948.55377	9.60594	47.0458

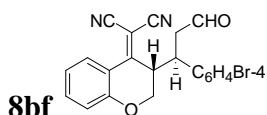
Totals : 2016.23505 28.84019

MWD1 D, Sig=320,16 Ref=360,100 (LUJUNJ3132-1000001.D)

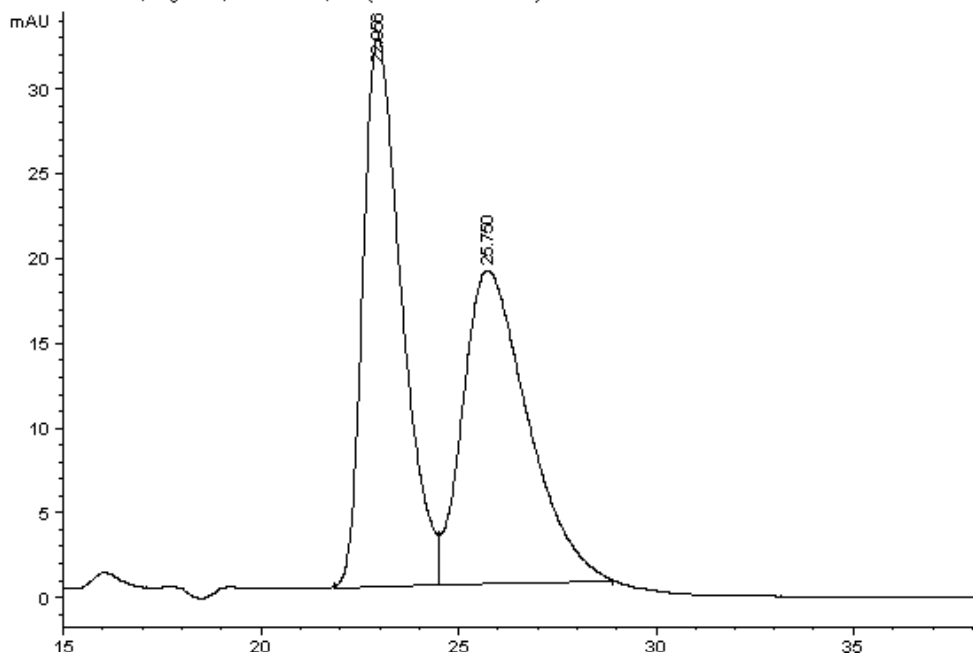


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.517	MM	0.9703	7.13688	1.22593e-1	1.2125
2	23.797	MM	1.7444	581.47253	5.55560	98.7875

Totals : 588.60941 5.67819



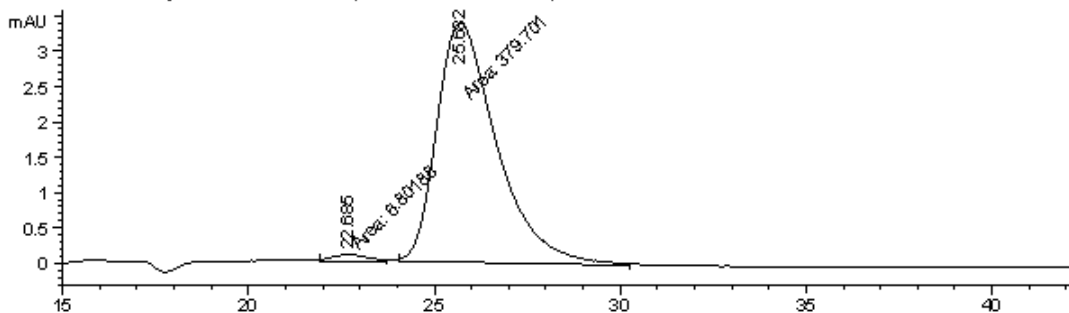
MWWD1 D, Sig=320,16 Ref=360,100 (LUJUNLJ312530.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	22.956	BV	0.9959	2203.47070	32.30103	50.9943
2	25.750	VB	1.3865	2117.54053	18.48512	49.0057

Totals : 4321.01123 50.78614

MWWD1 D, Sig=320,16 Ref=360,100 (LUJUN132-2000001.D)



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	22.685	MM	1.1010	6.80188	1.02965e-1	1.7599
2	25.682	MM	1.8665	379.70093	3.39057	98.2401

Totals : 386.50281 3.49353